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Effects of Mannan Oligosaccharides on Gas Emission, Protein and Energy Utilization, and Fasting Metabolism in Sheep

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Simple Summary: Mannan oligosaccharides (MOS) are a promising feed additive to improve animal health, immune capacity, and antioxidation. Based on the previous studies, we carried out three experiments to investigate the effects of MOS on the gas emission, protein and energy utilization, and fasting metabolism of sheep. The results showed that 2.0% MOS supplementation led to the lowest *in vitro* CO₂ production and lower CH₄ production and decreased *in vivo* intake. However, it also decreased urine nitrogen excretion and energy released as CH₄, and then improved the utilization of crude protein and energy of sheep. There were no differences in the parameters of respiration and energy metabolism of sheep under the fasting condition. The findings indicated that MOS slightly affected the gas emission and nutrients and energy utilization of sheep.

Abstract: This study investigated the effects of mannan oligosaccharides (MOS) on *in vitro* and *in vivo* gas emission, utilization of crude protein (CP) and energy, and relative parameters of sheep under fasting metabolism conditions. *In vitro* gas productions were evaluated over 12 h in sheep diets containing different amounts of MOS (from 0% to 6.0%/kg, the increment was 0.5%). A control experiment was used to assess the gas emission, utilization of CP and energy, and fasting metabolism in control sheep and sheep treated with 2.0% MOS over 24 days (d). The results showed that 2.0% MOS supplementation led to the lowest *in vitro* CO₂ production and less CH₄ production, while also leading to decrease *in vivo* nutrients intake, CP and energy excretion, digested and retained CP, and energy released as CH₄ (p < 0.05). Furthermore, 2.0% MOS supplementation appeared to decrease *in vivo* O₂ consumption and CH₄ production per metabolic body weight (BW^{0.75}), and increase the CP retention rate of sheep (p < 0.074). MOS did not affect other parameters, along with the same parameters of sheep under fasting metabolism conditions (p > 0.05). The findings indicate MOS has only slight effects on the gas emission and nutrients and energy metabolism of sheep.

Keywords: energy; gas; mannan oligosaccharides; protein; sheep

1. Introduction

Prebiotics are non-digestible food ingredients that benefit the host by selectively stimulating the activity of one or a limited number of bacteria in the intestine [1,2]. The predominant prebiotics include oligosaccharides such as fructooligosaccharides (FOS), inulin, mannan oligosaccharides (MOS), and xylooligosaccharides [1]. They have been termed a nutricine, meaning that they have no direct nutritive value, but maintain intestinal digestive and absorptive functions, thus improving the health and performance of farmed animals [3].

MOS are found in the outer layer of yeast, *Saccharomyces cerevisiae*, in the cell wall. As the most bioactive compounds, they contain both mannan proteins and complex carbohydrates, including β -glucans [4], and are widely used as a dietary supplement to boost the immune system and eliminate pathogens from the intestinal tract [5,6]. For instance, MOS bind to the mannose-specific lectin of Gram-negative pathogens that express Type-1 fimbriae (e.g., *Escherichia coli*), resulting in their excretion from the intestine [7]. The limited research focusing on MOS in ruminants found that MOS improved the immunoglobulin G (IgG) concentration of lamb's blood [8], enhanced the health of the ruminal epithelium of sheep [9], improved the colostrum quantity of cows [4], and increased the antioxidant capacity of sheep [10]. In previous study, β 1-4 galacto-oligosaccharides (GOS) reduced the CH₄ production in sheep [11]; however, very few scientists have studied the effect of MOS on the energy metabolism and gas emission of ruminants. So, we hypothesized that MOS could adjust the CO₂ and CH₄ emission in the rumen and improve the energy utilization of sheep. The purpose of this study was to investigate the effects of MOS on *in vitro* and *in vivo* gas production, respiration, protein digestion, energy metabolism, and fasting metabolism in sheep.

2. Materials and Methods

2.1. Experimental Design, Animals, and Housing

All experiments in this study were carried out in accordance with the approved guidelines of the Regulation of the Standing Committee of Gansu People's Congress. All experimental protocols and the collection of samples were approved by the Ethics Committee of Gansu Agriculture University under permission no. DK-005.

A single-factor experimental design was used for the *in vitro* gas production experiments. Thirteen different doses of MOS (Bio-Mos[®], Alltech, Nicholasville, KY, USA) were tested: 0, 0.5, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, and 6.0%/kg of basal diet (as fed basis). Each treatment was repeated three times. Four Chinese Northeast Merino rams with an eternal ruminal fistula and similar body weights (64.82 ± 3.17 kg) were used as ruminal fluid donors, and were fed basal diet and water ad libitum. An automatic recording device of trace gas production with six channels (Branch of Animal Husbandry, Jilin Academy of Agricultural Sciences, Gongzhuling, Jilin Province, China) equipped with gas flowmeters (CH₄ and CO₂, Sensors, Inc., Saline, MI, USA; H₂, CITY Technology, Inc., Hampshire, UK) was used to measure the gas flow. The *in vitro* tests lasted for 12 h.

