

## The Osteoblastic and Osteoclastic Interactions in Spinal Metastases Secondary to Prostate Cancer

Sathana Dushyanthen, Davina A.F. Cossigny and Gerald M.Y. Quan

Spinal Biology Research Laboratory, Department of Spinal Surgery, University of Melbourne Department of Surgery, Austin Health, Heidelberg Victoria, Australia.

**ABSTRACT:** Prostate cancer (PC) is one of the most common cancers arising in men and has a high propensity for bone metastasis, particularly to the spine. At this stage, it often causes severe morbidity due to pathological fracture and/or metastatic epidural spinal cord compression which, if untreated, inevitably leads to intractable pain, neurological deficit, and paralysis. Unfortunately, the underlying molecular mechanisms driving growth of secondary PC in the bony vertebral column remain largely unknown. Further investigation is warranted in order to identify therapeutic targets in the future. This review summarizes the current understanding of PC bone metastasis in the spine, highlighting interactions between key tumor and bone-derived factors which influence tumor progression, especially the functional roles of osteoblasts and osteoclasts in the bone microenvironment through their interactions with metastatic PC cells and the critical pathway RANK/RANKL/OPG in bone destruction.

**KEY WORDS:** prostate cancer, spine, metastasis, osteoblasts, osteoclasts, RANK, RANKL, OPG bone microenvironment

**CITATION:** Dushyanthen et al. The Osteoblastic and Osteoclastic Interactions in Spinal Metastases Secondary to Prostate Cancer. *Cancer Growth and Metastasis* 2013;6:61–80 doi:10.4137/CGM.S12769.

**RECEIVED:** July 10, 2013. **RESUBMITTED:** October 6, 2013. **ACCEPTED FOR PUBLICATION:** October 7, 2013.

**ACADEMIC EDITOR:** Marc D. Basson, Editor in Chief

**TYPE:** Review

**FUNDING:** This work was supported by the National Health and Medical Research Council of Australia (Fellowship No. 558418) and the Austin Health Medical Research Foundation.

**COMPETING INTERESTS:** Authors disclose no potential conflicts of interest.

**COPYRIGHT:** © the authors, publisher and licensee Libertas Academica Limited. This is an open-access article distributed under the terms of the Creative Commons CC-BY-NC 3.0 License.

**CORRESPONDENCE:** [gerald.quan@austin.org.au](mailto:gerald.quan@austin.org.au)

### Introduction

Prostate cancer (PC) is the fifth most commonly diagnosed cancer type globally and the second most common disease causing mortality in males.<sup>1</sup> Five-year survival rates for localized PC are high (almost 100%), however this drops to less than 33% in patients with metastatic disease.<sup>2</sup> Over 90% of patients with advanced PC develop bone metastases, with the most common site of establishment being the spine.<sup>3,4</sup> These patients are at risk of suffering due to severe symptoms such as bone pain and instability, pathological fracture, neurological deficit, and metastatic epidural spinal cord compression which may ultimately cause paralysis.<sup>5,6</sup> Despite advances in operative and non-operative management of symptomatic bone and spinal metastases from PC, treatment options are mainly palliative and are limited in efficacy in cases of severe bone loss and destruction, pathological fracture, and bony instability.

There is currently no effective therapy for advanced metastatic PC in terms of substantially improving morbidity and prolonging survival.<sup>7</sup> This review summarizes the interactions of the tumor with the bone microenvironment and the role of the bone remodeling cells, osteoblasts and osteoclasts, in the spread of metastatic PC to the bony skeleton and spine.

### Spinal Metastasis in PC

In the early stage of PC, localized PC cells are confined to the lobes of the prostate and, depending on the Gleason score (a histopathological method of grading PC), active surveillance, radical prostatectomy or radiotherapy are the current first line treatment options.<sup>6</sup> When the disease progresses, tumor cells develop a more aggressive phenotype which enables them to grow outside their initial localized surrounding, precipitated by genetic mutations as well as interactions



and stimulation from the local environment.<sup>3</sup> Paget<sup>8</sup> proposed the “seed and soil” theory in 1889, whereby neoplastic tumor cells metastasize only to specific environments based on suitability for growth.<sup>8,9</sup> Indeed, it has been suggested that the highly vascularized nature of the axial skeleton provides such an environment and the spine is the commonest site of skeletal metastasis secondary to PC.<sup>10</sup>

**The emerging importance of stem cells in PC associated bone metastasis.** PC was originally thought to be due to abnormal proliferating epithelial cells. However more recently has been shown to involve complex interactions between PC epithelial cells and the surrounding stromal tissue. Multiple signaling pathways provide cross-talk between these compartments via androgen receptors, tyrosine kinase receptor signaling, and immune surveillance. The cancer stem cell (CSC) hypothesis proposes that there is a small subpopulation of cancer cells that are more resistant to chemotherapy and toxicity.<sup>11</sup> These cells are able to drive tumor growth and metastasis, thus leading to relapse following treatment. CSCs are more resistant to stress induced by chemotherapy, with an increased expression of anti-apoptotic molecules and a reduced expression of pro-apoptotic genes.<sup>11</sup> Furthermore, it is suggested that these cells exhibit preferential activation of DNA damage checkpoint response and increased capacity for repair, thus chemotherapy may enrich the CSC population within tumors by eliminating sensitive cells.<sup>11</sup> Multipotent mesenchymal stem cells (MSCs) found within the bone marrow have recently been identified as playing an important role in supporting PC growth and survival in bone. These cells possess properties of self-renewal and repair, migrating towards active tumorigenesis and integrating into the niche contributing to the development of cancer-associated fibroblasts. Consequently, because of their tumor-tropic migratory response, MSCs are emerging as promising anti-cancer agents.<sup>12</sup> PC cells require an invasive capacity which is stimulated through type I collagen from bone marrow MSCs, inducing the secretions of proteases from PC cells.<sup>13</sup> In addition, stromal derived factor 1 $\alpha$  (SDF1 $\alpha$ ) receptor and CXC chemokine receptor-4 (CXCR4) are essential for metastatic spread, allowing tumor cells to chemoattract towards bone and easily access bone marrow cellular niches, ultimately promoting survival and growth. Furthermore, SDF1 $\alpha$  promotes tumor angiogenesis by attracting endothelial cells to the tumor.<sup>14</sup> Therefore, CSCs constitute a repository niche of androgen-insensitive and chemotherapy-resistant cells responsible for secondary metastatic progression. As such, they are an integral target for the cessation of PC progression.

**The role of epithelial to mesenchymal transition in metastatic progression.** PC is of epithelial origin and the process of epithelial to mesenchymal transition (EMT) is a critical step in the conversion of early stage cancer to an invasive and metastatic cancer in advanced disease progression.<sup>15</sup> EMT is a process whereby polarized epithelial cells convert into motile mesenchymal cells by undergoing phenotypic

transitions during malignant progression.<sup>16</sup> Consequently, these cells acquire stem cell-like properties which permit resistance to treatment, adhesive and invasive capacity, changes in morphology and cellular architecture, migratory potential, and the ability to undergo metastatic progression.<sup>16,17</sup> These cancer cells are thought to hijack core biological processes through key initiating events such as genetic mutation and activation of important oncogenic pathways involved in regulating cell proliferation and migration, cross-talk, angiogenesis, and epithelial-mesenchymal signaling.<sup>17</sup>

Pathological EMT in tumor cells occurs as a result of transcriptional reprogramming of abnormal survival signals via receptors such as fibroblast growth factor receptor (FGFR), platelet-derived growth factor receptor (PDGFR), transforming growth factor- $\beta$  receptor (TGF- $\beta$ R), and insulin-like growth factor-1 receptor (IGF-1R), as well as regulatory kinases such as phosphoinositide 3-kinase (PI3K), protein kinase B (AKT), and mammalian target of rapamycin (mTOR). These factors contribute to bone metastasis through molecular cross talk which is mediated by cytokines and other paracrine factors including vimentin, N-cadherin, platelet-derived growth factor (PDGF), nuclear factor kappa-light-chain-enhancer of activated B cells (NF- $\kappa$ B), and Notch-1.<sup>18</sup> These factors are expressed by the circulating tumor cells and are also able to acquire bone-like properties through transcriptional reprogramming of abnormal survival signals through independent activation of signaling pathways.<sup>17,18</sup>

Xu et al<sup>19</sup> reported that PC cells undergo EMT through cellular interactions with the host microenvironment, increasing the metastatic potential to bone.<sup>17,19</sup> A hallmark of EMT is the functional loss of epithelial cell markers and transmembrane adhesion glycoproteins E-Cadherin, cytokeratins, and  $\beta$ -catenin; factors which are proposed to suppress cancer progression.<sup>20</sup> In this process, mesenchymal cell markers, namely N-Cadherin, fibronectin and Vimentin, are produced. These factors are found at the invasive front between the tumor and interacting microenvironment, signaling pathways that facilitate migration and survival.<sup>21</sup> Zhau et al demonstrated the plasticity of cancer cells which, upon induction via these soluble factors, developed the ability to express markers found in osteoblasts; a phenomenon known as ‘osteomimicry.’<sup>22</sup> These factors include osteocalcin (OC), bone sialoprotein (BSP), osteopontin (OPN), non-collagenous bone matrix proteins, and receptor activator of nuclear factor kappa-B ligand (RANKL), which together allow cancer cells to survive and thrive within the bone microenvironment.<sup>22</sup>

Matrix metalloproteinases (MMPs), particularly membrane type 1 MMP (MT1-MMP), have been reported in PC EMT through both paracrine and autocrine pathways and are capable of cleaving E-cadherin, thus disrupting its function by interfering with cell adhesion and polarity of the cells.<sup>15</sup> Furthermore, secreted MMPs (MMP-2, -3, -9, 28) have been implicated in PC cell EMT.<sup>15</sup> MMP proteolysis serves a path-clearing mechanism in facilitating the movement of

cancer cells or groups of cells through the extracellular matrix (ECM).<sup>23</sup> Cleavage of certain ECM components leads to the unmasking of sites, generating fragments with new biological activities and modulating migration, growth and angiogenesis.<sup>23</sup>

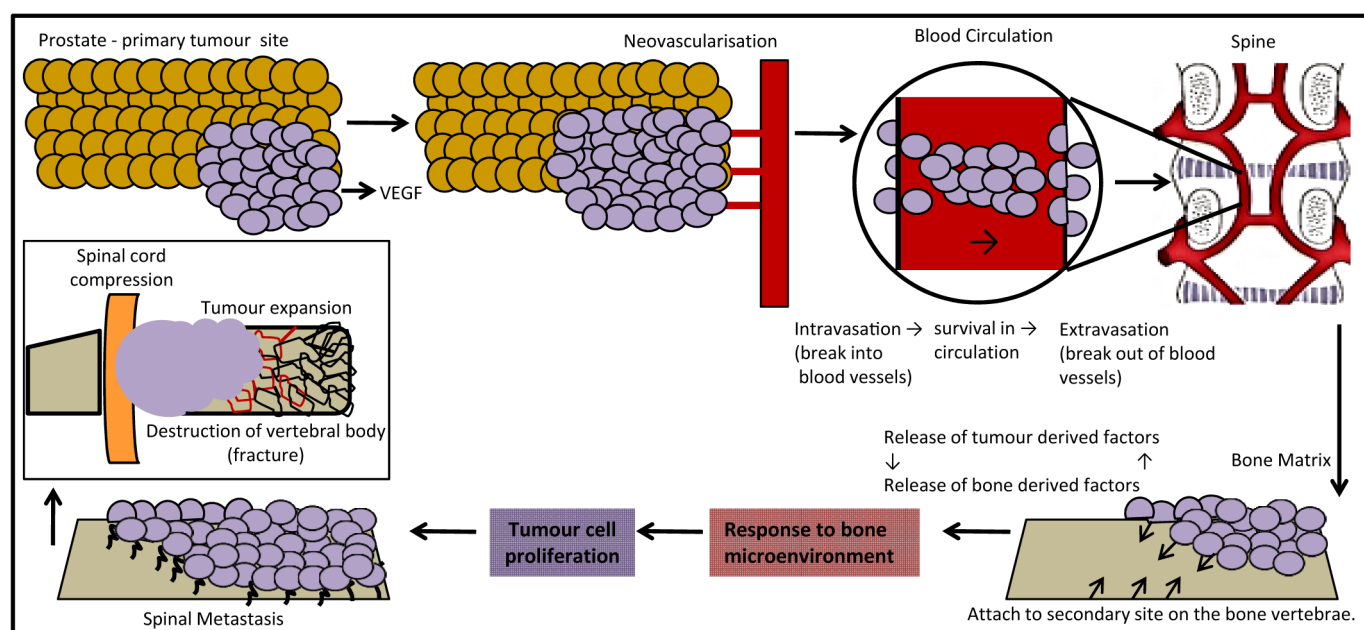
Bone enriched growth factors, cytokines, proteases, and components of the ECM in bone marrow, provide a hospitable environment for harboring circulating PC cells.<sup>24</sup> The process of metastasis is dependent on both the intrinsic properties of tumor cells as well as the response of the host bone microenvironment.<sup>25</sup> Studies using bone xenograft models have demonstrated that injection of PC cells adjacent to bone leads to cancer cell migration towards bone.<sup>26</sup> Thus, it appears that a bidirectional interaction between bone cells within the bone microenvironment and the PC cells exists, which facilitates tumor growth in the spine.<sup>27</sup> During PC progression to metastatic disease, cancer cells acquire characteristics that promote interactions with the local microenvironment.<sup>3</sup> The tumor releases vascular endothelial growth factor (VEGF), which initiates angiogenesis, the process of new blood vessel formation leading to an enhanced blood supply to the tumor.<sup>3</sup> The surrounding stromal cells produce growth factors and cytokines such as transforming growth factor- $\beta$  (TGF- $\beta$ ), insulin-like growth factor (IGF), fibroblast growth factor (FGF), PDGF, and bone morphogenetic proteins (BMPs) that can act directly on cancer cells as well as induce angiogenesis, and stimulating growth and spread of the tumor.<sup>26</sup> Disruptions to intracellular adhesion lead to detachment and dissemination of the tumor cells from the primary site, followed by local invasion into the

blood vessels and lymphatic system, extravasation, migration, and invasion of the bone marrow.<sup>2,28</sup> The tumor cells proliferate and colonize bone through migration and adherence, followed by degradation of the ECM.<sup>2</sup> Once PC cells enter the bone compartment, close interactions with factors of the surrounding bone cells, matrix, and the environment occur in a reciprocal fashion via cytokine mediators, which allow osteoblastic, osteoclastic, and mixed lesions to form.<sup>2</sup> The mechanism by which PC metastasizes to bone within the spine is illustrated in Figure 1.

A rich venous plexus known as Batson's plexus, surrounds the prostate and connects to the venous drainage of the spine.<sup>29</sup> This collection of veins provide a migratory route for tumor cells to localize in the lumbosacral region spinal metastases, which is a common occurrence in advanced PC.<sup>3</sup>

### Normal Bone Physiology and Remodeling

Bone physiology is a balanced and dynamic state consisting of continuous and coordinated cycles of bone formation by osteoblasts and bone resorption by osteoclasts, critical for maintenance of the structural integrity of bone.<sup>30</sup> Osteoblasts are derived from MSCs of bone marrow stroma and synthesize and secrete proteins that form the ECM of bone, facilitating the deposition and mineralization of bone while maintaining the structural integrity of the skeleton.<sup>31,32</sup> Formation of bone via osteoblasts involves the production of type 1 procollagen. This undergoes processing, modification, and secretion forming cross-linked arrangements whereby osteoblasts become



**Figure 1. Mechanisms involved in bone metastasis.** At the primary site within the prostate, the tumor secretes factors which promote growth and angiogenesis. It also secretes factors that allow detachment from primary site and migration into blood vessels. Cancer cells then migrate through the blood circulation and are attracted towards bone. Subsequent extravasation and growth within and invasion of bone occurs involving complex interactions between the tumor and the local bone microenvironment. This ultimately leads to osteoclast-induced bone destruction and pathological fracture, and the expanding tumor mass impinges on the adjacent neural structures and spinal cord, causing neurological deficit and paralysis.



