

Case Report

Dual EGFR and ABL Tyrosine Kinase Inhibitor Treatment in a Patient with Concomitant EGFR-Mutated Lung Adenocarcinoma and BCR-ABL1-Positive CML

Kousuke Watanabe¹, Hidenori Kage¹, Saki Nagoshi¹, Kazuhiro Toyama², Yoshiyuki Ohno³, Aya Shinozaki-Ushiku⁴, Kumi Nakazaki², Hiroshi Suzuki³, Mineo Kurokawa², and Takahide Nagase¹

¹Department of Respiratory Medicine, The University of Tokyo, Tokyo, Japan

²Department of Hematology and Oncology, The University of Tokyo, Tokyo, Japan

³Department of Pharmacy, The University of Tokyo Hospital, Faculty of Medicine, The University of Tokyo, Japan

⁴Department of Pathology, The University of Tokyo, Tokyo, Japan

Correspondence should be addressed to Hidenori Kage; kageh-tyk@umin.ac.jp

Received 12 September 2019; Accepted 28 February 2020; Published 19 March 2020

Academic Editor: Kaiser Jamil

Copyright © 2020 Kousuke Watanabe et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Tyrosine kinase inhibitor (TKI) combination is expected to increase in the era of precision medicine. TKI combination may be required to treat double primary cancers, each having a targetable gene, or to treat a single malignancy with multiple targetable genes. Here, we demonstrate the first report of dual EGFR and ABL TKI treatment in a patient with concomitant EGFR-mutated lung adenocarcinoma and BCR-ABL1-positive chronic myeloid leukemia (CML). A 60-year-old man with an 8-year history of CML was diagnosed as advanced EGFR-mutated lung adenocarcinoma. Complete molecular response of CML had been achieved by imatinib, and ABL-TKI had been switched to nilotinib four years previously due to muscle cramps. We discontinued nilotinib and started afatinib. Although partial response of lung adenocarcinoma was achieved, cytogenetic relapse of CML was observed following nilotinib discontinuation. We applied the previously described framework of cytochrome P450 3A4-mediated oral drug-drug interactions and selected gefitinib and nilotinib to treat both malignancies. We effectively and safely administered this combination for seven months. The present report is the first to demonstrate the safety and efficacy of dual EGFR and ABL TKI treatment in a patient with concomitant EGFR-mutated lung adenocarcinoma and CML.

1. Introduction

The prognosis of chronic myeloid leukemia (CML) has been dramatically improved by ABL tyrosine kinase inhibitor (TKI), with 5- and 8-year overall survival rate of 90% and 88%, respectively [1]. The prognosis of advanced EGFR-mutated lung adenocarcinoma has also been improved by EGFR-TKI, and the median progression free survival in the first-line setting is 10-12 months for the first and the second generation EGFR-TKIs and 18.9 months for the third generation EGFR-TKI osimertinib [2, 3].

TKI combination therapy is expected to increase in the era of precision medicine for two reasons. First, the cumula-

tive incidence of a new primary malignancy is expected to increase with improved prognosis of the first malignancy, and TKI combination may be required to treat both malignancies. Second, clinical sequencing will uncover multiple targetable genes in a single tumor, and TKI combination is currently being investigated as a rational treatment strategy to overcome TKI resistance [4].

Here, we report dual EGFR and ABL tyrosine kinase inhibitor treatment in a patient with advanced EGFR-mutated lung adenocarcinoma with an 8-year history of BCR-ABL1-positive CML. Limited information is available on the combination of EGFR-TKI and ABL-TKI, and the drug-drug interaction between EGFR-TKI and ABL-TKI is

of considerable importance for the concomitant use of both TKIs.

