

Review

# Stimuli-Responsive Poly(aspartamide) Derivatives and Their Applications as Drug Carriers

Guangyan Zhang <sup>1,2,\*</sup> , Hui Yi <sup>1</sup> and Chenhui Bao <sup>1</sup>

<sup>1</sup> School of Materials and Chemical Engineering, Hubei University of Technology, Wuhan 430068, China; yihuihbut@163.com (H.Y.); xiaochengzikid@163.com (C.B.)

<sup>2</sup> Hubei Provincial Key Laboratory of Green Materials for Light Industry, Hubei University of Technology, Wuhan 430068, China

\* Correspondence: zhangguangyan@whu.edu.cn

**Abstract:** Poly(aspartamide) derivatives, one kind of amino acid-based polymers with excellent biocompatibility and biodegradability, meet the key requirements for application in various areas of biomedicine. Poly(aspartamide) derivatives with stimuli-responsiveness can usually respond to external stimuli to change their chemical or physical properties. Using external stimuli such as temperature and pH as switches, these smart poly(aspartamide) derivatives can be used for convenient drug loading and controlled release. Here, we review the synthesis strategies for preparing these stimuli-responsive poly(aspartamide) derivatives and the latest developments in their applications as drug carriers.

**Keywords:** poly(aspartamide) derivatives; stimuli-responsive; drug carrier; nanoparticles; hydrogel; polymer-drug conjugate; drug-loading; controlled drug release



**Citation:** Zhang, G.; Yi, H.; Bao, C. Stimuli-Responsive Poly(aspartamide) Derivatives and Their Applications as Drug Carriers. *Int. J. Mol. Sci.* **2021**, *22*, 8817. <http://doi.org/10.3390/ijms22168817>

Academic Editors: Ádám Juhász and Edit Csapó

Received: 29 July 2021

Accepted: 13 August 2021

Published: 16 August 2021

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



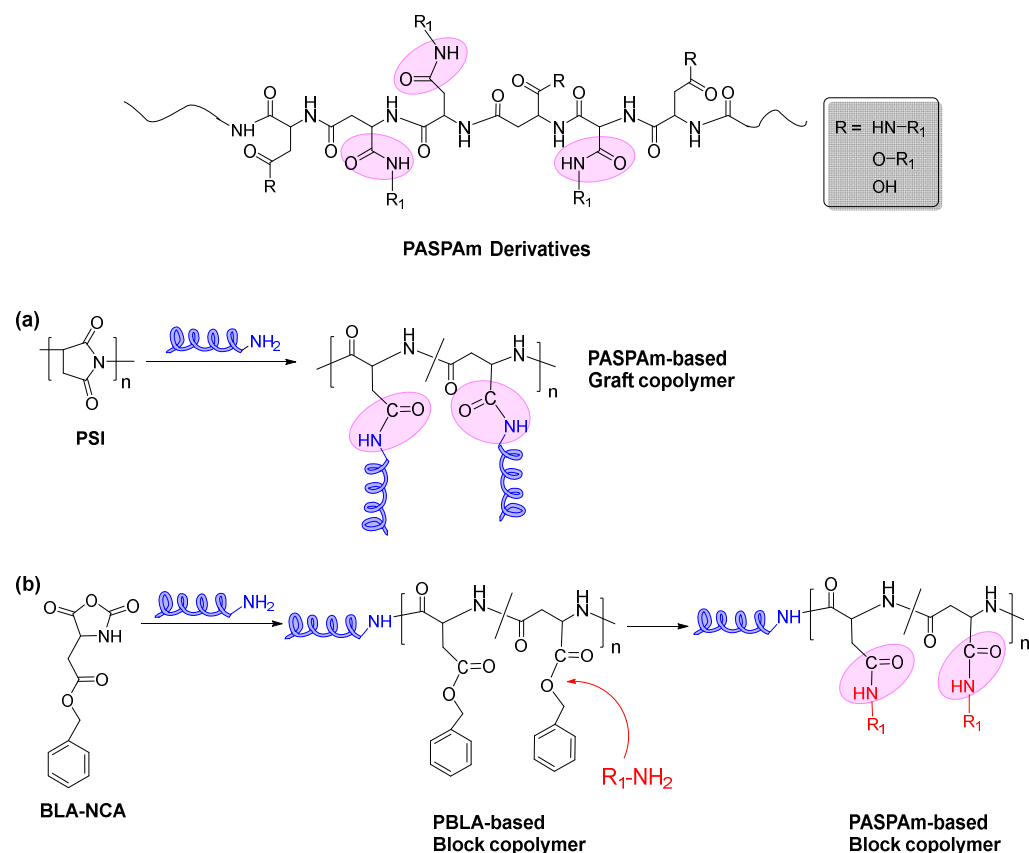
**Copyright:** © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Though the carbon-carbon backbone is conducive to the stability of polymers, it also limits the applications of these polymers in biomedical fields due to its low biocompatibility and non-biodegradability. In the past decades, the interest in amino acid-based biodegradable polymers has increased significantly in biomedicine due to their good biocompatibility, biodegradability and non-toxicity of their degradation products. For example, poly(glutamic acid) (PGA) and PGA-based polymers have been widely applied for controlled release of peptide, protein and anti-tumor drugs [1]; polylysine (PLL) and its derivatives have been extensively investigated in gene delivery systems [2]. Moreover, other amino-acid containing degradable polymers have also been studied in tissue engineering and biomedical imaging (e.g., magnetic resonance imaging and fluorescence imaging).

Poly(aspartic acid)-based polymers, whose backbone is composed of aspartic acid, are one of the most intensively studied amino acid-based polymers in drug [3,4]/gene [5–7] delivery systems and other biomedical applications [8] due to their simple synthesis method, excellent biocompatibility and biodegradability. The synthetic strategies of poly(aspartic acid)-based polymers mainly include the aminolysis reaction of poly(succinimide) (PSI) (Figure 1a) and the ring-opening polymerization (ROP) of benzyl-L-aspartate *N*-carboxyanhydride (BLA-NCA) (Figure 1b). For side chain-modified poly(aspartic acid)-based polymers, they can generally be prepared by the aminolysis reaction of PSI. For example, a side chain-modified poly(aspartamide) (PASPAm) derivative,  $\alpha,\beta$ -poly(*N*-2-hydroxyethyl)-*D,L*-aspartamide (PHEA), can be obtained by the aminolysis reaction between ethanolamine and PSI, and has been further modified and intensively studied as scaffolds [9], siRNA delivery systems [10,11] and drug carriers (e.g., hydrogel, nanoparticles) [12–16]. For the block copolymers containing poly(aspartic acid)-based segment, they usually can be synthesized by ROP of BLA-NCA. For instance, NK105 polymer, which is

composed of a poly(ethylene glycol) (PEG) block as the hydrophilic segment and modified poly(aspartic acid)-based block as the hydrophobic segment in which 50% carboxylic groups of PAsPAm were reacted with 4-phenyl-1-butanol by esterification reaction [17,18], has been used as delivery system in a paclitaxel-incorporating micellar nanoparticle anti-cancer formulation (NK105) that was approved for clinical trials (phase III) [19,20]. Besides NK105, poly(aspartic acid)-based polymers were also applied in two other formulations, NK911 [21] and NC-6300 [22,23]. Among the above three clinical formulations, NC-6300 that is an epirubicin (EPI) conjugated PEG-block-PAsPAm derived copolymer containing acid-labile hydrazone linkages, is the only one with stimuli-responsiveness. Results showed that NC-6300 exhibited a stronger anti-tumor effect and lower cardiotoxicity compared to free EPI, which may be due to its pH responsiveness.