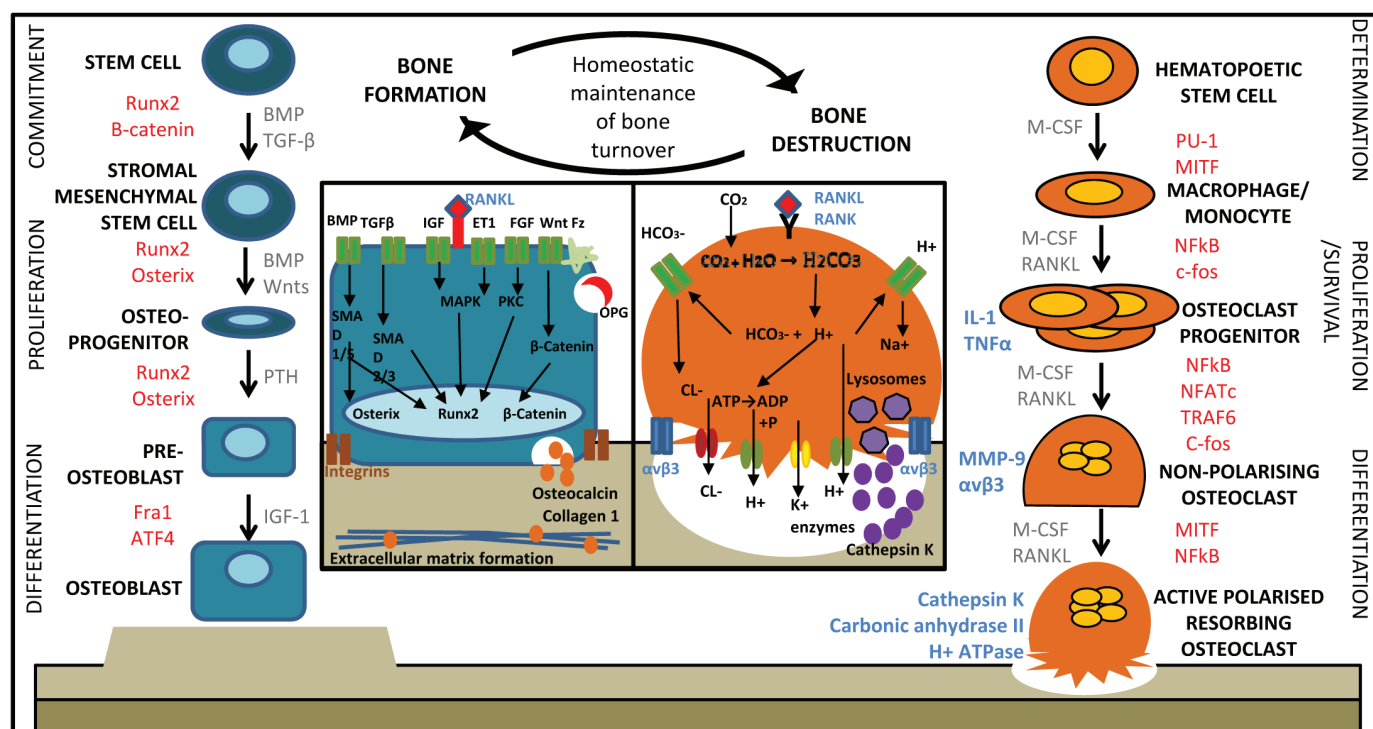
osteocytes once embedded in the bone. The newly formed bone matrix induced by osteoblasts is subsequently mineralized with the deposition of hydroxyapatite crystals to increase the resistance to compression.<sup>2</sup> Osteoblasts are also responsible for the regulation of osteoclastic bone resorption via the secretion of factors that induce osteoclast differentiation, activation, and survival.<sup>33,34</sup> The differentiation and growth of osteoblasts is regulated by complex signaling pathways including BMPs, IGFs, TGF- $\beta$ , and Wnt.<sup>2</sup> Differentiated osteoblasts secrete growth factors that become embedded in the bone matrix during formation.<sup>35</sup> These factors are released into the microenvironment during the bone resorption process.<sup>36</sup> Runt-related transcription factor 2 (Runx2) is the master transcription regulator of osteoblast differentiation and is over-expressed in PC metastasizing to bone.<sup>37</sup> It has the ability to increase oncogenic potential through the regulation of genes involved in metastasis and invasion, which facilitate interactions between the tumor and bone microenvironment causing severe osteoblastic lesions.<sup>2,38</sup> Runx2 functions in many regulatory processes such as the suppression of cell growth, epigenetic control of genes in mitosis, and bone turnover, which become unregulated in PC.<sup>38</sup>

Osteoclasts are derived from hematopoietic cells of the monocyte-macrophage myeloid lineage. These cells form through the cytoplasmic fusion of their mononuclear precursor resulting in multinucleated bone resorbing cells.<sup>3,39</sup> The differentiation and maturation of osteoclasts is regulated through cytokines released from osteoblasts.<sup>2</sup> The cytokine macrophage colony stimulating factor (M-CSF) controls proliferation and promotes survival of osteoclast precursors.<sup>40</sup> Furthermore, it stimulates activity in the mature resorbing cells such as proliferation, motility, and cytoskeletal organization.<sup>2</sup> Osteoclasts are involved in the process of bone resorption; the degradation of mineral matrices such as calcified cartilage and bone during growth and development, skeletal homeostasis, repair, and remodeling.<sup>34</sup> They tightly adhere and bind to the bone surface through polarization via  $\alpha v \beta 3$ ,  $\alpha v \beta 5$ , and  $\alpha 2 \beta 1$  integrins, using actin-rich podosomes to form sealed cytoplasmic extensions (transcytotic vesicle or vacuole) with the underlying bone matrix.<sup>28</sup> Within this zone, they form a ruffled border membrane, which increases surface area and enables secretion of acid proteases, lysosomal enzymes, protons, and Cathepsin K onto the extracellular compartment of the bone surface.<sup>2</sup> These secretions are essential for bone matrix solubilization and acid protease induced digestion of the organic matrix.<sup>2</sup> Tartrate resistant acid phosphatase 5b (TRAP5b) is integral to the bone matrix digestion process. TRAP5b mediates two functions: phosphatase activity, which facilitates enzymatic degradation of bone, and the generation of a reactive oxygen species, which degrades collagen.<sup>41</sup> Osteoclastic bone destruction results in disintegration of the collagen from within the matrix as well as the release of calcium, growth factors, and cytokines.<sup>42</sup> Osteoclastic action is highly up regulated in PC leading to excessive bone destruction.<sup>2,43</sup> The overall rate

of osteoclastic bone resorption can be regulated on multiple levels including the differentiation rate from the monocyte-macrophage precursor pool, regulation of mature osteoclast activity through key functional proteins, and through the mature-differentiated survival rate.<sup>44</sup> The origin, function, and regulation of osteoblasts and osteoclasts are described in Figure 2. These processes are tightly regulated in normal bone remodelling where there is equal bone destruction and bone formation, with no net change in overall bone mass. However, in the presence of PC cells, these process become highly disrupted and unregulated, leading to bone destruction and replacement by tumor.<sup>2</sup> The bone marrow consists of not only osteoblasts and osteoclasts, but also hematopoietic cells, adipocytes, and immune cells. Together, the bone matrix and abundant growth factors secreted by these cells make the bone microenvironment a complex and fertile environment for tumor growth.<sup>2</sup>

### Pathological Bone Turnover in PC Bone Metastasis

The bidirectional interactions of bone cells (osteoblasts and osteoclasts) with PC cells not only suggest that tumor-derived growth factors affect bone cells, but also that cells in the bone microenvironment stimulate metastatic tumor growth and progression. The pathological condition of bone is caused by dysfunctional signaling mechanisms for bone remodeling due to the presence and influence of the invading tumor, thus resulting in the destruction of bone. Within the bone matrix there are several factors, such as TGF- $\beta$ , IGF, FGF, and PDGF, which are stored during bone formation and released during resorption.<sup>45,46</sup> In PC, close cross-talk between the bone microenvironment and the tumor results in the release of these growth factors into the microenvironment, up-regulating differentiation and activity of osteoblasts, the bone forming cells, and subsequently osteoclasts (bone resorbing cells).<sup>47,48</sup> Increased production and release of these growth factors from dissolved bone during uncontrolled turnover and remodeling results in co-stimulation of bone and tumor cells. Consequently, enhanced tumor survival, growth, proliferation, as well as osteoclastogenesis and accelerated bone destruction, occurs.<sup>49</sup> Osteoblastic, osteoclastic, and mixed lesions can occur in PC skeletal metastases, due to an imbalance between osteoblastic mediated bone formation and osteoclast mediated bone resorption.<sup>50</sup> In particular, PC is known to often cause osteoblastic lesions<sup>51</sup> caused by over-expression of numerous pro-osteoblastic factors, such as RANKL, parathyroid hormone-related protein (PTHrP), BMPs, TGF- $\beta$ , IGFs, FGFs, interleukins (ILs), PDGF, and VEGF, leading to un-regulated osteoblastic function via the activation of proliferation and differentiation signaling pathways such as Wnt.<sup>51,52</sup> Furthermore, conditions within the bone environment, such as increased calcium release, hypoxia, and acidity, allow the tumor to thrive.<sup>28,53</sup> The newly formed bone contains ECM proteins such as Type I collagen, osteonectin, and BSP, which act as chemoattractants to PC cells



**Figure 2. The origin, function and regulation of osteoblasts & osteoclasts in bone physiology.** Bone turnover is maintained through the homeostatic balance between osteoblast-mediated bone formation and osteoclast-mediated bone resorption. Commitment to the monocyte/macrophage/osteoclast lineage leading to survival and proliferation is dependent on transcription factor PU-1 and signaling from M-CSF. Differentiation requires signaling from RANKL, c-Fos, NFATc, and NfκB. Polarization and attachment of the activated osteoclast to the bone surface occurs via  $\alpha v \beta 3$  integrin an intracellular signaling proteins TRAF6 and c-Src. The process of bone resorption is carried out by the effector proteins cathepsin K (involved in matrix degradation), carbonic anhydrase II, and H<sup>+</sup> ATPase. Osteoblasts are activated by several growth factors including BMPs, TGF- $\beta$ , IGFs, EGFs, FGFs, and Wnt.

migrating to bone.<sup>2</sup> The various factors which are implicated in PC bone metastasis are summarized in Table 1.

Bone destruction from osteoclast activity leads to the release of growth factors from the matrices which possess the ability to act on metastatic cancer cells.<sup>28,45</sup> These factors indirectly promote angiogenesis and stimulate tumor-derived osteolytic and osteoblastic factors, subsequently remodeling the skeleton and accommodating tumor growth.<sup>28,45</sup> TGF- $\beta$  is the most abundant growth factor in the PC bone micro-environment and regulates bone development and remodeling via the promotion of osteoprogenitor differentiation.<sup>83</sup> Osteoblasts deposit TGF- $\beta$  into the bone matrix where it is then released and activated through osteoclastic resorption.<sup>84</sup> In PC, TGF- $\beta$  mediates metastasis through the activation of EMT, tumor cell invasion, and suppression of immune surveillance by increasing angiogenesis.<sup>28,85</sup> It stimulates bone metastasis via the induction of proteolytic gene expression through PTHrP within tumor cells.<sup>86</sup> TGF- $\beta$ -induced PTHrP acts to increase osteoblastic production of RANKL, consequently stimulating osteoclast formation and activity thus promoting bone metastasis.<sup>28</sup> Additionally, there is an irregular control of the resorption pathway, leading to bone destruction and resulting in the release of several bone matrix factors such as

IGF-1, basic FGF, and PDGF which further stimulate tumor growth.<sup>28</sup> Figure 3 illustrates the cycle involving bone and tumor cell interactions in the bone microenvironment and the various factors involved.

### The RANK/RANKL/OPG Signaling Mechanism Involved in Bone Destruction

Local bone destruction is a significant aspect of PC bone metastasis in the spine and is mediated through osteoclasts. As such, the RANK-RANKL-OPG axis has been well established as a key regulatory pathway of bone activity in physiological conditions and is highly up-regulated in pathological conditions. Receptor activator of the nuclear factor kappa-B receptor (RANK), RANKL, and osteoprotegerin (OPG) are members of the tumor necrosis factor (TNF) family.<sup>28</sup> RANK is a membrane-bound TNF receptor expressed by osteoclast precursors and mature osteoclasts and controls the development and apoptotic processes in bone.<sup>30</sup> RANKL is a type II transmembrane protein and is expressed on the surface of bone marrow stromal cells and by mature osteoblasts.<sup>67,87</sup> RANKL is essential for differentiation, activation, and survival of osteoclasts.<sup>67</sup> The release of RANKL from tumor cells into the bone microenvironment has been suggested to simulate

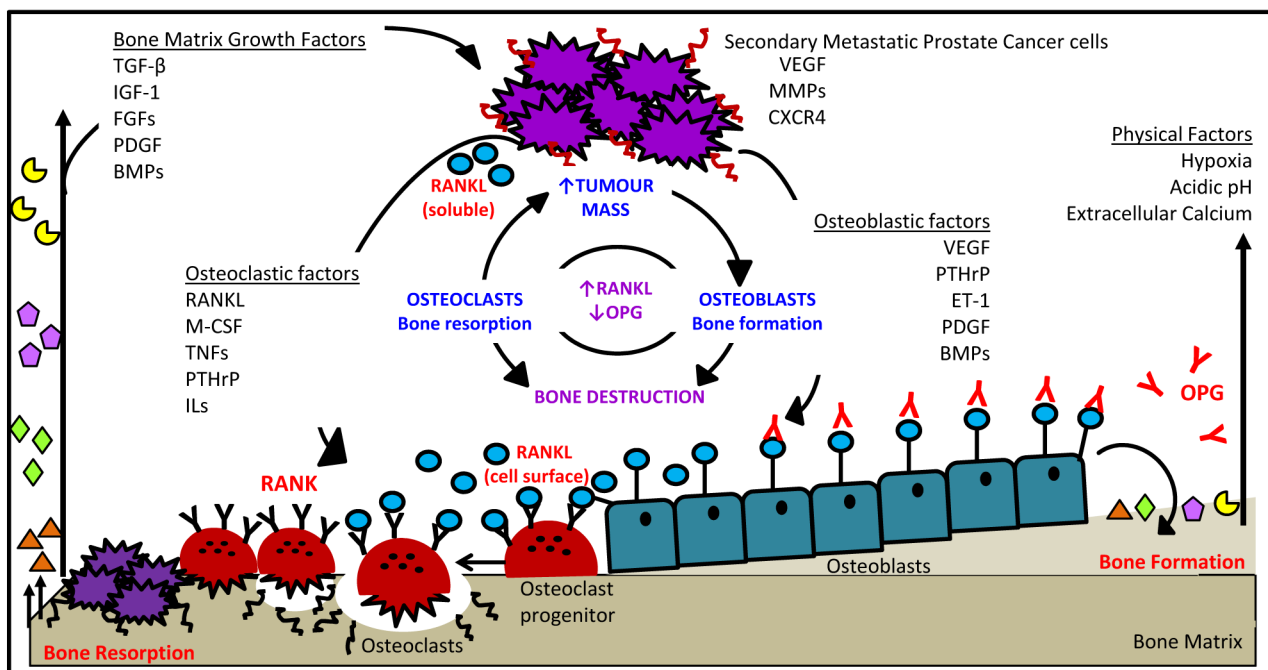
**Table 1.** The various mediators implicated in PC bone metastasis; their function and mechanism of action.