We have previously established a quantitative prediction framework of cytochrome P450 (CYP) 3A4-mediated oral drug-drug interactions [5, 6], and this method was applied to predict the drug-drug interaction between EGFR-TKI and ABL-TKI. Using this method, the increase of an area under the concentration-time curve (AUC) of CYP3A4 substrate by a CYP3A4 inhibitor can be calculated using the equation $1/(1 - CR_{CYP3A4} \times IR_{CYP3A4})$, where CR_{CYP3A4} is the ratio of contribution of CYP3A4 to clearance of a substrate drug after oral absorption and IR_{CYP3A4} is the time-averaged apparent inhibition ratio of the inhibitor. The CR_{CYP3A4} of a substrate is calculated based on the AUC increase observed in interaction studies with typical CYP3A4 inhibitors, such as ketoconazole and itraconazole. The IR_{CYP3A4} of an inhibitor is calculated based on the AUC increase of standard CYP3A4 substrate, such as midazolam. This framework can be applied to predict the magnitude of unknown CYP3A4-mediated drug-drug interactions.

The combination of gefitinib and nilotinib was selected based on the prediction of drug-drug interaction, and the patient was safely treated by the combination for seven months. As far as we know, this is the first report of dual EGFR and ABL tyrosine kinase inhibitor treatment for concomitant EGFR-mutated lung adenocarcinoma and BCR-ABL1-positive CML. The present case also demonstrates the usefulness of our prediction framework for CYP3A4-mediated drug-drug interactions in cancer therapeutics in general.

2. Case Presentation

A 60-year-old man with a 21-pack-year history of tobacco use presented with hoarseness due to left recurrent laryngeal nerve palsy. He had been diagnosed as BCR-ABL1-positive CML eight years previously and had achieved a complete molecular response using imatinib. The ABL-TKI had been switched to nilotinib four years previously due to imatinib-induced muscle cramps without relapse.

A computed tomography (CT) was performed to investigate the cause of the recurrent laryngeal nerve palsy and revealed a 4 cm cavitary pulmonary mass in the left lower lobe with mediastinal lymph node swelling (Figures 1(a) and 1(b)). Physical examination, complete blood cell count, and blood chemistry studies revealed no remarkable findings, while serum CEA level was elevated to 19.1 ng/ml.

Endobronchial ultrasonography-guided transbronchial needle aspiration of the mediastinal lymph node revealed adenocarcinoma cells (Figure 2(a)). The tumor was positive for thyroid transcription factor 1 (TTF-1) and EGFR mutation (exon 19 deletion), and a magnetic resonance imaging (MRI) revealed a single bone metastasis in the second lumbar vertebra. Separate tumor nodules in the left lower lobe were also noted by a chest CT scan (Figure 1(a)), and the patient was diagnosed as stage IVA (cT3N3M1b) lung adenocarcinoma.

Because the patient had maintained major molecular response of CML and no case report had ever described the

combination of EGFR-TKI and nilotinib, we discontinued nilotinib and started treatment with afatinib (Figure 3). After four months of afatinib treatment, partial response of the lung adenocarcinoma was confirmed by a CT scan (Figure 1(c)) with decreased CEA level (Figure 3). However, blood BCR-ABL1 level on the international scale (BCR-ABL1^{IS}) measured by the real-time quantitative reverse transcriptase polymerase chain reaction increased four months after the discontinuation of nilotinib (Figure 3). As the patient lost complete cytogenetic response (BCR-ABL1^{IS} increased to 3.2426%, and the percentage of BCR-ABL1-positive cells by fluorescence in situ hybridization of peripheral neutrophils was 13%), we discontinued afatinib and restarted nilotinib five months after its discontinuation.

At the time, three EGFR-TKIs (gefitinib, erlotinib, and afatinib) were approved in Japan as the first-line treatment of EGFR-mutant lung cancer; thus, pharmacokinetic drug-drug interaction between each EGFR-TKI and nilotinib was considered. Estimating the change in plasma concentrations of nilotinib and afatinib when given together was difficult, as both are substrates and inhibitors of P-glycoprotein (P-gp) [7–10]. Gefitinib and erlotinib are substrates for CYP3A4 [11, 12], and we predicted the increase of their plasma concentrations when coadministered with nilotinib, an inhibitor for CYP3A4, using the previously described quantitative prediction framework [5, 6]. The CR_{CYP3A4} of EGFR-TKIs were calculated based on the AUC increases reported in interaction studies with itraconazole or ketoconazole [13–16] (Table 1). The IR_{CYP3A4} of nilotinib was calculated to be 0.67 based on the 2.6-fold AUC increase of CYP3A4 substrate midazolam [17]. Using the equation $1/(1 - CR_{CYP3A4} \times IR_{CYP3A4})$, the AUCs of gefitinib and erlotinib were predicted to increase by 1.44- and 1.36-fold, respectively.