**Figure 1.** The synthetic pathways of PAsPAm-based (a) graft copolymers and (b) block copolymers.

Because stimuli-responsive polymers can respond to the changes of environmental factors (e.g., pH, temperature) by self-changing either their physical properties, chemical structures, or both, they exhibit attractive prospects in a variety of fields such as sensors, energy conversion and drug/gene delivery systems [24]. Therefore, various stimuli-responsive PAsPAm derivatives such as NC-6300 have been designed and synthesized over the last decades, and their potential applications in different fields were also investigated extensively. In this review, PAsPAm derivatives are defined as polymers composed wholly or partly of aspartamide units as shown in Figure 1.

Although some papers have reviewed the synthesis and applications of poly(aspartic acid) (PASP) [25] and PSI [26] derivatives, there is no comprehensive review summarizing the recent progress of “stimuli-responsive PAsPAm derivatives”. In this paper, we will provide an overview of the up-to-date developments on stimuli-responsive PAsPAm derivatives and their applications as drug carriers. First, the synthesis strategies of stimuli-responsive PAsPAm derivatives will be reviewed by the type of triggers. Next, the applications of stimuli-responsive PAsPAm derivatives for drug delivery will be in-

roduced. Finally, the further perspectives of stimuli-responsive PAsPAM derivatives will be discussed.

## 2. Synthesis

### 2.1. Temperature-Responsive Poly(aspartamide) Derivatives

Temperature is an important stimulus for stimuli-responsive polymers. Poly(*N*-isopropylacrylamide) (PNIPAAm) is one of the most investigated temperature-responsive polymers with a lower critical solution temperature (LCST) at around 32 °C. As is well known, the temperature-responsive behavior of PNIPAAm is ascribed to its *N*-isopropylamide pendants. Therefore, *N*-isopropylamide and various functional groups with similar structures were introduced for preparing temperature-responsive PAsPAM derivatives. For instance, two different temperature-responsive PAsPAM derivatives, PAIPAHA and PAATS, were prepared by the following procedures: (1) First, a certain amount of isopropylamine was reacted with PSI, and then (2) alkanolamines [27] or NaOH [28] were further used to open the remaining succinimide rings to obtain temperature responsiveness. Besides *N*-isopropylamide pendants, diisopropylamide [29] and isopropylethylenediamide [30,31] pendants were also introduced for the design of temperature-responsive PAsPAM derivatives.

In fact, in the absence of isopropylamide or diisopropylamide pendants, PAsPAM derivatives containing alkanolamides moieties also can possess temperature responsiveness. In 2003, a series of temperature-responsive PAsPAM(C<sub>5</sub>OH/C<sub>6</sub>OH) were synthesized via the aminolysis reaction of PSI with a mixture of 5-aminopentanol (NH<sub>2</sub>C<sub>5</sub>OH) and 6-aminohexanol (NH<sub>2</sub>C<sub>6</sub>OH) by Kobayashi's group [32]. Interestingly, the reaction product of PSI with NH<sub>2</sub>C<sub>5</sub>OH, denoted as PHPA, was completely soluble in water; while the reaction product of PSI with NH<sub>2</sub>C<sub>6</sub>OH was insoluble in water (test temperature: 0–100 °C). Since then, more and more temperature-responsive PAsPAM derivatives containing alkanolamides moieties were designed and synthesized [33–35]. For instance, adopting the similar strategy of Kobayashi's group, Chu et al. prepared a new family of temperature-responsive PAsPAM(C<sub>4</sub>OH/C<sub>6</sub>OH) with an LCST ranging from 28 to 53 °C using the mixture of 4-aminobutanol (NH<sub>2</sub>C<sub>4</sub>OH)/NH<sub>2</sub>C<sub>6</sub>OH instead of the mixture of NH<sub>2</sub>C<sub>5</sub>OH/NH<sub>2</sub>C<sub>6</sub>OH [36]. In 2014, phenethyl alcohol (phe) was grafted to the pentanolamide pendants of PHPA, and the obtained five phe-g-PHPA polymers all exhibited temperature responsiveness. The LCST of phe-g-PHPA can be tuned by adjusting the percentage of grafted phe moieties, and has a negative linear correlation with the grafting ratio of phe [37]. In 2015, a certain amount of 2-azidoethylamine (Az) was used for aminolysis reaction with PSI, and then excess NH<sub>2</sub>C<sub>5</sub>OH was further utilized to open the residual succinimide rings to prepare azide-functional PAsPAM derivative P(Asp-Az)<sub>x</sub>-HPA. Two P(Asp-Az)<sub>x</sub>-HPA polymers showed temperature-responsive behaviors in aqueous solution, and the molar ratio of Az/NH<sub>2</sub>C<sub>5</sub>OH in these two P(Asp-Az)<sub>x</sub>-HPA polymers was 39/61 and 56/44, respectively [38]. Additionally, the LCST of P(Asp-Az)<sub>x</sub>-HPA-based polymers can be further adjusted by grafting hydrophobic moieties, such as cinnamoyl group [33].

Moreover, ether oxygen also plays an important role for the design of temperature-responsive polymers. For example, oligo(ethylene glycol) (OEG) containing ether oxygen was usually introduced for preparing temperature-responsive polymers, such as OEG methacrylate based polymers [39,40] and polypeptides bearing OEG pendants [41,42]. In 2021, linear alkyl ether-type amine compounds, which contain one ether oxygen and varying carbon atoms, were successfully used for the synthesis of temperature-responsive PAsPAM derivatives bearing alkyl ether-type pendants [43]. The composition of temperature-responsive PAsPAM derivatives and their LCSTs are summarized in Table 1.

