

Effects of omalizumab therapy on peripheral nerve functions: short observational study

Goknur Ozaydin Yavuz¹, Abdullah Yilgör², Ibrahim Halil Yavuz¹, Aysel Milanlıoğlu², Vedat Çilingir², Aydın Çağaç², Murat Ozturk¹, Serap Gunes Bilgili¹

¹Department of Dermatology, Faculty of Medicine, Van Yuzuncu Yil University, Van, Turkey

²Department of Neurology, Faculty of Medicine, Van Yuzuncu Yil University, Van, Turkey

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Abstract

Introduction: Peripheral neuropathy (PN) is a common neurological condition causing symmetrical and diffuse damage in nerves. The etiology of PN includes systemic diseases, toxic exposure, medications, infections, and hereditary diseases. Omalizumab is a humanized monoclonal anti-IgE antibody that exerts its activity by binding to free IgE in circulation.

Aim: To investigate the relationship between omalizumab and peripheral neuropathy.

Material and methods: The study included 30 patients who underwent omalizumab therapy (Xolair) due to the diagnosis of chronic urticaria. A detailed neurological and physical examination was performed in each patient both before and 3 months after the therapy. Electrophysiological examination was also performed using a Medelec Synergy instrument.

Results: The 30 patients included 8 (26.7%) men and 22 (73.3%) women with a mean age of 37.5 ±14.14 years. No serious side effect of the medication was detected in any patient although local wound irritation occurred in 3 (10%) patients. Moreover, no change occurred in the pre-treatment Neuropathy Symptom Score (NSS) or Neurological Disability Score (NDS) of the patients and no pathological values that could result in neuropathy were observed during motor/sensory nerve conduction. However, significant changes were detected in the sensory and motor components of the nerves with regards to pre- and post-treatment values.

Conclusions: Omalizumab therapy caused no peripheral neuropathy in any of our patients but altered the latency, amplitude, and velocity values of the peripheral nerves.

Key words: chronic urticaria, omalizumab, neuropathy.

Introduction

Peripheral neuropathy (PN) is one of the most common neurological conditions, causing symmetrical and diffuse damage in nerves. The etiology of PN includes systemic diseases, toxic exposure, medications, infections, and hereditary diseases. The most common medications associated with PN include amiodarone, chloroquine, hydralazine, lithium, metronidazole, phenytoin, isoniazid, statins, and vincristine. In addition, biological agents including infliximab and adalimumab have been reported in recent case studies. The prevalence of PN has been reported to be as high as 2.4% in the general population and to be 26.4% in patients with diabetes mellitus [1–4].

Immunoglobulin E (IgE) plays a central role in the pathogenesis of allergic conditions. Therefore, anti-IgE therapies play a key role in the treatment of allergic diseases such as asthma [1–5]. Omalizumab is a humanized monoclonal anti-IgE antibody. Omalizumab exerts its activity by binding to free IgE in circulation, thereby inhibiting the binding of IgE to its high-affinity receptors (FcεRI) found on mast cells and basophils, ultimately reducing the expression of mediators in mast cells. Omalizumab is also an important treatment option particularly for severe asthma and resistant chronic urticaria [6–8].

On the other hand, although histamine is the most important mediator expressed in mast cells, neuropeptides such as nerve growth factor (NGF) are also expressed in these cells [9]. A previous study reported that

Address for correspondence: Goknur Ozaydin Yavuz MD, Department of Dermatology, Faculty of Medicine, Van Yuzuncu Yil University, 65100 Van, Turkey, phone: +90 505 4753362, +90 5054753362, fax: +90 5054753362, e-mail: goknuroz1@gmail.com

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mast cell activation led to an increase in the production and secretion of neuropeptides and the excitability of sensory nerves [10]. A recent study evaluated the effectiveness of omalizumab at 52 weeks and showed that omalizumab is a safe drug although it had several side effects including headache, injection site reaction, myalgia, lethargy, nausea, dizziness, weight gain, and arthralgia [11]. However, another study showed that omalizumab therapy resulted in optic neuritis when used for the treatment of bronchial asthma in 2 patients with Churg-Strauss syndrome [12]. Similarly, Lieberman *et al.* compared omalizumab therapy with placebo therapy and reported that local skin reactions occurred in 44% of the patients treated with omalizumab [13].

