





## Draft Genome Sequence of *Providencia rettgeri* APW139 S1, an NDM-18-Producing Clinical Strain Originating from Hospital Effluent in South Africa

Noyise B. Ntshobeni,a DMushal Allam,b Arshad Ismail,b Daniel G. Amoako,c DSabiha Y. Essack,c Hafizah Y. Cheniaa

ABSTRACT Providencia rettgeri is an opportunistic pathogen implicated in various clinical infections. Here, we report the genome sequence of a Providencia rettgeri strain isolated from hospital effluent in South Africa, which harbors the New Delhi metallo- $\beta$ -lactamase (NDM) variant 18 gene ( $bla_{\rm NDM-18}$ ). The 4,835,047-bp genome encodes a resistome and virulome that are of cardinal importance to Providencia infections.

rovidencia rettgeri, an opportunistic pathogen belonging to the family Enterobacteriaceae, is implicated in various diseases (1). Notably, many recent unique isolates have been found to be carbapenemase producers carrying the New Delhi metallo- $\beta$ lactamase (NDM) gene (2). The NDM-18 variant is identical to NDM-1 except for a tandem repeat of 5 amino acids (QRFGD, amino acid positions 44 to 48 of NDM-1) (3, 4). Moreover, to date, there are only a few genome sequences of *P. rettgeri* available in public databases. Here, we report the first draft genome sequence of a P. rettgeri isolate encoding the *bla*<sub>NDM-18</sub> gene.

P. rettgeri APW139\_S1 was isolated in 2017 from hospital effluent originating from the pediatric ward of a rural hospital in the uMgungundlovu district of KwaZulu Natal Province, South Africa. The API 20E system (bioMérieux, France) was used to confirm culture identity of P. rettgeri APW139\_S1. The APW139\_S1 strain was grown on nutrient agar (Oxoid, England) and incubated overnight at 37°C prior to DNA extraction. Genomic DNA was extracted from 1 CFU of a visibly pure culture of the isolate using the GenElute bacterial genomic DNA kit (Sigma-Aldrich, St. Louis, MO, USA). A pairedend library (2 imes 300 bp) was prepared using a Nextera XT DNA sample preparation kit, and whole-genome sequencing (WGS) was carried out on a MiSeq machine (Illumina, USA). The sequenced reads (1,476,722 reads) were quality trimmed and de novo assembled into 68 contigs (99 $\times$  coverage and an  $N_{50}$  value of 215,431 bp) using Genomics Workbench version 10.1 (CLC bio, Qiagen, Aarhus, Denmark). Default parameters were used for all software unless otherwise specified. The NCBI Prokaryotic Genome Annotation Pipeline version 4.3 was used for annotation (5). The genome had a 4,835,047-bp genome size, a GC content of 40.50%, a total of 4,566 genes, a total of 4,487 coding sequences (CDS), 4,408 CDS coding genes, 79 RNA genes, 9 rRNAs, and 66

CRISPRFinder (6) predicted two confirmed clustered regularly interspaced short palindromic repeat 1 (CRISPR1) arrays, both located on contig 2 (GenBank accession no. SHDG01000002).

The pathogenic bacterial database VFanalyzer (7) allowed identification of the critical virulence factors for adherence (ecpA, ecpC, papD, and fimD), invasion (ibeC, cheB, and motA), regulation (phoQ), toxin (rtxB, rtxD, and rtxE), efflux (farB), and

Citation Ntshobeni NB, Allam M, Ismail A, Amoako DG, Essack SY, Chenia HY. 2019. Draft genome sequence of Providencia rettgeri APW139\_S1, an NDM-18-producing clinical strain originating from hospital effluent in South Africa. Microbiol Resour Announc 8:e00259-19. https://doi.org/10.1128/MRA

Editor Julie C. Dunning Hotopp, University of Maryland School of Medicine

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Address correspondence to Hafizah Y. Chenia, Cheniah@ukzn.ac.za.

Received 12 March 2019 Accepted 2 May 2019 Published 23 May 2019

<sup>&</sup>lt;sup>a</sup>Discipline of Microbiology, College of Agriculture, Engineering and Science, University of KwaZulu-Natal, Durban, South Africa

bSequencing Core Facility, National Institute for Communicable Diseases, National Health Laboratory Service, Johannesburg, South Africa

cantimicrobial Research Unit, College of Health Sciences, University of KwaZulu-Natal, Durban, South Africa



endotoxin (*htrB* and *ipxK*), which contribute to the bacterium's ability to adhere, invade the host, mediate adaptation to the host, secrete toxins, mediate resistance, and cause autoimmune-based host responses, respectively (8, 9).

