

Full title:**Identification of regulatory factors promoting embryogenic callus formation in barley through transcriptome analysis****Authors:**

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Additional file 1**Table S1.** Statistics of the total reads from 5 libraries.

Sample		Raw_	Raw_	Valid_	Dedup	Valid_	Valid_	Valid_	Valid	Dedup
		reads	Bases (G)	reads	_reads	Q20 (%)	Q30 (%)	GC (%)	2raw (%)	2Valid (%)
IME_0h	1	51193136	7.68	50508092	41238767	99.94%	97.65%	54.0%	98.66%	81.65%
	2	51358534	7.70	50725082	41505310	99.94%	97.71%	54.0%	98.77%	81.82%
	3	52190674	7.83	51346982	41532379	99.94%	97.68%	54.0%	98.38%	80.89%
IME_24h	1	43390274	6.51	42482012	36191633	99.93%	97.49%	51.0%	97.91%	85.19%
	2	47504050	7.13	46361390	38961715	99.94%	97.51%	51.5%	97.59%	84.04%
	3	50787806	7.62	49289192	41412312	99.94%	97.43%	51.5%	97.05%	84.02%
IME_48h	1	51336244	7.70	50064794	41202782	99.92%	97.54%	51.5%	97.52%	82.30%
	2	54437730	8.17	52617624	44415455	99.94%	97.59%	51.0%	96.66%	84.41%
	3	53048612	7.96	51022778	43251413	99.94%	97.47%	51.5%	96.18%	84.77%
ME_0h	1	45827838	6.87	44726072	37049023	99.94%	97.43%	51.5%	97.60%	82.84%
	2	50216544	7.53	48998490	40579584	99.94%	97.40%	51.5%	97.57%	82.82%
	3	53234728	7.99	51294828	42127157	99.93%	97.65%	51.5%	96.36%	82.13%
ME_24h	1	51008886	7.65	49828700	42149102	99.93%	97.62%	51.5%	97.69%	84.59%
	2	53514406	8.03	51117256	43701976	99.94%	97.64%	51.5%	95.52%	85.49%
	3	50870118	7.63	49956734	42714327	99.93%	97.60%	51.5%	98.20%	85.50%

Sample: sample name; Raw_reads: total number of reads in raw data; Raw_bases (G): total data volume of offline raw data; Valid_reads: number of valid reads after removing joints, low-quality reads, etc.; Dedup_reads: the number of reads after UMI deduplication; Valid_Q20 (%): Q20 value of valid reads; Valid_Q30 (%): Q30 value of valid reads; Valid_GC (%): GC content of reads after UMI deduplication; Valid2raw (%): valid reads accounted for the percentage of raw reads; Dedup2Valid (%): the percentage of valid reads after deduplication of the reads in the genome.

Table S2. Summary of data cleaning and length distribution of tags

length range	transcripts	genes
>=20000	3 (0.00%)	396 (2.73%)
10000-20000	251 (0.15%)	611 (4.21%)
5000-10000	6,100 (3.66%)	1,803 (12.41%)
2000-5000	61,432 (36.82%)	4,537 (31.23%)
1000-2000	63,663 (38.16%)	3,484 (23.98%)
500-1000	25,153 (15.08%)	1,874 (12.90%)
300-500	6,025 (3.61%)	778 (5.35%)
<300	4,222 (2.53%)	1,046 (7.20%)
total number	166,849	14,529

Table S3 List of primers for plasmid construction.