**Table 1.** A summary of temperature-responsive PAsPAm derivatives.





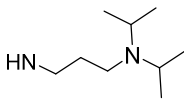
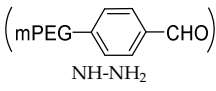
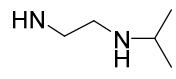
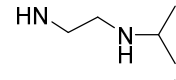
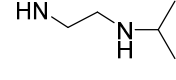
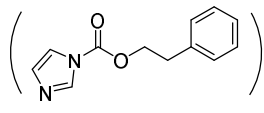
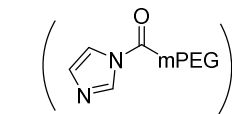
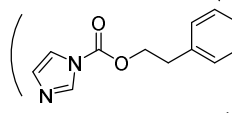
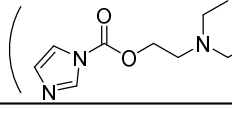

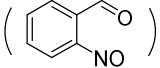
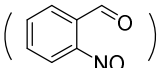
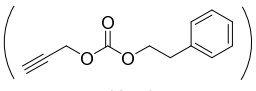
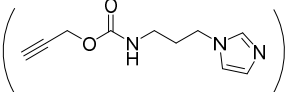
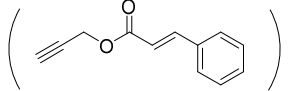
Temperature Responsive PAsPAm Derivatives	Composition of Side Chain		R <sub>1</sub> /R <sub>2</sub>	LCST (°C)	Ref
	R <sub>1</sub>	R <sub>2</sub>			
PAIPAHA		NH-(CH <sub>2</sub> ) <sub>6</sub> OH	55/45	30	[27]
PAIPAPA		NH-(CH <sub>2</sub> ) <sub>3</sub> OH	55/45	40	[27]
PAA-TS		OH	69/31 73/27 74/26 75/25 77/23 78/22 80/20	38 <sup>a</sup> 40 <sup>a</sup> 45 <sup>a</sup> 47 <sup>a</sup> 49 <sup>a</sup> 56 <sup>a</sup> 62 <sup>a</sup>	[28]
PSI-TS		-	69/31	34 <sup>a</sup>	[28]
mPEG-hyd-PDAH <sub>y</sub>		(mPEG-  ) NH-NH <sub>2</sub>	80/(9)20	40	[29]
PolyAspAm(HA/NIPEDA)		NH-(CH <sub>2</sub> ) <sub>5</sub> CH <sub>3</sub>	57/43 58/42 59/41 62/38	24 30 39 52	[30]
PolyAspAm(OA/NIPEDA)		NH-(CH <sub>2</sub> ) <sub>7</sub> CH <sub>3</sub>	57/43 59/41 60.5/39.5 62/38	22 36 43 49	[30]
PolyAspAm(LA/NIPEDA)		NH-(CH <sub>2</sub> ) <sub>11</sub> CH <sub>3</sub>	59/41 61/39 64/36 50/50	22 37 50 23	[30,31]
PAspAm(C5OH/C6OH)	NH-(CH <sub>2</sub> ) <sub>5</sub> OH	NH-(CH <sub>2</sub> ) <sub>6</sub> OH	60/40 70/30 80/20 30/70 25/75	29 37 44 53 40	[32]
PAspAm(C4OH/C6OH)	NH-(CH <sub>2</sub> ) <sub>4</sub> OH	NH-(CH <sub>2</sub> ) <sub>6</sub> OH	20/80 15/85 10/90	38 32 28	[36]
phe-g-PHPA	(  )	NH-(CH <sub>2</sub> ) <sub>5</sub> OH	(2)/100 (6)/100 (11)/100 (12)/100 (13)/100	54 32 20 14 9	[37]
Phe/DEAE-g-PHPA	(  )	NH-(CH <sub>2</sub> ) <sub>5</sub> OH	(0/22.2/30.3)/100	35.7 <sup>b</sup>	[34]
	(  )		(0/26.6/28.3)/100	31.5 <sup>b</sup>	
	(  )		(0/37.6/27.3)/100	22.5 <sup>b</sup>	
	(  )		(0/39.6/19.0)/100	29.5	
			(0/53.6/12.0)/100	45.8	
			(5/34.6/12.7)/100	28 <sup>b</sup>	
			(5/42.4/10.5)/100	15 <sup>b</sup>	

Table 1. Cont.

Temperature Responsive PAsPAM Derivatives	Composition of Side Chain		R <sub>1</sub> /R <sub>2</sub>	LCST (°C)	Ref
	R <sub>1</sub>	R <sub>2</sub>			
NB-g-PHPA		NH-(CH <sub>2</sub> ) <sub>5</sub> OH	(4)/100 (7.5)/100 (10)/100	32 21 11	[35]
NB-g-PHPA-g-mPEG	 NH-(CH <sub>2</sub> ) <sub>5</sub> OH	NH-mPEG	(12)98.5/1.5 (17)98.5/1.5	25 8	[35]
P(Asp-Az) <sub>x</sub> -HPA	NH-(CH <sub>2</sub> ) <sub>2</sub> N <sub>3</sub>	NH-(CH <sub>2</sub> ) <sub>5</sub> OH	40/60 39/61 56/44	63 58 29	[33] [38]
P(Asp-Az) <sub>x</sub> -HPA-PEA	 NH-(CH <sub>2</sub> ) <sub>2</sub> N <sub>3</sub>	NH-(CH <sub>2</sub> ) <sub>5</sub> OH	(5)39/61 (9.9)39/61	19 4	[38]
P(Asp-Az) <sub>x</sub> -HPA-IMZ	 NH-(CH <sub>2</sub> ) <sub>2</sub> N <sub>3</sub>	NH-(CH <sub>2</sub> ) <sub>5</sub> OH	(5)39/61 (9.7)39/61	22 10	[38]
P(HPA-Az)-CA	 NH-(CH <sub>2</sub> ) <sub>2</sub> N <sub>3</sub>	NH-(CH <sub>2</sub> ) <sub>5</sub> OH	(2.5)40/60 (4.3)40/60	47 40	[33]
PASP-OC <sub>x</sub>	NH-(CH <sub>2</sub> ) <sub>3</sub> OCH <sub>2</sub> CH <sub>3</sub> NH-(CH <sub>2</sub> ) <sub>2</sub> OCH <sub>2</sub> CH <sub>3</sub>	- -	100/- 100/-	54 87	[43]

<sup>a</sup> Determined at pH = 3.4; <sup>b</sup> Determined at pH = 7.4 PBS. mPEG: monomethoxy poly(ethyleneglycol).

## 2.2. pH-Responsive Poly(aspartamide) Derivatives

Poly(aspartic acid) is a water soluble polymer, and its conformation in aqueous solution can be affected by pH due to pH-induced deprotonation and protonation of its carboxylic acid pendants. For example, the  $\alpha$  helical conformation of poly(aspartic acid) that appeared in acidic solution was not observed in basic solution [44], which may be attributed to the negative charge repulsion caused by the deprotonation of pendant carboxylic acid groups in alkaline environment. Thus, one strategy for preparing pH-responsive PAsPAM derivatives is to introduce other pH-sensitive groups or non-ionic groups. The introduction of non-ionic groups can usually be used to adjust the phase transition pH value (pH<sub>t</sub>) [45].

1-(3-aminopropyl) imidazole (API) is one of the most commonly used reagents for introducing imidazole groups to PAsP or PBLA. For instance, Kim et al. grafted imidazole to PEG-b-PASP copolymer in which 60% carboxyl groups were reacted with the amine group of API forming amide linkages, resulting a pH-responsive zwitterionic PAsPAM derivatives [46]. In addition, pH-responsiveness also can be obtained by substituting the benzyl groups of PBLA with API [47]. Another way for introducing imidazole groups to synthesize pH-responsive PAsPAM derivatives is to directly react the succinimide rings of PSI with API [48–53] or histamine [54] through aminolysis reaction. Besides imidazole, tertiary amine [34,55–58] and other groups [30,59] are also intensively adopted for endowing pH responsiveness to PAsPAM derivatives.