FACTOR	FUNCTION
<b>Bone morphogenetic proteins</b> BMP-2 BMP-4 BMP-6 BMP-7	BMPs play an integral role in endochondral ossification, osteogenesis and bone repair through the initiation of osteoblast differentiation (through Runx2 induction). <sup>54</sup> In PC, BMPs are involved in proliferation, migration and invasion in of cancer cells in bone metastasis. Furthermore, BMPs promote VEGF expression, leading to angiogenesis and endothelial cell migration in PC cells. <sup>54</sup> The expression levels of BMPs increase while those of the cognate receptors decrease with progression of disease. As PC cells produce BMPs, and BMPs generally decrease the proliferation of PC cells, however in advanced progression, loss of BMP receptor expression permits PC cells escape from the physiologic constraint on cellular proliferation imposed by BMPs, leading to a more aggressive phenotype by altering the tumour microenvironment. <sup>55</sup>
<b>Endothelin-1</b>	Endothelin-1 is a small vasoconstricting peptide produced by the vascular endothelium, and has a key role in vascular homeostasis. ET-1 production is stimulated by cytokines (ILs), growth factors (TNF $\alpha$ , TGF- $\beta$ , PDGF), leading to growth and proliferation of tumours. <sup>56</sup> Activation of the ET A receptor by ET-1 promotes tumour cell growth and survival, mediating processes involved in tumour invasion and metastasis, angiogenesis and the inhibition of apoptosis. ET-1 may also increase osteoblast proliferation and bone formation by crosstalk with Wnt signaling, leading to the suppression of the inhibitor of Wnt signaling, Dickkopf-1 (DKK1). <sup>57</sup>
<b>Insulin-like growth factors</b> IGF-I IGF-II	IGFs are survival factors-potent mitogens and anti-apoptotic factors involved in cell proliferation and differentiation. <sup>58</sup> IGF-I and II are abundant in the bone matrix and are released during bone resorption. IGF-1R signalling plays a role in malignant cell transformation, cancer progression, and metastatic spread of different types of tumours. IGFs promote survival and angiogenesis in PC. <sup>58</sup>
<b>Interleukins</b> IL-1 $\beta$ IL-6 IL-8 IL-11	Interleukins are pro-inflammatory cytokines and a key regulator of immunosuppression in advanced cancer. It is involved in the regulation of proliferation, apoptosis, migration, invasion, and angiogenesis. <sup>59</sup>  IL-6 has been implicated in PC bone metastasis progression—stimulates RANKL production, chemotherapeutic resistance, and androgen independence (androgen receptor dysfunction). IL-6 induces AR expression, which leads to TGF $\beta$ activation and consequently MMP-9, resulting in invasion of PC cells. <sup>60</sup>
<b>Integrins</b>	Integrins are cell surface receptors for extracellular matrix proteins and play a key role in cell survival, proliferation, migration and gene expression, apoptosis, cell adhesion, angiogenesis and proteinase expression. <sup>61</sup> Integrin signalling has been shown to be de-regulated in PC. Integrins expressed in PC cells play an important role in colonization of prostate tumour cells in the bone. PC cells express the same integrins; avb3 and a2b1 which are express by osteoclast. These factors are able to facilitate tumour spreading in the bone, by aiding in attachment and migration to the bone matrix, facilitating extracellular matrix remodelling and cancer cell growth. <sup>62</sup>
<b>Cadherins</b>	Cadherin-11 mediates homophilic cell adhesion in a calcium dependent manner. Its expression is associated with osteoblast differentiation and may function in cell sorting, migration, and alignment during the maturation of osteoblast. <sup>63</sup>  E-cadherin, is prominently associated with intracellular adhesion mechanisms for tumour invasiveness, metastatic dissemination. <sup>64</sup>
<b>Matrix metalloproteinases</b> MMP-2 MMP-9 MMP-13	MMPs are proteolytic extracellular matrix-degrading enzymes, which are produced by both the cancer cells and the stromal cells. They degrade ECM components including; collagens, laminins, fibronectin, and vitronectin, which can clear a path to facilitate cell migration and invasion. <sup>65</sup> The expression of MMPs is regulated by many growth factors, such as TGF $\beta$ which up-regulates MMPs expression and activity such as migration, invasion, angiogenesis activation and growth. <sup>66</sup>
<b>RANK</b>	RANK is the receptor expressed on the surface of osteoclast precursors and mature osteoclasts. The receptor becomes activated upon binding of RANKL, activating several downstream signalling pathways for differentiation, maturation and survival of osteoclasts in the process of bone resorption. <sup>67</sup>
<b>RANKL</b>	RANKL is expressed by osteoblasts and bone marrow stromal cells. RANKL binding to RANK leads to differentiation of osteoclast precursors as well as to activation and survival of mature osteoclasts. RANKL is produced in two forms; a cell surface and a soluble molecule. Furthermore, it has been established that PC cells secrete soluble RANKL, which can act to directly activate osteoclasts. <sup>68</sup>
<b>Osteoprotegerin</b>	OPG is a soluble decoy receptor for RANKL (ligand) and acts by binding RANKL, thus, preventing it from binding and activating RANK receptor on osteoclasts. Thus, it inhibits osteoclastogenesis; osteoclast maturation, activation and survival and bone resorption. <sup>69</sup>
<b>Parathyroid hormone related peptide</b>	PTHrP is expressed by PC cells. This induces RANKL expression through osteoblasts differentiation, which further stimulates osteoclast activation and formation, leading to bone destruction. <sup>70</sup> This activity protects PC cells and osteoblasts from apoptosis. <sup>71</sup>
<b>Chemokines</b> CXCR4 CXCL12/SDF-1	Chemokines are primarily known in the regulation of the motility (chemotaxis) of hematopoietic cells (immune system cells) and their ability to stimulate directional migration of nearly all classes of leukocytes during inflammation through the activation of a group of cell surface receptors. Chemokines are believed to mediate cell adhesion, migration, invasion and angiogenesis during the metastasis of tumour cells to bone. Bone contains chemotactic factors that attract PC cells towards the microenvironment. CXCR4 (expressed on PC cells) and stromal-derived factor-1 (SDF-1), are elevated in metastatic PC cell lines and in bone metastasis. <sup>72,73</sup>
<b>Transforming growth factor</b>	TGF is a widely expressed and abundant growth factor involved in the regulation of cellular proliferation, chemotaxis, differentiation, immune response, and angiogenesis. Production of TGF- $\beta$ by PC-associated stroma has been shown to increase the growth and invasiveness of prostate epithelial cells. Bone is one of the largest reservoirs of TGF- $\beta$ 1, which is released from the bone matrix during bone resorption following PC cell metastasis. <sup>74</sup>

(continued)

Table 1. (Continued)

FACTOR	FUNCTION
<b>Platelet-derived growth factor</b>	PDGFs are potent stimulator of cell proliferation, migration, survival, chemotaxis, angiogenesis and transformation. PDGF is known to play a major role in cell-cell communication for normal development and also during pathogenesis. <sup>75</sup> PDGF from tumour cells regulates commitment of stromal mesenchymal cells to differentiate into osteoprogenitor cells and induces proliferation and migration of osteoblast cells, suggesting a role for PDGF in bone formation. <sup>76</sup> PDGF also stimulates bone resorption by increasing the number of osteoclasts and up-regulating matrix degrading enzyme expression. <sup>77</sup>
<b>Epidermal growth factor</b>	EGF is over-activated in PC and cross-talk with the androgen pathway is linked to progression from androgen-responsive disease to castrate resistant phenotypes. EGFR signalling activates androgen receptor pathway even in the circumstances of androgen deprivation. <sup>78</sup> Furthermore, EGFR itself may be under the regulation of androgen signalling pathway. EGF stored within the bone matrix, is released in the bone resorption. EGF thus, plays an important role in regulating cellular growth, proliferation, survival and function of PC cells. <sup>78</sup>
<b>Fibroblast growth factor</b>	FGFs have a broad range of biologic activities that play an important role in tumorigenesis including promotion of proliferation, motility, and angiogenesis and inhibition of cell death. <sup>79</sup> FGFs play a key role in the cellular growth and maintenance of normal prostatic epithelium and are expressed in normal prostatic stroma. FGFs are expressed as autocrine growth factors by prostate cancer cells and can also be expressed in the tumour microenvironment as paracrine growth factors FGFs e released from the bone matrix as a result of PC induced bone resorption, thus leading to up-regulation of signalling activity. <sup>80</sup>
<b>Vascular endothelial growth factor</b>	VEGF is a critical mediator of angiogenesis—formation of new capillaries from existing blood vessels and vasculature. It plays an important role for tumour growth and metastasis by providing oxygen and nutrients to the proliferating tumour cells. Furthermore, it increases vascular permeability allowing cancer cells to move and migrate through the blood vessel circulation. <sup>81</sup> VEGF signalling is important for osteoblast and osteoclast activity and survival. Osteoblasts and osteoclasts also express VEGF receptors (VEGFRs), which are up-regulated and over-activated in PC, leading to dys-regulated bone destruction. <sup>82</sup>



**Figure 3. The cycle involving bone and tumor cell interactions in the bone microenvironment.** Matrix metalloproteinases (MMPs), chemokines (CXCL12/CXCR4), and vascular endothelial growth factor (VEGF) are involved in tumor cell attraction and targeting of bone as well as facilitating survival and metastasis within the bone microenvironment. Physical factors including hypoxia, acid pH, and high extracellular calcium levels influence this process. The tumor secretes several osteoblastic and osteoclastic factors in response to the bone microenvironment. Tumor derived osteoclast-stimulatory factors include PTHrP and IL-1, IL-6, IL-8, IL-11, TNF, M-CSF, and RANKL. Bone-derived pro-osteoblastic growth factors released from the resorption process include TGF- $\beta$ , IGFs, EGFs, and FGFs, which in turn upregulate BMPs, VEGF, and RANKL. Chemokines, MMPs and cathepsin are secreted by the tumor, which in turn stimulates bone cells to release factors which promote further tumor growth in bone. OPG is downregulated due to the over-stimulation of RANKL. The increased RANKL-to-OPG ratio leads to increased osteoclast formation, survival, and activity, which increases the rate of bone remodeling and turnover.



osteoblast-derived activation of osteoclasts, leading to bone destruction in skeletal metastasis in PC.<sup>88,89</sup>

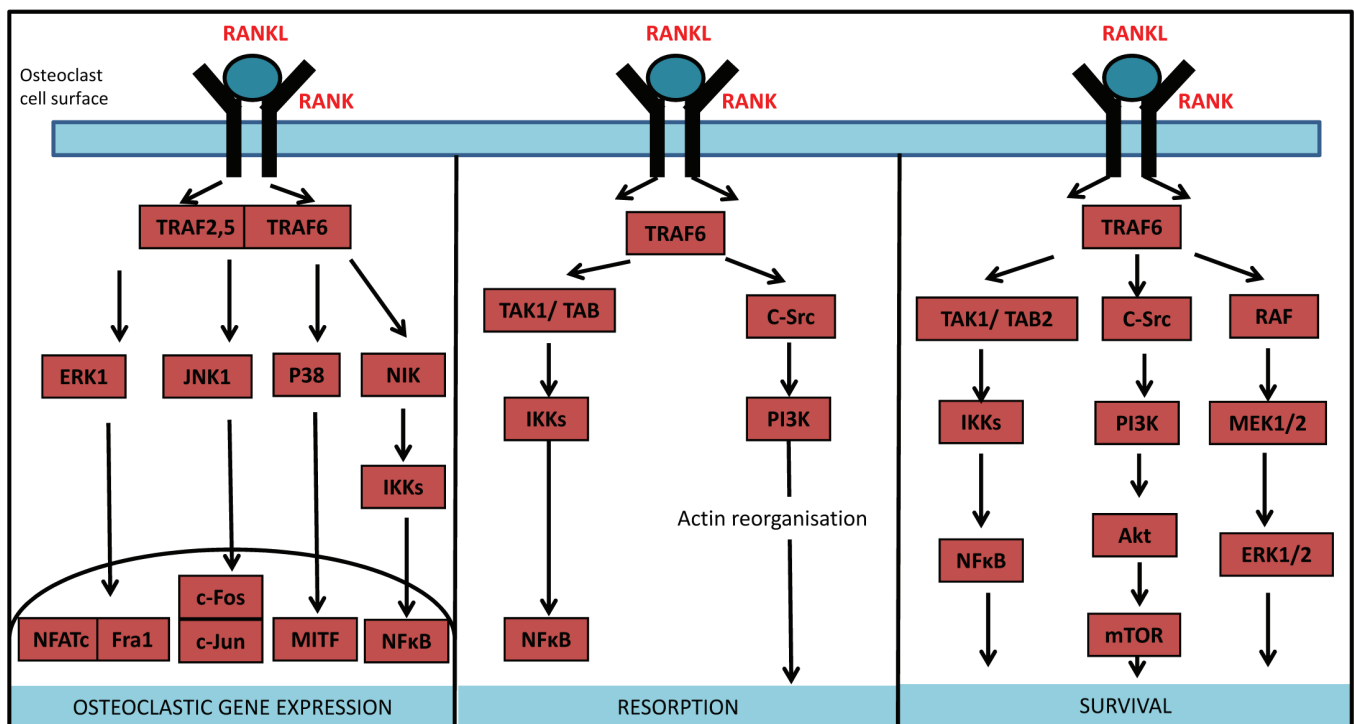
OPG is a naturally occurring protein that shares its homology with members of the TNF family.<sup>48,90</sup> Osteoblasts and stromal cells secrete OPG, which acts as a soluble decoy receptor and exerts its effects through the binding and subsequent sequestering of RANKL.<sup>91</sup> This prevents it from binding to RANK and activating downstream signaling events leading to osteoclast gene expression, differentiation, and survival, thus inhibiting bone destruction.<sup>28,67</sup> OPG and RANKL, therefore, share a synergistic relationship to maintain homeostasis.<sup>92</sup> While OPG has been shown to block the pathological increase in osteoclast activity and number, in cancer, RANKL expression is up regulated by stimulatory osteoclastic factors that are released from the tumor.<sup>67</sup> This leads to down-regulation of osteoblastic factors in the environment resulting in decreased OPG expression which causes unregulated and abnormal bone formation that ultimately leads to bone destruction.<sup>93</sup>

The RANK/RANKL/OPG system plays a critical role in osteoclastogenesis and bone destruction in PC.<sup>94</sup> The binding of osteoblasts causes RANKL to bind to the RANK receptor expressed on the cellular surface of osteoclast precursors leading to the commitment of the monocytic precursor to the osteoclastic lineage and the subsequent differentiation and activation of osteoclasts.<sup>95,96</sup> Following the activation of

the receptor (RANK), several downstream signaling cascades are initiated. These cascades control processes such as osteoclastogenic differentiation, cell survival and growth as well as actin reorganization, motility, and osteoclastic resorptive activity.<sup>5</sup> The NF- $\kappa$ B pathway initiates the transcription of a wide variety of genes encoding angiogenic factors, cytokines, cell adhesion molecules, and anti-apoptotic factors which are involved in tumor cell invasion, metastasis, survival, and chemo-resistance.<sup>86</sup> Due to the critical role of this pathway in PC, it provides an important target for therapeutic developments. The pathways in osteoclastogenesis through the RANK-RANKL-OPG triad system are illustrated in Figure 4.

### The Role of VEGF in Promoting Angiogenesis in PC Bone Metastasis

It is well established that tumors require a blood supply to establish, grow, and metastasize. Angiogenesis is the formation of new blood vessels from the existing vasculature within the growing tumor. It involves neovascularization of cancer cells through the division and migration of endothelial cells. This promotes the expansion, sustainable growth, progression, and metastasis of tumors to distant sites.<sup>97</sup> As tumor cells proliferate, the tumor mass expands beyond a capacity that can be supported by the existing vasculature. As a result, nutrient and oxygen levels fall, leading to an accumulation of metabolic waste. The tumor cells respond to this condition by secreting



**Figure 4. The process of osteoclastogenic differentiation through the RANK-RANKL-OPG system.** The binding of RANKL to RANK inhibits osteoclast apoptosis and promotes cell growth, proliferation, differentiation and survival, as well as cytoskeletal motility. TRAF-6 activation leads to downstream signaling of MAPKs involved in cell differentiation, proliferation, and survival in response to stress stimuli p38, JNK, ERKs, as well as NF $\kappa$ B complex. This leads to the activation of several downstream regulators of osteoclast formation and activation, including c-Fos, Fra-1, and NFATc1.





pro-angiogenic factors.<sup>98</sup> VEGF has been characterized as the most potent angiogenic factor. It functions in promoting endothelial cell migration and proliferation as well as increasing vascular permeability.<sup>99</sup>

The normal tumor environment is in a hypoxic state, which triggers VEGF expression through the hypoxia-inducible factor 1- $\alpha$  (HIF-1 $\alpha$ ) transcription complex as well as other growth factors such as TGF- $\beta$ , FGF, IGF, and PDGF.<sup>100</sup> These factors drive the switch from anti- to pro-angiogenic, providing a suitable blood supply for delivery of oxygen and nutrients to the PC cells and hence enabling them to proliferate.<sup>101</sup> The abnormal tumor vasculature increases permeability, facilitating tumor cell entry into the circulation as well as the production of tumor cell derived proteinases. These proteinases are involved in degradation in addition to enhancing invasion, ECM remodeling, adhesion, and proteolysis (through collagenases).<sup>102,103</sup> Hypoxia is the most potent inducer of VEGF production in bone.<sup>102</sup> Osteoblasts also express components of the HIF-1 $\alpha$  pathway, which acts to up-regulate VEGF expression in these cells.<sup>102</sup> This may be one mechanism that attracts the PC cells to bone, providing them with suitable means for establishing and proliferating at this site. Furthermore, angiogenesis is critical for bone formation by providing the vascularization required by bone-forming cells.<sup>54</sup> VEGF is expressed in osteoblasts and studies have shown that increased VEGF in osteoblasts is strongly correlated with osteogenesis (bone formation), stimulating chemotactic migration, and osteoblast differentiation.<sup>104</sup> Osteoblastogenic BMPs regulate VEGF expression and VEGF promotes osteoblastic activity.<sup>105</sup> VEGF has a direct role in osteoclastogenesis through promoting the activation and survival of osteoclasts with mature osteoclasts increasing expression of VEGF receptors.<sup>102</sup> VEGF also possesses the ability to up-regulate RANK on endothelial cells thereby increasing its sensitivity to RANKL and promoting angiogenesis.<sup>102</sup> As such, the up-regulated VEGF production in tumor cells and local bone cells may be responsible for close bidirectional attraction and interaction leading to excessive bone formation and active resorption, resulting in osteoblastic lesions and osteolytic destruction in PC.<sup>54</sup>

Angiogenesis inhibition via blocking the VEGF pathway has become an important therapeutic strategy.<sup>98</sup> Tumor cells as well as tumor cell lines have been reported to express both VEGF and VEGF receptors (VEGFRs).<sup>102</sup> Several antiangiogenic drugs have been introduced into the clinical setting including anti-VEGFA antibody, known as bevacizumab and multi-receptor tyrosine kinase inhibitors (RTKIs) such as, sorafenib and sunitinib.<sup>98</sup> VEGFR2 is a major endothelial cell signal mediator and a target for RTKIs.<sup>98</sup> The anti-VEGFR2 antibody has been shown to inhibit angiogenesis, reduce tumorigenicity and metastasis by inducing endothelial and tumor cell apoptosis, as well as inhibit MMP-9 production.<sup>106</sup> Recent studies have shown that suppression of PI3K/Akt/mTOR signaling using mTOR inhibitors reinstates sensitivity

of phosphatase and tensin homolog (PTEN) deficient cancer cells to tyrosine kinase inhibitor-mediated apoptosis. This may have therapeutic applications in the future.<sup>107</sup> The various growth factors implicated in PC bone metastasis and their current use or potential for therapeutic targets is summarized in Table 2.