Name	Sequence (5'-3')
<i>Fw-Axig1</i>	TATGACCATGATTACGAATTTCCTTCATCATCCTCCCCAG
<i>Rev-Axig1</i>	GTGACCTCTCTCACTTTCCCTTGATCAGCC
<i>Fw-HvWUS</i>	GGGAAAGTGAGAGAGGTCAC TAGCACAACAAAGTGGGGGT
<i>Rev-HvWUS</i>	ATGGCCGCGGGGACAATGACATCAACGAAAACGGGGTGGA
<i>Fw-Tnos</i>	TTGATGTCATTGTCCCGCGGCCATGCTAGAGTCC
<i>Rev-Tnos</i>	ATGTCGATAGGTCAGTGGATTTTGTTT
<i>Fw-PLTP</i>	CCAGTGACCTATCGACATGTGGGCTCCATT
<i>Rev-PLTP</i>	GAATGATTTCGTGCGCCACTGCCAACTTCT
<i>Fw-HvBBM</i>	CAGTGGCGACCGAATCATTCGCTAGCTTTGACTGCCCTGA
<i>Rev-HvBBM</i>	CCAGTGACCTTTAAGTGTCGAGTCCTCCTACTTCACCCGT
<i>Fw-PolyA</i>	CGACACTTAAAGGTCAGTGGATTTTGTTT
<i>Rev-PolyA</i>	CTTGTCATGCCTGCAGGTCGACCGCGGCCATGCTAGAGTCC
<i>Fw-ZmWUS</i>	AGTGAATGGCGGCCAGAGAGGTCAGTGCACAGGAG
<i>Rev-ZmWUS</i>	CGCGGTACATGCTCGGACAATGACAGCAACGCAC
<i>Fw-ZmBBM</i>	CTCCTCAAGGCGAATCAATCTAAGAAGAACTCAA
<i>Rev-ZmBBM</i>	CCCGGGGATCTTAAGTGTCGTTCCAGACAC

The parts marked in red represents the adapter sequences. Fw: forward primer. Rev: reverse primer.

Table S4 List of primers used in the qRT-PCR.

Name	Sequence (5'-3')
<i>QPCR-ACTIN-F</i>	GCTGAGCGGGAAATTGTAAG
<i>QPCR-ACTIN-R</i>	GATCATGGATGGCTGGAAGA
<i>QPCR-LEC1-F</i>	TACGCGCCAGGAAATAGTGG
<i>QPCR-LEC1-R</i>	CCGAAGGTCTGGTGGTTCTC
<i>QPCR-PLT5-F</i>	GGGGTTCGGGCCAGTAATT
<i>QPCR-PLT5-R</i>	CACCCCGCGAAGATGTAGAA
<i>QPCR-BBM-F</i>	CAGTGGTTTTTCTCGTGGCG
<i>QPCR-BBM-R</i>	GCGACTCTCCCTATCCTTGC
<i>QPCR-WUS-F</i>	CCATCCCCGGATCATGACATA
<i>QPCR-WUS-R</i>	CAGGTCGTAGGCTGCTTTGA
<i>QPCR-ARF11-F</i>	TCGTCACGTCAAAGAAGCTGA
<i>QPCR-ARF11-R</i>	CCTGGGCCTGTAGTAGACCA
<i>QPCR-ARF16B-F</i>	TGGTGTACAATGCCAGACACA
<i>QPCR-ARF16B-R</i>	TGGAAAACCAAGCAATCGCC
<i>QPCR-PIN1A-F</i>	GGATCTTCACGCCGGACCAG
<i>QPCR-PIN1A-R</i>	GAGATGAAGTGGAAGGACAGCA
<i>QPCR-SUVH2A-F</i>	ACCTTGCTCTCGAGAGGAGT
<i>QPCR-SUVH2A-R</i>	GCTGTGTCCTCTTCCATATC
<i>QPCR-SUVH3A-F</i>	ACCTTGCTCTCGAGAGGAGT
<i>QPCR-SUVH3A-R</i>	GCTGTGTCCTCTTCCATATT

F: forward primer. R: reverse primer.