Another strategy for preparing pH-responsive polymers is to introduce pH-cleavable linkages, which mainly include hydrazone [60–62], imine [63], acetal, orthoester and 2,3-dialkylmaleamic amide linkages. Hydrazone linkage [64] plays an important role in the

preparation of pH-responsive polymers. The most well-known pH-responsive PAsPAM derivative is NC-6300, in which EPI is bound to a PEG-PAsPAM block copolymer through hydrazone linkage [23]. Thus, EPI can be released under acidic environments, such as in lysosomes. Doxorubicin (DOX)-conjugated PAsPAM derivatives with hydrazone linkages were also designed and prepared by researchers [65–68]. Moreover, two PAsPAM cross-linkers containing hydrazide groups (PHHZA) or amine groups (PHEDA) were synthesized by Cha's team. PHHZA and PHEDA can react with the aldehyde groups of oxidized alginate forming alginate-based hydrogels via hydrazone and imine linkages, respectively [69]. In addition, Kataoka's team synthesized three pH-responsive charge-conversional PAsPAM derivatives containing *cis*-aconitic amide linkage (PAsp(DET-Aco) [70], PEG-PAsp(DET-Aco) [71]) or 2-propionic-3-methyl maleic amide linkage (PEG-PAsp(DET-PMM) [71]). The obtained three polymers showed negative charge at pH 7.4 due to the carboxylic group in *cis*-aconityl (Aco) or 2-propionic-3-methyl maleic (PMM) moieties, but exhibited positive charge at pH 5.5 because they can return to PAsp(DET) or PEG-PAsp(DET) via the breakup of *cis*-aconitic amide linkage or 2-propionic-3-methyl maleic amide linkage under acidic conditions. The pH-cleavable linkages used for preparing pH responsive PAsPAM derivatives are summarized in Table 2.

**Table 2.** A summary of pH-responsive PAsPAM derivatives containing pH-cleavable linkages.

pH Responsive PAsPAM Derivatives	Linkages	Formation of Linkages between		Ref
		A	B	
NC-6300	Hydrazone	Hydrazide group of PEG polyaspartate block copolymer	Ketone group of EPI	[23]
Cross-linked nanocapsules with PADH and PACA	Hydrazone	Hydrazide group of PADH	Aldehyde groups of PACA	[56]
	Hydrazone	Hydrazide group of PALHy	Ketone group of DOX	[65]
ALN-PEG/C-18/HYD-DOX-g-PASPAM	Hydrazone	Hydrazide group of PAsPAM	Ketone group of DOX	[66]
MPEG/CA10/DOX-g-PASPAM	Hydrazone	Hydrazide group of MPEG/Hyd/CA10-g-PASPAM	Ketone group of DOX	[67]
PA	Hydrazone	Hydrazide group of biotin-PEG/C18-PSI/Hydrazine	Ketone group of DOX	[68]
PEG/Hyd-Curcumin/C <sub>18</sub> -g-PSI (NFA-Cur)	Hydrazone	Hydrazide group of PEG/C18-g-PSI	Ketone group of Curcumin	[60]
Injectable PAsp hydrogel	Hydrazone	Hydrazide group of PAHy	Aldehyde groups of PAAld	[61]
PHHZA-linked alginate hydrogels	Hydrazone	Hydrazide group of PHHZA	Aldehyde groups of oxidized alginate	[69]
PHEDA-linked alginate hydrogels	Imine	Amino group of PHEDA	Aldehyde groups of oxidized alginate	[69]
Pasp-EDA-g-Ad/mPEG	Imine	Amino group of Pasp-EDA	Aldehyde group of 4-adamantane carboxylate benzaldehyde	[63]
PAsp(DET-Aco)	cis-Aconitic amide	Amino group of PAsp(DET)	Anhydride group of cis-aconitic anhydride	[70]
PEG-PAsp(DET-ACO)	cis-Aconitic amide	Amino group of PEG-PAsp(DET)	Anhydride group of cis-aconitic anhydride	[71]
PEG-PAsp(DET-PMM)	2-propionic-3-methyl maleic amide	Amino group of PEG-PAsp(DET)	Anhydride group of 2-propionic-3-methyl maleic anhydride	[71]
mPEG-g-P(ae-Asp)-Hap	$\beta$ -carboxylic amide	Amino group of PEG-g-P(ae-Asp)	Anhydride group of hexahydrophthalic anhydride	[72]

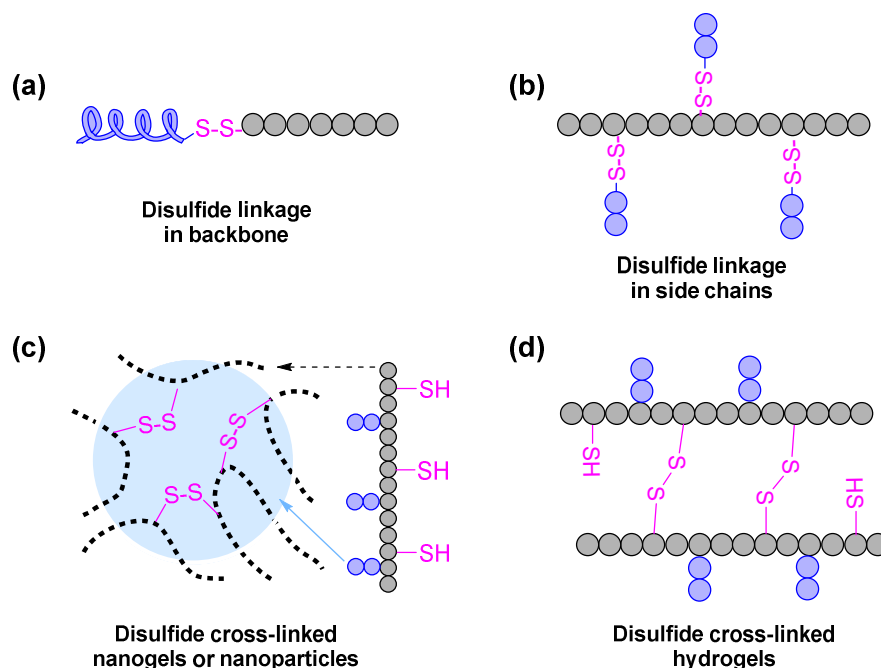


Besides the two above mentioned strategies, the unreacted succinimide rings in PAS-PAm derivatives also can be utilized to design pH-responsive controlled-release delivery vehicles, because the hydrophobic succinimide units can be hydrolyzed at pH above 7.0 to form hydrophilic aspartic acid units, thereby releasing the encapsulated materials due to the hydrophobic-to-hydrophilic change of PAS-PAm derivatives [73].

