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RESEARCH ARTICLE

Cerebrospinal fluid levels of neuron-specific enolase predict the severity of brain damage in newborns with neonatal hypoxic-ischemic encephalopathy treated with hypothermia

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Abstract

Objectives

To investigate whether cerebrospinal fluid levels of neuron-specific enolase (CSF-NSE) during the first 72 hours correlate with other tools used to assess ongoing brain damage, including clinical grading of hypoxic-ischemic encephalopathy (HIE), abnormal patterns in amplitude integrated electroencephalography (aEEG), and magnetic resonance imaging (MRI), as well as with the neurodevelopmental outcomes at two years of age.

Material and methods

Prospective observational study performed in two hospitals between 2009 and 2011. Forty-three infants diagnosed with HIE within 6 hours of life were included. HIE was severe in 20 infants, moderate in 12, and mild in 11. Infants with moderate-to-severe HIE received whole-body cooling. Both the HIE cohort and a control group of 59 infants with suspected infection underwent measurement of CSF-NSE concentrations at between 12 and 72 hours after birth. aEEG monitoring was started at admission and brain MRI was performed within the first 2 weeks. Neurodevelopment was assessed at 24 months.

Results

The HIE group showed higher levels of CSF-NSE than the control group: median 70 ng/ml (29; 205) vs 10.6 ng/ml (7.7; 12.9); p <0.001. Median levels of CSF-NSE in infants with severe, moderate, and mild HIE were 220.5 ng/ml (120.5; 368.8), 45.5 ng/ml (26, 75.3), and 26 ng/ml (18, 33), respectively. CSF-NSE levels correlated were significantly higher in infants with seizures, abnormal aEEG, or abnormal MRI, compared to those without abnormalities. Infants with an adverse outcome showed higher CSF-NSE levels than those with

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normal findings (p<0.001), and the most accurate CSF-NSE cutoff level for predicting adverse outcome in the whole cohort was 108 ng/ml and 50ng/ml in surviving infants.

Conclusions

In the era of hypothermia, CSF-NSE concentrations provides valuable information as a clinical surrogate of the severity of hypoxic-ischemic brain damage, and this information may be predictive of abnormal outcome at two years of age.

Introduction

Despite the advent of therapeutic hypothermia (TH) as the sole specific intervention shown to improve neurodevelopmental outcomes, hypoxic-ischemic encephalopathy (HIE) continues to contribute to major worldwide perinatal mortality and to long-term disability in full-term and near-term newborns [1, 2].

Early and accurate assessment of the severity of brain damage after a perinatal hypoxic—ischemic event remains one of the most difficult challenges in neonatal care. A variety of clinical, neuroimaging, and neurophysiological tools, and combinations of such, are used to predict long-term outcome [3–7]. However, current methods of assessing the risk of brain injury in the newborn have inherent limitations during the first hours of life, and uncertainty regarding the severity of ongoing brain damage and eventual neurological outcome persists during this early period [8, 9].

A biochemical index of brain injury would be highly desirable to increase the reliability of predictions of neurological sequelae after hypoxic-ischaemic brain injury. Several central nervous system-specific molecules have been investigated in blood serum and in cerebrospinal fluid (CSF) as possible quantitative indexes of perinatal brain injury [10–14].

Neuron specific enolase (NSE) is a well-known biomarker of neuronal injury, both in adults and in pediatric patients [14–17]. In newborns with HIE, the concentrations of NSE have been measured in blood serum and in CSF [18–29]. Serum NSE levels in the setting of HIE after asphyxia show vast variability, which entails significant limitations for its use as a viable biomarker [25]. This variability is probably related to the inconsistent and unpredictable bloodbrain-barrier permeability, the release of NSE from other extraneural sources in case of multiple organ injury, and confounding technical factors, such as the processing of the sample, the effect of hemolysis, the impact of the measuring method, and the of the reduction of concentrations as a function of storage time [30].

CSF levels of neuron-specific enolase (CSF-NSE) in infants with HIE have been shown to play a more consistent role as a surrogate marker of the extent of brain damage, and they provided good prediction of the outcome in infants with HIE in the pre-hypothermia era [20, 21, 26–28]. TH appears to lower CSF-NSE levels in infants with HIE; however, the predictive value of CSF-NSE for neurodevelopmental impairment at 12 months of age is not affected by cooling [29]. To our knowledge, no study has examined the correlation between CSF-NSE and other markers of ongoing brain damage during the acute phase of HIE, such us abnormal amplitude-integrated electroencephalogram (aEEG) and abnormal findings on cerebral MRI, nor its association with neurodevelopmental outcomes at 24 months in the hypothermia era.

The aim of this prospective cohort study of infants with HIE in the era of therapeutic hypothermia was to determine whether CSF-NSE correlates with the severity of HIE and other neonatal markers of ongoing brain damage, including aEEG and MRI, as well as with neurodevelopment at 2 years of age. This study shows that CSF-NSE within the first 72 hours may be a

useful biomarker for estimating ongoing brain damage and that it provides a surrogate outcome measure.

Material and methods

The study population included infants with HIE consecutively born at \geq 34 weeks gestational age with a birthweight \geq 1800 grams, admitted to two universitary third-level hospitals in Barcelona, Spain, between April 2009 and July 2011. Infants were considered to have HIE if they met the following three criteria: 1) at least one of the following clinical surrogates of hypoxic-ischemic insult: altered fetal heart rate pattern, sentinel event, or labor dystocia; 2) Apgar score \leq 5 at 5 and 10 minutes, need for resuscitation, including mask ventilation for more than 10 minutes after birth or endotracheal intubation, or acidosis (pH \leq 7.0 and/or base deficit \geq 16 mmol/L) in umbilical cord blood or within 60 minutes from birth; and 3) early neonatal encephalopathy defined as a syndrome of neurological dysfunction in the first 6 hours manifested by a subnormal level of consciousness with or without seizures (moderate or severe HIE) or palmary hyperexcitability, tremor, overactive myotatic reflexes, hypersensitivity to stimulation, or startle responses (mild HIE).

Newborns were excluded if a) they presented with congenital abnormalities, b) showed other identifiable etiologies of neurological dysfunction such as infection or genetic disease, or c) parental consent was not granted.

The severity of HIE was assessed immediately after admission, and always before starting TH, by three investigators (AGA, AA, GA); AGA was on call for the evaluation of infants of this project. Encephalopathy was classified as mild, moderate, or severe according to our semi-quantitative score which includes the aEEG findings [31, 32]. Newborns with moderate or severe HIE received whole-body cooling (Techotherm TSmed 200 N or Criticool, MTRE Ltd.) with rectal temperature maintained at 33–34 $^{\circ}$ C for 72 hours, following which they were slowly rewarmed (\leq 0.5 $^{\circ}$ C per hour). All cooled infants received sedation with fentanyl infusion throught the treatment.

Neuro-specific enolase in cerebrospinal fluid

A CSF sample was obtained by lumbar puncture at between 12 and 72 hours of life. The greater value for CSF-NSE was chosen if two lumbar punctures were performed. The CSF-NSE concentrations were measured blind to clinical data, including the grading of HIE as well as the aEEG traces and the neuroimaging findings.

CSF aliquots of 0.4 ml were distributed in plastic tubes, which were immediately frozen and stored at -80°C until analysed within 6 months after sampling. NSE concentrations in CSF were determined with an automated chemiluminescence immunoassay NSE Kritpor (Thermo Scientific® NSE Kryptor, BRAHMS GmbH., Germany). The coefficients of variation for interassay and intraassay variability were 2.8 to 6.2% and 2.2 to 9.7%, respectively. The sensitivity of this method was 1 ng/ml.

To compare CSF-NSE levels between patients with HIE and a control group, we included 59 patients without any abnormal neurological signs or confirmed central nervous system infections in whom lumbar puncture was performed as part of an early onset infection workup within 80 hours of life and for whom sufficient CSF was available for NSE analysis as part of a project on biomarkers in CSF.

Amplitude-integrated electroencephalography (aEEG)

aEEG recordings were immediately started at admission in the NICU and continued for at least 24 hours in infants with mild HIE and throughout TH and rewarming in infants with

moderate to severe HIE. The aEEG recordings were collected using CFM 6000 (Olympic Medical, Natus Inc, Seattle, WA, USA) or Nicolet One TM System (Viasys Healthcare, San Diego, California, USA) and digitally stored. The traces were blindly assessed by one researcher (GA) attending to the background pattern, cyclicity, and seizure activity. Classification of aEEG background patterns included the following categories: continuous (CNV), discontinuous (DCN), low voltage (LV), burst-supression (BS), and inactive or flat (FT) [33]. aEEG background trace was considered severely abnormal if BS, LV, or FT traces were observed. Electrical seizures were also noted.