In our patients, we also performed omalizumab therapy for the treatment of chronic urticaria. However, the complaints of pain and weakness in the extremities gradually increased in our patients and thus we could not be sure whether these complaints resulted from local irritation or a neurological condition caused by omalizumab, mainly because the drug was administered in two separate infusions with 150 mg flacons. Moreover, these conditions may be a result of peripheral nerve injury caused by the inhibition of mast cells that leads a reduction in the expression of neuromediators.

Aim

In this study, we aimed to evaluate the relationship between omalizumab and peripheral neuropathy.

Material and methods

The study included 30 patients who underwent omalizumab therapy (Xolair) due to the diagnosis of chronic urticaria. Age, gender, socioeconomic status, and family history were recorded for each patient. Omalizumab was subcutaneously administered at 300 mg/day for 28 days (total 4 times: 1, 29, 57, 85 days) in the Dermatology Department. To determine the presence of other factors that may affect peripheral nerve function, additional tests were performed, including complete blood count, sedimentation rate, liver and kidney function tests, urine analysis, thyroid hormones, vitamin B₁₂ level, folic acid level, and serologic tests. In addition, neuro-radiological imaging was performed as needed. Exclusion criteria included neurological symptoms and signs, diabetes mellitus, connective tissue disease, hepatic, renal, and thyroid diseases, amyloidosis, heart failure, alcohol abuse, corticosteroid use, cervical disc hernia, and malignancy. A detailed neurological and physical examination was performed in each patient before and three months after the therapy. Neurological symptoms were scored using the Neuropathy Symptom Score (NSS) and Neurological Disability Score (NDS). Following the neurological examination, electrophysiological exami-

nation was performed using a Medelec Synergy instrument (Oxford Instruments, Surrey, UK) with standard neurographic procedures, and the results were evaluated according to the American Diabetes Association (ADA) Diabetic Neuropathy Guidelines in the Neurology Department [14]. The measurements were performed 24 h before and 90 days after omalizumab therapy. Room temperature was kept at 22–24°C and the temperature of the extremity was kept at 34°C and it was heated as needed. Nerve conduction tests were performed in two motor and two sensory nerves (median and ulnar nerves) in the upper extremities and in two motor (tibial and common peroneal nerve) and two sensory nerves (sural and superficial peroneal sensory nerves) in the lower extremities. Pre- and post-treatment latency (ms), amplitude (mV), and velocity (m/s) values were compared for each nerve. Nerve conduction velocity was measured using the orthodromic method and nerve conduction was performed at supramaximal intensity to achieve the highest amplitude. Presence of an axonal pathology and demyelination in the nerves was defined as decreased sensory/motor nerve action potential amplitude and slowing of sensory/motor nerve conduction velocity. Polyneuropathy was defined as the presence of two or more abnormalities detected in electrophysiological examination. The study was approved by the local ethics committee and informed consent was obtained from each patient (Number: Y.Y.U: 2017/ 02).

Statistical analysis

Data were analyzed using IBM SPSS for Windows, Version 20.0 (Armonk, NY: IBM Corp.). Normal distributions of variables were determined by histogram and/or by the Kolmogorov-Smirnov/Shapiro-Wilk test. Descriptive statistics were expressed as mean, standard deviation (SD), median, minimum, and maximum. Numerical variables were compared using the paired sample *t*-test for data with normal distribution and the Wilcoxon signed-rank test for data with non-normal distribution. Spearman's correlation coefficient was used to assess the correlation between variables. A *p*-value of < 0.05 was considered significant.