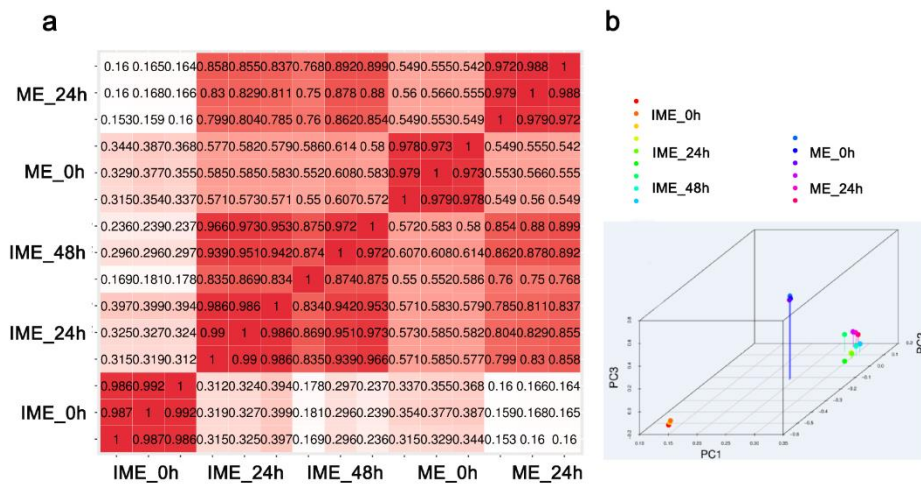


Fig. S1 Pearson correlation between samples. **a:** Correlation heat map between samples. **b:** Principal component analysis of the three-dimensional map.