Neuroimaging studies

All MRI studies were performed in one of the two participating centers (HSJD). Brain magnetic resonance imaging (MRI) was performed using a 1.5 Tesla unit (General Electric) with a specific neonatal head coil. The imaging protocol has been published elsewhere [34]. Minimally, axial and sagittal T1-weighted, axial and coronal T2-weighted, and axial diffusion-weighted images were available for all patients. Some patients underwent an "early" study performed around the fourth day of life, and a "late" study during the second week of life. For infants with only one study, this was considered, while for infants with both an early and a late study, only the score of the late study was considered in establishing the correlation with CSF-NSE values.

Two researchers (TA, AGA) masked to clinical data and CSF-NSE levels reviewed MRIs. Images were scored according to the scheme previously reported by Rutherford et al [35, 36]. This score grades the severity of damage in four brain regions: the posterior limb of internal capsule (PLIC), the basal ganglia and thalami, the white matter, and the cortex. According to the findings, each site except for the PLIC is graded as normal, or as mildly, moderately, or severely affected. The PLIC is graded as normal, equivocal, or absent. Discrepancies in the scoring of the images were discussed and resolved by consensus. Moderate-to-severe injury was defined as moderate-to-severe damage in any of the regions examined. Infants were grouped into 4 patterns of injury: normal, basal ganglia/thalami injury (including basal ganglia/thalami ± cortical injury without white matter injury), watershed pattern (which included moderate to severe white matter injury without any other damage), and global injury (including minimally basal ganglia/thalami and white matter injury).

Neurodevelopmental outcome

Neurodevelopmental assessments were made in all surviving infants at 24 months of age (median 24.3 months, IQR 23.3, 26.7 months, min-max 13–35 months) using the Bayley Scales of Infant and Toddler Development, Third Edition (BSID-III), which contains 3 individual developmental scores: a cognitive composite score, a language composite score (with receptive and expressive scores), and a motor composite score (with gross and fine motor sub-scores). These 3 scales have a mean score of 100 ± 15 . A developmental quotient score < 70 (>2 SD below the mean) indicates significant delay, and a score of < 85 (>1 SD below the mean) indicates at least mild-to-moderate delay [37].

Cerebral palsy was defined according to the Surveillance of CP in Europe [38]. Adverse outcome was defined as death, cerebral palsy, or Bayley-III cognitive, language, or motor scores <85.

Statistical analysis

Regarding perinatal, neurological, and outcome findings, qualitative variables were summarized with absolute and relative frequencies, and qualitative variables with median and

interquartile range (IQR) or mean (standard deviation). Continuous variables were compared using Mann-Whitney's U test or Kruskal-Wallis test, as appropriate. Categorical variables were compared using the Chi-squared test or Fisher's exact test.

CSF-NSE values were log-transformed to approximate a normal distribution and to achieve homogeneity of variance. Then differences between groups were tested by analysis of variance, with Tukey *post hoc* pairwise analysis. Spearman correlation coefficients (r_s) were used to assess the correlation between quantitative variables. Regression models were used to evaluate the relationship between biomarker levels and the presence and severity of HIE (controls, and mild, moderate, and severe HIE), including interaction for the age at which lumbar puncture was performed. In order to estimate the predictive capacity of the CSF-NSE regardless of the degree of HIE and the age at lumbar puncture, the CSF-NSE-outcome association was estimated by multiple regression, adjusting for these covariates. To determine the optimal cutoff value for CSF-NSE, defined as the highest value of sensitivity multiplied by specificity, receiver operating characteristic curve (ROC) analysis for CSF-NSE levels was made. The area under the curve, sensitivity (S), specificity (Sp), positive predictive value (PPV), and negative predictive value (NPV), including 95% confidence intervals (95% CIs) were expressed. All reported P values were 2-sided, and a P value of <0.05 was considered as indicating statistical significance. Statistical analyses were performed using SPSS verson 20 (IBM, Armonk, NY, USA).

We planned to recruit 50 subjects over a 2-year period. Accounting for 10–15% patient loss for analysis due to clinical instability of the patient, haemorrhage, or insufficient CSF, *a priori* power calculations were based on analysis of data from 40 subjects. This sample size provided at least 90% statistical power (two-sided alpha risk 5%) to detect a minimum expected effect size of 50 points (SD 48) of difference in biomarker levels between patients with adverse neurological outcome and those with non-adverse neurological outcome.

Ethical considerations

Written information was given to the parents and written consent was obtained at the bedside on admission of each infant after explanation of the study and before its onset. The research was conducted according to the Declaration of Helsinki principles and those of the human studies committee that approved the study (Clinic Hospital Protocol Number: HBC/2010/6049).

Results

Fifty-nine consecutive patients with HIE were recruited during the study period but 6 were excluded due to perinatal infection with *Listeria monocytogenes* (2 infants), multiple hemorrhagic strokes (1 infant), spinal cord injury (1 infant), late admission at 5 days of age (1 infant), or lack of parental informed consent (1 infant). Of the 53 candidate patients, CSF-NSE could not be performed in 10 infants due to clinical instability, haemorrhage, or insufficient CSF.

CSF-NSE was performed at a median age of 48 hours (15, 73) in the group of infants with HIE and 32 hours (24, 58) in the control group; p = 0.344. Compared to controls, patients with HIE had fewer spontaneous deliveries (7% vs 61%), lower cord pH values (6.94 \pm 0.16 vs 7.22 \pm 0.11), and lower Apgar scores at 5 minutes (5 (3, 6) vs 10 (8, 10)). Birthweight was lower in the HIE group compared to controls: 3004 ± 587 grams vs 3266 ± 489 grams (p = 0.034). No differences were found between the HIE and control groups regarding gestational age or gender.

The main perinatal characteristics of the 43 recruited infants with HIE, according to the severity of encephalopathy, are shown in <u>Table 1</u>. All infants with moderate or severe HIE were cooled, except for 3 infants with severe HIE, due to late transfer beyond 12 hours of life

Table 1. Perinatal, neurological and outcome findings of the 43 HIE infants enrolled in the study.

	Total HIE N = 43	Mild HIE N = 11	Moderate HIE N = 12	Severe HIE N = 20	P value
Gestational age, weeks	38 (37, 40)	38 (37, 40)	39 (37, 41)	38 (36, 40)	0.685
Weight, grams	3000 (2530, 3500)	3160 (2600, 3700)	3115 (2423, 3498)	2835 (2500, 3355)	0.346
Sex (female)	18/43 (42)	4/11 (36)	5/12 (42)	9/20 (45)	0.897
Intrauterine growth retardation	4/43 (9)	0/11	2/12 (17)	2/20 (10)	-
Sentinel event	16/43 (37)	4/11 (36)	4/12 (33)	8/20 (40)	0.929
Spontaneous delivery	3/43 (7)	1/11 (9)	1/12 (8)	1/20 (5)	-
Umbilical cord pH ^a	6.97 (6.81, 7.03)	7.00 (6.99, 7.05)	6.90 (6.79, 7.00)	6.94 (6.81, 7.06)	0.190
Apgar score at 5 minutes	5 (3, 6)	6 (4, 7)	5.5 (4, 7)	4 (2, 5)	0.051
Advanced resuscitation ^b	31/43 (72)	6/11 (55)	8/12 (67)	17/20 (85)	-
Electrical seizures during TH	24/43 (56)	0/11 (0)	6/12 (50)	18/20 (90)	< 0.001
Abnormal aEEG BGP in first 6 hours of life	20/39 (51)	0/8 (0)	6/12 (50)	14/19 (74)	< 0.001
Abnormal aEEG BGP in first 80 hours of life ^c	24/43 (56)	0/11 (0)	6/12 (50)	18/20 (90)	< 0.001
MRI Score	2.5 (0, 9.3)	0 (0, 1)	0 (0, 4)	9.5 (6.5, 11)	< 0.001
Moderate-severe injury (MRI)	20/38 (51)	1/11 (9)	3/11 (27)	16/16 (100)	< 0.001
Global injury pattern (MRI)	17/38 (45)	0/11 (0)	2/11 (18)	15/16 (94)	< 0.001
Death	14/43 (33)	0/11	1/12 (8)	13/20 (65)	< 0.001
Adverse outcome ^c	22/42 (52)	0/11	3/11 (27)	19/20 (95)	< 0.001
Cerebral palsy	5/28 (18)	0/11	1/10 (10)	4/7 (57)	0.006
CP and/or any abnormal BSID-III score < 85 ^d	8/28 (29)	0/11	2/10 (20)	6/7 (86)	< 0.001
CP and/or any abnormal BSID-III score < 70 ^d	5/28 (18)	0/11	0/10	3/7 (43)	0.006
BSID-III Score (Motor) ^e	94 (83, 99)	99 (94, 103)	96 (90, 98)	79 (45, 88)	0.009
BSID-III Score (Cognitive) ^e	98 (86, 105)	105 (95, 105)	105 (90, 110)	80 (45, 85)	0.006
BSID-III Score (Language) ^e	91 (83, 100)	97 (91, 100)	91 (88, 103)	77 (45, 83)	0.019