Interestingly, it has been reported that some temperature-responsive PAS-PAm derivatives also exhibit pH responsiveness even without introducing pH-responsive functional groups or pH-cleavable linkages. The pH responsiveness of temperature-responsive PAS-PAm derivatives may be related to the presence of hydroxyl groups in alkanolamide pendants [36] or the end groups (carboxyl and amine groups) of PAS-PAm backbone [43].

### 2.3. Redox-Responsive Poly(aspartamide) Derivatives

PASPAm derivatives generally can be endowed with redox-responsiveness by introducing disulfide linkage in their backbone (Figure 2a) or side chains (Figure 2b). Chu et al. [47] used cystamine-modified mPEG as a macroinitiator to initiate the polymerization of BLA-NCA, and then obtained a redox-responsive mPEG-block-PASPAm derivative, which contains a disulfide linkage between mPEG segment and PAS-PAm segment. Redox-responsive PAS-PAm derivatives, which have mPEG pendants with disulfide linkages, can also be synthesized through grafting 3,3'-dithiodipropionic acid-modified mPEG [74] or the aminolysis reaction between PSI and amino-terminated disulfide functionalized mPEG [48]. Moreover, some cationic polymers [75,76] or other compounds [77] can also be functionalized with cystamine and then used to design reduction-responsive PAS-PAm derivatives for DNA/RNA delivery. Of course, cystamine also can be used as a cross-linker for preparing reduction-responsive nanogels [78] or nanoparticles [33] (Figure 2c). In addition to S-S linkage, Se-Se linkage is also a good strategy for designing redox-responsive PAS-PAm derivatives [79].



**Figure 2.** The redox-responsive PAS-PAm derivatives containing disulfide linkages (a) in backbone and (b) in side chains, and disulfide cross-linked (c) nanogels or nanoparticles and (d) hydrogels.

In addition, thiol-disulfide inter-conversion can also be used for preparing redox-responsive PAS-PAm hydrogels [80–83] or stabilized disulfide cross-linked nanoparticles [84]. For instance, PSI first reacted with cysteamine, and then further reacted with other amine compounds (e.g., *N,N*-dimethyl-2-aminoethyl [85], 6-monodeoxy-6-amino-



beta-cyclodextrin hydrochloride [86]) to obtain thiolated PASPAm derivatives. The obtained thiolated PASPAm derivatives can be cross-linked by the oxidation of the thiol groups in their cysteamide pendants and formed reduction-responsive hydrogels (Figure 2d).

#### 2.4. Other Stimuli-Responsive Poly(aspartamide) Derivatives

Besides the above three stimuli-responsive (temperature, pH and redox) PASPAm derivatives, PASPAm derivatives with other stimuli responsiveness (including light responsiveness and carbon dioxide (CO<sub>2</sub>) responsiveness) have also been investigated in the past two decades.

CO<sub>2</sub> is an inexpensive gas and also an important metabolite in cells. It is reported that polymers containing dialkylamine [87,88], amidine [89] or guanidine [90] moieties can exhibit reversible CO<sub>2</sub>-responsive behavior because CO<sub>2</sub> can be bonded onto these polymers in the presence of water. For example, Kim's group prepared three CO<sub>2</sub>-responsive PASPAm derivatives, PHEA-HIS [54], PHEA-Larg and PolyAspAm(OA/Larg) [91]. The swelling degree of cross-linked PHEA-HIS hydrogel decreased to about 18 when CO<sub>2</sub> was bubbled into the solution containing PHEA-HIS hydrogel samples, and returned to about 50 after bubbling N<sub>2</sub> gas. PHEA-Larg hydrogels and PolyAspAm(OA/Larg) nanoparticle also exhibited similar reversible volume change behavior by CO<sub>2</sub>/N<sub>2</sub> purge.

Because light is a clean stimulus and can be easily triggered on or off, light-responsive polymers have shown huge potential for a variety of applications. For designing light-responsive polymers, photo-cleavage groups such as *o*-nitrobenzyl [92] were generally introduced. For instance, *o*-nitrobenzyl (NB) alcohol [35] was first activated by *N,N'*-carbonyldiimidazole and then grafted onto the pendant pentanolamide moieties of PHPA. When the obtained light-responsive NB-*g*-PHPA was treated under UV light at 365 nm for 30 min, the photo-cleavage *o*-nitrobenzyl carbonate ester linkage was broken and byproduct *o*-nitrosobenzaldehyde was detected.

In addition, light can also be used for cross-linking to stabilize PASPAm-based micelles. For instance, light responsive coumarin moieties were introduced to the side chains of PASPAm derivative, and the formed micelles can be conveniently cross-linked and stabilized by exposing it to the light of 365 nm for 30 min [93]. Photo-crosslinking can also be achieved by introducing methacryloyl [9,94] or acryloyl [95] moieties to the ethanolamide or hydrazide pendants of PASPAm.

Dual or multi-responsive PASPAm derivatives can also be prepared by the combination of above-mentioned strategies for endowing various stimuli-responsiveness. Therefore, we will not go into the details here.

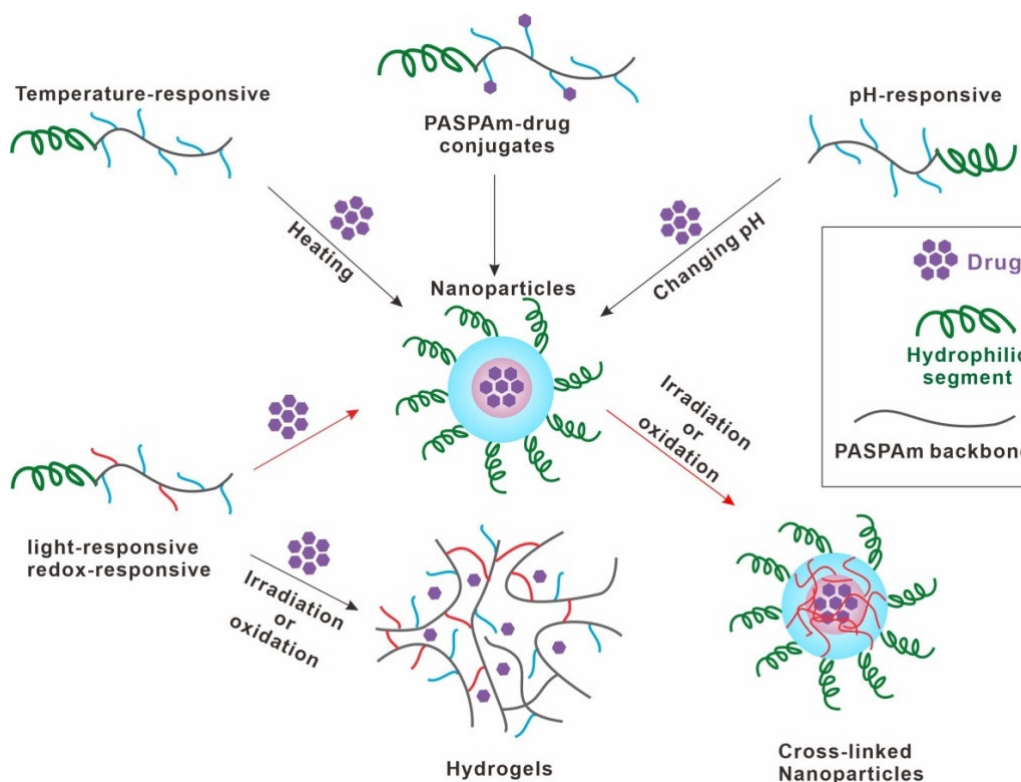
### 3. Applications

Due to their good biocompatibility and biodegradability, PASPAm derivatives could be regarded as potential and promising materials and can be utilized in various areas of biomedical applications, such as drug carriers. As a drug carrier, the process of drug loading and drug release are both important. PASPAm derivatives with stimuli-responsiveness can usually show convenient drug loading methods or excellent controlled drug release performances. Therefore, this part focuses on the applications of stimuli-responsive PASPAm derivatives as drug carriers from two aspects: drug loading and drug release.