A control experiment was used to assess in vivo CO₂, CH₄, and NH₃ production, O₂ consumption, urine nitrogen excretion, total heat output, and the apparent digestibility and retention rates of crude protein (CP) and energy in control sheep and sheep treated with 2.0% MOS (because adding 2.0% MOS to the basal diet led to the lowest CO_2 and lower CH_4 production in an *in vitro* experiment; then, this dose was chosen to apply in an *in vivo* experiment). The experimental group consisted of eight healthy rams (*Dorper* $\circ X Small tail Han-yang <math> \varphi$; four rams per group) with similar body weights $(42.50 \pm 3.24 \text{ kg})$. The test period included a 14-day (d) acclimation period, a 6-d digestive and metabolic experiment, a 60-hour (h) fasting acclimation period (the respiratory quotient (RQ) approximately was below 0.71 when sheep fasted for over 60 h), and a 24-h fasting metabolism experiment. All rams were housed individually in a respiratory chamber (Branch of Animal Husbandry, Jilin Academy of Agricultural Sciences, Gongzhuling, Jilin Province, China). Each chamber was equipped with a feeder and a drinker that provided ad libitum access to feed and water. Each chamber was also equipped with automatic gas flowmeters (CH₄ and CO₂, Sensors, Inc., Saline, MI, USA; O₂, Advanced Micro Instruments, Inc., Costa Mesa, CA, USA; NH₃, Industrial Health & Safety Instrumentation, Inc., St. Petersburg, FL, USA). Before rams went into the respiratory chamber, the 6.14-L calibrating CO₂ was pumped into the respiratory chamber to test the recovery rate, and repeated three times; the result of the gas recovery tests was 98.22% (Table S1). The data on gas production and O_2 consumption were calibrated by the gas recovery rate of each chamber.

2.2. Experimental Diets

The basal diet was formulated to meet or exceed the recommendations for all nutrients of ram sheep under China Agricultural Industry Standard NY/T816-2004 (Table 1).

Items	Concentrate		
Ingredient (%)			
Corn	24.90		
Soybean meal	12.72		
Chinese wildrye	60.00		
Calcium hydrophosphate	0.78		
Limestone	0.80		
Salt	0.40		
Additive premix ¹	0.40		
Chemical composition (%) ²			
DM	85.71		
DE (MJ/kg) ³	9.25		
CP	9.46		
DE/CP (MJ/g)	0.10		
NDF	46.06		
ADF	23.75		
Ca	0.65		
Р	0.40		

Table 1. Ingredient and	d chemical com	position of	the basal diet.
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¹ Additive premix includes mineral elements (mg/kg): S, 200; Fe, 25; Zn, 40; Cu, 8; I, 0.3; Mn, 40; Se, 0.2; Co, 0.1; vitamins (IU/kg): vitamin A, 940; vitamin E, 20. ² Concentrations of dry matter (DM), crude protein (CP), Ca, and P were measured in accordance with AOAC (DM: method 930.15, CP: method 990.03, Ca: method 978.02, P: method 946.06) [12], and concentrations of neutral detergent fiber (NDF) and acid detergent fiber (ADF) were measured in accordance with Goering and Soest [13]. ³ DE = digestible energy.

2.3. In Vitro Procedures

Rumen contents from each donor sheep were obtained immediately before morning feeding by soft tube and syringe via ruminal fistula and strained through four layers of cheesecloth. Then, the fluid from each sheep was pooled and mixed with culture medium solution in a 1:2 ratio (vol/vol) at 39 °C in accordance with the description by Menke et al. [14], and the mixed solution was also constantly filled with N_2 to maintain anaerobic condition. Before incubation, in order to eliminate the metering error, 0.2843% calibrating CO₂ was pumped into the channel, and then the gas flowmeter was calibrated to 0.2843%. All in vitro cultures contained 2 g of experimental diets, which were carefully weighed into the fermentation channel of the automatic recording device of trace gas production, while the control channel did not receive any diet in order to calibrate the gas production from the ruminal fluid. Additionally, different doses of MOS were added into the channel in terms of experimental treatment. Then, each channel received 150 mL of mixed solution (including 50 mL of ruminal fluid and 100 mL of culture medium solution), and was maintained at 39 °C for 12 h of incubating. Since there were six channels for incubation, the culture was divided into nine periods, which meant that control, 0%, 0.5%, 1.0%, 1.5%, and 2.0% MOS treatments were carried out during the first period; control, 2.5%, 3.0%, 3.5%, 4.0%, and 4.5% MOS treatments were carried out during the second period; control, 5.0%, 5.5%, and 6.0% MOS treatments were carried out during the third period; and then, this procedure was repeated two times during the fourth to ninth periods. The productions of CO₂, CH₄, and H₂ were recorded by a gas flowmeter automatically per every 6 min. Then, the gas production was calculated per min from a total of 12 h of incubated gas production, after which the gas production per every 24 h was calculated.

2.4. In Vivo Procedures

In the acclimation period and digestive and metabolic experiments, all sheep received a basal diet or 2.0% MOS-treated diet randomly. Since they were housed in the respiratory chamber, the productions of CO_2 , CH_4 , NH_3 , and H_2 along with the consumption of O_2 were measured using the gas flowmeter automatically per every 27 min.

The weight of feed intake and residues were recorded daily carefully. During the digestive and metabolic experiments, 10% of total diet, 10% of total feces output, and 5% of total urine output (5 mL of sulfuric acid added into total urine daily before collection to prevent nitrogen losses) were sampled for 6 sequential d and stored at -20 °C. At the end of the data collection period, the diets, fecal samples, and urine samples were thawed and pooled for each sheep. For nitrogen analysis, 3% of total feces output was sampled daily and stored in wide-mouth bottles containing 20 mL of 10% sulfuric acid for nitrogen fixation, and these samples were also pooled for 6 d individually.