Quantitative variables are expressed as median (interquartile range), and categorical values are expressed as n/N (%). Continuous variables were compared using Kruskal-Wallis test. Categorical variables were compared using the Chi-squared test. P value of <0.05 was considered as indicating statistical significance. aEEG: amplitude integrated electroencephalogram; BGP: background pattern; BSID-III: Bayley Scales of Infant and Toddler Development Third Edition; CP: cerebral palsy; MRI: magnetic resonance imaging; TH: therapeutic hypothermia.

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in 2 infants and to critically ill status in one case. Infants with mild HIE were not cooled. MRI was performed in 37 patients, as five patients died before an MR scan could be performed (4 severe and one moderate HIE). Both aEEG background pattern and MRI findings were associated with severity of HIE (Table 1). Brain MRI was performed at a median age of 10 days; IQR 4.4, 13.9.

Fourteen infants (33%) did not survive the neonatal period, one with moderate HIE and 13 with severe HIE. In the only patient with moderate HIE who died, death was due to severe coagulopathy. In the 13 infants with severe HIE, death was preceded by an end-of-life decision (EoL). The EoL decision was considered for those patients with persistent severe encephalopathy (coma) in combination with persistent severe altered aEEG and severe neuroimaging findings (brain ultrasound scans and/or MRI). CSF-NSE levels were not considered in making an end-of-life decision.

Of the 29 neonates who survived, 24 were evaluated with BSID-III; additionally, 4 were contacted by phone, with parents reporting that they had a normal outcome. One infant were lost

^aData were not available in 1, 2, and 3 infants with mild, moderate, and severe HIE, respectively.

^bAdvanced resuscitation: tracheal intubation, chest compressions and/or adrenaline.

^cAdverse outcome: death or cerebral palsy and/or BSID-III score <85.

^dOutcome in 28/29 surviving infants.

^eBSID-III test was performed in 24 out of 29 surviving infants

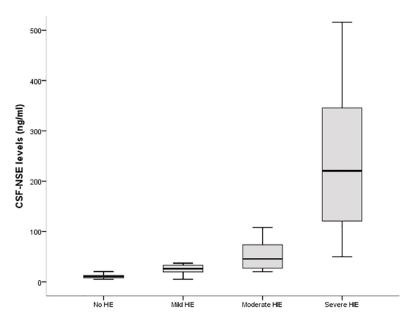


Fig 1. Cerebrospinal fluid levels of neuron-specific enolase in 59 controls and 43 infants with HIE, according to the severity of the encephalopathy. CSF-NSE levels in the control group were 10.6~(7.7; 12.9). CSF-NSE levels in 20, 12, and 11 infants with severe, moderate, and mild HIE, respectively, were 220.5~ng/ml~(120.5; 368.8), 45.5~ng/ml~(26, 75.3), and 26~ng/ml~(18, 33). No statistically significant differences were found between any of the grades of HIE compared to controls or between any of the grades of HIE (analysis of variance after log-transformation of CSF-NSE values, with Tukey *post hoc* pairwise analysis); p<0.05. HIE: hypoxic-ischaemic encephalopathy; CSF-NSE: cerebrospinal fluid-neuron specific enolase.

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to follow-up. Of the overall cohort, 22 infants (54%) had adverse outcomes, and 8 infants out of the 29 neonates who survived had CP and/or BSID-III Score < 85. Table 1 shows the outcome results according to the severity of HIE.

CSF-NSE and neurological variables

The HIE group showed significantly higher levels of CSF-NSE than the control group: median 70 ng/ml (29; 205) vs 10.6 (7.7; 12.9); p <0.001. According to the severity of HIE, all subgroups presented higher CSF-NSE levels than the control population (p<0.001). Infants with severe HIE had higher levels of CSF-NSE than those with moderate HIE (p<0.001), and infants with moderate HIE had higher levels than those with mild HIE (p = 0.024). CSF-NSE levels in infants with severe, moderate, and mild HIE were: 220.5 ng/ml (120.5; 368.8), 45.5 ng/ml (26, 75.3) and 26 ng/ml (18, 33), respectively (Fig 1). All severe cases had CSF-NSE levels above 109 ng/ml except for two infants with levels of 50 ng/ml and 70 ng/ml, respectively. All mild cases had a level below 38 ng/ml.

Regression analysis showed that age at lumbar puncture did not influence the association between CSF-NSE levels and the presence or severity of HIE.

CSF-NSE concentrations were higher in infants with abnormal neurological findings including seizures, aEEG background activity, and MRI, compared to those without abnormalities (p<0.001) (Table 2).

Regarding MRI injury, the 20/38 infants that had moderate-to-severe injury had higher CSF-NSE levels compared to the infants who did not have injury (15/38) or those with mild injury (3/38): 133 (73; 268) vs 28.5 (20.8; 36.3) (p<0.001). The pattern of damage on MRI was predominantly global (17/38), and only 2 and 3 infants out of the 38 infants with MRI had solely basal ganglia/thalamus and watershed injury, respectively. Infants with global injury had

Variables		Yes		P value	
	n	CSF-NSE (ng/ml)	n	CSF-NSE (ng/ml)	
Moderate or severe encephalopathy	32	116.5 (56, 267.8)	11	26 (18, 33)	< 0.001
Electrical seizures during TH	24	133 (71.8, 267.8)	19	28 (21, 37)	< 0.001
Abnormal aEEG BGP in first 6 hours of life	20	133 (77.8, 368.8)	19	33 (25, 59)	< 0.001
Abnormal aEEG BGP in first 80 hours of life	24	183.5 (87,5; 294.5)	19	32 (23; 50)	< 0.001
Moderate-severe injury (MRI)	20	133 (72.5, 267.8)	18	28.5 (20.8, 36.3)	< 0.001
Global injury pattern (MRI)	17	166 (96.5, 274.5)	21	29 (22, 46)	< 0.001
Cerebral palsy	5	268 (130; 290)	23	33 (23; 59)	0.002
CP and/or BSID-III Score < 85 ^a	8	157.5 (56, 277.8)	20	28.5 (21.5, 37)	0.001
CP and/or BSID-III Score < 70 ^a	5	268 (130, 290)	23	33 (23, 59)	0.002
Adverse outcome ^b	22	203 (112.3; 322.3)	20	29 (21.5; 36.8)	< 0.001
Death	14	218.5 (121.5, 483)	29	34 (25, 73.5)	< 0.001

Table 2. Cerebrospinal fluid levels of neuron-specific enolase of 43 infants with HIE according to their neurological findings and outcomes.

CSF-NSE values are expressed as median (interquartile range). Mann-Whitney U test was used to analyze the differences in CSF-NSE levels between groups. P value of <0.05 was considered as indicating statistical significance. aEEG: amplitude integrated electroencephalogram; BGP: background pattern; BSID-III: Bayley Scales of Infant and Toddler Development Third Edition; CP: cerebral palsy; MRI: magnetic resonance imaging; TH: therapeutic hypothermia.

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higher CSF-NSE levels: 166 (96.5, 275) vs 29 (22, 46) (p<0.001). CSF-NSE levels showed a positive correlation with the MRI score: r_s 0.824, p<0.001.

CSF-NSE and outcome

Infants with an adverse outcome showed higher CSF-NSE levels than those with normal findings (p< 0.001) (Fig 2 and Table 2). ROC analysis revealed a CSF-NSE cutoff value of maximum predictive accuracy of 108 mg/dL (AUC 0.97, S 0.86, Sp 1.0, PPV 1.0, NPV 0.87) for prediction of adverse outcome. Among surviving infants, CSF-NSE optimal cutoff-point to predict CP and/or BSID-III score <85 was 50 mg/dL (AUC 0.93, S 1.0, Sp 0.8, PPV 0.67, NPV 1.0) (Table 3).