### The Function of Growth Factors in PC Bone Metastasis

Several growth factors have been implicated in PC bone metastasis. The mineralized bone matrix contains a wide range of growth factors, where IGF-1 is one of the most abundant.<sup>122</sup> IGF-1 is involved in regulating cell differentiation, proliferation, migration, and apoptosis. Mitogenic effects of IGFs are mediated through binding with IGF-1 receptor and activation of the PI-3 kinase/Akt signaling pathway. Once metastasis has been established in the bone, osteoclastic bone resorption is activated. Growth factors such as IGF-1 and TGF- $\beta$  are released into the bone marrow cavity where they interact with and influence metastatic tumor cells.<sup>122</sup> The IGF-1R is an RTK that, upon activation by IGF-1, shows mitogenic and anti-apoptotic effects.<sup>122</sup> Increased IGF-1R signaling results in activation of growth-promoting intracellular signaling pathways, including the ras-raf-MAPK (mitogen-activated protein kinase) and PI3K cascades.<sup>123</sup> IGFs are proposed to bypass inhibition by stimulation of autocrine androgen synthesis, survival signaling, and enhancement of androgen receptor nuclear localization by stabilizing microtubules. IGF-1 is crucial for osteoblast differentiation and found abundantly in the bone microenvironment.<sup>124</sup>

BMPs are members of the TGF- $\beta$  superfamily and are potent inducers of osteoblast differentiation.<sup>125</sup> BMPs initiate new bone formation by recruiting progenitor stem cells and initiating their growth and differentiation into bone. They are also involved in stimulating cancer cell migration.<sup>126</sup> In addition, BMP mRNA and protein expression has been found in human PC cell lines from bone metastasis specimens.<sup>54</sup> Furthermore, receptors have been expressed in the cancer cell lines and tissue.<sup>54</sup> In vivo, inhibition of BMP activity by noggin inhibits the development of osteoblastic lesions and reduces the osteoblastic component in mixed lytic/blastic lesions.<sup>54</sup> Osteoblast-derived BMP-2 can activate the Akt, MAPK, and ERK pathways, which in turn induce IKK $\alpha$ /b phosphorylation and NF- $\kappa$ B activation, resulting in the activation of  $\beta$ 1 and  $\beta$ 3 integrins and contributing to the migration of PC cells.<sup>125</sup> BMP signaling also activates the intracellular receptor type I kinase, followed by phosphorylation of Smad, which translocates to the nucleus and induces the expression of genes important for bone formation.<sup>125</sup> BMPs are implicated in homing, migration and invasion of PC cells to bone that ultimately leads to promotion and formation of osteoblastic lesions.<sup>127</sup>

TGF- $\beta$ 1 is a growth factor that regulates cell proliferation, differentiation, chemotaxis, immune response, and angiogenesis.<sup>74</sup> Production of TGF- $\beta$  by PC-associated

**Table 2.** Growth factors implicated in Prostate Cancer bone metastasis. Role in tumour stimulation and proliferation and current and potential therapeutic targets.

GROWTH FACTOR	RECEPTOR	EXPERIMENTS	FINDINGS	DRUG AVAILABILITY	REF
<b>Insulin-like growth factor</b>	IGF-1R	PC xenograft models <i>In vitro</i> <i>In vivo</i> studies	Involved in anti-apoptosis, cell differentiation and proliferation, transformation and angiogenesis (bone metastasis). Upregulates expression of both RANK/RANKL stimulates differentiation of osteoclast and represses OPG.	CIXTUMUMAB(IMC-A12): IGF-1R inhibitor-fully human monoclonal antibody (Phase II)	108 109
IGF-I & IGF-II		Clinical studies: Phase I/II/III	Reduces IGF-1R activation by preventing IGF-I and IGF-II interaction with its receptor. Promotes receptor internalization and degradation.	GANITUMAB(AMG 479): IGF-1R inhibitor- fully human antibody (IgG1)	110 111
<b>Epidermal growth factor</b>	EGFR	Transwell or Boyden chamber Dunn chamber 3D invasion <i>In vivo</i> invasion	Modulation of cell proliferation and survival for tumorigenesis. Inhibition of downstream signalling pathways.	GEFITINIB & ERLOTINIB: Oral anilinoquinazoline compound that blocks EGFR tyrosine kinase activity (Phase I/II with radiation)	113 114
FGF				TRASTUZUMAB – Phase I LAPATINIB – Phase II CETUXIMAB –Phase Ia/IIb PANITUMAB – Phase 1	115 116
<b>Fibroblast growth factor</b>	FGFR1-4	Transwell or Boyden chamber Wound healing assay	Initiation of growth, vascularization, and progression of tumours through proliferation and evasion of cell death. Enhance angiogenesis through paracrine action on endothelial and other stromal cells.  bFGF is highly angiogenic and regulates protease expression, e.g. urokinase-type plasminogen activator, collagenase, which may be involved in metastatic cascade.	AZ8010: ATP-competitive FGFR tyrosine kinase inhibitor  PEG-interferon alfa-2b BGJ398 E-3810 TSU-68 Dovitinib lactate (TKI258) BIBF 1120	80
FGF8 & FGF9					
bFGF					
FGF-2					
<b>Platelet derived growth factor</b> (PDGF-D)	Cognate α-PDGFR β-PDGFR	<i>In vivo</i> studies  Clinical studies: Phase I & II	Promotes tumorigenesis, supports stromal cell recruitment and malignant transformation.  Proliferation of connective tissue, monocyte/macrophage and smooth muscle cell chemotaxis, angiogenesis.	CEDIRANIB (AZD2171) IMATINIB Tyrosine kinase inhibitor CP-673,451: PDGFR-β Inhibitor Anti-angiogenesis, growth inhibition	117
<b>Vascular endothelial growth factor</b> (VEGF)	VEGFR-1 VEGFR-3  VEGFR-2	Transwell or Boyden chamber 3D culture Orthotopic PC xenograft	Induction of endothelial cell apoptosis and reduction of endothelial cell MMP-9 production	BEVACIZUMAB: Approved SORAFENIB: Approved  Anti-angiogenic	108
<b>Transforming growth factor</b> (TGF-β)	TGF-βR1 TGF-βR2	Transwell or Boyden chamber Dunn chamber Wound healing assay	Sensitivity to TGF-B inhibition is lost with tumour progression. Causes osteoblast and osteoclast recruitment, differentiation, bone matrix production (ECM), cell growth angiogenesis, immunosuppression.	AXITINIB (AG-013736) VATALANIB (PTK787)	106
<b>Bone morphogenetic proteins</b> BMP-2 BMP-6	BMPR-I BMPR-II	<i>In vitro</i> studies <i>In vivo</i> studies	Proliferation, apoptosis, invasion differentiation and migration of cellular targets. Acts in conjunction with RANKL to form mixed osteolytic/blastoc lesions.	NICOTUZUMAB : Inhibits TGF-B activity leading to tumour regression.  AP 12009, GC1008, Lucanix  Targets: BMP-7, 9, 15 Tumour suppression through increased Noggin activity which prevents osteoblastic/osteoclastic activity.	111 118 119 120 121



stroma increases the growth and invasiveness of prostate epithelial cells.<sup>74</sup> TGF- $\beta$  has been shown to promote osteoblastic bone metastases in experimental models.<sup>128</sup> Bone is an abundant reservoir for TGF- $\beta$ 1, which is released from the bone matrix during bone remodeling following PC cell migration and establishment.<sup>74</sup> TGF- $\beta$  is activated by cleavage of its precursor by osteoclast-derived or PC secreted proteases. TGF- $\beta$  signaling occurs through a transmembrane receptor serine–threonine complex which consists of type I and II receptor kinases. The binding of TGF- $\beta$ 1 to the type II receptor leads to the formation of a heterodimeric complex with the type I receptor, resulting in phosphorylation.<sup>74,128</sup> The receptor-associated Smads are subsequently recruited to the activated receptor complex. The phosphorylated regulatory Smad2/3 interact with the co-Smad, Smad4. This complex then translocates into the nucleus, binding to specific DNA sequences, consequently recruiting co-activators or co-repressors to regulate the transcription of TGF- $\beta$  target genes.<sup>128</sup> Activated TGF- $\beta$ 1 also induces activation of SMAD-independent pathways such as PI3K, AKT, and MAPK. Thus, TGF- $\beta$  is a potential therapeutic target in advanced PC using TGF- $\beta$  receptor type I (TGF- $\beta$ RI) kinase inhibitors.<sup>2</sup>

Communication between epithelial and stromal compartments via FGF family signaling pathways have important roles in homeostasis of the normal prostate.<sup>129</sup> Upon binding of FGF to its receptor, FGFR dimerization is induced leading to phosphorylation and activation of various downstream signaling pathways including MAPK-PI3K/AKT, phospholipase-C (PLC), and signal transducer and activator of transcription (STATs).<sup>80</sup> FGFs are expressed as autocrine growth factors by PC cells, however they can also be expressed in the tumor microenvironment as paracrine growth factors.<sup>80</sup> They promote cancer progression by increasing proliferation and preventing cell death. FGFs are well known angiogenic factors stimulating angiogenesis through paracrine actions on endothelial and other stromal cells in the tumor microenvironment. FGF signaling is therefore a promising therapeutic target in aggressive PC. Several FGFR small-molecule inhibitors have entered clinical trials with the action of inhibiting multiple tyrosine kinases.<sup>80</sup>

De-regulation of EGF receptor (EGFR) is often associated with carcinogenesis which can be caused by its over-expression, gene amplification, mutations, or deletions. Increased expressions of EGFR and its ligands (EGF and TGF- $\beta$ ) have been described in prostate tumors, and autocrine activation of EGFR signaling regulates the growth of androgen-independent PC.<sup>12</sup> EGFRs belong to the human epidermal receptor (HER) axis, which binds the EGF, activating downstream signaling events through PI3 kinase and MAPK.<sup>12</sup> This activates the Akt family of kinases and STATs, resulting in downstream events that regulate cellular proliferation, growth, survival, and migration.<sup>78</sup> Normanno and Gullick demonstrated that factors released under

the control of EGFR signaling modulate bone remodeling through the induction of RANKL in bone marrow stromal cells, thus activating osteoclasts.<sup>130</sup> EGFR signaling also regulates the ability of MSCs to release growth factors that promote neo-angiogenesis and tumor cell migration.<sup>12</sup> A number of anti-EGFR drugs are currently awaiting Food and Drug Administration approval or are under evaluation in clinical trials.

### The Role of PTHrP in PC Bone Metastasis

PTHrP was initially identified as a product associated with humoral hypercalcemia of malignancy resulting from human tumors.<sup>131</sup> It binds to the type 1 PTH receptor (PTH1R) and functions through endocrine, autocrine, paracrine, and intracrine actions to modulate development, growth, and differentiation of normal cells as well as inducing metastasis of malignant cells via cytokine interaction.<sup>132</sup> Studies of the downstream signaling events of PTH-1R have focused mainly on the cAMP-dependent protein kinase A (PKA) and protein kinase C (PKC) pathways. At the cellular level, PKA and MAPK-ERK (extracellular signal-regulated kinase) pathways govern the majority of the effects induced by PTH and PTHrP on osteoblasts. The MAPK cascade is also called the ERK cascade and found to be involved in cell adhesion, proliferation, differentiation, viability, and apoptosis.<sup>133</sup> Activation of the MAPK cascade involves phosphorylation of MAPKKK (Raf-1), MAPKK, and MAPK. Active MAPK subsequently phosphorylates transcription factors, other downstream kinases, and substrates leading to biologic activity. PTHrP regulates MAPK<sup>133</sup> and is critical in the regulation of cellular functions including migration, angiogenesis, survival, and calcium release and transport in osteoclasts.<sup>134</sup>

Recently, PTHrP has been found to be important in androgen dependent resistance of PC tumor cells to apoptosis as well as tumor proliferation and progression.<sup>135</sup> Over-expression of PTHrP is involved in the promotion of metastasis to bone, especially the initial osteolysis and subsequent osteoblastic phase.<sup>136</sup> PTHrP enhances bone remodeling and the release of several biological factors such as VEGF, ILs, and Endothelin-1 as well as various growth factors including IGFs, FGFs, TGF- $\beta$  and PGDF, providing a fertile environment for tumor growth.<sup>52</sup> PTHrP also induces osteoblast differentiation and protects PC cells and osteoblasts from apoptosis. It participates in the indirect activation of osteoclasts through stimulation of osteoblastic RANKL production and decreasing OPG leading to the activation of osteoclastogenesis.<sup>48,132</sup> This leads to the process of bone destruction in osteolytic metastasis whereby the bone matrix releases stored immobilizing growth factors, such as TGF- $\beta$ , which is activated and released as a result of osteoclast activated resorption activity.<sup>28</sup> The release of growth factors from the matrix further stimulate the cancer cells to up-regulate production of PTHrP.<sup>137</sup>



## The Key Role of MMPs and Chemokines in PC Bone Metastasis

Tumor metastasis involves the interactions between invading cancer cells and the surrounding stromal tissue. Such interactions promote degradation of the ECM via specialized proteolytic enzymes, which are produced by both the cancer cells and the stromal cells.<sup>138</sup> MMPs are potent zinc and calcium dependent proteolytic enzymes that play a vital role in this process of degradation and digestion of structural components of the ECM. This is an essential step for tumor invasion, migration, metastatic progression, and tumorigenesis.<sup>139-141</sup> Moreover, MMPs are involved in the cleavage and release of growth factors, cell surface receptors, cell adhesion molecules, and chemokines from the matrix as a result of degradation, leading to enhanced tumor growth and establishment of a more aggressive phenotype due to acquired resistance.<sup>142,143</sup> In particular, MMP-7 is involved in the cleavage of RANKL from the osteoblast cell surface, forming a soluble form of RANKL. Among the other MMPs, MMP-2 (gelatinase A) and MMP-9 (gelatinase B), referred to as type IV collagenases or gelatinases, are specifically associated with PC metastasis.<sup>138</sup> MMP-2 and MMP-9 induce tumor angiogenesis,<sup>138</sup> with MMP-9 found to play a vital role in tumor induced osteoclast recruitment.<sup>142</sup> Dong et al demonstrated that MMP-9 activity was originating from newly recruited osteoclasts during the early colonization of the bone marrow spaces by tumor cells.<sup>142,144</sup> MT1-MMP is a membrane-anchored protease capable of activating pro-MMP-2 on the cell surface, as well as promoting tumor angiogenesis and growth. Additionally, it degrades several components of the ECM, including type 1 collagen, which is the most abundant protein in bone. It has been proposed that MT1-MMP is able to function as enzymes that shed and release non-ECM substrates, particularly RANKL, from the tumor cells, which activates osteoclastic bone degradation.<sup>144,145</sup> As such, MMPs are potential therapeutic targets against cancer using endogenous and synthetic tissue inhibitors of metalloproteinase (TIMPs).<sup>146</sup>