Results

The 30 patients included 8 (26.7%) men and 22 (73.3%) women with a mean age of 37.5 ±14.14 years (Table 1). No serious side effect of the medication was observed in any patient although local wound irritation occurred in 3 (10%) patients. No change occurred in the pre-treatment NSS and NDS scores of the patients. Moreover, no pathological values that could result in neuropathy were observed during motor/sensory nerve conduction. Nevertheless, significant changes were detected in the sensory and motor components of the nerves with regards to pre- and post-treatment values.

A comparison of pre- and post-treatment latency, amplitude, and velocity values of the sensory component of the median nerves indicated no significant difference between the pre- and post-treatment latency and velocity values ($p > 0.05$). However, post-treatment amplitude values (mean: 23.03 ± 4.71) were significantly lower than pre-treatment values (mean: 24.61 ± 5.55) ($p = 0.024$) (Table 2).

No significant difference was found between the pre- and post-treatment latency and velocity values of the sensory component of the ulnar nerves ($p > 0.05$). Nevertheless, post-treatment amplitude values (mean: 18.71 ± 5.34) were significantly lower than pre-treatment values (mean: 20.11 ± 5.61) ($p = 0.030$) (Table 3).

In the sensory component of the sural nerves, no significant difference was found between the pre- and post-treatment amplitude values ($p > 0.05$). However, post-treatment latency values (mean: 2.12 ± 0.24) were significantly higher than pre-treatment values (mean: 1.96 ± 0.21) ($p = 0.009$), whereas post-treatment velocity values (mean: 46.34 ± 3.60) were significantly lower than pre-treatment values (mean: 48.14 ± 7.09) ($p = 0.001$) (Table 4).

In the sensory component of the superficial peroneal nerves, no significant difference was found between pre- and post-treatment latency values ($p > 0.05$), whereas post-treatment amplitude and velocity values (mean: 14.00 ± 3.02 and 47.20 ± 3.36 , respectively) were significantly lower than pre-treatment values (mean: 16.45 ± 4.05 and 49.34 ± 3.36 , respectively) ($p < 0.001$ for both) (Table 5).

On the other hand, a comparison of pre- and post-treatment latency, amplitude, and velocity values of the motor component of the median nerves indicated no significant difference between the pre- and post-treatment latency values ($p > 0.05$) although post-treatment amplitude and velocity values (mean: 8.40 ± 1.54 and 57.97 ± 4.42 , respectively) were significantly lower than pre-treatment values (mean: 9.21 ± 2.08 and 59.42 ± 4.61 , respectively) ($p = 0.007$ and 0.049 , respectively) (Table 6).

Table 1. Characteristics of patients

Parameter	N	%
Gender:		
Male	8	26.67
Female	22	73.33
Marital status:		
Married	23	76.67
Single/other	7	23.33
Education status:		
Primary school	6	20.00
Middle school	8	26.67
High school	9	30.00
University	7	23.33
Income status:		
Low income	15	50.00
Middle income	11	36.67
High income	4	13.33
Age*	37.50 \pm 14.14	35.50
Disease duration [months]*	12.67 \pm 8.05	10.50
Cumulative dose (for each patient)	1200 mg	

*Results are expressed as mean \pm SD and median data instead of N and %.

Moreover, no significant difference was found between the pre- and post-treatment latency and amplitude values of the motor component of the ulnar nerves ($p > 0.05$). However, post-treatment velocity values (mean: 57.62 ± 4.13) were significantly lower than pre-treatment values (mean: 59.04 ± 4.02) ($p = 0.002$).