After the approval by the institutional review board, the combination therapy by gefitinib and nilotinib was initiated (Figure 3). Gefitinib was selected for the combination with nilotinib, because the recommended dose of gefitinib (250 mg per day) is one-third of the maximum tolerated dose [18], whereas the recommended dose for erlotinib (150 mg per day) is the maximum tolerated dose [19]. Nilotinib is also a substrate of CYP3A4, and its CR_{CYP3A4} was calculated to be 0.67 based on its 3-fold AUC increase by CYP3A4 inhibitor ketoconazole [20]. Although gefitinib has not been shown to inhibit CYP3A4, the dose of nilotinib was decreased from 600 mg to 400 mg per day to avoid unexpected side effects of the TKI combination.

The primary lung tumor increased in size after discontinuation of afatinib (Figure 1(d)) but decreased again two months after gefitinib (Figure 1(e)). Major molecular response of CML was achieved five months after the readministration of nilotinib. The combination of gefitinib and nilotinib was safely administered for seven months. Only grade 1 skin rash and diarrhea were observed during the TKI combination according to the Common Terminology Criteria for Adverse Events (CTCAE) version 5 [21]. Although major molecular response of CML was sustained, the lung adenocarcinoma became resistant to gefitinib six months after the combination therapy (Figure 1(f)). The rebiopsy of the left hilar lymph node revealed small cell transformation without