Evidence from bone xenograft models demonstrate that PC cells injected adjacent to the bone migrate toward bone, suggesting that there is chemo-attraction between the tumor and the bone microenvironment. Chemokines interact with cell surface receptors promoting oncogenic and cellular transformation by acting as growth factors that increase proliferation and tumor angiogenesis.<sup>72</sup> CXCL12 (also known as SDF-1) from bone stromal cell binds to CXCR4 receptor and activates downstream signaling through the PI3K/Akt and MAPK/MEK pathways. CXCR4 is widely expressed in the cellular environment and interacts only with the SDF-1 ligand. It is involved in the immune response, protecting the tumor from the host response as well as activating key survival pathways such as anti-apoptosis, leading to growth and proliferation of tumor cells.<sup>147</sup> CXCR4 has been implicated in the “homing”-directed

dissemination of circulating cancer cells to microenvironments of high chemokine concentration such as lymph nodes and bone marrow.<sup>148</sup> During this process, the circulating cancer cells mimic hematopoietic and immune cells in terms of localizing to high CXCL12-expressing sites, firm adhesion to endothelial cells, transmigration across the blood vessel wall, and migration towards the chemokine source.<sup>148</sup> The binding of CXCL12 from the bone microenvironment to CXCR4 expressed on the surface of PC cells activates cell signaling pathways and leads to the expression and secretion of proteases.<sup>148</sup> SDF-1 is thought to recruit osteoclast precursors expressing CXCR4 to induce MMPs (2,9,14), which promote proteolytic breakdown of the ECM. In addition, it triggers the angiogenic switch through up regulation of VEGF.<sup>149,150</sup> The SDF1a/CXCR4 signaling pathway plays a major role in migration by promoting adhesion, invasion, proliferation, and cancer growth in bone.<sup>34,39</sup> As such, receptor antagonists present potential targets to reduce growth and proliferation of tumors.<sup>151</sup>

A major characteristic of bone-homing PC cells is their capacity to release copious levels of IL-6, which aids in facilitating bone invasion and growth of metastatic lesions.<sup>152</sup> Additionally, osteoblasts also express IL-6 receptors. As such, IL-6 is a major pleiotropic, pro-inflammatory cytokine playing a role in immune response, cell differentiation, hematopoiesis, wound repair, and bone remodeling.<sup>152</sup> It has an effect on cell proliferation, apoptosis, and angiogenesis through activation of Janus kinase (Jak), STAT factor 3, phosphatidylinositol-3-kinase MAP/k, and PI3-K-Akt.<sup>59</sup> IL-6 also stimulates osteoclastic bone resorption by inducing RANKL expression in osteoblastic cells thereby affecting the growth of PC cells in a paracrine and autocrine manner.<sup>153</sup> IL-6 activates the androgen receptor in a ligand-independent manner and contributes to PC progression in castrated patients. Additionally, IL-6 may protect PC cells from cell death induced by certain chemotherapeutic agents through activation of STAT3 and anti-apoptotic proteins such as bclXL. Elevated IL-6 levels in patients with castration-resistant prostate cancer (CRPC) are independently associated with decreased survival.<sup>153</sup> Therapies that can counteract the up-regulated IL-6 activity on tumor growth and survival may benefit patients with advanced PC. The various chemokines and cytokines that are currently being investigated in PC bone metastasis for potential therapeutic targets are summarized in Table 3.

## Emerging Therapies for PC Spinal Metastasis

Skeletal metastases as a consequence of advanced PC often cause severe and debilitating symptoms for patients that are refractory to current treatments thereby highlighting the need for more effective and novel therapies. Denosumab (XGEVA, Amgen) is a monoclonal human antibody which binds to and neutralizes RANKL, preventing its binding to the RANK receptor. Administered subcutaneously, it inhibits the activation and activity of osteoclasts and has

**Table 3.** Chemokines and cytokines implicated in Prostate Cancer bone metastasis. Role in tumour stimulation and proliferation as well as current and potential for therapeutic targets.

CHEMOKINE	RECEPTOR	EXPERIMENTS	FINDINGS	DRUG AVAILABILITY	REF
<b>Stromal derived factor-1</b>	CXCR-4	Transwell assay (Boyden chamber)	Involved in migration, MMP expression growth, survival, invasion and angiogenesis.	PLERIXAFOR (AMD3100): Receptor antagonists, that block binding pocket of CXCR4.	154
	CXCR-7	<i>In vitro</i> studies	Enhanced proliferation and recruitment of immune cells that promote tumour growth	Inhibition of CXCR4 sensitises PC to chemotherapy	155
CXCL-12		<i>In vivo</i> studies			108
CXCL-11		-Invasion assay -Recruitment of bone marrow-derived cells		MDX-1338, BTK140, CTCE-9908: CXCL12 analogues	
		[Phase I/II clinical studies]		Inhibition of tumour growth Inhibition of VEGF and angiogenesis	
CCL2	CCR-2	[Phase II clinical studies]	Promote the migration of monocytes and macrophages to sites of inflammation.	CARLUMAB(CNTO888): anti-human monoclonal antibody	108
CCL22	CCR-4	Transwell assay (Boyden chamber)	Involved in migration of cancer cells. Recruitment of Natural Killer cells and other immune cells (T cells).	MLN1202: CCR2 blocker MOGAMULIZUMAB KW-0761): Human monoclonal antibody targeting CCR4	156
CX3CL1 fractalkine	CX3CL1-R	<i>In vivo</i> studies	Mediates adhesion of PC cells.	None	157
CXCL-1	CXCR-1	Transwell assay (Boyden chamber)	Induces proliferation and growth of PC cells through chemotaxis.	None	158
CXCL-5	CXCR-2	Wound-healing assay <i>In vivo</i> studies (Nude mice)		None	159
CXCL-6		[Phase I clinical studies]		Anti-CXCL8 antibodies (ABX-IL8): Inhibit angiogenesis, MMP production and reduces tumour growth	160
CXCL-8					
<b>Macrophage colony stimulating factor</b>	CSF-1R	Transwell assay (Boyden chamber) Dunn chamber 3D invasion <i>In vivo</i> invasion	Induces systemic immune responses (TNF and ILs).	ARRY-382: csf-1 inhibitor Prevents osteoclast (macrophage) differentiation and inhibit tumour cell proliferation	108
		[Phase I/II clinical studies]	Regulates the survival, proliferation, differentiation and function of cells in the monocyte lineage including macrophages and osteoclasts.	PLX3397: Inhibition of PC tumour growth	161
<b>M-CSF</b>					
<b>Interleukins</b>	IL-1B	<i>In vivo</i> studies (Murine models)	Involved in immune response.	SILTUXIMAB (CNTO 328): Chimeric Human monoclonal antibody against IL-6	162
	IL-6	<i>In vitro</i> studies	Potentiated by hypoxic environments and chemically induced stresses including exposure to chemotherapy agents. Induce tumour cell proliferation and through growth factor stimulation, pro-angiogenic and anti-apoptotic effects.		153
IL-8 (CXCL-8)	IL-6R (CD 126)	Xenograft models			163
IL-11	CXCR-1 CXCR-2	[Phase I/II clinical studies]			164
	IL-11R				

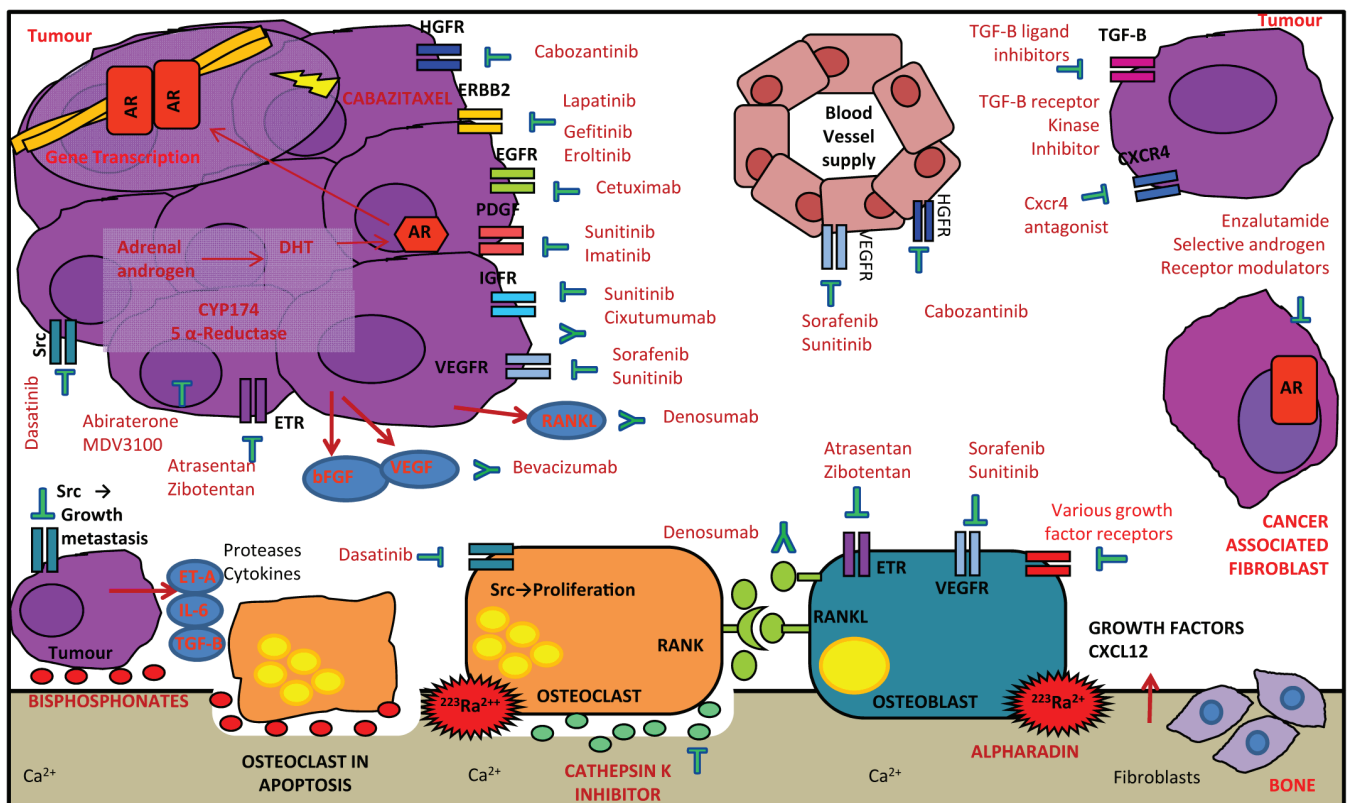


been found to be clinically effective in inhibiting subsequent bone resorption.<sup>165,166</sup> Zoledronic acid (Zometa, Novartis) is a widely used bisphosphonate for bone metastases, acting to decrease fracture risk via inhibition of the catalytic action of farnesyl pyrophosphate synthase. This leads to a reduction in osteoclastic activity and induction of apoptosis, inhibiting bone resorption as well as inhibiting tumor cell adhesion to bone.<sup>147,167</sup> The structural similarity of bisphosphonates to inorganic phosphate assists them in incorporating into bone and binding to hydroxyapatite. Once inside the bone, they act to decrease bone resorption by decreasing the availability of hydroxyapatite crystals and osteoclast-mediated resorption,<sup>70</sup> and inhibiting recruitment, differentiation, attachment and survival of osteoclasts.<sup>70</sup> Furthermore, they act on osteoblasts to indirectly inhibit osteoclast differentiation and activation<sup>168</sup> and inhibit RANKL expression in PC cells, diminishing osteoclast activity even more.<sup>70</sup>

An international phase III double-blind study comparison conducted between denosumab and zoledronic acid in PC patients reported that denosumab was superior in delaying metastatic skeletal complications by an average of more than 3 months.<sup>34</sup> Additionally, bone turnover markers were highly suppressed with denosumab treatment. However, the results of this study concluded that there were no differences

in overall survival time between patient treatments.<sup>169</sup> Another study tested the effectiveness of denosumab in prolonging bone metastasis-free survival in men with advanced CRPC. These results revealed a risk reduction of 15% and an extension of metastasis free-survival of 4.2 months. In addition, risk of symptomatic bone-metastases was 33% lower in the denosumab group.<sup>87</sup> Several systemic agents that possess selectivity for anti-neoplastic effects on bone metastases are currently being tested for palliative efficacy and effect on prolonging survival in patients with advanced PC. These therapies include endothelin receptor antagonists (atrasentan and zibotentan), proto-oncogene Src (c-Src tyrosine kinase) inhibitors, and radiopharmaceuticals (strontium-89 and samarium-153).<sup>87</sup> More recently, alpharadin (radium-223), an alpha radiation-emitting agent, has been of interest due to its ability to target bone metastasis, which is attributable to its similar chemical and physical properties to calcium.<sup>170</sup> The various therapeutic targets currently employed for the treatment of PC bone metastasis are illustrated in Figure 5.

Endothelins and their receptors are emerging as potential targets for PC bone metastasis. The unregulated endothelin pathway in PC plays a vital role in the proliferation, growth, and invasion via up-regulation of tumor proteases



**Figure 5. Therapeutic targets for the various mediators involved in PC bone metastasis.** Potential targets for PC skeletal metastases include the cancer cells themselves, the tumoral blood supply, cancer-associated fibroblasts, osteoclasts and osteoblasts. Important PC pathways may be blocked by neutralizing antibodies, receptor antagonists and inhibitors.



(MMPs) as well as urokinase-type plasminogen activators, inhibition of apoptosis, and promotion of VEGF-induced angiogenesis in tumors.<sup>56</sup> Furthermore, endothelins promote abnormal osteogenesis in the bone microenvironment, proliferation of osteoblasts, bone remodeling, and release of growth factors that stimulate survival of cancer cells.<sup>56,171,172</sup> Endothelin-1 (ET-1) is a potent vasoconstrictor. Activation of the endothelin receptor A (ET<sub>A</sub>) can lead to induction of a survival pathway, whereas activation of the endothelin receptor B (ET<sub>B</sub>) results in clearance of circulating ET-1 as well as stimulation of apoptosis.<sup>173</sup> In PC, ET-1 activates the ET<sub>A</sub> receptor, mediating the signaling cascade which promotes tumor survival, growth, angiogenesis, and invasion, thus inhibiting apoptosis.<sup>173</sup> Once ET-1 binds to and activates ET<sub>A</sub>, interactions and activation of a G-protein triggers the parallel activation of several signal-transduction pathways. This includes phospholipase C activity, which causes an increase in intracellular Ca<sup>2+</sup> levels, protein kinase C, EGF-R, PI3K, and ras/raf/MAPK pathways.<sup>56</sup> This cascade of events ultimately induces nuclear

transcription of several proto-oncogenes, including c-myc, c-fos, and c-jun, which possess the capability of influencing cell growth and proliferation.<sup>56</sup> A selective ET<sub>A</sub> antagonist known as Atrasentan has been the focus of pre-clinical trials where it has been shown to prevent osteoblastic lesions in mouse models.<sup>56</sup> Zibotentan is an oral ET<sub>A</sub> specific antagonist shown to inhibit cell invasion, proliferation, and metastasis.<sup>57</sup> A phase II trial of patients with CRPC bone metastasis showed improvements in overall survival.<sup>57</sup> The current and potential therapeutic targets for PC bone metastasis are discussed in Table 4.

Better understanding of the underlying molecular mechanisms contributing to and which drive metastasis of PC cells to secondary skeletal sites, particularly the spine, is critical in order to prevent or improve the devastating complications related to pathological fracture and metastatic epidural spinal cord compression. Targeting critical pathways involved in pathological bone turnover may inhibit these skeletal complications and thus improve pain, function, and quality of life in patients with advanced metastatic PC.