In the motor component of the superficial peroneal nerves, post-treatment latency values (mean: 3.90 ± 0.45) were significantly higher than pre-treatment values

Table 2. Comparison of pre- and post-treatment latency, amplitude, and velocity values of the sensory component of the median nerves

Variable	Mean \pm SD	Median	Minimum	Maximum	P-value
Latency-pre	2.17 \pm 0.23	2.13	1.75	2.75	0.278 ^b
Latency-post	2.23 \pm 0.31	2.13	1.80	3.00	
Amplitude-pre	24.61 \pm 5.55	25.50	13.50	35.00	0.024 ^a
Amplitude-post	23.03 \pm 4.71	23.10	13.50	31.90	
Velocity-pre	55.67 \pm 5.81	55.60	40.00	63.40	0.821 ^b
Velocity-post	55.28 \pm 5.24	55.95	42.00	63.90	

^aPaired samples t-test, ^bWilcoxon signed-rank test, pre – pre-treatment, post – post-treatment.

Table 3. Comparison of pre- and post-treatment latency, amplitude, and velocity values of the sensory component of the ulnar nerves

Variable	Mean ± SD	Median	Minimum	Maximum	P-value
Latency-pre	1.89 ±0.19	1.90	1.50	2.45	0.436 ^b
Latency-post	1.91 ±0.22	1.83	1.60	2.40	
Amplitude-pre	20.11 ±5.61	18.95	11.90	34.00	0.030 ^b
Amplitude-post	18.71 ±5.34	17.45	12.00	31.00	
Velocity-pre	54.76 ±2.99	54.90	50.00	61.10	0.209 ^a
Velocity-post	53.94 ±3.60	52.75	44.90	60.60	

^aPaired samples t-test, ^bWilcoxon signed-rank test, pre – pre-treatment, post – post-treatment.

Table 4. Comparison of pre- and post-treatment latency, amplitude, and velocity values of the sensory component of the sural nerves

Variable	Mean ± SD	Median	Minimum	Maximum	P-value
Latency-pre	1.96 ±0.21	2.00	1.55	2.42	0.009 ^b
Latency-post	2.12 ±0.24	2.10	1.55	2.55	
Amplitude-pre	15.39 ±3.30	14.55	9.40	22.20	0.053 ^b
Amplitude-post	14.48 ±3.32	14.20	8.00	21.80	
Velocity-pre	48.14 ±7.09	48.85	16.80	58.10	0.001 ^a
Velocity-post	46.34 ±3.60	45.80	41.90	56.90	

^aPaired samples t-test, ^bWilcoxon signed-rank test, pre – pre-treatment, post – post-treatment.

Table 5. Comparison of pre- and post-treatment latency, amplitude, and velocity values of the sensory component of the superficial peroneal nerves

Variable	Mean ± SD	Median	Minimum	Maximum	P-value
Latency-pre	2.00 ±0.28	2.00	1.50	2.55	0.497
Latency-post	2.05 ±0.28	2.05	1.60	2.90	
Amplitude-pre	16.45 ±4.05	16.20	9.20	23.50	< 0.001
Amplitude-post	14.00 ±3.02	14.30	9.00	21.80	
Velocity-pre	49.34 ±4.61	48.25	41.90	58.10	0.002
Velocity-post	47.20 ±3.36	47.45	41.30	55.90	

^aPaired samples t-test, ^bWilcoxon signed-rank test, pre – pre-treatment, post – post-treatment.

Table 6. Comparison of pre- and post-treatment latency, amplitude, and velocity values of the motor component of the median nerves

Variable	Mean ± SD	Median	Minimum	Maximum	P-value
Latency-pre	2.76 ±0.35	2.73	2.15	3.70	0.052 ^b
Latency-post	2.91 ±0.47	2.80	2.20	3.95	
Amplitude-pre	9.21 ±2.08	8.70	6.60	15.80	0.007 ^b
Amplitude-post	8.40 ±1.54	8.10	5.70	13.50	
Velocity-pre	59.42 ±4.61	60.15	51.00	69.40	0.049 ^a
Velocity-post	57.97 ±4.42	58.60	50.00	64.90	

^aPaired samples t-test, ^bWilcoxon signed-rank test, pre – pre-treatment, post – post-treatment.