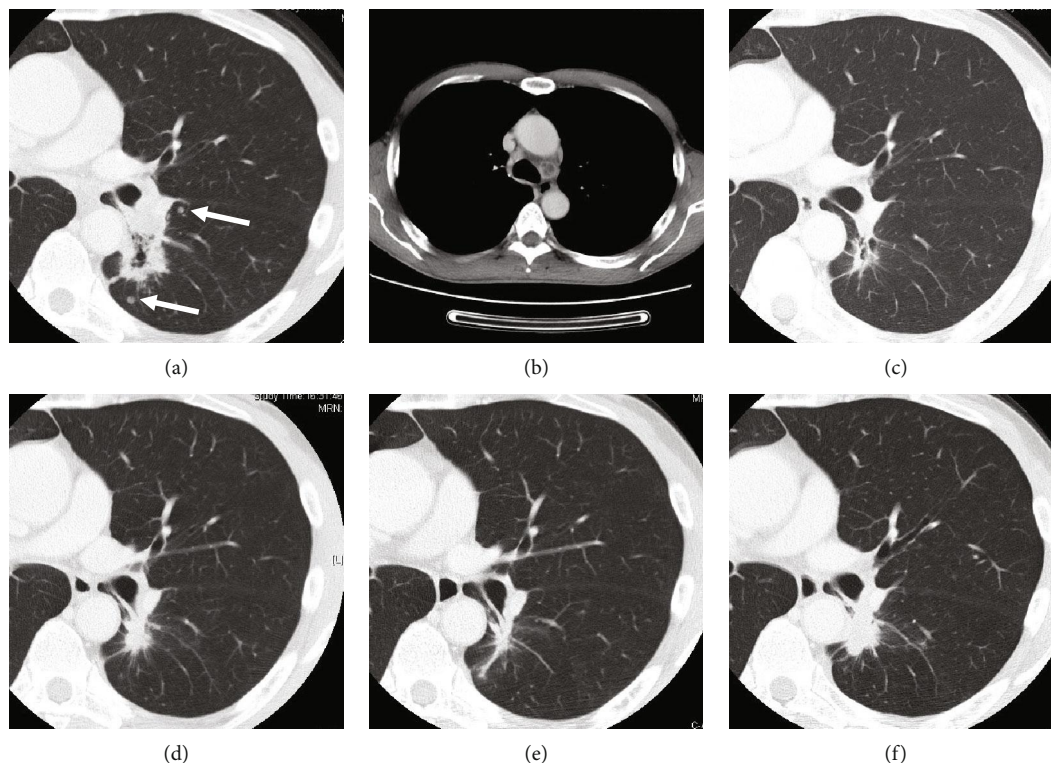


FIGURE 1: Chest CT scan. Chest CT scan performed on admission (a, b) four months after afatinib treatment (c) before the initiation of gefitinib (d) two months after gefitinib treatment (e) and six months after gefitinib treatment (f). Arrows indicate separate tumor nodules in the same lobe as the primary tumor.

secondary EGFR T790M mutation (Figure 2(b)). The small cell carcinoma cells were diffusely positive for synaptophysin and CD56 and focally positive for chromogranin A (Figures 2(c)–2(e)).

3. Discussion

CML is a myeloproliferative neoplasm characterized by BCR-ABL1 fusion gene. The prognosis of CML has been dramatically improved by ABL-TKI, and the occurrence of a new primary malignancy can be a serious problem during the clinical course. There are limited case reports of concomitant EGFR-mutated lung adenocarcinoma and CML. Kaneshiro et al. report a case of recurrent EGFR-mutated lung adenocarcinoma in a patient receiving nilotinib for CML. In that report, nilotinib was discontinued, and the patient was successfully treated by gefitinib monotherapy without CML relapse [22]. To our knowledge, the present case is the first report of the dual EGFR and ABL TKI treatment in a patient with concomitant EGFR-mutated lung adenocarcinoma and BCR-ABL1-positive CML.

Data are conflicting whether the incidence of second malignancy is increased or decreased in CML patients receiving ABL-TKI. Verma et al. report that the risk of secondary malignancy was lower than expected in 1445 patients treated with ABL-TKI [23]. A study based on the Swedish CML registry reports increased incidence of a second malignancy with a standardized incidence ratio of 1.52 [24]. Data from cancer

registries in Japan show that the incidence of a second malignancy is the same as that in the general population [25].

No standard therapy exists in advanced lung cancer patients with myeloid malignancy. The combination of dasatinib with EGFR-TKIs erlotinib or gefitinib has been reported as phase I/II clinical trials for advanced non-small-cell lung cancer [26, 27]. However, there has been no report on the concomitant use of EGFR-TKI and ABL-TKI in a patient with concomitant lung adenocarcinoma and CML, and only the alternating erlotinib and imatinib therapy has been reported in a case of concomitant EGFR-mutated lung adenocarcinoma and c-kit-mutated gastrointestinal stromal tumor [28]. Ogata et al. report that EGFR-TKI induced severe neutropenia in a case of EGFR-mutated lung adenocarcinoma with chronic myelomonocytic leukemia, suggesting that EGFR-TKI may cause severe hematological side effects in patients with hematological malignancy [29]. In the current case, the combination of gefitinib and nilotinib was safely administered for seven months.

CYP3A4 is the most abundant CYP enzyme in the liver and intestine, and approximately 50% of the currently available drugs are metabolized by CYP3A4 [6]. The drug-drug interaction studies are usually conducted in the course of drug development. However, *in vivo* quantitative data are often lacking for most drug combinations.