#### 3.1. Drug Loading

Amphiphilic polymers, which can self-assemble to form nanoparticles including micelles and vesicles, are one of the most important classes of polymers used to deliver hydrophobic drugs such as paclitaxel (PTX) and doxorubicin (DOX). For preparing drug-loaded nanoparticles, one of the important methods is dialysis. However, the dialysis method often takes a long time (at least a few hours) and requires the use of toxic organic solvents (e.g., DMSO, CH<sub>3</sub>OH) to dissolve the polymeric carrier [93], and the drug loading efficiency is usually low. Thus, a simple, easy-to-implement drug loading strategy with high efficiency is needed for preparing drug-loaded nanoparticles.

Temperature and pH are often used as triggers for drug loading. For temperature responsive polymers, the general drug loading process is as follows: (1) prepare polymer aqueous solution of a certain concentration in PBS at a temperature (e.g., 4 °C) lower than LCST; (2) add high-concentration drug solution (e.g., 20 mg/mL) to the prepared temperature responsive polymer aqueous solution; (3) heat the mixture of drug solution and polymer aqueous solution immediately with about 1 min incubation by soaking into a water bath or other ways (temperature higher than LCST, such as 60 °C); (4) slowly cool down the mixture to room temperature and then filter the mixture to remove non-entrapped hydrophobic drug. The process of preparing drug-loaded nanoparticles from the temperature-responsive polymer aqueous solution is called the quick heating method. For example, Jiang et al. reported that PTX was successfully loaded into micelles formed by temperature-responsive PAsPAM derivatives through quick heating method, avoiding the use of toxic organic solvents [35,96,97]. Using temperature-responsive PEGylation phe-g-PHPA as drug carrier, the PTX-loading capacity was high, up to 29%, with a high loading efficiency of 99% [97]. Using pH-responsive PAsPAM derivatives, the drug also can be loaded into micelles by quickly changing the pH of the polymer aqueous solution from below to above  $pH_t$ . For instance, the aqueous mixture containing pH-responsive phe/DEAE-g-PHPA-g-mPEG and doxorubicin hydrochloride was prepared at pH value below  $pH_t$  first (pH = 4.0), and then a certain volume of weakly alkaline PBS solution (pH = 8.2) was added to adjust the pH above  $pH_t$ . After filtering to remove non-entrapped DOX, DOX-loaded micelles prepared with pH-responsive phe/DEAE-g-PHPA-g-mPEG were obtained [34]. Therefore, temperature-responsive and pH-responsive PAsPAM derivatives play an important role in the drug loading process because they can achieve a rapid drug loading procedure (Figure 3).



**Figure 3.** Typical drug loading procedures of various stimuli-responsive PAsPAM derivatives.

Hydrogels are also an important class of polymers that can be used as drug carriers. Cross-linking is a well-known strategy for preparing drug-loaded hydrogels or nanogels. The cross-linking reaction can occur between two polymers containing reactive functional groups [13,69,81], or between polymer and small molecule cross-linker [98–101]. Of course,

enzymes can also be used as cross-linker to promote the cross-linking reaction of PASPAM derivatives for the preparation of nanogels [102]. For PASPAM derivatives without stimuli-responsiveness, another option is the precipitation of the polymer in physiological medium to form gel-like depot. The drug and polymer can be dissolved in an organic solvent with high miscibility with water, and then the resulting mixture solution was slowly injected into physiological medium to allow the formation of drug-loaded hydrogels. According to this strategy, Fiorica et al. successfully prepared a sulpiride-loaded gel-like depot using PASPAM-poly lactide copolymer, in which *N*-methyl-2-pyrrolidone was employed to dissolve sulpiride (an antipsychotic drug) and PASPAM-poly lactide copolymer due to its low systemic toxicity [14]. This sulpiride-loaded depot can prolong a sustained release of sulpiride *in vitro* for about one week.

For PASPAM derivatives that respond to stimuli, even if organic solvents are not used, drug-loaded hydrogels can also be prepared, which will reduce the potential risks caused by the use of organic solvents. For instance, Cao et al. successfully prepared ketoprofen-loaded hydrogels by irradiation of the mixture of light-responsive PASP-Hy-AC aqueous solution and ketoprofen (a nonsteroidal anti-inflammatory drug) under long-wavelength UV for 1 h, and the obtained ketoprofen-loaded hydrogel exhibited the sustained ketoprofen release for about 50 h [95]. Redox-responsive PASPAM derivatives containing thiol groups can also be used for drug-loading. For example, PASP-CEA was dissolved in the PBS containing ofloxacin (a second-generation fluoroquinolone antibiotic), and then oxidant solution was added to ofloxacin/PASP-CEA PBS solution to form ofloxacin-loaded hydrogel via the oxidation of the thiol groups [83]. Moreover, some temperature-responsive polymer aqueous solutions can turn into a gel state in a short time (a few minutes) when heated. Thus, the formation of drug-loaded hydrogels based on temperature-responsive polymers, such as chitosan/ $\beta$ -glycerophosphate [103,104] and Pluronic F127 [105], can be triggered by the temperature difference between the external environment and the body. However, there is no report on the use of temperature-responsive PASPAM derivatives for preparing drug loaded hydrogels, although there are many reported temperature-responsive PASPAM derivatives.

Polymer-drug conjugates composed of drug molecules covalently linked to polymeric carriers are also an important part of drug delivery systems. Because the drug and the polymer carrier are covalently linked, polymer-drug conjugates generally do not require response to external stimuli to achieve drug loading.

### 3.2. Drug Release

As drug carriers, it is also very important that they can release drugs to the desired locations in response to specific stimuli. Thus, stimuli-responsive polymers also play a key role in the drug release process. For drug release, pH and redox are the most investigated triggers due to the acidic environment around the tumor (pH range of 5.5–7.0) [106] and in the endosome (pH  $\approx$  5.0) [107] and the presence of reduced glutathione (GSH) [108].