After the digestive and metabolic experiments, sheep fasted approximately over 60 h until the RQ was below 0.71. Then, the sheep fasting metabolism experiment was carried out. The procedures of gas production and O_2 consumption recording, feces output sampling, and urine output sampling was as previously stated, but the recording and sampling times were 24 h.

2.5. Chemical Analysis

The diets and fecal samples were dried at 65 °C for 72 h in a forced-air oven, and then ground through a 1-mm screen using a Wiley Mill (Ogaw Seiki Co., Ltd., Tokyo, Japan). The gross energy of diet, feces, and urine samples were measured using a calorimeter (C2000, IKA[®]-Werke GmbH & Co. KG, Staufen, Germany). The nitrogen contents of the diet, feces containing 10% sulfuric acid, and urine samples were determined by the Kjeldahl method (AOAC, method 990.03) [12].

2.6. Calculations and Statistical Analysis

A gas flowmeter records the dynamic gas production or consumption; therefore, the total gas production or consumption is calculated using the following equations:

In vitro gas production (mL/24 h) = average production (mL/min) \times 60 \times 24;

In vivo gas production/consumption (L/24 h) = average production/consumption (L/min) \times 60 \times 24.

Parameters related to energy metabolism are calculated using the following equations [15]:

Total heat output (J) = $16.175 \times O_2$ (L) + $5.021 \times CO_2$ (L) - $2.167 \times CH_4$ (L) - $5.987 \times Urine$ nitrogen (g).

Oxidation of protein (g) = Urine nitrogen (g) \times 6.25.

 CO_2 production from the oxidation of protein (L) = Urine nitrogen (g) × 4.754.

 O_2 consumption from the oxidation of protein (L) = Urine nitrogen (g) \times 5.923.

Heat output from the oxidation of protein (J) = Urine nitrogen (g) \times 113.76.

Data from the *in vitro* experiment were analyzed by one-way analysis of variance (SPSS 19.0, IBM Co. Limited, Chicago, IL, USA) using the following model:

$$X_{ij} = \mu + \alpha_i + e_{ij} \tag{1}$$

where X_{ij} is the observation of the dependent variable (i = 1 to 13, j = 1 to 3), μ is the population mean, α_i is the random effect of treatment, and e_{ij} is the random error associated with the observation.

Data from the *in vivo* experiments were analyzed by an independent samples *t* test.

Significance was determined at $p \le 0.05$ and tendency at 0.05 using Tukey's multiple comparison tests.

3. Results

3.1. In Vitro CO₂, CH₄ and H₂ Production

The 24-h *in vitro* CO₂, CH₄, and H₂ production under different amounts of adding MOS are shown in Table 2. MOS influenced *in vitro* CO₂ and CH₄ productions significantly (p < 0.05). An MOS of 3.5% led to the highest production of CO₂ and CH₄, while 2.0% MOS resulted in the lowest amount of CO₂ emissions, and 0.5% and 5.0% MOS resulted in the lowest CH₄ emissions (p < 0.05). No differences in H₂ production were observed between different MOS doses (p > 0.05).

Table 2. *In vitro* CO₂, CH₄, and H₂ production with different amounts of adding mannan oligosaccharides (MOS).

MOS (%) ¹	CO ₂ (mL/24 h)	CH ₄ (mL/24 h)	H ₂ (mL/24 h)
0	161.52 ^{cde}	27.87 ^{bc}	0.11
0.5	153.03 ^{cde}	26.21 ^c	0.06
1.0	155.61 ^{cde}	28.29 ^{bc}	0.04
1.5	145.64 ^{de}	28.50 ^{bc}	0.10
2.0	143.65 ^e	27.65 ^{bc}	0.10
2.5	169.92 ^{abc}	30.80 ^{abc}	0.05
3.0	169.16 ^{abc}	30.88 ^{abc}	0.06
3.5	189.27 ^a	36.58 ^a	0.08
4.0	186.53 ^{ab}	34.66 ^{ab}	0.09
4.5	172.96 ^{abc}	28.92 ^{abc}	0.11
5.0	162.40 ^{cde}	26.64 ^c	0.10
5.5	166.69 ^{bcd}	27.73 ^{bc}	0.09
6.0	162.60 ^{cde}	27.12 ^{bc}	0.10
SEM ²	2.05	0.58	0.01
<i>p</i> -value	< 0.001	0.001	0.841

¹ MOS doses were 0, 0.5, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, and 6.0% added to a 2-g basal diet. Mean results of *in vitro* gas production are shown for yield per 24 h (n = 3 per treatment, *in vitro* gas production (mL/24 h) = average production (mL/min) × 60 × 24). ² SEM: standard error of mean. Values within a column with different superscripts differ significantly at p < 0.05.

3.2. In Vivo Gas Production, CP, and Energy Digestion and Retention of Sheep

The CO₂, CH₄, and NH₃ production, O₂ consumption, urine nitrogen excretion, total heat output, and relative parameters regarding the energy metabolism of the control and 2.0% MOS-treated sheep are shown in Table 3. MOS did not affect RQ, CO₂, CH₄, and NH₃ production, O₂ consumption, and total heat output (p > 0.05). However, MOS decreased the urine nitrogen excretion and parameters of protein oxidation related to the energy metabolism of sheep (p < 0.05). Furthermore, MOS tended to decrease O₂ consumption and CH₄ production per metabolic body weight (BW^{0.75}), along with the ratio of heat output from protein oxidation to total heat output (p < 0.074).