We have previously established a quantitative prediction framework of CYP3A4-mediated oral drug interactions. The estimated AUC increases were within a range of 0.5- to 2.0-fold of the observed AUC increases in 57 out of 60 drug

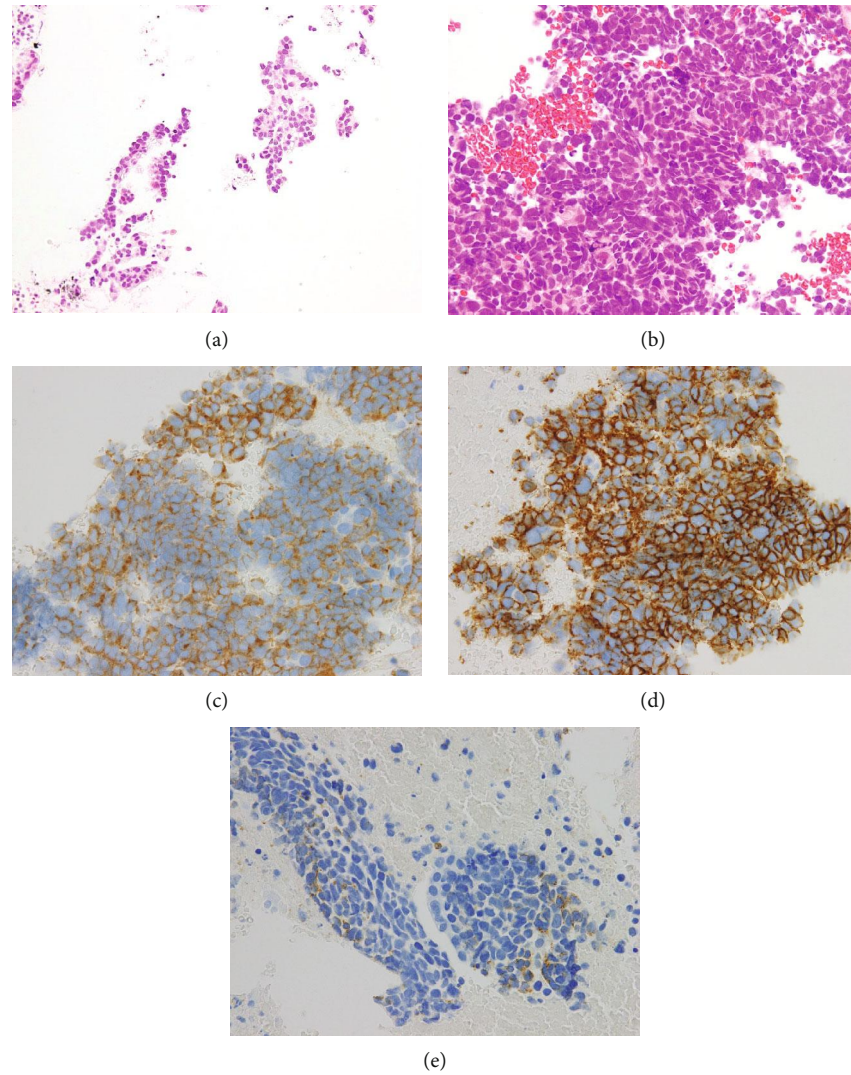


FIGURE 2: Microscopic features of lung cancer. (a) Adenocarcinoma cells from the first biopsy of a mediastinal lymph node. (b) Small cell transformation from the second biopsy of the hilar lymph node (hematoxylin and eosin stain; original magnifications: (a) ×200; (b) ×400). (c-e) Immunohistochemistry of small cell carcinoma cells with neuroendocrine markers. Synaptophysin (c), CD56 (d), and chromogranin A (e) (c-e, ×400).

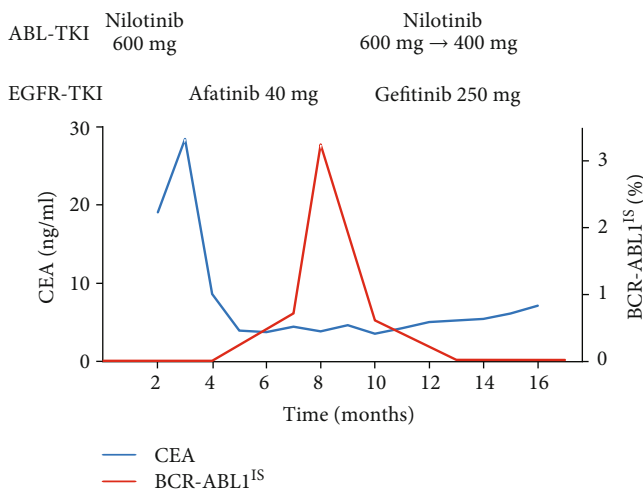


FIGURE 3: Serum CEA levels and BCR-ABL1 transcript levels on the international scale (BCR-ABL1^{IS}) during the treatment.