**Table 4.** The current treatments and therapies for bone metastasis in Prostate Cancer.

INTERVENTION	MECHANISM OF ACTION	EVIDENCE
<p><b>Bisphosphonates</b></p> <p>(Zoledronic acid: Zometa, Novartis Pharmaceuticals)</p> <p>Zoledronic acid has been widely utilized for the prevention of skeletal-related events (SREs) in patients with bone metastases who have undergone hormonal therapy.</p>	<p>Bisphosphonates inhibit osteoclast activity in PC bone metastasis. The suggested MOA is inhibition of tumour cell adhesion and invasion of the extracellular bone matrix and/or antiangiogenic effects.<sup>174</sup></p> <p>Bisphosphonates are pyrophosphates which have two phosphonate groups, and bind with high affinity to calcium which is abundantly found in the bone. Once ingested by osteoclasts, bisphosphonates induce apoptosis in these cells and thus prevent further bone loss.<sup>70,168</sup></p> <p>ZA contains nitrogen in its structure, which is effective against a variety of cancers in both osteolytic and osteoblastic lesions.<sup>175</sup></p>	<p>In a Phase III trial, ZA significantly reduced the incidence of SRE by 36% and delayed the first SRE by more than 5 months compared with the placebo.<sup>174</sup></p>
<p><b>Denosumab</b></p> <p>(PROLIA, amgen inc. Xgeva, amgen inc.)</p> <p>Fda approved in 2011 as a treatment to increase bone mass in patients at high risk for fracture receiving androgen deprivation therapy (ADT), reduced the incidence of vertebral fracture</p>	<p>Denosumab is a monoclonal antibody that binds to RANKL, a protein involved in the formation, function, and survival of osteoclasts, causing inhibition of osteoclastic bone resorption.</p>	<p>Showed a superior effect compared to zoledronic acid in the prevention of SREs (e.g., fracture, spinal cord compression and radiation or surgery to bone) in a large Phase III study.<sup>176,177</sup></p>
<p><b>Radionucleotides</b></p> <p>Xofigo (Radium: Ra 223 dichloride)</p> <p>FDA approved in 2013 to treat men with symptomatic late-stage (metastatic) castration-resistant prostate cancer, that has spread to bones.</p>	<p>Radium 223 is an alpha-emitter that releases a large helium nucleus, causing more biologic damage but over a much shorter path length and with the possibility of killing tumour cells and reducing tumour burden.<sup>178</sup></p> <p>Radionuclides residing in family IIA of the periodic table carry the same divalent charge as elemental calcium and are incorporated into bone matrix directly. Others are chelated to organic phosphates, which are incorporated into the matrix.<sup>178</sup></p> <p>Other approved radionucleotides: Strontium-89 Samarium-153 Phosphorus-32</p>	<p>A Phase II clinical trial of patients with symptomatic, hormone-refractory PC, showed an improvement in survival, PSA levels, and alkaline phosphatase levels compared with placebo.</p> <p>A median overall survival of 14 vs. 11.2 months was reported. The time to a SRE was also significantly longer for patients (13.6 vs. 8.4 months; P = .00046).</p> <p>The time to disease progression based on PSA and alkaline phosphatase levels was significantly superior.<sup>178</sup></p>

(continued)



Table 4. (Continued)

INTERVENTION	MECHANISM OF ACTION	EVIDENCE
<b>SRC kinase inhibition</b> Dasatinib (SPRYCEL, Bristol-Myers Squibb) Saracatinib (AZD0530) Bosutinib (SKI-606, Wyeth)	Src is active or overexpressed during prostate tumor growth and metastasis. Src signaling is required during osteoclast maturation and activation and plays a role in the formation and maintenance of PC bone metastases. <sup>179</sup> Intracellular tyrosine kinase inhibitors transduce signals from a range of upstream proteins, including receptors for EGF, PDGF, VEGF that promote several mechanism involved in metastasis, like cell proliferation and survival, cell adhesion, migration, invasion, and dissemination to distant organs. <sup>179</sup> Dasatinib is one of the several inhibitors It has been shown to suppress markers of bone turnover-decrease proliferation of immature osteoblasts while enhancing their differentiation. <sup>179</sup>	In chemotherapy-naïve patients with metastatic CRPC, 20/41 had a 35% decrease in uNTX compared with baseline. In addition, 21/42 had a decrease in BAP. Furthermore, among patients who underwent bone scans, 11/22 were stable at 12 weeks and 3/9 were stable at 24 weeks. Prolonged PSA doubling time in 32/39 was observed. <sup>180</sup>
<b>Angiogenesis inhibitors</b> (Avastin, Roche/Genentech)  Sunitinib (Sutent, Pfizer)  Sorafenib (Nexavar, Bayer)  Cabozantinib (XL184, Exelixis)  AVE0005, Sanofi-Aventis  Tasquinimod (Active Bech)	Bevacizumab is a humanized monoclonal antibody that neutralizes VEGF-A activity, leading to the suppression of cell proliferation, angiogenesis and invasions in the bone. <sup>107</sup>  Small molecule tyrosine kinase inhibitors (TKIs) with activity against VEGFR, amongst other pro-angiogenic targets, have shown activity in a number of tumor types. Sunitinib, an oral tyrosine kinase inhibitor (TKI) of VEGFR and PDGFR.  Sorafenib multi-targeted Tyrosine Kinase Inhibitor.  Cabozantinib is a small molecule inhibitor of MET and VEGFR2 shown to suppress metastasis, angiogenesis and tumour growth in preclinical models.  Aflibercept, also known as VEGF-Trap, is a protein composed of the extracellular domains of VEGFR-1 and -2 fused with the constant region (Fc) of the human IgG1 antibody. Aflibercept acts as a decoy receptor, preventing VEGF from binding to VEGFRs. <sup>107</sup>  Tasquinimod is an oral agent with anti-angiogenic and potential anti-neoplastic activities, and has been shown to decrease blood vessel density. <sup>70</sup>	Bevacizumab monotherapy did not show significant activity in CRPC, however the combination of bevacizumab plus docetaxel and estramustine chemotherapy resulted in a 50% PSA decline in 75% of patients and a partial radiological response in 23 of 39 (59%) of patients with measurable disease in a phase II study. <sup>107</sup> Phase II studies investigating sunitinib monotherapy showed evidence of radiological responses in patients with mCRPC in the absence of PSA decline. <sup>107</sup> Several phase II studies investigating sorafenib monotherapy in mCRPC have demonstrated modest activity with some discordance between PSA and radiological responses Cabozantinib has shown a significant survival advantage in a phase III study treating patients with medullary thyroid cancer. A phase I study investigated the combination of Aflibercept and docetaxel in patients with advanced solid tumour including PC. Preliminary evidence of anti-tumor activity was observed in multiple tumour types. <sup>107</sup>  In a randomized phase II trial in chemotherapy-naïve patients with mCRPC, tasquinimod significantly improved progression-free survival compared to placebo (7.6 months vs. 3.3 months, p = 0.0042). <sup>107</sup>
<b>Abiraterone acetate</b> (Zytiga Tablets, Janssen Biotech, Inc.)  FDA approved in 2012, in combination with prednisone for the treatment of patients with metastatic castration-resistant prostate cancer.	Abiraterone acetate is a potent selective inhibitor of CYP17-hydroxylase and C17,20-lyase, enzymes necessary for the synthesis of androgens from steroid precursors. <sup>181</sup> It catalyzes the sequential reactions of the conversion of pregnenolone and progesterone to their 17- $\alpha$ -hydroxy derivatives and the subsequent formation of DHEA and androstenedione. As DHEA and androstenedione are androgens and precursors of testosterone, inhibition of CYP17 activity by abiraterone decreases circulating levels of testosterone. <sup>181</sup>	In Phase III studies of 1195 patients, AA plus prednisone (797 patients), compared to placebo plus prednisone (398 patients), prolonged overall survival among patients with metastatic CRPC who had disease progression after docetaxel-based chemotherapy. The median overall survival was 14.8 months in the AA plus prednisone group vs. 10.9 months in the control. <sup>182</sup>
<b>Androgen receptor antagonist</b>  Enzalutamide/Xtandi  MDV3100	In addition, MDV3100 is an oral androgen receptor antagonist It directly inhibits AR by binding the receptor irreversibly. This interaction impairs AR nuclear translocation, DNA binding, and recruitment of co-activators. <sup>116</sup> Preclinical studies have demonstrated that MDV3100 binds to the AR receptor with substantially higher affinity compared to Bicalutamide (a clinical AR modulator), resulting in more complete suppression of the androgen receptor pathway. <sup>116</sup>	In a Phase I/II study, MDV3100 showed anti-tumor activity in patients with metastatic CRPC. 56% of 140 patients in the trial demonstrated decreases in serum PSA of 50% or more. 61 out of 109 patients had stabilized bone disease after treatment. <sup>183</sup>





## Author Contributions

Wrote the first draft of the manuscript: SD, DC, GQ. Contributed to the writing of the manuscript: SD, DC, GQ. Agree with manuscript results and conclusions: SD, DC, GQ. Jointly developed the structure and arguments for the paper: SD, DC, GQ. Made critical revisions and approved final version: SD, DC, GQ. All authors reviewed and approved of the final manuscript.

## DISCLOSURES AND ETHICS

As a requirement of publication the authors have provided signed confirmation of their compliance with ethical and legal obligations including but not limited to compliance with ICMJE authorship and competing interests guidelines, that the article is neither under consideration for publication nor published elsewhere, of their compliance with legal and ethical guidelines concerning human and animal research participants (if applicable), and that permission has been obtained for reproduction of any copyrighted material. This article was subject to blind, independent, expert peer review. The reviewers reported no competing interests.

## REFERENCES

1. Siegel R, Naishadham D, Jemal A. Cancer statistics, 2012. *CA Cancer J Clin.* 2012;62(1):10–29.
2. Jin JK, Dayyani F, Gallick GE. Steps in prostate cancer progression that lead to bone metastasis. *Int J Cancer.* 2011;128(11):2545–2561.
3. Ye L, Kynaston HG, Jiang WG. Bone metastasis in prostate cancer: molecular and cellular mechanisms (Review). *Int J Mol Med.* 2007;20(1):103–111.
4. Bubendorf L, Schopfer A, Wagner U, et al. Metastatic patterns of prostate cancer: an autopsy study of 1,589 patients. *Hum Pathol.* 2000;31(5):578–583.
5. Thobe MN, Clark RJ, Bainer RO, Prasad SM, Rinker-Schaeffer CW. From Prostate to Bone: Key Players in Prostate Cancer Bone Metastasis. *Cancers (Basel).* 2011;3(1):478–493.
6. Fizazi K, Carducci M, Smith M, et al. Denosumab versus zoledronic acid for treatment of bone metastases in men with castration-resistant prostate cancer: a randomised, double-blind study. *Lancet.* 2011;377(9768):813–822.
7. Siegel R, DeSantis C, Virgo K, et al. Cancer treatment and survivorship statistics, 2012. *CA Cancer J Clin.* 2012;62(4):220–241.
8. Paget S. The distribution of secondary growths in cancer of the breast. 1889. *Cancer Metastasis Rev.* 1989;8(2):98–101.
9. Fokas E, Engenhardt-Cabillic R, Daniilidis K, Rose F, An HX. Metastasis: the seed and soil theory gains identity. *Cancer Metastasis Rev.* 2007;26(3–4):705–715.
10. Goya M, Ishii G, Miyamoto S, et al. Prostate-specific antigen induces apoptosis of osteoclast precursors: potential role in osteoblastic bone metastases of prostate cancer. *Prostate.* 2006;66(15):1573–1584.
11. Wang L, Huang X, Zheng X, et al. Enrichment of prostate cancer stem-like cells from human prostate cancer cell lines by culture in serum-free medium and chemoradiotherapy. *Int J Biol Sci.* 2013;9(5):472–479.
12. Borghese C, Cattaruzza L, Pivetta E, et al. Gefitinib inhibits the cross-talk between mesenchymal stem cells and prostate cancer cells leading to tumor cell proliferation and inhibition of docetaxel activity. *J Cell Biochem.* 2013;114(5):1135–1144.
13. Sariisik E, Docheva D, Padula D, et al. Probing the interaction forces of prostate cancer cells with collagen I and bone marrow derived stem cells on the single cell level. *PLoS One.* 2013;8(3):e57706.
14. Mognetti B, Montagna GL, Perrelli MG, Pagliaro P, Penna C. Bone marrow mesenchymal stem cells increase motility of prostate cancer cells via production of stromal cell-derived factor-1alpha. *J Cell Mol Med.* 2013;17(2):287–292.
15. Cao J, Chiarelli C, Richman O, et al. Membrane type 1 matrix metalloproteinase induces epithelial-to-mesenchymal transition in prostate cancer. *J Biol Chem.* 2008;283(10):6232–6240.
16. Behnsawy HM, Miyake H, Harada K, Fujisawa M. Expression patterns of epithelial-mesenchymal transition markers in localized prostate cancer: significance in clinicopathological outcomes following radical prostatectomy. *BJU Int.* 2013;111(1):30–37.
17. Armstrong AJ, Freedland SJ, Garcia-Blanco M. Epithelial-mesenchymal transition in prostate cancer: providing new targets for therapy. *Asian J Androl.* 2011;13(2):179–180.
18. Sethi S, Macoska J, Chen W, Sarkar FH. Molecular signature of epithelial-mesenchymal transition (EMT) in human prostate cancer bone metastasis. *Am J Transl Res.* 2010;3(1):90–99.
19. Xu J, Wang R, Xie ZH, et al. Prostate cancer metastasis: role of the host microenvironment in promoting epithelial to mesenchymal transition and increased bone and adrenal gland metastasis. *Prostate.* 2006;66(15):1664–1673.
20. Kim CJ, Sakamoto K, Tambe Y, Inoue H. Opposite regulation of epithelial-to-mesenchymal transition and cell invasiveness by periostin between prostate and bladder cancer cells. *Int J Oncol.* 2011;38(6):1759–1766.
21. Zhu ML, Kyriianou N. Role of androgens and the androgen receptor in epithelial-mesenchymal transition and invasion of prostate cancer cells. *FASEB J.* 2010;24(3):769–777.
22. Zhau HE, Odero-Marah V, Lue HW, et al. Epithelial to mesenchymal transition (EMT) in human prostate cancer: lessons learned from ARCaP model. *Clin Exp Metastasis.* 2008;25(6):601–610.
23. Radisky ES, Radisky DC. Matrix metalloproteinase-induced epithelial-mesenchymal transition in breast cancer. *J Mammary Gland Biol Neoplasia.* 2010;15(2):201–212.
24. Zhang C, Soori M, Miles FL, et al. Paracrine factors produced by bone marrow stromal cells induce apoptosis and neuroendocrine differentiation in prostate cancer cells. *Prostate.* 2011;71(2):157–167.
25. Kambara T, Oyama T, Segawa A, Fukabori Y, Yoshida K. Prognostic significance of global grading system of Gleason score in patients with prostate cancer with bone metastasis. *BJU Int.* 2010;105(11):1519–1525.
26. Feldman BJ, Feldman D. The development of androgen-independent prostate cancer. *Nat Rev Cancer.* 2001;1(1):34–45.
27. Huang SF, Kim SJ, Lee AT, et al. Inhibition of growth and metastasis of orthotopic human prostate cancer in athymic mice by combination therapy with pegylated interferon-alpha-2b and docetaxel. *Cancer Res.* 2002;62(20):5720–5726.
28. Kingsley LA, Fournier PG, Chirgwin JM, Guise TA. Molecular biology of bone metastasis. *Mol Cancer Ther.* 2007;6(10):2609–2617.
29. Batson OV. The function of the vertebral veins and their role in the spread of metastases. 1940. *Clin Orthop Relat Res.* 1995(312):4–9.
30. Mundy GR. Metastasis to bone: causes, consequences and therapeutic opportunities. *Nat Rev Cancer.* 2002;2(8):584–593.
31. Keller ET, Brown J. Prostate cancer bone metastases promote both osteolytic and osteoblastic activity. *J Cell Biochem.* 2004;91(4):718–729.
32. Patel N, Ngo K, Hastings J, Ketchum N, Sepahpanah F. Prevalence of prostate cancer in patients with chronic spinal cord injury. *PM R.* 2011;3(7):633–636.
33. Nieder C, Haukland E, Pawinski A, Dalhaug A. Pathologic fracture and metastatic spinal cord compression in patients with prostate cancer and bone metastases. *BMC Urol.* 2010;10:23.
34. Teitelbaum SL, Ross FP. Genetic regulation of osteoclast development and function. *Nat Rev Genet.* 2003;4(8):638–649.
35. Raubenheimer EJ, Noffke CE. Pathogenesis of bone metastasis: a review. *J Oral Pathol Med.* 2006;35(3):129–135.
36. Morrissey C, Vessella RL. The role of tumor microenvironment in prostate cancer bone metastasis. *J Cell Biochem.* 2007;101(4):873–886.
37. Roodman GD. Mechanisms of bone metastasis. *Discov Med.* 2004;4(22):144–148.
38. Arya M, Bott SR, Shergill IS, Ahmed HU, Williamson M, Patel HR. The metastatic cascade in prostate cancer. *Surg Oncol.* 2006;15(3):117–128.
39. Yoneda T. Cellular and molecular mechanisms of breast and prostate cancer metastasis to bone. *Eur J Cancer.* 1998;34(2):240–245.
40. Hayashida T, Jinno H, Kitagawa Y. [Epithelial-mesenchymal transition and tumor microenvironment propose the seed and soil theory of the present age]. *Nippon Rinsho.* 2012;70 Suppl 7:203–207. Japanese.
41. Zhao H. Membrane trafficking in osteoblasts and osteoclasts: new avenues for understanding and treating skeletal diseases. *Traffic.* 2012;13(10):1307–1314.
42. Trilling GM, Cho H, Ugas MA, et al. Spinal metastasis in head and neck cancer. *Head Neck Oncol.* 2012;4:36.
43. Wang L, Rahman S, Lin CY, et al. A novel murine model of human renal cell carcinoma spinal metastasis. *J Clin Neurosci.* 2012;19(6):881–883.
44. Guise TA. Bone loss and fracture risk associated with cancer therapy. *Oncologist.* 2006;11(10):1121–1131.
45. Casimiro S, Guise TA, Chirgwin J. The critical role of the bone microenvironment in cancer metastases. *Mol Cell Endocrinol.* 2009;310(1–2):71–81.
46. Goya M, Yonou H, Saito S. [Mechanism of the development of osteoblastic bone metastasis of prostate cancer]. *Nippon Rinsho.* 2011;69 Suppl 5:638–644. Japanese.
47. Richards J, Lim AC, Hay CW, et al. Interactions of abiraterone, eplerenone, and prednisolone with wild-type and mutant androgen receptor: a rationale for increasing abiraterone exposure or combining with MDV3100. *Cancer Res.* 2012;72(9):2176–2182.
48. Schulze J, Weber K, Baranowsky A, et al. p65-Dependent production of interleukin-1beta by osteolytic prostate cancer cells causes an induction of chemokine expression in osteoblasts. *Cancer Lett.* 2012;317(1):106–113.
49. Carlin BI, Andriole GL. The natural history, skeletal complications, and management of bone metastases in patients with prostate carcinoma. *Cancer.* 2000;88(12 Suppl):2989–2994.
50. Yang J, Fizazi K, Peleg S, et al. Prostate cancer cells induce osteoblast differentiation through a Cbfa1-dependent pathway. *Cancer Res.* 2001;61(14):5652–5659.
51. Akech J, Wixted JJ, Bedard K, et al. Runx2 association with progression of prostate cancer in patients: mechanisms mediating bone osteolysis and osteoblastic metastatic lesions. *Oncogene.* 2010;29(6):811–821.