The GoSeqlt tools Web platform (10) predicted an acquired antibiotic resistome, including genes coding for aminoglycoside [aadA1 and aac(6)-lb3], fluoroquinolone/quinolone [aac(6)-lb-cr], macrolide [mph(E) and msrE], rifampin (ARR-2), sulfonamide (sul1), tetracycline [tet(59)], trimethoprim (dfrA14), and  $\beta$ -lactam ( $bla_{OXA-2}$ ,  $bla_{OXA-10}$ ,  $bla_{NDM-18}$ , and  $bla_{PER-7}$ ) resistance. In silico WGS analysis indicated that the subclass B1 metallo- $\beta$ -lactamase NDM-18 was plasmid encoded and contained on contig\_68 (GenBank accession no. SHDG01000068). Comparative BLAST analysis of contig\_68 indicated its similarity (query length, 100%; identity,  $\geq$ 99.66%) to  $bla_{NDM-1}$ -harboring plasmids of other Enterobacteriaceae species, such as Klebsiella pneumoniae plasmids pA1705 (GenBank accession no. MH909349), p362713 (accession no. MH909347), and p309074 (accession no. MH909346), all from China. To the best of our knowledge, this is the first report of the NDM-18 variant in P. rettgeri. The valuable information offered by genomics allows a better understanding of the resistance and pathogenicity of multidrug-resistant (MDR) P. rettgeri strains.

This study was approved by the Biomedical Research Ethics Committee (BE521/16), College of Health Sciences, University of KwaZulu-Natal.

**Data availability.** This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession no. SHDG00000000. The version described in this paper is the first version, SHDG01000000. The raw reads have been submitted to the SRA under accession no. SRR8591527.

## **ACKNOWLEDGMENTS**

The research reported in this publication was funded by the South African Research Chairs Initiative of the Department of Science and Technology and National Research Foundation of South Africa (grant no. 80869 and 98342) by the South African Medical Research Council under a Self-Initiated Research Grant, and by the College of Health Sciences, University of KwaZulu-Natal, South Africa.

## **REFERENCES**

- Tshisevhe VS, Lekalakala MR, Tshuma N, Janse van Rensburg S, Mbelle N. 2016. Outbreak of carbapenem-resistant *Providencia rettgeri* in a tertiary hospital. S Afr Med J 107:31–33. https://doi.org/10.7196/SAMJ.2017 .v107i1.12002.
- Marquez-Ortiz RA, Haggerty L, Olarte N, Duarte C, Garza-Ramos U, Silva Sanchez J, Castro BE, Sim EM, Beltran M, Moncada MV, Valderrama A, Castellanos JE, Charles IG, Vanegas N, Escobar-Perez J, Petty NK. 2017. Genomic epidemiology of NDM-1-encoding plasmids in Latin American clinical isolates reveals insights into the evolution of multidrug resistance. Genome Biol Evol 9:1725–1741. https://doi.org/10.1093/gbe/evx115.
- 3. Somboro AM, Osei Sekyere J, Amoako DG, Essack SY, Bester LA. 2018. Diversity and proliferation of metallo-β-lactamases: a clarion call for clinically effective metallo-β-lactamase inhibitors. Appl Environ Microbiol 84:e00698-18. https://doi.org/10.1128/AEM.00698-18.
- Wu W, Feng Y, Tang G, Qiao F, McNally A, Zong Z. 2019. NDM metalloβ-lactamases and their bacterial producers in health care settings. Clin Microbiol Rev 32:e00115-18. https://doi.org/10.1128/CMR.00115-18.
- Tatusova T, Dicuccio M, Badretdin A, Chetvernin V, Nawrocki P, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic

- Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/qkw569.
- Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a Web tool to identify clustered regularly interspaced short palindromic repeats. Nucleic Acids Res 35:W52–W57. https://doi.org/10.1093/nar/gkm360.
- Liu B, Zheng D, Jin Q, Chen L, Yang J. 2019. VFDB 2019: a comparative pathogenomic platform with an interactive Web interface. Nucleic Acids Res 47:D687–D692. https://doi.org/10.1093/nar/gky1080.
- Ribet D, Cossart P. 2015. How bacterial pathogens colonize their hosts and invade deeper tissues. Microbes Infect 17:173–183. https://doi.org/ 10.1016/i.micinf.2015.01.004.
- Clegg S, Murphy CN. 2016. Epidemiology and virulence of Klebsiella pneumoniae. Microbiol Spectr 4. https://doi.org/10.1128/microbiolspec .UTI-0005-2012.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. J Antimicrob Chemother 67:2640–2644. https://doi.org/10.1093/jac/dks261.

Volume 8 Issue 21 e00259-19