TABLE 1: Contribution ratio of CYP3A4 to clearance of EGFR-TKIs after oral absorption (CR_{CYP3A4}).

	Calculated CR _{CYP3A4}	Standard inhibitor*	AUC fold change by standard inhibitor	Reference
Gefitinib	0.46	Itraconazole	1.78	[13]
Erlotinib	0.40	Ketoconazole	1.68	[14]
Afatinib	0**			
Osimertinib	0.20	Itraconazole	1.24	[15]

*IR_{CYP3A4} values for itraconazole and ketoconazole are 0.95 and 1.00, respectively (Ref. [5]). **Substrate for the P-glycoprotein (P-gp). Metabolism by cytochrome P-450 is of negligible (Ref. [16]).

combinations [5, 6]. By using this framework, the contribution of CYP3A4 to clearance of EGFR-TKIs after oral absorption can be quantitatively compared between different drugs.

For example, osimertinib has lower value of CR_{CYP3A4} than the first generation EGFR-TKIs (gefitinib and erlotinib) (Table 1), and its plasma concentration is predicted to be less influenced by a CYP3A4 inhibitor.

Dasatinib is an inhibitor of CYP3A4 and its IR_{CYP3A4} is calculated to be 0.16 based on the 1.2-fold AUC increase of CYP3A4 substrate simvastatin [30]. Using the equation $1/(1 - CR_{CYP3A4} \times IR_{CYP3A4})$, the AUC of erlotinib is predicted to increase only by 1.07. In agreement with the prediction, the AUC of erlotinib was not affected by dasatinib in the phase I/II clinical trial of dasatinib and erlotinib for advanced non-small-cell lung cancer [26].

In summary, we have reported for the first time the safety and efficacy of dual EGFR and ABL tyrosine kinase inhibitor treatment in a patient with concomitant EGFR-mutated lung adenocarcinoma and BCR-ABL1-positive CML. The combination of gefitinib and nilotinib was safely administered for seven months. The limitation of the present report is that we did not measure the actual plasma concentrations of gefitinib with or without nilotinib. Further reports are needed to establish the safety and efficacy of TKI combination therapy.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

K.W. and Y.O. received lecture fees from Chugai Pharmaceutical Company. H.K. and T.N. received lecture fees from AstraZeneca, Chugai Pharmaceutical Company, and Boehringer Ingelheim. H.S. received research grants from Chugai Pharmaceutical Company and Boehringer Ingelheim. M.K. received consulting fees from Daiichi Sankyo Company. M.K. received honoraria from Chugai Pharmaceutical Company, Boehringer Ingelheim, Takeda Pharmaceutical Company, Yakult Honsha Company, Bristol-Myers Squibb, and Otsuka Pharmaceutical Company. M.K. received research grants from Chugai Pharmaceutical Company, Takeda Pharmaceutical Company, Pfizer, Novartis Pharma, Bristol-Myers Squibb, and Otsuka Pharmaceutical Company. K.T. received lecture fees from Chugai Pharmaceutical Company, Daiichi Sankyo Company, and Bristol-Myers Squibb.