52. Liao J, Li X, Koh AJ, et al. Tumor expressed PTHrP facilitates prostate cancer-induced osteoblastic lesions. *Int J Cancer*. 2008;123(10):2267–2278.
53. Reichert JC, Quent VM, Burke LJ, Stansfield SH, Clements JA, Hutmacher DW. Mineralized human primary osteoblast matrices as a model system to analyse interactions of prostate cancer cells with the bone microenvironment. *Biomaterials*. 2010;31(31):7928–7936.
54. Dai J, Kitagawa Y, Zhang J, et al. Vascular endothelial growth factor contributes to the prostate cancer-induced osteoblast differentiation mediated by bone morphogenetic protein. *Cancer Res*. 2004;64(3):994–999.
55. Lee GT, Jung YS, Ha YS, Kim JH, Kim WJ, Kim IY. Bone morphogenetic protein-6 induces castration resistance in prostate cancer cells through tumor infiltrating macrophages. *Cancer Sci*. 2013;104(8):1027–1032.
56. Carducci MA, Jimeno A. Targeting bone metastasis in prostate cancer with endothelin receptor antagonists. *Clin Cancer Res*. 2006;12(20 Pt 2):6296s–6300s.
57. James ND, Caty A, Payne H, et al. Final safety and efficacy analysis of the specific endothelin A receptor antagonist zibotentan (ZD4054) in patients with metastatic castration-resistant prostate cancer and bone metastases who were pain-free or mildly symptomatic for pain: a double-blind, placebo-controlled, randomized Phase II trial. *BJU Int*. 2010;106(7):966–973.
58. Neuhaus ML, Platz EA, Till C, et al. Insulin-like growth factors and insulin-like growth factor-binding proteins and prostate cancer risk: results from the prostate cancer prevention trial. *Cancer Prev Res (Phila)*. 2013;6(2):91–99.
59. Malinowska K, Neuwirt H, Cavarretta IT, et al. Interleukin-6 stimulation of growth of prostate cancer in vitro and in vivo through activation of the androgen receptor. *Endocr Relat Cancer*. 2009;16(1):155–169.
60. Wang X, Lee SO, Xia S, et al. Endothelial cells enhance prostate cancer metastasis via IL-6 → androgen receptor → TGF-β → MMP-9 signals. *Mol Cancer Ther*. 2013;12(6):1026–1037.
61. Goel HL, Alam N, Johnson IN, Languino LR. Integrin signaling aberrations in prostate cancer. *Am J Transl Res*. 2009;1(3):211–220.
62. van den Hoogen C, van der Horst G, Cheung H, Buijs JT, Pelger RC, van der Pluijm G. Integrin alpha expression is required for the acquisition of a metastatic stem/progenitor cell phenotype in human prostate cancer. *Am J Pathol*. 2011;179(5):2559–2568.
63. Chu K, Cheng CJ, Ye X, et al. Cadherin-11 promotes the metastasis of prostate cancer cells to bone. *Mol Cancer Res*. 2008;6(8):1259–1267.
64. Mao Q, Zheng X, Yang K, et al. Suppression of migration and invasion of PC3 prostate cancer cell line via activating E-cadherin expression by small activating RNA. *Cancer Invest*. 2010;28(10):1013–1018.
65. Nguyen HL, Zucker S, Zarrabi K, Kadam P, Schmidt C, Cao J. Oxidative stress and prostate cancer progression are elicited by membrane-type 1 matrix metalloproteinase. *Mol Cancer Res*. 2011;9(10):1305–1318.
66. Yu L, Wang CY, Shi J, et al. Estrogens promote invasion of prostate cancer cells in a paracrine manner through up-regulation of matrix metalloproteinase 2 in prostatic stromal cells. *Endocrinology*. 2011;152(3):773–781.
67. Dougall WC. Molecular pathways: osteoclast-dependent and osteoclast-independent roles of the RANKL/RANK/OPG pathway in tumorigenesis and metastasis. *Clin Cancer Res*. 2012;18(2):326–335.
68. Santini D, Schiavon G, Vincenzi B, et al. Receptor activator of NF-κB (RANK) expression in primary tumors associates with bone metastasis occurrence in breast cancer patients. *PLoS One*. 2011;6(4):e19234.
69. Vandyke K, Jackson P, Rowe A, Russell PJ, Blair JM. Androgen decreases osteoprotegerin expression in prostate cancer cells. *Prostate Cancer Prostatic Dis*. 2007;10(2):160–166.
70. Prakash G, Gautam G. Optimal bone health management strategies in patients with prostate cancer. *Indian J Urol*. 2013;29(2):89–99.
71. Boras-Granic K, Wolsolmerski JJ. PTHrP and breast cancer: more than hypercalcemia and bone metastases. *Breast Cancer Res*. 2012;14(2):307.
72. Singh RK, Sudhakar A, Lokeshwar BL. Role of Chemokines and Chemokine Receptors in Prostate Cancer Development and Progression. *J Cancer Sci Ther*. 2010;2(4):89–94.
73. Salazar N, Castellán M, Shirodkar SS, Lokeshwar BL. Chemokines and chemokine receptors as promoters of prostate cancer growth and progression. *Crit Rev Eukaryot Gene Expr*. 2013;23(1):77–91.
74. Wan X, Li ZG, Yingling JM, et al. Effect of transforming growth factor beta (TGF-β) receptor I kinase inhibitor on prostate cancer bone growth. *Bone*. 2012;50(3):695–703.
75. Ustach CV, Taube ME, Hurst NJ Jr, et al. A potential oncogenic activity of platelet-derived growth factor d in prostate cancer progression. *Cancer Res*. 2004;64(5):1722–1729.
76. Conley-LaComb MK, Huang W, Wang S, et al. PTEN regulates PDGF ligand switch for beta-PDGFR signaling in prostate cancer. *Am J Pathol*. 2012;180(3):1017–1027.
77. Cheng J, Ye H, Liu Z, et al. Platelet-derived growth factor-BB accelerates prostate cancer growth by promoting the proliferation of mesenchymal stem cells. *J Cell Biochem*. 2013;114(7):1510–1518.
78. Traish AM, Morgentaler A. Epidermal growth factor receptor expression escapes androgen regulation in prostate cancer: a potential molecular switch for tumour growth. *Br J Cancer*. 2009;101(12):1949–1956.
79. Wang J, Yu W, Cai Y, Ren C, Ittmann MM. Altered fibroblast growth factor receptor 4 stability promotes prostate cancer progression. *Neoplasia*. 2008;10(8):847–856.
80. Feng S, Shao L, Yu W, Gavine P, Ittmann M. Targeting fibroblast growth factor receptor signaling inhibits prostate cancer progression. *Clin Cancer Res*. 2012;18(14):3880–3888.
81. Wang Q, Diao X, Sun J, Chen Z. Stromal cell-derived factor-1 and vascular endothelial growth factor as biomarkers for lymph node metastasis and poor cancer-specific survival in prostate cancer patients after radical prostatectomy. *Urol Oncol*. 2013;31(3):312–317.
82. Lee RJ, Smith MR. Targeting MET and vascular endothelial growth factor receptor signaling in castration-resistant prostate cancer. *Cancer J*. 2013;19(1):90–98.
83. Park JI, Lee MG, Cho K, et al. Transforming growth factor-beta1 activates interleukin-6 expression in prostate cancer cells through the synergistic collaboration of the Smad2, p38-NF-κB, JNK, and Ras signaling pathways. *Oncogene*. 2003;22(28):4314–4332.
84. Festuccia C, Angelucci A, Gravina GL, et al. Osteoblast-derived TGFβ1 modulates matrix degrading protease expression and activity in prostate cancer cells. *Int J Cancer*. 2000;86(6):888.
85. Tenta R, Sourla A, Lembessis P, Luu-The V, Koutsilieris M. Bone microenvironment-related growth factors, zoledronic acid and dexamethasone differentially modulate PTHrP expression in PC-3 prostate cancer cells. *Horm Metab Res*. 2005;37(10):593–601.
86. Codony-Servat J, Marin-Aguilera M, Visa L, et al. Nuclear factor-κB and interleukin-6 related docetaxel resistance in castration-resistant prostate cancer. *Prostate*. 2013;73(5):512–521.
87. Paller CJ, Carducci MA, Philips GK. Management of bone metastases in refractory prostate cancer—role of denosumab. *Clin Interv Aging*. 2012;7:363–372.
88. Azim HA, Kamal NS, Azim HA, Jr. Bone metastasis in breast cancer: the story of RANKL-ligand. *J Egypt Natl Canc Inst*. 2012;24(3):107–114.
89. Brown JM, Corey E, Lee ZD, et al. Osteoprotegerin and rank ligand expression in prostate cancer. *Urology*. 2001;57(4):611–616.
90. Zhang J, Dai J, Qi Y, et al. Osteoprotegerin inhibits prostate cancer-induced osteoclastogenesis and prevents prostate tumor growth in the bone. *J Clin Invest*. 2001;107(10):1235–1244.
91. Fohr B, Dunstan CR, Seibel MJ. Clinical review 165: Markers of bone remodeling in metastatic bone disease. *J Clin Endocrinol Metab*. 2003;88(11):5059–5075.
92. Glass DA 2nd, Bialek P, Ahn JD, et al. Canonical Wnt signaling in differentiated osteoblasts controls osteoclast differentiation. *Dev Cell*. 2005;8(5):751–764.
93. Mak KK, Bi Y, Wan C, et al. Hedgehog signaling in mature osteoblasts regulates bone formation and resorption by controlling PTHrP and RANKL expression. *Dev Cell*. 2008;14(5):674–688.
94. Lescarbeau RM, Seib FP, Prewitz M, Werner C, Kaplan DL. In vitro model of metastasis to bone marrow mediates prostate cancer castration resistant growth through paracrine and extracellular matrix factors. *PLoS One*. 2012;7(8):e40372.
95. Tsingotjidou AS, Zotalis G, Jackson KR, et al. Development of an animal model for prostate cancer cell metastasis to adult human bone. *Anticancer Res*. 2001;21(2A):971–978.
96. Virk MS, Lieberman JR. Tumor metastasis to bone. *Arthritis Res Ther*. 2007;9 Suppl 1:S5.
97. Theoleyre S, Wittrant Y, Tat SK, Fortun Y, Redini F, Heymann D. The molecular triad OPG/RANK/RANKL: involvement in the orchestration of pathophysiological bone remodeling. *Cytokine Growth Factor Rev*. 2004;15(6):457–475.
98. Zhang K, Waxman DJ. Impact of tumor vascularity on responsiveness to antiangiogenesis in a prostate cancer stem cell-derived tumor model. *Mol Cancer Ther*. 2013;12(5):787–798.
99. Pan L, Baek S, Edmonds PR, et al. Vascular endothelial growth factor (VEGF) expression in locally advanced prostate cancer: secondary analysis of radiation therapy oncology group (RTOG) 8610. *Radiat Oncol*. 2013;8:100.
100. Wang Q, Diao X, Sun J, Chen Z. Regulation of VEGF, MMP-9 and metastasis by CXCR4 in a prostate cancer cell line. *Cell Biol Int*. 2011;35(9):897–904.
101. Weber DC, Tille JC, Combesure C, et al. The prognostic value of expression of HIF1α, EGFR and VEGF-A, in localized prostate cancer for intermediate- and high-risk patients treated with radiation therapy with or without androgen deprivation therapy. *Radiat Oncol*. 2012;7:66.
102. Clarkin CE, Gerstenfeld LC. VEGF and bone cell signalling: an essential vessel for communication? *Cell Biochem Funct*. 2013;31(1):1–11.
103. Karsenty G, Wagner EF. Reaching a genetic and molecular understanding of skeletal development. *Dev Cell*. 2002;2(4):389–406.
104. Wang Y, Wan C, Deng L, et al. The hypoxia-inducible factor alpha pathway couples angiogenesis to osteogenesis during skeletal development. *J Clin Invest*. 2007;117(6):1616–1626.
105. Deckers MM, van Bezooijen RL, van der Horst G, et al. Bone morphogenetic proteins stimulate angiogenesis through osteoblast-derived vascular endothelial growth factor A. *Endocrinology*. 2002;143(4):1545–1553.