Although polymer-drug conjugates are usually not required to respond to external stimuli during drug loading, they are often required to respond to external stimuli during the drug release process to deliver the drug to the desired location. NC-6300, which has already undergone Phase I (19 subjects) [109] and Phase 1b (29 subjects) [110] clinical trials, is the most famous stimuli-responsive polymer-drug conjugate based on PASPAM. The EPI conjugated with hydrazone linkage in NC-6300 can be released under acidic environment around tumors, so it exhibits selective tumor accumulation. Clinical trial results showed that NC-6300 was well tolerated in patients with various solid tumors including urothelial carcinoma, breast cancer, cholangiocarcinoma and leiomyosarcom, and its toxicity was lower than conventional EPI formulations. The maximum tolerated dose and recommended phase 2 dose of NC-6300 obtained in Phase 1b were 185 mg/m<sup>2</sup> and 150 mg/m<sup>2</sup>, respectively, which are higher than conventional EPI dose. Besides EPI, DOX was another widely investigated anti-tumor drug in polymer-drug conjugates as a drug delivery system. For instance, Lim et al. conjugated DOX to alendronate (ALN)-modified

PASPAM derivatives with hydrazone linkages to obtain polymer-DOX conjugate ALN-PEG/C18/HYD-DOX-g-PASPAM, and prepared bone targeting nanoparticles. Results showed that 75% of the conjugated DOX was released from the prepared polymer-DOX conjugate at pH 5.0 due to the cleavage of hydrazone bonds, and the volume of tumor decreased significantly to 1550 mm<sup>3</sup> after treatment with ALN-PEG/C18/HYD-DOX-g-PASPAM (control sample in PBS, 3850 mm<sup>3</sup>) [66]. The pH-controlled release of DOX in other PASPAM-DOX conjugates containing hydrazone linkages were also studied, results showed that the amount of DOX released at pH 5.0 was significantly more than that at pH 7.4 in the same time period [65,67,68]. Chang et al. conjugated PTX to the end group (-NH<sub>2</sub>) of the backbone of PEG-b-PBLA via a reduction-responsive disulfide linkage using 3,3'-dithiodipropionic acid as a coupling agent, and results showed that the release rate of PTX in an environment with 10 mM GSH (mimicking the environment in tumor cells) was significantly higher than that in an environment with 2 μM GSH (mimicking the environment in human blood) [111]. Therefore, the introduction of stimuli-responsive cleavable linkages is a key strategy for the controlled drug release of polymer-drug conjugates.

For nanoparticle drug delivery systems prepared from amphiphilic polymers, the response to environmental stimuli generally can break the hydrophobicity-hydrophilicity balance, thereby disturbing the stability of the drug-loaded nanoparticles to achieve the aim of drug release. Gu et al. prepared DOX-loaded nanoparticles by dialysis using temperature-responsive PAIPAAH as polymeric carrier, and their DOX release profiles evaluated at different temperature showed that the release amount and rate of DOX at 37 °C (human body temperature, 45% within 10 h) were both higher than that at 25 °C (room temperature, 35% after 110 h) [27]. This may be caused by the LCST of PAIPAAH (30 °C) being between room temperature and body temperature. When the environmental temperature is higher than the LCST of PAIPAAH, the formed nanoparticles were disturbed and became unstable due to the weakened hydrogen-bonding or hydrophobic interactions between PAIPAAH and DOX. Thus, temperature can be used as a switch to control the drug release. The response to pH or GSH can also be used to realize the controlled release of drug from nanoparticles. Cai et al. prepared a disulfide cross-linked micellar nanodrug loaded with sorafenib from pH/reduction dual-responsive mPEG-PAsp (MEA&DIP), and realized controlled release of sorafenib using pH and GSH as the switch [112]. Due to the disulfide cross-linking, this sorafenib-loaded nanodrug can remain stable in blood circulation, but it can also rapidly release sorafenib inside cancer cells due to GSH-induced disulfide bond breakage and the protonation of DIP (2-aminoethyl-diisopropylamide) pendants in an acidic environment (leading to a hydrophobic-to-hydrophilic change). Light with specific wavelength also can be used as a stimulus for controlled drug release. After 15 min of irradiation at 365 nm, PTX-loaded nanoparticles containing photo-cleavable o-nitrobenzyl carbonate ester linkages showed an significantly faster PTX release behavior [35].

For hydrogels, drug release generally can be controlled by the degree of swelling, and redox is one of the most investigated triggers due to the reductive environment in tumor tissue (10 mM GSH). Krisch et al. chose fluorescein isothiocyanate-dextran as model drug to investigate the drug release behavior. Results reveal that a reductive environment can promote the release of model drug from S-S cross-linked PASPAM hydrogel, because the degree of swelling increased (10 mM DTT in PBS) due to the cleavage of S-S linkages [81]. Park et al. prepared DOX-loaded PASPAM nanogels, the release rate of DOX in reductive environment was significantly faster than that in non-reductive environment due to the disintegration of nanogels [78].

#### 4. Degradation

The degradability of PASPAM derivatives is based on the hydrolysis process of amide linkages in their backbone. Many studies have confirmed that the backbone of PASPAM derivatives can be degraded by bacteria (i.e., strain KP-2: JCM10638, 15 days) [113,114], enzyme (i.e., hydrolase purified from *Sphingomonas* sp. KT-1, β-amide linkage) [115,116] or activated sludge (28 days) [117,118] after a few days. For PASPAM derivatives that



respond to stimuli, the degradation site and behavior are more diverse. For instance, Juriga et al. [119,120] investigated the degradability of cross-linked PAsPAM hydrogels containing disulfide linkages with different enzymes (trypsin-EDTA, dispase and collagenase I). Collagenase I showed the best effect for the degradation of hydrogel disk, followed by trypsin-EDTA and then dispase, because collagenase I cleaved the disulfide linkages in the hydrogel. Interestingly, the degradation of PAsPAM derivatives containing pH-responsive amino groups can be achieved without enzyme due to the nucleophilic attack of the N in  $\text{CONH}_{\text{side}}$  at the C in  $\text{CONH}_{\text{main}}$  [121], and can be affected by their pendant moieties. Naito et al. [7] reported that the bearing pendants with a longer alkyl spacer (i.e., 3-aminopropylamide (3 carbons), 4-aminobutylamide (4 carbons)) showed more strongly suppressed degradation than that with a shorter alkyl spacer (i.e., 2-aminoethylamide (2 carbons)). Although the degradation of PAsPAM derivatives has been widely investigated, so far there have been no in vivo studies on their degradation.

## 5. Conclusions

With the ever-increasing demand for novel smart materials in biomedical fields, the development of stimuli-responsive PAsPAM derivatives has attracted more and more attention due to their biocompatibility and biodegradability. From a synthesis point of view, most stimuli-responsive PAsPAM derivatives can be easily prepared by introducing functional groups or moieties that respond to various stimuli, but it is worth noting that temperature-responsive PAsPAM derivatives can also be synthesized without introducing specific temperature-responsive moieties. In addition, some temperature-responsive PAsPAM derivatives possess pH responsiveness even without introducing pH-responsive functional groups or pH-cleavable linkages. Thus, the pH response mechanism of temperature-responsive PAsPAM derivatives needs to be studied in depth in the future.