CSF-NSE levels showed a moderate negative correlation with the three BSDI-III domains (motor, cognitive, and language composite scores): r_s -0.650, p = 0.001; r_s -0.641, p = 0.001; and r_s -0.604, p = 0.002 respectively.

Regression analysis showed that age at lumbar puncture nor the severity of HIE did not interact with the association between CSF-NSE level and outcome variables.

The predictive values for the neurological variables included in the study (severity of HIE, aEEG background pattern, MRI findings, and the CSF-NSE cutoff-point of 108 ng/ml) are shown in Table 4.

Discussion

The study shows that CSF-NSE within the first 72 hours may be a useful biomarker for estimating ongoing brain damage, based on its correlation with the clinical grading of encephalopathy, aEEG tracings, and MRI findings. More importantly, NSE concentrations in CSF can provide a surrogate outcome measure.

Several methods have been used for early estimation of the cerebral damage after hypoxicischemic injury and prediction of outcome. The most readily available tools include clinical grading of the encephalopathy, electrophysiological information such us assessment of

^aOutcome in 28/29 surviving infants

^bAdverse outcome: death or cerebral palsy and/or BSID-III score <85.

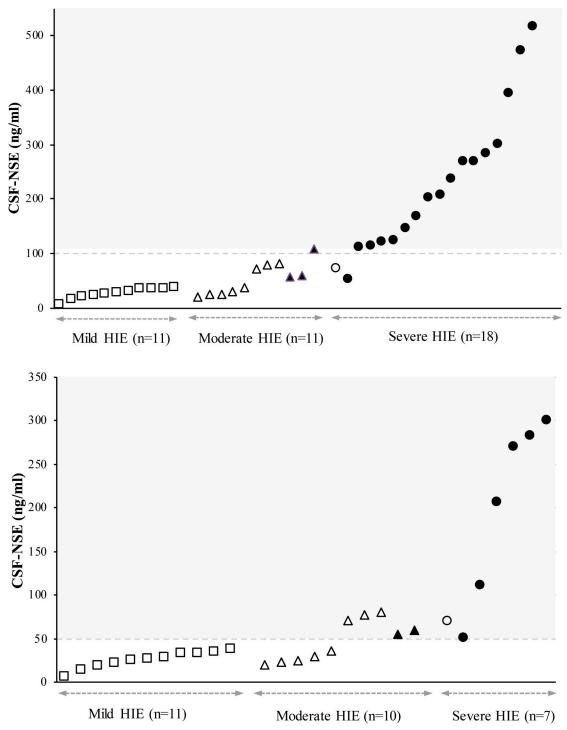


Fig 2. Cerebrospinal fluid levels of neuron-specific enolase of 43 patients according to their neurological findings and outcomes. Each CSF-NSE value represents a square-shaped infant, trigulum, or circle according to the severity of HIE (mild, moderate, or severe). Filled shapes represent infants with adverse outcomes (death or cerebral palsy and/or BSID-III score < 85). Empty dots represent those infants without adverse outcome. The figure above shows the entire cohort of 43 children, and the one below shows the 29 surviving infants. Two infants with very severe HIE with extreme CSF-NSE values (2855 ng/ml and 1490 ng/ml) and one infant lost in the follow-up are not represented in the figures. The dashed line represents the cutoff value of 108 mg/dL and 50 mg/dl for adverse outcome, according to the maximum predictive accuracy in the receiver operating curve analysis, for the total cohort (figure above) and only the surviving infants (figure below), respectively. HIE: hypoxic-ischaemic encephalopathy; CSF-NSE: cerebrospinal fluid-neuron specific enolase; aEEG: amplitude integrated electroencephalogram; BGP: background

pattern; BSID-III: Bayley Scales of Infant and Toddler Development Third Edition; CP: cerebral palsy; MRI: magnetic resonance imaging.

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electrical activity by means of an amplitude-integrated EEG and evoked responses, neuroimaging, and cerebral blood flow studies [7, 9, 39–41].

However, all these evaluations have some limitations during the first 72 hours. During this time frame, the level of alertness is often confounded by the use of sedatives during cooling and/or antiepileptic drugs if clinical or electrical seizures occur, rendering clinical evaluation less reliable [42, 43]. In addition, qualitative evaluation of significant brain injury on both diffusion-weighted and conventional MRI during the first 48h may be underestimated [2, 43, 44]. Other quantitative techniques such as quantitative diffusion parameters and proton magnetic resonance spectroscopy have been used to overcome these limitations [45]. However, in clinical practice their use in infants with perinatal HIE during this narrow short time frame is limited. Sick infants may not tolerate transfer to radiology rooms, and early MRI is challenging as it requires specialized interpretive expertise.

Amplitude-integrated EEG background patterns in neonatal encephalopathy correlate closely with neurological outcome [46]. However, their predictive value depends on how long the background patterns are altered in relation to the hypoxic-ischemic insult, and the greater predictive value for adverse outcome peaks at 48–72 hours of life in cooled infants [41].

Table 3. Receiver operating characteristic curve analysis of cerebrospinal fluid levels of neuron-specific enolase and neurological findings.

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Variable	N	Cutoff-point	Area under the ROC (95% CI)	Sensitivity (95% CI)	Specificity (95% CI)	PPV (95%CI)	NPV (95%CI)
Moderate-to-severe HIE	32	50	0.91 (0.83,0.99)	0.81 (0.64,0.93)	1.00 (0.72,-)	1.00 (0.86,1.00)	0.65 (0.42,-)
Severe HIE	20	110	0.98 (0.95,1.00)	0.90 (0.68,0.98)	1.00 (0.85,-)	1.0 (0.82,1.00)	0.92 (0.73,-)
Electrical seizures during TH	24	50	0.85 (0.71,0.99)	0.92 (0.73,0.99)	0.79 (0.54,0.94)	0.85 (0.64,0.98)	0.88 (0.65,0.97)
Abnormal aEEG BGP in first 6 hours of life	20	77	0.87 (0.75,0.99)	0.80 (0.56,0.94)	0.89 (0.67,0.99)	0.89 (0.65,0.97)	0.80 (0.58,0.97)
Abnormal aEEG BGP in first 80 hours of life	24	77	0.92 (0.83,1)	0.83 (0.63,0.95)	0.95 (0.74,0.99)	0.95 (0.76,0.99)	0.82 (0.60,0.99)
Moderate-severe injury (MRI)	20	80	0.93 (0.85,1.00)	0.75 (0.51,0.91)	1.00 (0.81,-)	1.00 (0.79,1.00)	0.78 (0.55,-)
Global injury pattern (MRI)	17	80	0.95 (0.88,1.00)	0.82 (0.57,0.96)	0.95 (0.76,0.99)	0.93 (0.69,0.99)	0.87 (0.65,0.99)
Cerebral palsy	5	205	0.95 (0.84,1.00)	0.80 (0.28,0.99)	1.00 (0.85;-)	1.00 (0.50,1.00)	0.96 (0.69,-)
CP and/or BSID-III Score < 85 ^a	8	50	0.93 (0.83,1.00)	1.00 (0.63,-)	0.80 (0.56,0.94)	0.67 (0.39,-)	1.00 (0.77,1.00)
CP and/or BSID-III Score < 70 ^a	5	205	0.95 (0.84.1.00)	0.80 (0.28,0.99)	1.00 (0.85,-)	1.00 (0.49,1.00)	0.96 (0.69,-)
Adverse outcome ^b	22	108	0.97 (0.93,1.00)	0.86 (0.65,0.97)	1.0 (0.83,-)	1.0 (0.82,1.0)	0.86 (0.66,-)
Death	14	108	0.91 (0.83,0.99)	1.00 (0.77,-)	0.83 (0.64,0.94)	0.74 (0.51,-)	1.00 (0.85,1.00)

aEEG: amplitude integrated electroencephalogram; BGP: background pattern; BSID-III: Bayley Scales of Infant and Toddler Development Third Edition; CI: confidence interval; CP: cerebral palsy; MRI: magnetic resonance imaging; NPV: negative predictive value; PPV: positive predictive value; ROC: Receiver operating curve; TH: therapeutic hypothermia.

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^aOutcome in 28/29 surviving infants.