References

- [1] R. Hehlmann, M. C. Müller, M. Lauseker et al., "Deep molecular response is reached by the majority of patients treated with imatinib, predicts survival, and is achieved more quickly by optimized high-dose imatinib: results from the randomized CML-study IV," *Journal of Clinical Oncology*, vol. 32, no. 5, pp. 415–423, 2014.
- [2] K. Park, E. H. Tan, K. O'Byrne et al., "Afatinib versus gefitinib as first-line treatment of patients with EGFR mutation-positive non-small-cell lung cancer (LUX-Lung 7): a phase 2B, open-label, randomised controlled trial," *The Lancet Oncology*, vol. 17, no. 5, pp. 577–589, 2016.
- [3] J. C. Soria, Y. Ohe, J. Vansteenkiste et al., "Osimertinib in untreated EGFR-mutated advanced non-small-cell lung cancer," *The New England Journal of Medicine*, vol. 378, no. 2, pp. 113–125, 2018.
- [4] K. Azuma, T. Hirashima, N. Yamamoto et al., "Phase II study of erlotinib plus tivantinib (ARQ 197) in patients with locally advanced or metastatic EGFR mutation-positive non-small-cell lung cancer just after progression on EGFR-TKI, gefitinib or erlotinib," *ESMO Open*, vol. 1, no. 4, 2016.
- [5] Y. Ohno, A. Hisaka, and H. Suzuki, "General framework for the quantitative prediction of CYP3A4-mediated oral drug interactions based on the AUC increase by coadministration of standard drugs," *Clinical Pharmacokinetics*, vol. 46, no. 8, pp. 681–696, 2007.
- [6] A. Hisaka, M. Kusama, Y. Ohno, Y. Sugiyama, and H. Suzuki, "A proposal for a pharmacokinetic interaction significance classification system (PISCS) based on predicted drug exposure changes and its potential application to alert classifications in product labelling," *Clinical Pharmacokinetics*, vol. 48, no. 10, pp. 653–666, 2009.
- [7] European Medicines Agency, "European public assessment report (EPAR) product information for nilotinib," September 2019, <https://www.ema.europa.eu/en/medicines/human/EPAR/tasigna>.
- [8] United States Food and Drug Administration, "FDA drug label for nilotinib," September 2019, https://www.accessdata.fda.gov/drugsatfda_docs/label/2018/022068s029lbl.pdf.
- [9] European Medicines Agency, "European public assessment report (EPAR) product information for afatinib," September 2019, <https://www.ema.europa.eu/en/medicines/human/EPAR/giotrif>.
- [10] United States Food and Drug Administration, "FDA drug label for afatinib," September 2019, https://www.accessdata.fda.gov/drugsatfda_docs/label/2018/201292s014lbl.pdf.
- [11] European Medicines Agency, "European public assessment report (EPAR) product information for gefitinib," September 2019, <https://www.ema.europa.eu/en/medicines/human/EPAR/iressa>.
- [12] European Medicines Agency, "European public assessment report (EPAR) product information for erlotinib," September 2019, <https://www.ema.europa.eu/en/medicines/human/EPAR/tarceva>.
- [13] H. C. Swaisland, M. Ranson, R. P. Smith et al., "Pharmacokinetic drug interactions of gefitinib with rifampicin, itraconazole and metoprolol," *Clinical Pharmacokinetics*, vol. 44, no. 10, pp. 1067–1081, 2005.
- [14] A. Rakhit, M. P. Pantze, S. Fettner et al., "The effects of CYP3A4 inhibition on erlotinib pharmacokinetics: computer-based simulation (SimCYP) predicts in vivo metabolic inhibition," *European Journal of Clinical Pharmacology*, vol. 64, no. 1, pp. 31–41, 2008.
- [15] K. Vishwanathan, P. A. Dickinson, K. So et al., "The effect of itraconazole and rifampicin on the pharmacokinetics of osimertinib," *British Journal of Clinical Pharmacology*, vol. 84, no. 6, pp. 1156–1169, 2018.
- [16] P. Stopfer, K. Marzin, H. Narjes et al., "Afatinib pharmacokinetics and metabolism after oral administration to healthy male volunteers," *Cancer Chemotherapy and Pharmacology*, vol. 69, no. 4, pp. 1051–1061, 2012.
- [17] H. Zhang, J. Sheng, J. H. Ko et al., "Inhibitory effect of single and repeated doses of nilotinib on the pharmacokinetics of CYP3A substrate midazolam," *Journal of Clinical Pharmacology*, vol. 55, no. 4, pp. 401–408, 2015.