The effects of MOS on sheep dry matter intake (DMI), CP, and energy digestion and retention are shown in Table 4. The supplementation of MOS decreased dry matter (DM), CP, and energy intake, CP and energy excreted in feces and urine, digested and retained CP, and the energy released as CH₄ in sheep (p < 0.05). The addition of MOS did not affect the apparent digestibility of CP and energy, retention of energy, digestible energy (DE), and metabolizable energy (ME) (p > 0.05). However, MOS tended to increase the retention rate of CP (p < 0.054).

Item ¹		Control	MOS-Treated	SEM ⁹	<i>p</i> -Value
RQ ²		0.86	0.87	0.002	0.328
CO ₂ production	(L/24 h)	375.05	372.29	4.04	0.111
CO ₂ production	(L/24 h/kg BW ^{0.75}) ³	20.77	20.58	0.20	0.641
CH ₄ production	(L/24 h)	21.64	20.22	0.40	0.600
err ₄ production	(L/24 h/kg BW ^{0.75})	1.20	1.12	0.02	0.072
NH ₃ production	(L/24 h)	32.58	33.43	0.61	0.923
ini is production	(L/24 h/kg BW ^{0.75})	1.80	1.85	0.03	0.507
O_2 consumption	(L/24 h)	438.37	430.69	4.96	0.074
O ₂ consumption	(L/24 h/kg BW ^{0.75})	24.27	23.81	0.25	0.358
Urine nitrogen excretion	(g/24 h)	14.87 ^a	13.56 ^b	0.30	0.028
offile introgen excietion	(g/24 h/kg BW ^{0.75})	0.82 ^a	0.75 ^b	0.02	0.035
Tatal bast subset 4	(MJ/24 h)	8.65	8.57	0.10	0.694
Total heat output ⁴	(MJ/24 h/kg BW ^{0.75})	0.48	0.47	0.01	0.553
Oridation of muchain 5	(g/24 h)	92.93 ^s	84.78 ^b	1.88	0.028
Oxidation of protein ⁵	(g/24 h/kg BW ^{0.75})	5.15 ^a	4.70 ^b	0.11	0.035
CO ₂ production from the	(L/24 h)	70.69 ^a	64.48 ^b	1.43	0.028
oxidation of protein ⁶	(L/24 h/kg BW ^{0.75})	3.92 ^a	3.57 ^b	0.08	0.035
O ₂ consumption from the	(L/24 h)	88.07 ^a	80.34 ^b	1.78	0.028
oxidation of protein ⁷	(L/24 h/kg BW ^{0.75})	4.88 ^a	4.45 ^b	0.10	0.035
Heat output form the	(MJ/24 h)	1.69 ^a	1.54 ^b	0.03	0.028
oxidation of protein ⁸	(MJ/24 h/kg BW ^{0.75})	0.094 ^a	0.086 ^b	0.002	0.035
The ratio of heat output from to total heat ou	the oxidation of protein	19.55	18.12	0.39	0.069

Table 3. In vivo gas production and energy metabolism of control and MOS-treated sheep.

¹ Sheep were fed diets containing 0% and 2.0% MOS (*n* = 4 per treatment). Mean results of CO₂, CH₄, and NH₃ production, O₂ consumption, urine nitrogen excretion, total heat output, and relative parameters regarding energy metabolism are shown for the 6-d collection phase of the study for each treatment. The *in vivo* gas production or consumption was calculated using the following equation: *in vivo* gas production/consumption (L/24 h) = average production/consumption (L/min) × 60 × 24. ² RQ = respiratory quotient. ³ BW^{0.75} = Metabolic body weight. ⁴ Total heat output (J) = 16.175 × O₂ (L) + 5.021 × CO₂ (L) – 2.167 × CH₄ (L) – 5.987 × Urine nitrogen (g). ⁵ Oxidation of protein (g) = Urine nitrogen (g) × 6.25. ⁶ CO₂ production from the oxidation of protein (L) = Urine nitrogen (g) × 4.754. ⁷ O₂ consumption from the oxidation of protein (L) = Urine nitrogen (g) × 113.76. ⁹ SEM: standard error of the mean. Values within a row with different superscripts differ significantly at *p* < 0.05.

Table 4. In vivo nutrients and energy digestion and retention of control and MOS-treated sheep.

Ite	em ¹	Control	MOS-Treated	SEM ¹¹	<i>p</i> -Value
Dry matter inta	Dry matter intake (DMI, kg/24 h)		1.48 ^b	0.02	< 0.001
Crude pi	rotein (CP)				
	(g/24 h)	182.96 ^a	163.56 ^b	2.27	< 0.001
CP intake	$(g/24 h/kg BW^{0.75})^2$	10.07 ^a	9.22 ^b	0.13	< 0.001
	(g/24 h)	46.90 ^a	38.78 ^b	2.03	0.044
CP in feces	(g/24 h/kg BW ^{0.75})	2.51	2.12	0.12	0.107
	(g/24 h)	92.93 ^a	84.78 ^b	1.88	0.028
CP in urine	(g/24 h/kg BW ^{0.75})	9.40	9.41	0.19	0.976
\mathbf{D}^{*} (1 $\mathbf{C}\mathbf{D}^{3}$	(g/24 h)	136.05 ^a	124.78 ^b	2.14	0.007
Digested CP ³	(g/24 h/kg BW ^{0.75})	7.56	7.09	0.12	0.058
n	(g/24 h)	43.12 ^a	40.00 ^b	0.66	0.016
Retained CP ⁴	(g/24 h/kg BW ^{0.75})	2.34	2.28	0.03	0.339
Apparent dig	Apparent digestibility (%) ⁵		76.40	1.02	0.336
Retention rate (%) ⁶		23.56	24.42	0.22	0.054