^bAdverse outcome: death or cerebral palsy and/or BSID-III score <85. CSF-NSE values are expressed in ng/ml

Table 4.	Predictive	values of the	e different	neurological	tools related	to the outcome.

Adverse outcome	Sensitivity	Specificity	PPV	NPV	Odds Ratio (95%CI)
Severe HIE	0.86	0.95	0.95	0.86	120.3 (11.5;1262.8)
Moderate-to-severe	1.00	0.55	0.71	1.00	54.5 (2.9;1021.5)
Abnormal aEEG BGP in first 80 hours of life	0.86	0.75	0.79	0.83	19 (3.9;92.6)
Moderate-severe injury (MRI)	0.94	0.85	0.84	0.94	90.7 (8.5;964.0)
Global injury pattern (MRI)	0.82	0.9	0.88	0.86	42 (6.2,286.7)
CSF-NSE > 108 ng/ml	0.82	1.00	1.00	0.83	168.6 (8.5,3347.3)

aEEG: amplitude integrated electroencephalogram; BGP: background pattern; CI: confidence interval; MRI: magnetic resonance imaging; NPV: negative predictive value; PPV: positive predictive value; ROC: Receiver operating curve; TH: therapeutic hypothermia.

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Despite the limitations in accurate assessment of brain damage during the first 72 hours after the hypoxic-ischemic insult, this challenge has to be addressed in order to provide early neuroevolutive prognostic information to parents [9].

The concept of "window of opportunity" is used in severe brain injury to indicate as the period of time in which a critically ill neonate might die after removal from the ventilator. After this window, the infant would no longer be dependent on such treatment and would probably survive with subjectively intolerable disability. This window of opportunity is a determining factor in the urgency of having neurological outcome prognostic indicators during the first 72 hours [47].

To improve early identification of infants at risk of neuroevolutive sequelae, reliable early biomarkers of brain damage are a valuable clinical tool to improve accurate identification of infants at risk after a hypoxic-ischemic insult.

Neuron specific enolase (NSE) is a well-known biochemical marker of neuronal injury in adults [16, 17], and pediatric patients [14, 15]. NSE is stable in biological fluids, it originates predominantly in the cytoplasm of neuroendocrine cells and neurons, and its measurement does not elicit immunologic cross-reactivity with non-neuronal enolase.

For clinical purposes, measurement of this potentially sensitive biomarker for brain damage in peripheral blood may have advantages compared to CSF. Blood sampling is less invasive, and can be performed more often, even when the patient is unstable. However, serum concentration both in the pre-hypothermia era and in cooled infants has shown inconsistent results in relation to the severity of brain injury assessed by clinical grading of HIE, neuroimaging, and outcome beyond 12 months of age [18-25]. There are several possible explanations for these controversial results. First, brain injury does not necessarily result in consistent, predictable blood-brain-barrier disruption. Second, increased serum NSE is not necessarily of CNS origin. Several tissues, including muscle, kidney, heart, adipose tissue, adrenal glands, lymphocytes, red blood cells, and platelets contain significant amounts of NSE [48–52]. A hypoxicischemic event can result in multiple organ injury, including all those tissues and disseminated intravascular coagulation. Finally, technical confounders must be taken into account. These include the processing of the sample, the effects of the time on the sampling stored since after 6 months at -80°C there might be a significant decrease in concentrations in CSF, and, particularly, the effect of hemolysis [53]. Moreover, the choice of measuring methods may have influence on the levels of NSE [54], so care should be taken in using our specified cut-off level of NSE for determination of prognosis if another NSE determination method was used.

The main advantages of CSF sampling is the fact that the brain is in direct contact with the CSF and the damaged brain tissue releases high quantities of proteins into this fluid. In

^aAdverse outcome: death or cerebral palsy and/or BSID-III score <85.

addition, there are no sources of NSE other than neurons [30]. Hence, the consistent value of CSF-NSE levels in the first 96 hours of life seen in the prehypothemia era are not surprising, and this biomarker reflects brain damage in accordance with the clinical severity of HIE, as do MRI and aEEG findings [14, 20, 26, 28].

To our knowledge only one study has been published on CSF-NSE that included cooled infants with HIE, but this study did not examine the relation between CSF-NSE concentrations and aEEG and MRI findings as surrogates for the severity of hypoxic-ischemic event [29]. We examined the relationship between CSF-NSE and relevant clinical variables (seizures), in addition to aEEG, MRI, and neurodevelopmental outcomes. Our results are consistent with the previously reported value of the CSF-NSE to identify severe brain damage after sudden and unexpected postnatal collapse [55], and with the relationship between CSF-NSE levels and infarction volume and topology, as well as neurodevelopment at 2 years of age in infants with neonatal arterial ischemic stroke [56].

One strength of our study is that it offers cutoff points that are relevant in clinical practice. CSF-NSE levels ≥ 108 ng/ml showed the greatest specificity and positive predictive value for identifying infants at increased risk of adverse outcome. Considering only those who survived, optimal cutoff point to predict abnormal outcome was 50 ng/ml. CSF-NSE levels may be particularly useful when additional information is needed about brain injury and prognosis, especially if there are inconsistencies between prognostic tools, such as neurological examination and neurophysiological studies, and also if an end-of-life decision is considered within the first 72 hours of life [8].

Biochemical markers of brain injury, such as NSE, are released in a time sequence related to brain damage. In animal models of focal and global ischemia, an early peak of serum NSE levels reflects the ischemia-induced cytoplasmic loss of NSE in neurons that is detectable before irreversible neuronal damage occurs [57, 58]. This has also been corroborated in human adults with stroke, where, after an initial rise within 3 hours, NSE concentrations in blood serum decrease followed by a secondary increase until day 5, with concentrations after 24 hours reflecting the volume of infarcted brain areas [59, 60].

Several studies have determined serum NSE at different times in the first days of life. These studies have shown that serum NSE concentrations are relatively stable and higher during the first 24 hours after birth, decreasing thereafter [24, 61]. It is not known how soon NSE can be detected in the CSF after brain injury, nor are the kinetics of its release and clearance in this biological fluid well understood. Therefore, the time point of maximum diagnostic efficacy is unknown [49]. The scheduling of CSF sampling is generally decided arbitrarily, but in most studies lumbar puncture has been performed within the first 72 hours of life [21, 26, 28, 62]. Despite a range of 12–80 hours in our study, we did not find that the age at lumbar puncture was an interaction factor between the CSF-NSE levels and the variables analyzed. Whether serial measurements might provide indirect information regarding ongoing neuronal injury and might therefore help increase accuracy in predicting patient outcome is unknown. However, lumbar puncture is an interventional procedure, and it is not free from risks. It should not be performed when the infant is unstable or suffers from significant coagulopathy, which makes it difficult to obtain serial samples.

Our determination of CSF-NSE concentrations at a median age of 48 hours after birth may mirror irreversible neuronal damage as it is correlated with surrogate biomarkers of brain damage and the outcomes in this study. The reliability of CSF-NSE to predict subsequent adverse neurological outcomes was similar to the reliability of aEEG and neuroimaging. Therefore, CSF-NSE testing is suggested as both a valuable diagnostic tool in clinical management of the infant with HIE and a prognostic parameter during the course of the encephalopathy. In addition to helping to predict neurodevelopmental outcomes, biochemical brain

damage biomarkers such as CSF-NSE should gain momentum in clinical research on HIE. We feel that they can be particularly useful to identify newborns at 48 hours who are at risk of adverse prognosis and potential candidates to receive adjuvant neuroprotective therapies to promote neuronal regeneration and structural recovery from injury during the tertiary phase of HIE (as is the case with stem cell transplantation and anti-inflammatory agents and gangliosides). Furthermore, this biochemical marker could be useful for rigorous monitoring of therapeutic interventions aimed at ameliorating brain damage.

Our study has some limitations. Like many longitudinal studies on neurobiological markers that may reflect the severity and progression of brain injury, the inability to obtain or analyze some CSF samples was a concern. Given the low incidence and prevalence of HIE, one for every 1000 births in our setting [63], the number of recruited infants was not huge. Larger validation studies would be advisable. Our study was not powered to evaluate death or disability separately, and since most infants with severe damage die, larger studies are needed to assess the relationship between biomarkers and outcome, independent of mortality. This is especially important for the group of infants with moderate HIE—the group with the greatest prognostic uncertainty. In our series 3 of the 32 infants with moderate-to-severe HIE were not cooled; all three had severe HIE. When analyses were performed eliminating these three patients, no difference in results emerged, even when including optimal cut-off values for CSF-NSE (ROC analysis) (\$1 and \$2 Tables). Finally, Since it is not ethical to perform lumbar puncture in healthy babies, we decided to use babies with suspected infection as control subjects. This approach has already been used in previous studies with the same purpose as ours, and we believe that CSF-NSE values are highly unlikely to be influenced by the consideration of infectious risk since their neurological status and CSF findings were normal [21, 26, 27].