(mean: 3.68 ± 0.44) ($p = 0.005$), whereas post-treatment amplitude and velocity values (mean: 3.47 ± 0.93 and 48.35 ± 4.14 , respectively) were significantly lower than pre-treatment values (mean: 3.98 ± 1.06 and 49.64 ± 3.46 , respectively) ($p < 0.004$ and 0.036 , respectively).

In the motor component of the tibial nerves, no significant difference was found between pre- and post-treatment latency values ($p > 0.05$). However, post-treatment amplitude and velocity values (mean: 7.63 ± 1.30 and 44.87 ± 3.20 , respectively) were significantly lower than pre-treatment values (mean: 8.64 ± 2.00 and 46.60 ± 3.45 , respectively) ($p < 0.002$ and 0.006 , respectively).

Discussion

The results indicated that omalizumab did not cause peripheral neuropathy but altered the latency, amplitude, and velocity values of the peripheral nerves. To our knowledge, this is the first study in the literature investigating the relationship between omalizumab and peripheral neuropathy.

We evaluated the effect of omalizumab in patients with chronic urticaria. Kim *et al.* also evaluated the effect of omalizumab in patients with chronic spontaneous urticaria and reported that 61.75% of the patients were women. Similarly, women also constituted the majority of our patients (73.3%), which implies that chronic urticaria has a female preponderance [15].

Omalizumab has been shown to be a safe drug in numerous studies. However, a number of side effects have been associated with omalizumab, including anaphylaxis, urticaria, eosinophilic granulomatosis with polyangiitis, susceptibility to parasitic infections, injection site reactions, cardiovascular diseases, and serum sickness [13, 16–20]. On the other hand, a previous study reported that headache and disturbance of sleep were the most common neurological side effects of omalizumab [19]. Similarly, Corren *et al.* reviewed more than 7,500 patients undergoing omalizumab therapy and found that headache was the most common neurological side effect and also noted that omalizumab led to musculoskeletal disturbances including low back pain, arthralgia, pain in the extremities, and myalgia [17]. In our study, no complaint of headache was found in any patient, which could be ascribed to the small patient series in our study.

Literature reviews indicate that there are a limited number of studies investigating the effect of omalizumab on the nerves. Jachiet *et al.* reported that omalizumab therapy resulted in optic neuritis in 2 patients [12]. In contrast, Kalteren *et al.* evaluated a patient with ocular myasthenic syndrome and reported that all the symptoms were resolved after the treatment of the syndrome with omalizumab therapy [21]. On the other hand, Alvarez-Lario *et al.* evaluated the effectiveness of biological treatment and reported that the side effects of the

treatment resulted in peripheral neuropathy associated with Guillain-Barré syndrome in 21 patients. The authors considered that peripheral neuropathy resulted from the increased susceptibility to infections caused by biological agents [22]. As shown in these studies, omalizumab typically increases susceptibility to infections. However, no infection associated with omalizumab was observed in our patients. On the other hand, although no peripheral neuropathy occurred in any of our patients, we consider that omalizumab has the potential to affect peripheral nerves since it has been shown to increase susceptibility to infections, to alter the growth factors and neuropeptides expressed in mast cells, and to trigger the auto-immune process, as observed in a patient described by Kalteren *et al.* In addition, depending on the finding that omalizumab changed the latency, amplitude, and velocity values of peripheral nerves in our patients, we believe that omalizumab is likely to increase the severity of neuropathy when used in combination with a drug that affects peripheral nerves.

Our study was limited since it was a single-center study and had a relatively small number of patients. Moreover, since we assessed the neurological symptoms of the patients before and three months after the treatment, different outcomes could have been detected if the symptoms had also been assessed at 1 year after the treatment.

Conclusions

Omalizumab therapy is becoming gradually popular, with a growing side-effect profile. In this study, we investigated the relationship between omalizumab and neuropathy. However, further studies are needed to shed light on our findings.

Conflict of interest

The authors declare no conflict of interest.

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