Item	L	Control	MOS-Treated	SEM ¹¹	<i>p</i> -Value
Energ	у				
Energy intake	(MJ/24 h)	35.39 ^a	31.64 ^b	0.44	< 0.001
Energy intake	(MJ/d/kg BW ^{0.75})	1.96 ^a	1.73 ^b	0.03	< 0.001
Enorgy in faces	(MJ/24 h)	12.69 ^a	10.05 ^b	0.61	0.028
Energy in feces	(MJ/d/kg BW ^{0.75})	0.74 ^a	0.48 ^b	0.03	< 0.001
Energy in urine	(MJ/24 h)	0.46 ^a	0.42 ^b	0.01	0.028
Energy in unite	(MJ/d/kg BW ^{0.75})	0.03	0.02	0.001	0.301
Energy in CH₄	(MJ/24 h)	0.86 ^a	0.80 ^b	0.01	0.015
Energy in CI1 ₄	(MJ/d/kg BW ^{0.75})	0.05	0.04	0.001	0.094
Discostible on energy (DE) 7	(MJ/24 h)	22.70	21.59	0.46	0.231
Digestible energy (DE) ⁷	(MJ/d/kg BW ^{0.75})	1.22	1.25	0.02	0.549
Metabolizable energy (ME)	(MJ/24 h)	21.40	20.39	0.45	0.270
8 8	(MJ/d/kg BW ^{0.75})	1.15	1.18	0.02	0.470
Apparent digestibility (%) 9		64.34	68.59	1.52	0.165
Retention rate (%) ¹⁰		60.63	64.32	1.48	0.216

Table 4. Cont.

¹ Sheep were fed 0% and 2.0% MOS (n = 4 per treatment). Mean results of DMI, CP, and energy digestion and retention are shown for the 6-d collection phase of the study for each treatment. ² BW ^{0.75} = Metabolic body weight. ³ Digested CP (g) = CP intake – CP in feces. ⁴ Retained CP (g) = CP intake – CP in feces – CP in urine. ⁵ CP apparent digestibility (%) = (CP intake – CP in feces)/(CP intake). ⁶ CP retention rate (%) = (CP intake – CP in feces – CP in urine)/(CP intake). ⁷ Digestible energy (MJ) = energy intake – energy in feces. ⁸ Metabolizable energy (MJ) = energy intake – energy in feces – energy in transparent digestibility (%) = (energy intake – energy in feces)/(energy intake). ¹⁰ Energy retention rate (%) = (energy intake – energy in CH₄) / (energy intake). ¹¹ SEM: standard error of the mean. Values within a row with different superscripts differ significantly at p < 0.05.

3.3. In Vivo Gas Production and Energy Metabolism of Sheep under Fasting Metabolism Conditions

There were no differences in RQ, CO₂ production, O₂ consumption, urine nitrogen excretion, total heat output, and relative parameters regarding energy metabolism between the control and MOS-treated sheep under fasting metabolism conditions (p > 0.05, Table 5).

Table 5. *In vivo* gas production and energy metabolism of control and MOS-treated sheep under fasting metabolism conditions.

Item ¹		Control	MOS-treated	SEM ⁹	<i>p</i> -Value
RQ ²	RO ²		0.69	0.006	0.267
CO_2 production	(L/24 h)	69.00	75.03	3.93	0.849
CO ₂ production	(L/24 h/kg BW ^{0.75}) ³	3.83	4.15	0.22	0.494
O_2 consumption	(L/24 h)	103.45	109.57	5.24	0.921
O ₂ consumption	(L/24 h/kg BW ^{0.75})	5.74	6.06	0.29	0.617
Uning nitragon avanation	(g/24 h)	3.28	2.95	1.52	0.828
Urine nitrogen excretion	(g/24 h/kg BW ^{0.75})	0.18	0.35	0.08	0.490
T (11 ()) 4	(MJ/24 h)	2.03	1.99	0.18	0.783
Total heat output ⁴	(MJ/24 h/kg BW ^{0.75})	0.11	0.11	0.004	0.788
0.11/2 ((g/24 h)	20.53	18.42	3.87	0.828
Oxidation of protein ⁵	(g/24 h/kg BW ^{0.75})	1.26	1.11	0.26	0.824
CO ₂ production from the oxidation of	(L/24 h)	15.61	14.01	2.94	0.828
protein ⁶	(L/24 h/kg BW ^{0.75})	0.96	0.85	0.20	0.824
O ₂ consumption from the oxidation	(L/24 h)	19.45	17.45	3.67	0.828
of protein ⁷	(L/24 h/kg BW ^{0.75})	1.19	1.05	0.24	0.824
Heat output form the oxidation of	(MJ/24 h)	0.37	0.34	0.07	0.828
protein ⁸	(MJ/24 h/kg BW ^{0.75})	0.02	0.02	0.005	0.824
The ratio of heat output from the oxidation of protein to total heat output (%)		19.84	19.09	4.95	0.953

¹ Sheep were under the fasting metabolism conditions (*n* = 4 per treatment). Mean results of CO₂ production, O₂ consumption, urine nitrogen excretion, total heat output, and relative parameters regarding the energy metabolism of control and MOS-treated sheep are shown for the 24-h collection phase of the study for each treatment. The *in vivo* gas production or consumption was calculated using following equation: *in vivo* gas production/consumption (L/24 h) = average production/consumption (L/min) × 60 × 24. ² RQ = respiratory quotient. ³ BW^{0.75} = Metabolic body weight. ⁴ Total heat (J) = 16.175 × O₂ (L) + 5.021 × CO₂ (L) – 5.987 × Urine N (g). ⁵ Oxidation of protein (g) = Urine nitrogen (g) × 6.25. ⁶ CO₂ production from the oxidation of protein (L) = Urine nitrogen (g) × 4.754. ⁷ O₂ consumption from the oxidation of protein (L) = Urine nitrogen (g) × 113.76. ⁹ SEM: standard error of the mean.