Conclusion

HIE is a rare but potentially devastating disease associated with high morbidity and mortality. Our study suggests that NSE in CSF measured in infants with HIE at a median age of 48 hours of life might be a useful tool to identify those newborns with suspicion of severe brain injury based on clinical assessment, aEEG, and MRI. Furthermore, in the era of hypothermia, this biomarker appears to be able to distinguish infants will fully recover from those who are likely to present an adverse outcome.

Supporting information

S1 Table. Cerebrospinal fluid levels of neuron-specific enolase of the 40 cooled infants with HIE according to their neurological findings and outcomes. The three infants that did not undergo hypothermia treatment had severe HIE and were not included in this analysis (NSE values were 166, 281, and 2855 ng/ml). CSF-NSE values are expressed as median (interquartile range). Mann-Whitney U test was used to analyze the differences in CSF-NSE levels between groups. P value of <0.05 was considered as indicating statistical significance. aEEG: amplitude integrated electroencephalogram; BGP: background pattern; BSID-III: Bayley Scales of Infant and Toddler Development Third Edition; CP: cerebral palsy; MRI: magnetic resonance imaging; TH: therapeutic hypothermia. ^aOutcome in 28/29 surviving infants. ^bAdverse outcome: death or cerebral palsy and/or BSID-III score <85. (DOCX)

S2 Table. Receiver operating characteristic curve analysis of cerebrospinal fluid levels of neuron-specific enolase and neurological findings of the 40 cooled infants. The three infants that did not undergo hypothermia treatment had severe HIE and were not included in

this analysis (NSE values were 166, 281 and 2855 ng/ml). aEEG: amplitude integrated electro-encephalogram; BGP: background pattern; BSID-III: Bayley Scales of Infant and Toddler Development Third Edition; CP: cerebral palsy; MRI: magnetic resonance imaging; PPV: positive predictive value; NPV: negative predictive value; ROC: Receiver operating curve; TH: therapeutic hypothermia. ^aOutcome in 28/29 surviving infants. ^bAdverse outcome: death or cerebral palsy and/or BSID-III score <85. CSF-NSE values are expressed in ng/ml. (DOCX)

S1 Dataset.

(XLSX)

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References

- Tagin MA, Woolcott CG, Vincer MJ, Whyte RK, Stinson DA. Hypothermia for neonatal hypoxic ischemic encephalopathy: an updated systematic review and meta-analysis. Archives of pediatrics & adolescent medicine. 2012; 166(6):558–66. https://doi.org/10.1001/archpediatrics.2011.1772 PMID: 22312166.
- Bonifacio SL, deVries LS, Groenendaal F. Impact of hypothermia on predictors of poor outcome: how
 do we decide to redirect care? Seminars in fetal & neonatal medicine. 2015; 20(2):122–7. Epub 2015/
 01/13. https://doi.org/10.1016/j.siny.2014.12.011 PMID: 25577654; PubMed Central PMCID:
 PMC4375009.
- Nanavati T, Seemaladinne N, Regier M, Yossuck P, Pergami P. Can We Predict Functional Outcome in Neonates with Hypoxic Ischemic Encephalopathy by the Combination of Neuroimaging and Electroencephalography? Pediatr Neonatol. 2015; 56(5):307–16. Epub 2015/04/12. https://doi.org/10.1016/j. pedneo.2014.12.005 PMID: 25862075; PubMed Central PMCID: PMC4529385.
- 4. Polat M, Simsek A, Tansug N, Sezer RG, Ozkol M, Baspinar P, et al. Prediction of neurodevelopmental outcome in term neonates with hypoxic-ischemic encephalopathy. European journal of paediatric neurology: EJPN: official journal of the European Paediatric Neurology Society. 2013; 17(3):288–93. Epub 2012/12/13. https://doi.org/10.1016/j.ejpn.2012.11.004 PMID: 23231917.

- Shellhaas RA, Kushwaha JS, Plegue MA, Selewski DT, Barks JD. An Evaluation of Cerebral and Systemic Predictors of 18-Month Outcomes for Neonates With Hypoxic Ischemic Encephalopathy. Journal of child neurology. 2015; 30(11):1526–31. Epub 2015/03/01. https://doi.org/10.1177/0883073815573319 PMID: 25724376; PubMed Central PMCID: PMC4551612.
- Chang T, du Plessis A. Neurodiagnostic techniques in neonatal critical care. Curr Neurol Neurosci Rep. 2012; 12(2):145–52. Epub 2012/02/10. https://doi.org/10.1007/s11910-012-0254-y PMID: 22318538.
- van Laerhoven H, de Haan TR, Offringa M, Post B, van der Lee JH. Prognostic tests in term neonates with hypoxic-ischemic encephalopathy: a systematic review. Pediatrics. 2013; 131(1):88–98. Epub 2012/12/19. https://doi.org/10.1542/peds.2012-1297 PMID: 23248219.
- Rasmussen LA, Cascio MA, Ferrand A, Shevell M, Racine E. The complexity of physicians' understanding and management of prognostic uncertainty in neonatal hypoxic-ischemic encephalopathy. J Perinatol. 2018. Epub 2018/12/21. https://doi.org/10.1038/s41372-018-0296-3 PMID: 30568164.
- Natarajan N, Pardo AC. Challenges in neurologic prognostication after neonatal brain injury. Semin Perinatol. 2017; 41(2):117–23. Epub 2017/02/01. https://doi.org/10.1053/j.semperi.2016.11.008 PMID: 28139254.
- Bennet L, Booth L, Gunn AJ. Potential biomarkers for hypoxic-ischemic encephalopathy. Seminars in fetal & neonatal medicine. 2010; 15(5):253–60. Epub 2010/07/22. https://doi.org/10.1016/j.siny.2010. 05.007 PMID: 20646975; PubMed Central PMCID: PMC2930085.
- Lv H, Wang Q, Wu S, Yang L, Ren P, Yang Y, et al. Neonatal hypoxic ischemic encephalopathy-related biomarkers in serum and cerebrospinal fluid. Clinica chimica acta; international journal of clinical chemistry. 2015; 450:282–97. Epub 2015/09/01. https://doi.org/10.1016/j.cca.2015.08.021 PMID: 26320853.
- Douglas-Escobar M, Weiss MD. Hypoxic-ischemic encephalopathy: a review for the clinician. JAMA pediatrics. 2015; 169(4):397–403. Epub 2015/02/17. https://doi.org/10.1001/jamapediatrics.2014.3269 PMID: 25685948.
- Chalak LF, Sanchez PJ, Adams-Huet B, Laptook AR, Heyne RJ, Rosenfeld CR. Biomarkers for severity
 of neonatal hypoxic-ischemic encephalopathy and outcomes in newborns receiving hypothermia therapy. J Pediatr. 2014; 164(3):468–74.e1. Epub 2013/12/18. https://doi.org/10.1016/j.jpeds.2013.10.067
 PMID: 24332821; PubMed Central PMCID: PMC4006934.
- Ramaswamy V, Horton J, Vandermeer B, Buscemi N, Miller S, Yager J. Systematic review of biomarkers of brain injury in term neonatal encephalopathy. Pediatric neurology. 2009; 40(3):215–26. Epub 2009/02/17. https://doi.org/10.1016/j.pediatrneurol.2008.09.026 PMID: 19218035.
- 15. Berger RP, Bazaco MC, Wagner AK, Kochanek PM, Fabio A. Trajectory analysis of serum biomarker concentrations facilitates outcome prediction after pediatric traumatic and hypoxemic brain injury. Developmental neuroscience. 2010; 32(5–6):396–405. Epub 2010/09/18. https://doi.org/10.1159/000316803 PMID: 20847541; PubMed Central PMCID: PMC3215242.
- 16. Rundgren M, Karlsson T, Nielsen N, Cronberg T, Johnsson P, Friberg H. Neuron specific enolase and S-100B as predictors of outcome after cardiac arrest and induced hypothermia. Resuscitation. 2009; 80 (7):784–9. Epub 2009/05/27. https://doi.org/10.1016/j.resuscitation.2009.03.025 PMID: 19467754.
- Selakovic V, Raicevic R, Radenovic L. The increase of neuron-specific enolase in cerebrospinal fluid and plasma as a marker of neuronal damage in patients with acute brain infarction. J Clin Neurosci. 2005; 12(5):542–7. Epub 2005/06/01. https://doi.org/10.1016/j.jocn.2004.07.019 PMID: 15921910.
- Nagdyman N, Grimmer I, Scholz T, Muller C, Obladen M. Predictive value of brain-specific proteins in serum for neurodevelopmental outcome after birth asphyxia. Pediatr Res. 2003; 54(2):270–5. Epub 2003/05/09. https://doi.org/10.1203/01.pdr.0000072518.98189.a0 PMID: 12736385.
- Celtik C, Acunas B, Oner N, Pala O. Neuron-specific enolase as a marker of the severity and outcome of hypoxic ischemic encephalopathy. Brain & development. 2004; 26(6):398–402. Epub 2004/07/28. https://doi.org/10.1016/j.braindev.2003.12.007 PMID: 15275704.
- Tekgul H, Yalaz M, Kutukculer N, Ozbek S, Kose T, Akisu M, et al. Value of biochemical markers for outcome in term infants with asphyxia. Pediatric neurology. 2004; 31(5):326–32. Epub 2004/11/03. https://doi.org/10.1016/j.pediatrneurol.2004.05.004 PMID: 15519113.
- Ezgu FS, Atalay Y, Gucuyener K, Tunc S, Koc E, Ergenekon E, et al. Neuron-specific enolase levels and neuroimaging in asphyxiated term newborns. Journal of child neurology. 2002; 17(11):824–9. Epub 2003/02/15. https://doi.org/10.1177/08830738020170111301 PMID: 12585722.
- 22. Roka A, Kelen D, Halasz J, Beko G, Azzopardi D, Szabo M. Serum S100B and neuron-specific enolase levels in normothermic and hypothermic infants after perinatal asphyxia. Acta paediatrica. 2012; 101 (3):319–23. Epub 2011/10/11. https://doi.org/10.1111/j.1651-2227.2011.02480.x PMID: 21981269.
- 23. Massaro AN, Chang T, Baumgart S, McCarter R, Nelson KB, Glass P. Biomarkers S100B and neuron-specific enolase predict outcome in hypothermia-treated encephalopathic newborns*. Pediatr Crit Care Med. 2014; 15(7):615–22. Epub 2014/04/30. https://doi.org/10.1097/pcc.000000000000155 PMID: 24777302; PubMed Central PMCID: PMC4376375.