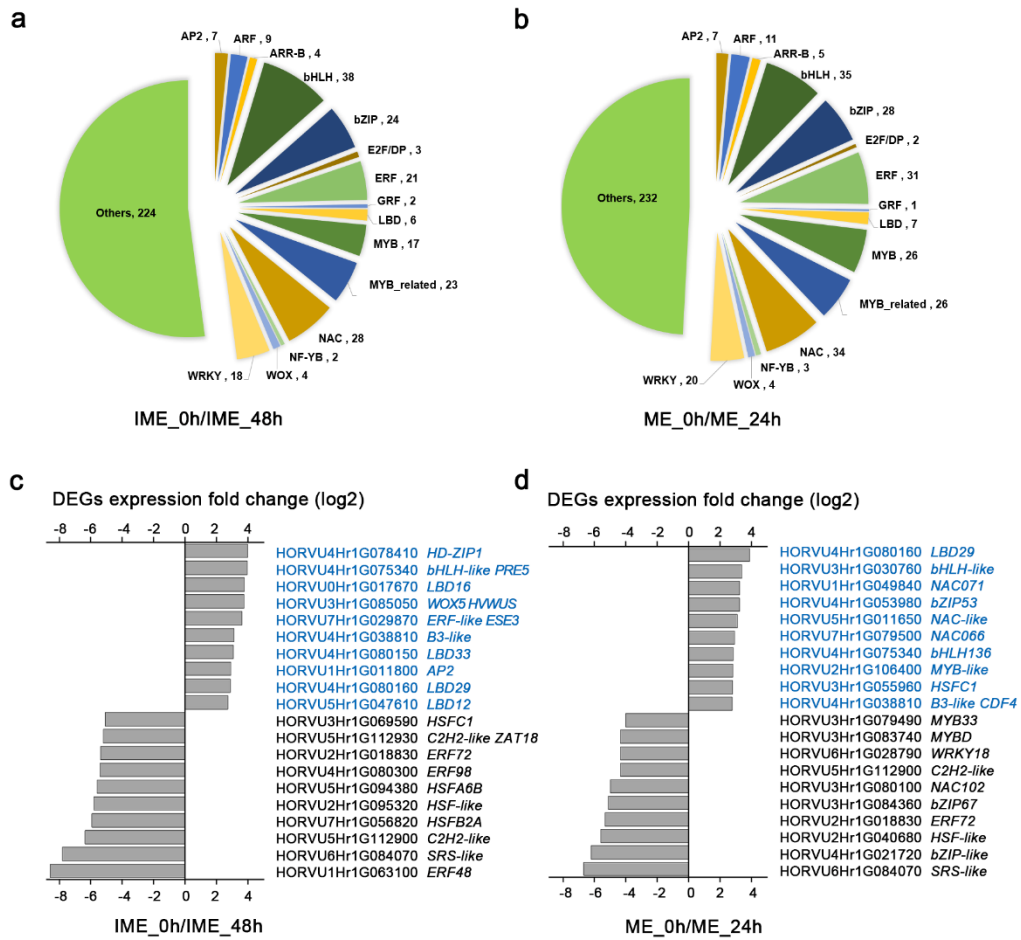


Fig. S2 Expression of a set of callus-inducing medium (CIM)--induced transcription factors during immature and mature embryo-derived callus formation. **a-b:** Pie chart of differentially expressed transcription factors in IME_0h/IME_48h and ME_0h/ME_24h. Numbers represent the gene members associated with a given TF family. **c:** The top 10 differentially expressed TFs in IME_0h/IME_48h ranked by fold change. Genes marked in blue are upregulated TFs, and transcription factors marked in black are downregulated. **d:** The top 10 differentially expressed TFs in ME_0h/ ME_24h.

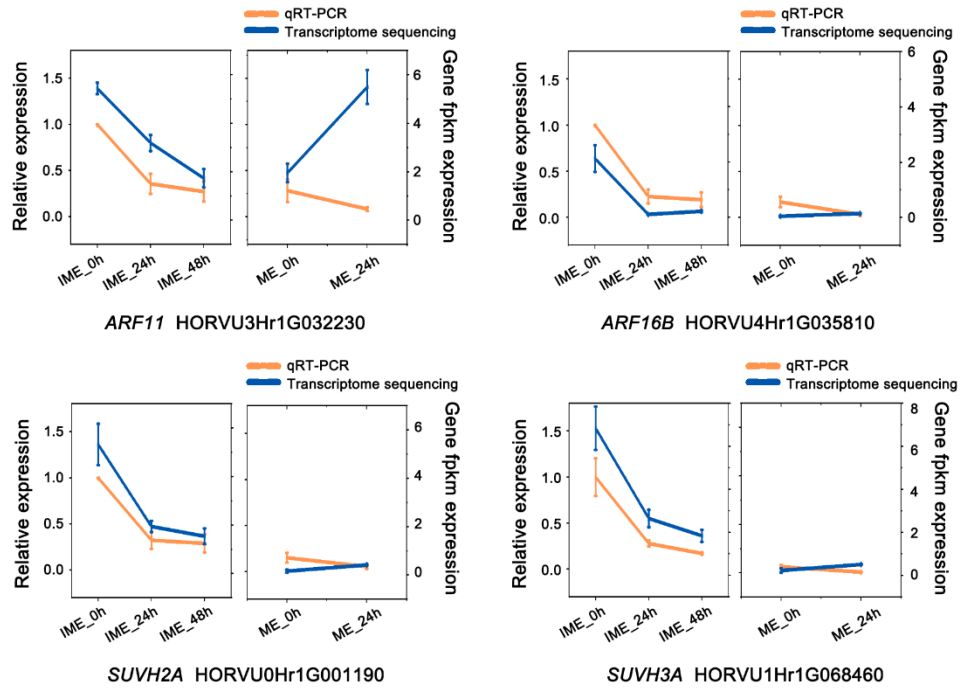


Fig. S3 Transcript levels of *ARF11*, *ARF16B*, *SUVH2A* and *SUVH3A* in the five samples as revealed by qRT-PCR and RNA-seq data. The data shown are means $\pm$ S.D. of three biological replicates.