- [18] J. Baselga, D. Rischin, M. Ranson et al., “Phase I safety, pharmacokinetic, and pharmacodynamic trial of ZD1839, a selective oral epidermal growth factor receptor tyrosine kinase inhibitor, in patients with five selected solid tumor types,” *Journal of Clinical Oncology*, vol. 20, no. 21, pp. 4292–4302, 2002.
- [19] M. Hidalgo, L. L. Siu, J. Nemunaitis et al., “Phase I and pharmacologic study of OSI-774, an epidermal growth factor receptor tyrosine kinase inhibitor, in patients with advanced solid malignancies,” *Journal of Clinical Oncology*, vol. 19, no. 13, pp. 3267–3279, 2001.
- [20] C. Tanaka, O. Q. P. Yin, T. Smith et al., “Effects of rifampin and ketoconazole on the pharmacokinetics of nilotinib in healthy participants,” *Journal of Clinical Pharmacology*, vol. 51, no. 1, pp. 75–83, 2011.
- [21] National Cancer Institute, “Common terminology criteria for adverse events. v5.0,” September 2019, https://ctep.cancer.gov/protocolDevelopment/electronic_applications/ctc.htm#ctc_50.
- [22] K. Kaneshiro, K. Takatsuki, K. Kawase, M. Matsumoto, and Y. Minami, “A case of lung adenocarcinoma complicated with chronic myeloid leukemia with a successful response to gefitinib,” *Japanese Journal of Lung Cancer*, vol. 57, no. 6, pp. 787–790, 2017.
- [23] D. Verma, H. Kantarjian, S. S. Strom et al., “Malignancies occurring during therapy with tyrosine kinase inhibitors (TKIs) for chronic myeloid leukemia (CML) and other hematologic malignancies,” *Blood*, vol. 118, no. 16, pp. 4353–4358, 2011.
- [24] N. Gunnarsson, L. Stenke, M. Höglund et al., “Second malignancies following treatment of chronic myeloid leukaemia in the tyrosine kinase inhibitor era,” *British Journal of Haematology*, vol. 169, no. 5, pp. 683–688, 2015.
- [25] T. Nakazato, N. Iriyama, M. Tokuhira et al., “Incidence and outcome of second malignancies in patients with chronic myeloid leukemia during treatment with tyrosine kinase inhibitors,” *Medical Oncology*, vol. 35, no. 7, 2018.
- [26] E. B. Haura, T. Tanvetyanon, A. Chiappori et al., “Phase I/II study of the Src inhibitor dasatinib in combination with erlotinib in advanced non-small-cell lung cancer,” *Journal of Clinical Oncology*, vol. 28, no. 8, pp. 1387–1394, 2010.
- [27] M. L. Johnson, G. J. Riely, N. A. Rizvi et al., “Phase II trial of dasatinib for patients with acquired resistance to treatment with the epidermal growth factor receptor tyrosine kinase inhibitors erlotinib or gefitinib,” *Journal of Thoracic Oncology*, vol. 6, no. 6, pp. 1128–1131, 2011.
- [28] T. Miyoshi, R. Mori, S. Amano et al., “Efficacy of erlotinib and imatinib in a patient with a rectal gastrointestinal stromal tumor and synchronous pulmonary adenocarcinoma: a case report,” *The Journal of Medical Investigation*, vol. 63, no. 1.2, pp. 144–148, 2016.
- [29] H. Ogata, I. Okamoto, G. Yoshimoto et al., “Chronic myelomonocytic leukemia blast crisis in a patient with advanced non-small cell lung cancer treated with EGFR tyrosine kinase inhibitors,” *Respiratory Investigation*, vol. 55, no. 2, pp. 181–183, 2017.
- [30] European Medicines Agency, “European public assessment report (EPAR) product information for dasatinib,” February 2020, <https://www.ema.europa.eu/en/medicines/human/EPAR/sprycel>.