4. Discussion

In this study, the MOS addition range from 0.5% to 2.0% decreased in vitro CO₂ and CH₄ production. Basically, carbohydrate was degraded by microbes in the rumen to produce approximately 65.5% CO₂, 28.8% CH₄, and small quantities of N₂, O₂, and H₂ [16]. Although some bicarbonates from saliva through the ruminal wall produce CO₂, the degradation of carbohydrates by ruminal microbes is the major pathway for CO_2 production [16]. In the rumen, methanogenesis evolves from carbon oxide, acetic acids, and methanol by methanogens. For most methanogens, methanogenesis from CO_2 and H_2 is the sole energy source. CH_4 production is considerably enhanced by the CO_2 present in the headspace, and there is an equilibrium established between the dissolved CO_2 in the media and the partial pressure of CO_2 in headspace gas. Meanwhile, the higher CO_2 concentration in the headspace would result in a greater dissolved CO_2 concentration in the media [17,18]. The result of the current study was in accordance with the previous conclusion that a positive relationship existed between CO_2 and CH₄; however, some scientists believed that there was a negative relationship, because CO₂ and H_2 are in general the precursors for CH_4 formation in the rumen [19,20]. This experiment illustrated that in an *in vitro* condition, MOS as an additive had only slight effects on ruminal fermentation, the partial pressure of CO₂, and methanogens and methanogenesis, and adding 0.5–2.0% MOS decreased CO₂ production and led to lower CH₄ production. It might be that MOS doses ranging from 0.5% to 2.0% were appropriate doses for ruminal microbes' fermentation pathway, and most of the produced CO_2 was potentially absorbed by the rumen wall to synthesize some nutrients, such as aspartic acid and isoleucine, and a lower partial pressure of CO_2 and dissolved CO_2 led to lower methanogenesis. Furthermore, higher doses of MOS meant higher oligosaccharides transferred into rumen as a substrate for microbes' fermentation, so, more substrates produced more CO₂, and instead promoted methanogenesis. However, MOS doses ranging from 2.5% to 4.5% generated the highest CO₂ and CH₄ emissions, but MOS doses ranging 5.0% to 6.0% resulted in lower CO₂ and CH₄ production. Further investigation is needed to confirm whether moderate doses of MOS improve carbohydrate fermentation in the rumen and much higher doses of MOS inhibit ruminal fermentation because more soluble carbohydrates led to lower pH and were harmful to microbes [20]. In addition, the fate of CO_2 is more complicated because of the pooling and recycling of animal metabolic carbon as urea and bicarbonates in saliva with that produced by the rumen organisms [21]. Furthermore, the process of methanogenesis is affected by many environmental factors, including the carbohydrate type and digestion passage rate. Feed additives such as yeast have the ability to shift H₂ utilization from methanogenesis to reductive acetogenesis through the homoacetogenic bacteria that can produce acetate from CO_2 and H_2 [22], along with internal factors [23]. So, more research about MOS on ruminal fermentation and the microbial population need to be undertaken in the future.

4.2. In Vivo Gas Production, CP, and Energy Digestion and Retention of Sheep

In the present study, the supplementation of MOS decreased ingestion, CP, and energy excreted in feces and urine, digested and retained CP, energy released as CH_4 , along with the same parameters per $BW^{0.75}$, but MOS did not affect the apparent digestibility of CP and energy. Surprisingly, MOS decreased the ingestion of DM in sheep; the reason may be that there were low replicates in each treatment, indicating that individual differences may be a key factor in the results. Additional experiments with more sheep are needed to verify these effects of MOS.

Although MOS decreased the DMI of sheep, it decreased the CP and energy excretion in feces and urine at the same time; as a result, no differences were observed on the digestion and retention of CP and energy, even though higher digestibility and retention rates occurred in MOS-treated sheep. It indicates that MOS negatively influences the nutrients and energy intake of sheep; however, it makes sheep improve the utilization of nutrients and energy. Accordingly, because of improving the utilization of energy both in the rumen and the whole body of sheep, the methanogenesis in rumen was restricted, and resulted in less energy released as CH₄. A previous study found that GOS significantly increased the DE of sheep fed with Italian ryegrass hay and concentrate (3:2, on a DM basis), but did not affect nitrogen digestion and retention [24]. Other researchers reported that MOS did not affect the apparent digestibility and retention rate of nutrients in cattle and sheep [4,10]; these findings are in accordance with that of Goiri et al. [25], who pointed out that chitosan did not influence the nutrients' apparent digestibility. However, another study indicated that MOS enhanced the health of the ruminal epithelium of sheep by reducing the thickness of the stratum corneum, and it might have increased the nutrients digestibility of CP in Murrah buffalo calves. These findings indicated that MOS improved the CP utilization via decreased urine nitrogen excretion. So, more research about the effects of MOS on nutrients and energy metabolism in rumen also need to be undertaken in the future.