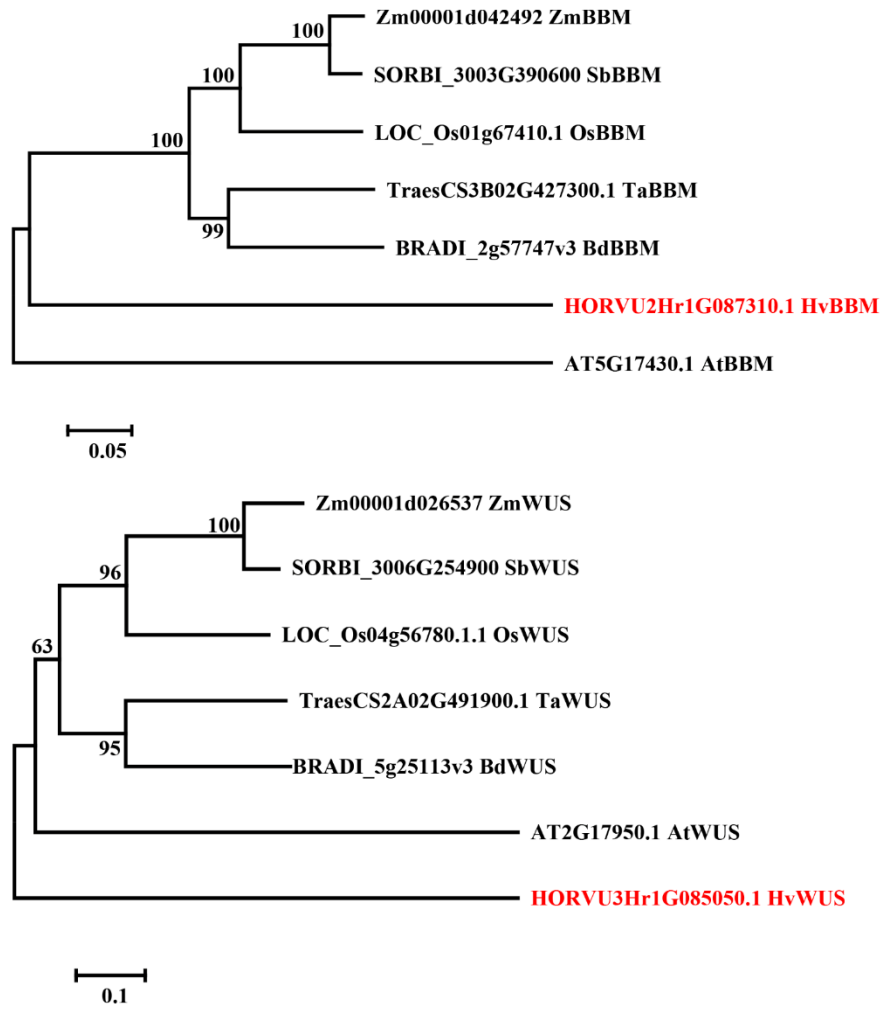


Fig. S4 Phylogenetic tree of *BBM* and *WUS* genes in barley and other species. The phylogenetic tree was constructed in MEGA 4 by the Neighbor-Joining method. The gene IDs are *HvBBM* (HORVU2Hr1G087310.1), *HvWUS* (HORVU3Hr1G085050.1) from *Hordeum vulgare*, *ZmBBM* (Zm00001d042492), *ZmWUS* (Zm00001d026537) from *Zea mays*, *SbBBM* (SORBI_3003G390600), *SbWUS* (SORBI_3006G254900) from *Sorghum bicolor*, *OsBBM* (LOC_Os01g67410.1), *OsWUS* (LOC_Os04g56780.1) from *Oryza sativa*, *TaBBM* (TraesCS3B02G427300.1), *TaWUS* (TraesCS2A02G491900.1) from *Triticum aestivum*, *BdBBM* (BRADI_2g57747v3), *BdWUS* (BRADI_5g25113v3) from *Brachypodium distachyon*, and *AtBBM* (AT5G17430.1), *AtWUS* (AT2G17950.1) from *Arabidopsis thaliana*.