- Kelen D, Andorka C, Szabo M, Alafuzoff A, Kaila K, Summanen M. Serum copeptin and neuron specific enolase are markers of neonatal distress and long-term neurodevelopmental outcome. PloS one. 2017; 12(9):e0184593. Epub 2017/09/21. https://doi.org/10.1371/journal.pone.0184593 PMID: 28931055; PubMed Central PMCID: PMC5607206.
- Massaro AN, Wu YW, Bammler TK, Comstock B, Mathur A, McKinstry RC, et al. Plasma Biomarkers of Brain Injury in Neonatal Hypoxic-Ischemic Encephalopathy. J Pediatr. 2018; 194:67–75 e1. Epub 2018/ 02/27. https://doi.org/10.1016/j.jpeds.2017.10.060 PMID: 29478510.
- Garcia-Alix A, Cabanas F, Pellicer A, Hernanz A, Stiris TA, Quero J. Neuron-specific enolase and myelin basic protein: relationship of cerebrospinal fluid concentrations to the neurologic condition of asphyxiated full-term infants. Pediatrics. 1994; 93(2):234

 –40. PMID: 7510064.
- Blennow M, Savman K, Ilves P, Thoresen M, Rosengren L. Brain-specific proteins in the cerebrospinal fluid of severely asphyxiated newborn infants. Acta paediatrica. 2001; 90(10):1171–5. Epub 2001/11/ 08. https://doi.org/10.1080/080352501317061594 PMID: 11697430.
- 28. Thornberg E, Thiringer K, Hagberg H, Kjellmer I. Neuron specific enolase in asphyxiated newborns: association with encephalopathy and cerebral function monitor trace. Arch Dis Child Fetal Neonatal Ed. 1995; 72(1):F39–42. Epub 1995/01/01. https://doi.org/10.1136/fn.72.1.f39 PMID: 7743283; PubMed Central PMCID: PMC2528422.
- Sun J, Li J, Cheng G, Sha B, Zhou W. Effects of hypothermia on NSE and S-100 protein levels in CSF in neonates following hypoxic/ischaemic brain damage. Acta paediatrica. 2012; 101(8):e316–20. Epub 2012/03/29. https://doi.org/10.1111/j.1651-2227.2012.02679.x PMID: 22452413.
- Garca-Alix A, Quero J. Brain-specific proteins as predictors of outcome in asphyxiated term infants. Acta paediatrica. 2001; 90(10):1103–5. Epub 2001/11/08. https://doi.org/10.1080/080352501317061459 PMID: 11697416.
- Alsina M, Martin-Ancel A, Alarcon-Allen A, Arca G, Gaya F, Garcia-Alix A. The Severity of Hypoxic-Ischemic Encephalopathy Correlates With Multiple Organ Dysfunction in the Hypothermia Era. Pediatr Crit Care Med. 2017; 18(3):234–40. https://doi.org/10.1097/PCC.000000000001068 PMID: 28107263.
- 32. Carreras N, Alsina M, Alarcon A, Arca-Diaz G, Agut T, Garcia-Alix A. Efficacy of passive hypothermia and adverse events during transport of asphyxiated newborns according to the severity of hypoxic-ischemic encephalopathy. J Pediatr (Rio J). 2018; 94(3):251–7. https://doi.org/10.1016/j.jped.2017.05.
 009 PMID: 28822711.
- Hellström-Westas L DVL, Rosen I. An atlas of amplitude integrated EEGs in the newborn. 2nd ed ed. London2008 2008.
- **34.** Agut T, Leon M, Rebollo M, Muchart J, Arca G, Garcia-Alix A. Early identification of brain injury in infants with hypoxic ischemic encephalopathy at high risk for severe impairments: accuracy of MRI performed in the first days of life. BMC pediatrics. 2014; 14:177. Epub 2014/07/10. https://doi.org/10.1186/1471-2431-14-177 PMID: 25005267; PubMed Central PMCID: PMC4113122.
- Martinez-Biarge M, Diez-Sebastian J, Kapellou O, Gindner D, Allsop JM, Rutherford MA, et al. Predicting motor outcome and death in term hypoxic-ischemic encephalopathy. Neurology. 2011; 76 (24):2055–61. Epub 2011/06/15. https://doi.org/10.1212/WNL.0b013e31821f442d PMID: 21670434; PubMed Central PMCID: PMC3111238.
- Rutherford MA. The asphyxiated term infant. 2002. In: MRI of the Neonatal Brain [Internet]. UK: Saunders. Available from: http://www.mrineonatalbrain.com/index.php.
- Rescorla LA. Assessment of young children using the Achenbach System of Empirically Based Assessment (ASEBA). Ment Retard Dev Disabil Res Rev. 2005; 11(3):226–37. Epub 2005/09/15. https://doi.org/10.1002/mrdd.20071 PMID: 16161094.
- Surveillance of cerebral palsy in Europe: a collaboration of cerebral palsy surveys and registers. Surveillance of Cerebral Palsy in Europe (SCPE). Dev Med Child Neurol. 2000; 42(12):816–24. Epub 2000/12/29. https://doi.org/10.1017/s0012162200001511 PMID: 11132255.
- Merchant N, Azzopardi D. Early predictors of outcome in infants treated with hypothermia for hypoxicischaemic encephalopathy. Dev Med Child Neurol. 2015; 57 Suppl 3:8–16. Epub 2015/03/25. https:// doi.org/10.1111/dmcn.12726 PMID: 25800487.
- Merhar SL, Chau V. Neuroimaging and Other Neurodiagnostic Tests in Neonatal Encephalopathy. Clinics in perinatology. 2016; 43(3):511–27. Epub 2016/08/16. https://doi.org/10.1016/j.clp.2016.04.009
 PMID: 27524451.
- Del Rio R, Ochoa C, Alarcon A, Arnaez J, Blanco D, Garcia-Alix A. Amplitude Integrated Electroencephalogram as a Prognostic Tool in Neonates with Hypoxic-Ischemic Encephalopathy: A Systematic Review. PloS one. 2016; 11(11):e0165744. Epub 2016/11/02. https://doi.org/10.1371/journal.pone. 0165744 PMID: 27802300; PubMed Central PMCID: PMC5089691.