In the current study, treatment with 2.0% MOS resulted in less DM ingestion compared to the control. This led to less O_2 consumption and CO_2 production, because the digestion of feed and metabolism of nutrients in MOS-treated sheep consumed less O_2 and produced less CO_2 . In *in vivo* conditions, CO_2 production from respiration is much larger than ruminal fermentation, and MOS only adjusts ruminal fermentation slightly; so, the decreasing tendency of CH_4 is observed from sheep in the respiratory chamber, not CO_2 production. Thus, it is possible that the basic vital activities of sheep were not affected by MOS. Respiration is a kind of rhythmic vital activity; its frequency and amplitude are determined by the body condition, which depends on nervous humoral regulation. Normally, feed intake cannot change the basic respiration frequency and amplitude, not to mention that there was only a small quantity of MOS; the results of the current study are also in consonance with the physiological theory.

Food ingestion affects the production of body heat in animals; however, for example, animals continuously produce heat and lose it to their surroundings, either directly by radiation, conduction, and convection, or indirectly by the evaporation of water [27]. In the current study, MOS decreased the urine nitrogen excretion and relative protein oxidation in sheep; however, the ratio of heat output from protein oxidation in the body to total heat output was decreased at the same time, indicated that the basal metabolism process is hardly influenced by MOS.

4.3. In Vivo Gas Production and Energy Metabolism of Sheep under Fasting Metabolism Conditions

MOS did not affect the CO₂ production, O₂ consumption, urine nitrogen excretion, total heat output, and relative parameters regarding the energy metabolism of sheep under fasting metabolism conditions in the current study. In a fasting animal, the quantity of heat produced is equal to the energy of the tissue catabolized. When measured under specific conditions, the energy is known as the animal's basal metabolism. A fasting animal must oxidize reserves of nutrients to provide the energy needed for essential processes such as respiration and circulation of the blood [27]. MOS cannot modify primary vital activities; no differences were observed in CO₂ production, O₂ consumption, urine nitrogen excretion, total heat output, and relative parameters regarding energy metabolism in 2.0% MOS-treated sheep and non-MOS fed sheep under fasting metabolism conditions.

5. Conclusions

The supplementation of MOS did not affect *in vitro* H_2 production, *in vivo* CO_2 , CH_4 , and NH_3 production, O_2 consumption, total heat output, the apparent digestibility of CP and energy, the retention rates of energy of sheep, and the respiration and energy metabolism of sheep under fasting metabolism conditions. However, the addition of 2.0% MOS to the sheep diet led to the lowest *in vitro* CO_2 production and less CH_4 production. Furthermore, treatment with 2.0% MOS decreased the intake of DM, CP, and energy, along with the CP and energy in feces and urine, digested and retained CP, and released the energy as CH_4 . Treatment with 2.0% MOS appeared to increase the retention rate of CP in sheep. The results suggest that MOS only slightly affects ruminal fermentation and metabolism in sheep, and the effects were not strong enough to lead to substantial changes.

Supplementary Materials: The following are available online at http://www.mdpi.com/2076-2615/9/10/741/s1, Table S1: Gas recovery rate by calibrating CO₂.

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References

- 1. Adhikari, P.A.; Kim, W.K. Overview of prebiotics and probiotics: Focus on performance, gut health and immunity—a review. *Ann. Anim. Sci.* **2017**, *17*, 949–966. [CrossRef]
- 2. Gibson, G.R.; Roberfroid, M.B. Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *J. Nutr.* **1995**, *125*, 1401–1412. [CrossRef] [PubMed]
- 3. Halas, V.; Nochta, I. Mannan oligosaccharides in nursery pig nutrition and their potential mode of action. *Animals* **2012**, *2*, 261–274. [CrossRef] [PubMed]
- 4. Westland, A.; Martin, R.; White, R.; Martin, J.H. Mannan oligosaccharide prepartum supplementation: Effects on dairy cow colostrum quality and quantity. *Animal* **2017**, *11*, 1779–1782. [CrossRef] [PubMed]
- Najdegerami, E.H.; Tokmachi, A.; Bakhshi, F. Evaluating the Effects of Dietary Prebiotic Mixture of Mannan Oligosaccharide and Poly-β-Hydroxybutyrate on the Growth Performance, Immunity, and Survival of Rainbow Trout, *Oncorhynchus mykiss* (Walbaum 1792), Fingerlings. *J. World Aquacult. Soc.* 2017, 48, 415–425. [CrossRef]
- Fernandez, F.; Hinton, M.; van Gils, B. Dietary mannan-oligosaccharides and their effect on chicken caecal microflora in relation to Salmonella Enteritidis colonization. *Avian Pathol.* 2002, *31*, 49–58. [CrossRef] [PubMed]
- 7. Thomas, W.E.; Nilsson, L.M.; Forero, M.; Sokurenko, E.V.; Vogel, V. Shear-dependent 'stick-and-roll' adhesion of type 1 fimbriated *Escherichia coli*. *Mol. Microbiol.* **2004**, *53*, 1545–1557. [CrossRef] [PubMed]
- 8. Demirel, G.; Turan, N.; Tanor, A.; Kocabagli, N.; Alp, M.; Hasoksuz, M.; Yilmaz, H. Effects of dietary mannanoligosaccharide on performance, some blood parameters, IgG levels and antibody response of lambs to parenterally administered *E. coli* O157:H7. *Arch. Anim. Nutr.* **2007**, *61*, 126–134. [CrossRef] [PubMed]
- 9. Diaz, T.G.; Branco, A.F.; Jacovaci, F.A.; Jobim, C.C.; Bolson, D.C.; Daniel, J.L.P. Inclusion of live yeast and mannan-oligosaccharides in high grain-based diets for sheep: Ruminal parameters, inflammatory response and rumen morphology. *PLoS ONE* **2018**, *13*, e0193313.
- Zheng, C.; Li, F.D.; Hao, Z.L.; Liu, T. Effects of adding mannan oligosaccharides on digestibility and metabolism of nutrients, ruminal fermentation parameters, immunity, and antioxidant capacity of sheep. *J. Anim. Sci.* 2018, *96*, 284–292. [CrossRef]
- 11. Mwenya, B.; Santoso, B.; Sar, C.; Gamo, Y.; Kobayashi, T.; Arai, I.; Takahashi, J. Effects of including β-1,4 galacto-oligosaccharides, lactic acid bacteria or yeast culture on methanogenesis as well as energy and nitrogen metabolism in sheep. *Anim. Feed Sci. Tech.* **2004**, *115*, 313–326. [CrossRef]
- 12. AOAC. Official Methods of Analysis, 17th ed.; Association of Offical Analytical Chemists: Arlington, VA, USA, 2000.
- 13. Goering, H.R.; Soest, P.J.V. *Forage Fiber Analysis. Agricultural Handbook No.* 379; Agricultural Research Service-US Department of Agriculture: Washington, DC, USA, 1970.
- 14. Menke, K.H.; Raab, L.; Salewski, A.; Steingass, H.; Fritz, D.; Schneider, W. The estimation of the digestibility and metabolizable energy content of ruminant feedingstuffs from the gas production when they are incubated with rumen liquor *in vitro*. *J. Agr. Sci.* **1979**, *93*, 217–222. [CrossRef]
- 15. Brouwer, E. Report of Subcommittee on Constants and Factors. In *Proceedings of the 3rd Symposium on Energy Metabolism, Troon, Scotland, May 1964;* EAAP Publ.: Academic Press: London, UK, 1965; pp. 441–443.
- 16. Aschenbach, J.R.; Penner, G.B.; Stumpff, F.; Gäbel, G. Ruminant Nutrition Symposium: Role of fermentation acid absorption in the regulation of ruminal pH. *J. Anim. Sci.* **2011**, *89*, 1092–1107. [CrossRef] [PubMed]