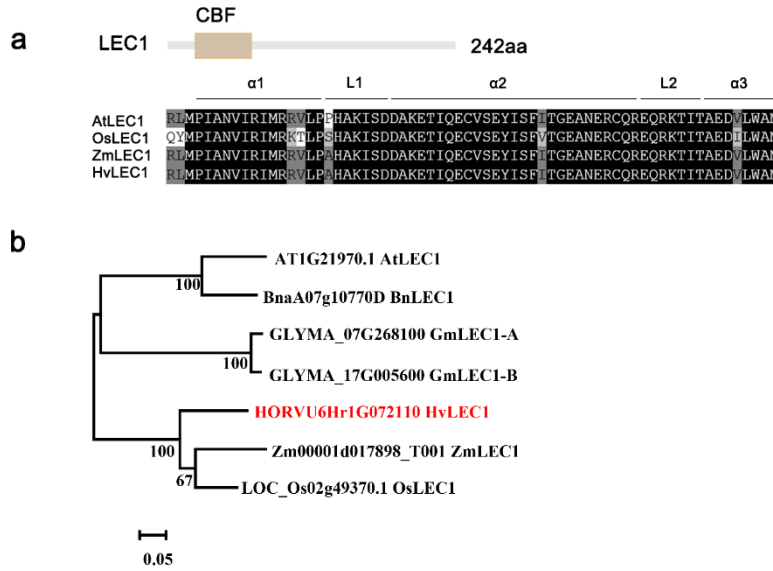


Fig. S5 Expression analysis of candidate *LEC1* gene during callus formation. **a:** Sequence alignment and domain analysis of the LEC1 in *Arabidopsis*, rice, maize and barley. **b:** Phylogenetic tree of LEC1 among barley and other species. The phylogenetic tree was constructed in MEGA 4 by the Neighbor-Joining method. The gene IDs are *HvLEC1* (HORVU6Hr1G072110) from *Hordeum vulgare*, *AtLEC1* (AT1G21970.1) from *Arabidopsis thaliana*, *BnLEC1* (BnaA07g10770D) from *Brassica napus*, *GmLEC1-A* (GLYMA_07G268100), *GmLEC1-B* (GLYMA_17G005600) from *Glycine max*, and *ZmLEC1* (Zm00001d017898_T001) from *Zea mays*, and *OsLEC1* (LOC_Os02g49370.1) from *Oryza sativa*.

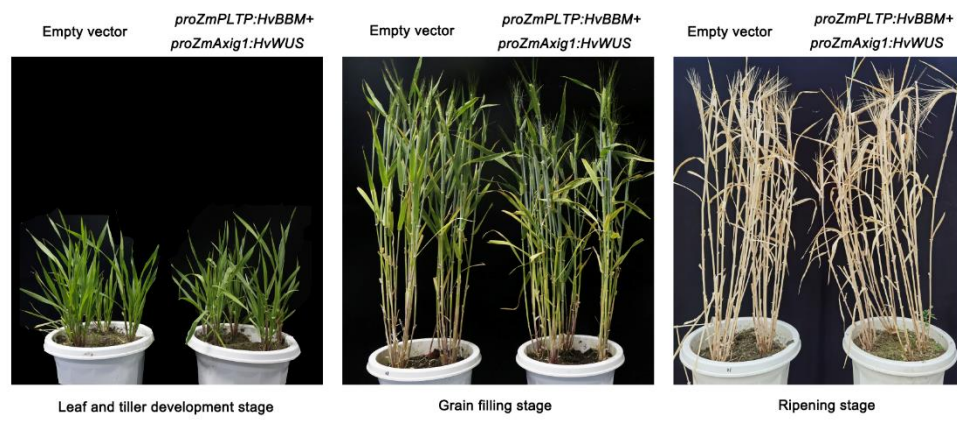


Fig. S6 Phenotypes of plants regenerated from callus transformed with empty vectors and with *WUS* and *BBM* genes.

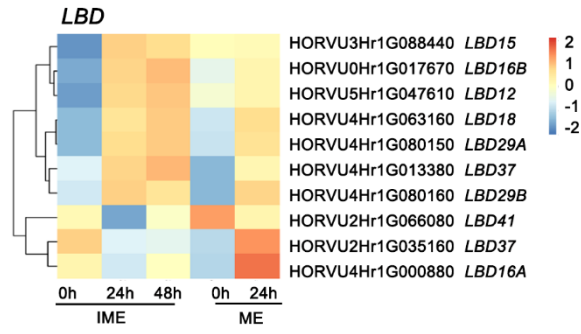


Fig. S7 Expression analysis of candidate *LBD* genes potentially associated with callus formation in barley.

The expression levels were visualised by using OmicStudio tools at <https://www.omicstudio.cn/tool>

based on RNA-seq datasets (Additional file 4). Numbers beneath the heat map indicate the relative expression intensities, and the higher expression intensities are indicated by more reddish colors.