- 42. Shankaran S, Laptook AR, Tyson JE, Ehrenkranz RA, Bann CM, Das A, et al. Evolution of encephalopathy during whole body hypothermia for neonatal hypoxic-ischemic encephalopathy. J Pediatr. 2012; 160(4):567–72.e3. Epub 2011/11/05. https://doi.org/10.1016/j.jpeds.2011.09.018 PMID: 22050871; PubMed Central PMCID: PMC3299861.
- 43. Gunn AJ, Wyatt JS, Whitelaw A, Barks J, Azzopardi D, Ballard R, et al. Therapeutic hypothermia changes the prognostic value of clinical evaluation of neonatal encephalopathy. J Pediatr. 2008; 152 (1):55–8, 8 e1. Epub 2007/12/25. https://doi.org/10.1016/j.jpeds.2007.06.003 PMID: 18154900.
- Boudes E, Tan X, Saint-Martin C, Shevell M, Wintermark P. MRI obtained during versus after hypothermia in asphyxiated newborns. Arch Dis Child Fetal Neonatal Ed. 2015; 100(3):F238–42. Epub 2015/01/22. https://doi.org/10.1136/archdischild-2014-306550 PMID: 25605620.
- 45. Charon V, Proisy M, Bretaudeau G, Bruneau B, Pladys P, Beuchee A, et al. Early MRI in neonatal hypoxic-ischaemic encephalopathy treated with hypothermia: Prognostic role at 2-year follow-up. European journal of radiology. 2016; 85(8):1366–74. Epub 2016/07/18. https://doi.org/10.1016/j.ejrad.2016.05. 005 PMID: 27423674.
- 46. Weeke LC, Boylan GB, Pressler RM, Hallberg B, Blennow M, Toet MC, et al. Role of EEG background activity, seizure burden and MRI in predicting neurodevelopmental outcome in full-term infants with hypoxic-ischaemic encephalopathy in the era of therapeutic hypothermia. European Journal of Paediatric Neurology. 2016; 20(6):855–64. https://doi.org/https://doi.org/10.1016/j.ejpn.2016.06.003.
- Wilkinson D. The window of opportunity for treatment withdrawal. Archives of pediatrics & adolescent medicine. 2011; 165(3):211–5. Epub 2011/03/09. https://doi.org/10.1001/archpediatrics.2011.4 PMID: 21383270.
- Haimoto H, Hosoda S, Kato K. Differential distribution of immunoreactive S100-alpha and S100-beta proteins in normal nonnervous human tissues. Lab Invest. 1987; 57(5):489–98. Epub 1987/11/01. PMID: 3316838.
- 49. Isgro MA, Bottoni P, Scatena R. Neuron-Specific Enolase as a Biomarker: Biochemical and Clinical Aspects. Advances in experimental medicine and biology. 2015; 867:125–43. Epub 2015/11/05. https://doi.org/10.1007/978-94-017-7215-0_9 PMID: 26530364.
- Polglase GR, Ong T, Hillman NH. Cardiovascular Alterations and Multiorgan Dysfunction After Birth Asphyxia. Clinics in perinatology. 2016; 43(3):469–83. Epub 2016/08/16. https://doi.org/10.1016/j.clp.2016.04.006 PMID: 27524448; PubMed Central PMCID: PMC4988334.
- Pelinka LE, Hertz H, Mauritz W, Harada N, Jafarmadar M, Albrecht M, et al. Nonspecific increase of systemic neuron-specific enolase after trauma: clinical and experimental findings. Shock (Augusta, Ga). 2005; 24(2):119–23. Epub 2005/07/27. https://doi.org/10.1097/01.shk.0000168876.68154.43 PMID: 16044081.
- 52. Kaiser E, Kuzmits R, Pregant P, Burghuber O, Worofka W. Clinical biochemistry of neuron specific enolase. Clinica chimica acta; international journal of clinical chemistry. 1989; 183(1):13–31. Epub 1989/07/31. https://doi.org/10.1016/0009-8981(89)90268-4 PMID: 2548772.
- Ramont L, Thoannes H, Volondat A, Chastang F, Millet MC, Maquart FX. Effects of hemolysis and storage condition on neuron-specific enolase (NSE) in cerebrospinal fluid and serum: implications in clinical practice. Clinical chemistry and laboratory medicine. 2005; 43(11):1215–7. Epub 2005/10/20. https://doi.org/10.1515/CCLM.2005.210 PMID: 16232088.
- 54. Rundgren M, Cronberg T, Friberg H, Isaksson A. Serum neuron specific enolase—impact of storage and measuring method. BMC Res Notes. 2014; 7:726. Epub 2014/10/17. https://doi.org/10.1186/1756-0500-7-726 PMID: 25319200; PubMed Central PMCID: PMC4216829.
- **55.** Echeverria-Palacio CM, Agut T, Arnaez J, Valls A, Reyne M, Garcia-Alix A. Neuron-Specific Enolase in Cerebrospinal Fluid Predicts Brain Injury After Sudden Unexpected Postnatal Collapse. Pediatric neurology. 2019. Epub 2019/04/27. https://doi.org/10.1016/j.pediatrneurol.2019.02.020 PMID: 31023601.
- 56. Arca G, Arnaez J, Agut T, Nunez C, Stephan-Otto C, Valls A, et al. Neuron-specific enolase is correlated with lesion topology, relative infarct volume and outcome of symptomatic NAIS. Arch Dis Child Fetal Neonatal Ed. 2019. Epub 2019/06/16. https://doi.org/10.1136/archdischild-2018-316680 PMID: 31201253.
- Barone FC, Clark RK, Price WJ, White RF, Feuerstein GZ, Storer BL, et al. Neuron-specific enolase increases in cerebral and systemic circulation following focal ischemia. Brain research. 1993; 623 (1):77–82. Epub 1993/09/24. https://doi.org/10.1016/0006-8993(93)90012-c PMID: 8221097.
- Horn M, Seger F, Schlote W. Neuron-specific enolase in gerbil brain and serum after transient cerebral ischemia. Stroke. 1995; 26(2):290–6; discussion 6–7. Epub 1995/02/01. https://doi.org/10.1161/01.str. 26.2.290 PMID: 7831703.
- 59. Wunderlich MT, Ebert AD, Kratz T, Goertler M, Jost S, Herrmann M. Early neurobehavioral outcome after stroke is related to release of neurobiochemical markers of brain damage. Stroke. 1999; 30 (6):1190–5. Epub 1999/06/04. https://doi.org/10.1161/01.str.30.6.1190 PMID: 10356098.

- 60. Wunderlich MT, Lins H, Skalej M, Wallesch CW, Goertler M. Neuron-specific enolase and tau protein as neurobiochemical markers of neuronal damage are related to early clinical course and long-term outcome in acute ischemic stroke. Clin Neurol Neurosurg. 2006; 108(6):558–63. Epub 2006/02/07. https://doi.org/10.1016/j.clineuro.2005.12.006 PMID: 16457947.
- 61. Massaro AN, Chang T, Kadom N, Tsuchida T, Scafidi J, Glass P, et al. Biomarkers of brain injury in neonatal encephalopathy treated with hypothermia. J Pediatr. 2012; 161(3):434–40. Epub 2012/04/13. https://doi.org/10.1016/j.jpeds.2012.02.047 PMID: 22494878; PubMed Central PMCID: PMC3580861.
- 62. Garcia-Alix A, Arnaez J, Cortes V, Girabent-Farres M, Arca G, Balaguer A. Neonatal hypoxic-ischaemic encephalopathy: most deaths followed end-of-life decisions within three days of birth. Acta paediatrica. 2013; 102(12):1137–43. https://doi.org/10.1111/apa.12420 PMID: 24102859.
- 63. Arnaez J, Garcia-Alix A, Arca G, Caserio S, Valverde E, Moral MT, et al. Population-Based Study of the National Implementation of Therapeutic Hypothermia in Infants with Hypoxic-Ischemic Encephalopathy. Ther Hypothermia Temp Manag. 2018; 8(1):24–9. Epub 2017/08/12. https://doi.org/10.1089/ther.2017.0024 PMID: 28800288.