- 17. Patra, A.K.; Yu, Z.T. Effects of gas composition in headspace and bicarbonate concentrations in media on gas and methane production, degradability, and rumen fermentation using *in vitro* gas production techniques. *J. Dairy Sci.* **2013**, *96*, 4592–4600. [CrossRef] [PubMed]
- 18. Alford, J.S., Jr. Measurement of dissolved carbon dioxide. Can. J. Microbiol. 1976, 22, 52–56. [CrossRef]
- 19. Sirohi, S.K.; Pandey, N.; Singh, B.; Puniya, A.K. Rumen methanogens: A review. *Indian J. Microbiol.* **2010**, *50*, 253–262. [CrossRef] [PubMed]
- 20. Parra-Garcia, A.; Elghandour, M.M.M.Y.; Greiner, R.; Barbabosa-Pilego, A.; Camacho-Diaz, L.M.; Salem, A.Z.M. Effects of *Moringa oleifera* leaf extract on ruminal methane and carbon dioxide production and fermentation kinetics in a steer model. *Environ. Sci. Pollut. Res.* **2019**, *26*, 15333–15344. [CrossRef] [PubMed]
- 21. Soest, P.V. Nutritional Ecology of the Ruminant, 2nd ed.; Cornell University Press: New York, NY, USA, 1994.
- Elghandour, M.M.Y.; Kholif, A.E.; López, S.; Mendoza, G.D.; Odongo, N.E.; Salem, A.Z.M. *In vitro* Gas, Methane, and Carbon Dioxide Productions of High Fibrous Diet Incubated With Fecal Inocula From Horses in Response to the Supplementation With Different Live Yeast Additives. *J. Equine Vet. Sci.* 2016, 38, 64–71. [CrossRef]
- Zhao, L.P.; Meng, Q.X.; Li, Y.; Wu, H.; Huo, Y.L.; Zhang, X.Z.; Zhou, Z.M. Nitrate decreases ruminal methane production with slight changes to ruminal methanogen composition of nitrate-adapted steers. *BMC Microbiol.* 2018, *18*, 21. [CrossRef]
- 24. Pen, B.; Takaura, K.; Yamaguchi, S.; Asa, R.; Takahashi, J. Effects of *Yucca schidigera* and *Quillaja saponaria* with or without β1–4 galacto-oligosaccharides on ruminal fermentation, methane production and nitrogen utilization in sheep. *Anim. Feed Sci. Tech.* **2007**, *138*, 75–88. [CrossRef]
- 25. Goiri, I.; Oregui, L.M.; Garcia-Rodriguez, A. Use of chitosans to modulate ruminal fermentation of a 50:50 forage-to-concentrate diet in sheep. *J. Anim. Sci.* **2010**, *88*, 749–755. [CrossRef] [PubMed]
- 26. Sharma, A.N.; Kumar, S.; Tyagi, A.K. Effects of mannan-oligosaccharides and *Lactobacillus acidophilus* supplementation on growth performance, nutrient utilization and faecal characteristics in Murrah buffalo calves. *J. Anim. Physiol. Anim. Nutr.* **2018**, *102*, 679–689. [CrossRef] [PubMed]
- 27. McDonald, P.; Edwards, R.A.; Greenhalgh, J.F.D.; Morgan, C.A. *Animal Nutrition*; Pearson Education Limited: Upper Saddle River, NJ, USA, 2002.



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