106. Sweeney P, Karashima T, Kim SJ, et al. Anti-vascular endothelial growth factor receptor 2 antibody reduces tumorigenicity and metastasis in orthotopic prostate cancer xenografts via induction of endothelial cell apoptosis and reduction of endothelial cell matrix metalloproteinase type 9 production. *Clin Cancer Res.* 2002;8(8):2714–2724.
107. Mukherji D, Temraz S, Wehbe D, Shamseddine A. Angiogenesis and anti-angiogenic therapy in prostate cancer. *Crit Rev Oncol Hematol.* 2013;87(2):122–131.
108. Roussos ET, Condeelis JS, Patsialou A. Chemotaxis in cancer. *Nat Rev Cancer.* 2011;11(8):573–587.
109. Fahrenholtz CD, Beltran PJ, Burnstein KL. Targeting IGF-IR with ganitumab inhibits tumorigenesis and increases durability of response to androgen-deprivation therapy in VCaP prostate cancer xenografts. *Mol Cancer Ther.* 2013;12(4):394–404.
110. Dean JP, Sprenger CC, Wan J, et al. Response of the insulin-like growth factor (IGF) system to IGF-IR inhibition and androgen deprivation in a neoadjuvant prostate cancer trial: effects of obesity and androgen deprivation. *J Clin Endocrinol Metab.* 2013;98(5):E820–828.
111. Russell PJ, Bennett S, Stricker P. Growth factor involvement in progression of prostate cancer. *Clin Chem.* 1998;44(4):705–723.
112. Kojima S, Inahara M, Suzuki H, Ichikawa T, Furuya Y. Implications of insulin-like growth factor-I for prostate cancer therapies. *Int J Urol.* 2009;16(2):161–167.
113. Guturi KK, Mandal T, Chatterjee A, et al. Mechanism of beta-catenin-mediated transcriptional regulation of epidermal growth factor receptor expression in glycogen synthase kinase 3 beta-inactivated prostate cancer cells. *J Biol Chem.* 2012;287(22):18287–18296.
114. Kumar R, Srinivasan S, Pahari P, Rohr J, Damodaran C. Activating stress-activated protein kinase-mediated cell death and inhibiting epidermal growth factor receptor signaling: a promising therapeutic strategy for prostate cancer. *Mol Cancer Ther.* 2010;9(9):2488–2496.
115. Peraldo-Neia C, Migliardi G, Mello-Grand M, et al. Epidermal Growth Factor Receptor (EGFR) mutation analysis, gene expression profiling and EGFR protein expression in primary prostate cancer. *BMC Cancer.* 2011;11:31.
116. Fu W, Madan E, Yee M, Zhang H. Progress of molecular targeted therapies for prostate cancers. *Biochim Biophys Acta.* 2012;1825(2):140–152.
117. Najy AJ, Jung YS, Won JJ, et al. Cediranib inhibits both the intraosseous growth of PDGF D-positive prostate cancer cells and the associated bone reaction. *Prostate.* 2012;72(12):1328–1338.
118. Virk MS, Alaei F, Petrigliano FA, et al. Combined inhibition of the BMP pathway and the RANK-RANKL axis in a mixed lytic/blastic prostate cancer lesion. *Bone.* 2011;48(3):578–587.
119. Virk MS, Petrigliano FA, Liu NQ, et al. Influence of simultaneous targeting of the bone morphogenetic protein pathway and RANK/RANKL axis in osteolytic prostate cancer lesion in bone. *Bone.* 2009;44(1):160–167.
120. Feeley BT, Krenke L, Liu N, et al. Overexpression of noggin inhibits BMP-mediated growth of osteolytic prostate cancer lesions. *Bone.* 2006;38(2):154–166.
121. Hau P, Jachimczak P, Schlingensiepen R, et al. Inhibition of TGF-beta2 with AP 12009 in recurrent malignant gliomas: from preclinical to phase I/II studies. *Oligonucleotides.* 2007;17(2):201–212.
122. Nordstrand A, Lundholm M, Larsson A, Lerner UH, Widmark A, Wikström P. Inhibition of the Insulin-Like Growth Factor-1 Receptor Enhances Effects of Simvastatin on Prostate Cancer Cells in Co-Culture with Bone. *Cancer Microenviron.* 2013.
123. Tolmachev V, Malmberg J, Hofstrom C, et al. Imaging of insulinlike growth factor type 1 receptor in prostate cancer xenografts using the affibody molecule 111In-DOTA-ZIGF1R:4551. *J Nucl Med.* 2012;53(1):90–97.
124. Dayyani F, Varkaris A, Araujo JC, et al. Increased serum insulin-like growth factor-1 levels are associated with prolonged response to dasatinib-based regimens in metastatic prostate cancer. *Prostate.* 2013;73(9):979–985.
125. Susperregui AR, Viñals F, Ho PW, Gillespie MT, Martin TJ, Ventura F. BMP-2 regulation of PTHrP and osteoclastogenic factors during osteoblast differentiation of C2C12 cells. *J Cell Physiol.* 2008;216(1):144–152.
126. Haudenschild DR, Palmer SM, Moseley TA, You Z, Reddi AH. Bone morphogenetic protein (BMP)-6 signaling and BMP antagonist noggin in prostate cancer. *Cancer Res.* 2004;64(22):8276–8284.
127. Yoshikawa H, Nakase T, Myoui A, Ueda T. Bone morphogenetic proteins in bone tumors. *J Orthop Sci.* 2004;9(3):334–340.
128. Mishra S, Tang Y, Wang L, et al. Blockade of transforming growth factor-beta (TGFbeta) signaling inhibits osteoblastic tumorigenesis by a novel human prostate cancer cell line. *Prostate.* 2011;71(13):1441–1454.
129. Corn PG, Wang F, McKeehen W, Navone N. Targeting Fibroblast Growth Factor Pathways in Prostate Cancer. *Clin Cancer Res.* 2013.
130. Normanno N, Gullick WJ. Epidermal growth factor receptor tyrosine kinase inhibitors and bone metastases: different mechanisms of action for a novel therapeutic application? *Endocr Relat Cancer.* 2006;13(1):3–6.
131. Asadi F, Farraj M, Sharifi R, Malakouti S, Antar S, Kukreja S. Enhanced expression of parathyroid hormone-related protein in prostate cancer as compared with benign prostatic hyperplasia. *Hum Pathol.* 1996;27(12):1319–1323.
132. Asadi F, Faraj M, Malakouti S, Kukreja SC. Effect of parathyroid hormone related protein, and dihydrotestosterone on proliferation and ornithine decarboxylase mRNA in human prostate cancer cell lines. *Int Urol Nephrol.* 2001;33(3):417–422.
133. Chen C, Koh AJ, Datta NS, et al. Impact of the mitogen-activated protein kinase pathway on parathyroid hormone-related protein actions in osteoblasts. *J Biol Chem.* 2004;279(28):29121–29129.
134. Iddon J, Bundred NJ, Hoyland J, et al. Expression of parathyroid hormone-related protein and its receptor in bone metastases from prostate cancer. *J Pathol.* 2000;191(2):170–174.
135. Asadi FK, Kukreja SC, Boyer B, Valess AM, Cook JL. E1A oncogene expression inhibits PTHrP P3 promoter activity and sensitizes human prostate cancer cells to TNF-induced apoptosis. *Int Urol Nephrol.* 2010;42(4):971–978.
136. Bhatia V, Saini MK, Shen X, et al. EB1089 inhibits the parathyroid hormone-related protein-enhanced bone metastasis and xenograft growth of human prostate cancer cells. *Mol Cancer Ther.* 2009;8(7):1787–1798.
137. Silva I, Branco JC. Rank/Rankl/opg: literature review. *Acta Reumatol Port.* 2011;36(3):209–218.
138. Zhang L, Shi J, Feng J, Klocker H, Lee C, Zhang J. Type IV collagenase (matrix metalloproteinase-2 and -9) in prostate cancer. *Prostate Cancer Prostatic Dis.* 2004;7(4):327–332.
139. Reis ST, Pontes-Junior J, Antunes AA, et al. MMP-9 overexpression due to TIMP-1 and RECK underexpression is associated with prognosis in prostate cancer. *Int J Biol Markers.* 2011;26(4):255–261.
140. Reis ST, Antunes AA, Pontes J Jr, et al. Underexpression of MMP-2 and its regulators, TIMP2, MT1-MMP and IL-8, is associated with prostate cancer. *Int Braz J Urol.* 2012;38(2):167–174.
141. Prata J, Lian JB, Javed A, et al. Regulatory roles of Runx2 in metastatic tumor and cancer cell interactions with bone. *Cancer Metastasis Rev.* 2006;25(4):589–600.
142. Dong Z, Bonfil RD, Chinni S, et al. Matrix metalloproteinase activity and osteoclasts in experimental prostate cancer bone metastasis tissue. *Am J Pathol.* 2005;166(4):1173–1186.
143. Gupta A, Cao W, Chellaiah MA. Integrin alphavbeta3 and CD44 pathways in metastatic prostate cancer cells support osteoclastogenesis via a Runx2/Smad 5/receptor activator of NF-kappaB ligand signaling axis. *Mol Cancer.* 2012;11:66.
144. Bonfil RD, Dong Z, Trindade Filho JC, et al. Prostate cancer-associated membrane type 1-matrix metalloproteinase: a pivotal role in bone response and intraosseous tumor growth. *Am J Pathol.* 2007;170(6):2100–2111.
145. Sabbota AL, Kim HR, Zhe X, Fridman R, Bonfil RD, Cher ML. Shedding of RANKL by tumor-associated MT1-MMP activates Src-dependent prostate cancer cell migration. *Cancer Res.* 2010;70(13):5558–5566.
146. Ibrahim T, Flamini E, Mercatali L, Sacanna E, Serra P, Amadori D. Pathogenesis of osteoblastic bone metastases from prostate cancer. *Cancer.* 2010;116(6):1406–1418.
147. Aeschlimann D, Evans BA. The vital osteoclast: how is it regulated? *Cell Death Differ.* 2004;11 Suppl 1:S5–7.
148. Bonfil RD, Chinni S, Fridman R, Kim HR, Cher ML. Proteases, growth factors, chemokines, and the microenvironment in prostate cancer bone metastasis. *Urol Oncol.* 2007;25(5):407–411.
149. Matsuo K, Irie N. Osteoclast-osteoblast communication. *Arch Biochem Biophys.* 2008;473(2):201–209.
150. Wang J, Sun Y, Song W, et al. Diverse signaling pathways through the SDF-1/CXCR4 chemokine axis in prostate cancer cell lines leads to altered patterns of cytokine secretion and angiogenesis. *Cell Signal.* 2005;17(12):1578–1592.
151. Kakinuma T, Hwang ST. Chemokines, chemokine receptors, and cancer metastasis. *J Leukoc Biol.* 2006;79(4):639–651.
152. Tawara K, Oxford JT, Jorczyk CL. Clinical significance of interleukin (IL)-6 in cancer metastasis to bone: potential of anti-IL-6 therapies. *Cancer Manag Res.* 2011;3:177–189.
153. Hudes G, Tagawa ST, Whang YE, et al. A phase 1 study of a chimeric monoclonal antibody against interleukin-6, siltuximab, combined with docetaxel in patients with metastatic castration-resistant prostate cancer. *Invest New Drugs.* 2013;31(3):669–676.
154. Taichman RS, Cooper C, Keller ET, Pienta KJ, Taichman NS, McCauley LK. Use of the stromal cell-derived factor-1/CXCR4 pathway in prostate cancer metastasis to bone. *Cancer Res.* 2002;62(6):1832–1837.
155. Domanska UM, Timmer-Bosscha H, Nagengast WB, et al. CXCR4 inhibition with AMD3100 sensitizes prostate cancer to docetaxel chemotherapy. *Neoplasia.* 2012;14(8):709–718.
156. Loberg RD, Ying C, Craig M, Yan L, Snyder LA, Pienta KJ. CCL2 as an important mediator of prostate cancer growth in vivo through the regulation of macrophage infiltration. *Neoplasia.* 2007;9(7):556–562.
157. Loberg RD, Ying C, Craig M, et al. Targeting CCL2 with systemic delivery of neutralizing antibodies induces prostate cancer tumor regression in vivo. *Cancer Res.* 2007;67(19):9417–9424.
158. Mian BM, Dinney CP, Bermejo CE, et al. Fully human anti-interleukin 8 antibody inhibits tumor growth in orthotopic bladder cancer xenografts via down-regulation of matrix metalloproteinases and nuclear factor-kappaB. *Clin Cancer Res.* 2003;9(8):3167–3175.



159. Huang S, Mills L, Mian B, et al. Fully humanized neutralizing antibodies to interleukin-8 (ABX-IL8) inhibit angiogenesis, tumor growth, and metastasis of human melanoma. *Am J Pathol.* 2002;161(1):125–134.
160. Yang XD, Corvalan JR, Wang P, Roy CM, Davis CG. Fully human anti-interleukin-8 monoclonal antibodies: potential therapeutics for the treatment of inflammatory disease states. *J Leukoc Biol.* 1999;66(3):401–410.
161. Xu J, Escamilla J, Mok S, et al. CSF1R signaling blockade stanches tumor-infiltrating myeloid cells and improves the efficacy of radiotherapy in prostate cancer. *Cancer Res.* 2013;73(9):2782–2794.
162. Wilson C, Maxwell PJ, Longley DB, Wilson RH, Johnston PG, Waugh DJ. Constitutive and treatment-induced CXCL8-signalling selectively modulates the efficacy of anti-metabolite therapeutics in metastatic prostate cancer. *PLoS One.* 2012;7(5):e36545.
163. Karkera J, Steiner H, Li W, et al. The anti-interleukin-6 antibody siltuximab down-regulates genes implicated in tumorigenesis in prostate cancer patients from a phase I study. *Prostate.* 2011;71(13):1455–1465.
164. Fizazi K, De Bono JS, Flechon A, et al. Randomised phase II study of siltuximab (CNTO 328), an anti-IL-6 monoclonal antibody, in combination with mitoxantrone/prednisone versus mitoxantrone/prednisone alone in metastatic castration-resistant prostate cancer. *Eur J Cancer.* 2012;48(1):85–93.
165. Roux S, Mariette X. RANK and RANKL expression in giant-cell tumour of bone. *Lancet Oncol.* 2010;11(6):514.
166. Cross SS, Harrison RF, Balasubramanian SP, et al. Expression of receptor activator of nuclear factor kappa beta ligand (RANKL) and tumour necrosis factor related, apoptosis inducing ligand (TRAIL) in breast cancer, and their relations with osteoprotegerin, oestrogen receptor, and clinicopathological variables. *J Clin Pathol.* 2006;59(7):716–720.
167. Asahi H, Mizokami A, Maeda Y, Komatsu K, Koshida K, Namiki M. Bisphosphonate therapy for hormone refractory prostate cancer with bone metastasis. *J Urol.* 2003;169(1):281–282.
168. Uemura H, Yanagisawa M, Ikeda I, et al; Yokohama Bone Metastasis Study Group. Possible anti-tumor activity of initial treatment with zoledronic acid with hormonal therapy for bone-metastatic prostate cancer in multicenter clinical trial. *Int J Clin Oncol.* 2013;18(3):472–477.
169. Fizazi K, Lipton A, Mariette X, et al. Randomized phase II trial of denosumab in patients with bone metastases from prostate cancer, breast cancer, or other neoplasms after intravenous bisphosphonates. *J Clin Oncol.* 2009;27(10):1564–1571.
170. So A, Chin J, Fleshner N, Saad F. Management of skeletal-related events in patients with advanced prostate cancer and bone metastases: Incorporating new agents into clinical practice. *Can Urol Assoc J.* 2012;6(6):465–470.
171. Fizazi K, Miller K. Specific endothelin-A receptor antagonism for the treatment of advanced prostate cancer. *BJU Int.* 2009;104(10):1423–1425.
172. Warren R, Liu G. ZD4054: a specific endothelin A receptor antagonist with promising activity in metastatic castration-resistant prostate cancer. *Expert Opin Investig Drugs.* 2008;17(8):1237–1245.
173. Nelson JB, Udan MS, Guruli G, Pflug BR. Endothelin-1 inhibits apoptosis in prostate cancer. *Neoplasia.* 2005;7(7):631–637.
174. Pouessel D, Culine S. Complete clinical and biological response to zoledronic acid in castrate-resistant prostate cancer metastatic to bone. *Anticancer Drugs.* 2012;23(1):141–142.
175. Kamiya N, Suzuki H, Endo T, et al. Additive effect of zoledronic acid on serum prostate-specific antigen changes for hormone-sensitive prostate cancer patients with bone metastasis treated by combined androgen blockade. *Int J Urol.* 2012;19(2):169–173.
176. Snedecor SJ, Carter JA, Kaura S, Botteman MF. Denosumab versus zoledronic acid for treatment of bone metastases in men with castration-resistant prostate cancer: a cost-effectiveness analysis. *J Med Econ.* 2013;16(1):19–29.
177. Todenhofer T, Hennenlotter J, Schmiedel BJ, et al. Alterations of the RANKL pathway in blood and bone marrow samples of prostate cancer patients without bone metastases. *Prostate.* 2013;73(2):162–168.
178. Tomblyn M. The role of bone-seeking radionuclides in the palliative treatment of patients with painful osteoblastic skeletal metastases. *Cancer Control.* 2012;19(2):137–144.
179. Saad F, Lipton A. SRC kinase inhibition: targeting bone metastases and tumor growth in prostate and breast cancer. *Cancer Treat Rev.* 2010;36(2):177–184.
180. Yu EY, Massard C, Gross ME, et al. Once-daily dasatinib: expansion of phase II study evaluating safety and efficacy of dasatinib in patients with metastatic castration-resistant prostate cancer. *Urology.* 2011;77(5):1166–1171.
181. Derleth CL, Yu EY. Targeted therapy in the treatment of castration-resistant prostate cancer. *Oncology (Williston Park).* 2013;27(7):620–628.
182. de Bono JS, Logothetis CJ, Molina A, et al. Abiraterone and increased survival in metastatic prostate cancer. *N Engl J Med.* 2011;364(21):1995–2005.
183. Scher HI, Beer TM, Higano CS, et al. Antitumor activity of MDV3100 in castration-resistant prostate cancer: a phase 1–2 study. *Lancet.* 2010;375(9724):1437–1446.