From an application point of view, stimuli-responsive PAsPAM-based drug delivery systems including nanoparticles, hydrogels and polymer-drug conjugates are reviewed in this paper from two aspects: drug loading and controlled release. External stimuli can not only be utilized to achieve convenient drug loading but also controlled drug release. Temperature and pH generally can be used as triggers to realize drug loading for nanoparticles, while thiol groups and light with specific wavelength usually can be used as triggers to prepare drug-loaded hydrogels. Although temperature has been reported to trigger the preparation of drug-loaded hydrogels by other temperature-responsive materials, there is no report on the use of temperature-responsive PAsPAM derivatives to prepare drug-loaded hydrogels. Considering the advantages of the stimuli-triggered drug loading method (avoiding the use of toxic organic solvents and high drug loading efficiency), such studies may be an interesting area in drug delivery systems. For controlled drug release, it can be triggered by most of stimuli, such as temperature, pH, GSH and light. NC-6300, which exhibits selective release of EPI into tumor tissue, is a typical representative of stimuli-responsive polymer-drug conjugates for drug delivery based on PAsPAM derivatives.

In conclusion, stimuli-responsive PAsPAM derivatives hold great prospects as drug carriers. Although some considerable progress has been made, there are still some issues that have not yet been clarified in terms of response mechanisms. Taking account of the growing interest in stimuli-responsive polymers, more interesting stimuli-responsive PAsPAM derivatives may be designed and applied for drug delivery in the future.

**Author Contributions:** Conceptualization, G.Z.; writing—original draft preparation, G.Z., H.Y. and C.B.; writing—review and editing, G.Z., H.Y. and C.B. All authors have read and agreed to the published version of the manuscript.

**Funding:** This work was funded by the Research Foundation of Hubei Provincial Key Laboratory of Green Materials for Light Industry and Collaborative Innovation Center of Green Light-weight Materials and Processing (201907B08).

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Data Availability Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Park, S.B.; Sung, M.H.; Uyama, H.; Han, D.K. Poly(glutamic acid): Production, composites, and medical applications of the next-generation biopolymer. *Prog. Polym. Sci.* **2021**, *113*, 28. [[CrossRef](#)]
2. Patil, N.A.; Kandasubramanian, B. Functionalized polylysine biomaterials for advanced medical applications: A review. *Eur. Polym. J.* **2021**, *146*, 24. [[CrossRef](#)]
3. Aderibigbe, B.A.; Mhlwatika, Z.; Nwamadi, M.; Balogun, M.O.; Matshe, W.M.R. Synthesis, characterization and in vitro analysis of polymer-based conjugates containing dihydrofolate reductase inhibitors. *J. Drug Deliv. Sci. Technol.* **2019**, *50*, 388–401. [[CrossRef](#)]
4. Alven, S.; Aderibigbe, B.A.; Balogun, M.O.; Matshe, W.M.R.; Ray, S.S. Polymer-drug conjugates containing antimalarial drugs and antibiotics. *J. Drug Deliv. Sci. Technol.* **2019**, *53*, 12. [[CrossRef](#)]
5. Craparo, E.F.; Drago, S.E.; Mauro, N.; Giammona, G.; Cavallaro, G. Design of New Polyaspartamide Copolymers for siRNA Delivery in Antiasthmatic Therapy. *Pharmaceutics* **2020**, *12*, 89. [[CrossRef](#)] [[PubMed](#)]
6. Kravicz, M.H.; Balogh, D.T.; Kar, M.; Wedepohl, S.; Bentley, M.; Calderon, M. Influence of Alkyl Chains of Modified Polysuccinimide-Based Polycationic Polymers on Polyplex Formation and Transfection. *Macromol. Biosci.* **2019**, *19*, 14. [[CrossRef](#)]
7. Naito, M.; Otsu, Y.; Kamegawa, R.; Hayashi, K.; Uchida, S.; Kim, H.J.; Miyata, K. Tunable nonenzymatic degradability of N-substituted polyaspartamide main chain by amine protonation and alkyl spacer length in side chains for enhanced messenger RNA transfection efficiency. *Sci. Technol. Adv. Mater.* **2019**, *20*, 105–115. [[CrossRef](#)] [[PubMed](#)]
8. Yan, S.; Chen, S.; Gou, X.; Yang, J.; An, J.; Jin, X.; Yang, Y.-W.; Chen, L.; Gao, H. Biodegradable Supramolecular Materials Based on Cationic Polyaspartamides and Pillar 5 arene for Targeting Gram-Positive Bacteria and Mitigating Antimicrobial Resistance. *Adv. Funct. Mater.* **2019**, *29*, 1904683. [[CrossRef](#)]
9. Fiorica, C.; Palumbo, F.S.; Pitarresi, G.; Giammona, G. Photocrosslinkable polyaspartamide/poly lactide copolymer and its porous scaffolds for chondrocytes. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2017**, *76*, 794–801. [[CrossRef](#)] [[PubMed](#)]
10. Lopukhov, A.V.; Yang, Z.; Haney, M.J.; Bronich, T.K.; Sokolsky-Papkov, M.; Batrakova, E.V.; Klyachko, N.L.; Kabanov, A.V. Mannosylated Cationic Copolymers for Gene Delivery to Macrophages. *Macromol. Biosci.* **2021**, *21*, 2000371. [[CrossRef](#)]
11. Perrone, F.; Craparo, E.F.; Cemazar, M.; Kamensek, U.; Drago, S.E.; Dapas, B.; Scaggiante, B.; Zanconati, F.; Bonazza, D.; Grassi, M.; et al. Targeted delivery of siRNAs against hepatocellular carcinoma-related genes by a galactosylated polyaspartamide copolymer. *J. Control. Release* **2021**, *330*, 1132–1151. [[CrossRef](#)]
12. Craparo, E.F.; Drago, S.E.; Giammona, G.; Cavallaro, G. Production of polymeric micro- and nanostructures with tunable properties as pharmaceutical delivery systems. *Polymer* **2020**, *200*, 9. [[CrossRef](#)]
13. Fiorica, C.; Ventura, C.A.; Pitarresi, G.; Giammona, G. Polyaspartamide based hydrogel with cell recruitment properties for the local administration of hydrophobic anticancer drugs. *React. Funct. Polym.* **2019**, *138*, 9–17. [[CrossRef](#)]
14. Fiorica, C.; Palumbo, F.S.; Pitarresi, G.; Giorgi, M.; Calascibetta, F.; Giammona, G. In-situ forming gel-like depot of a polyaspartamide-poly lactide copolymer for once a week administration of sulphiride. *J. Pharm. Pharmacol.* **2015**, *67*, 78–86. [[CrossRef](#)] [[PubMed](#)]
15. Zhang, Y.; Ma, S.; Liu, X.; Xu, Y.; Zhao, J.; Si, X.; Li, H.; Huang, Z.; Wang, Z.; Tang, Z.; et al. Supramolecular Assembled Programmable Nanomedicine As In Situ Cancer Vaccine for Cancer Immunotherapy. *Adv. Mater.* **2021**, *33*, 2007293. [[CrossRef](#)]
16. Craparo, E.F.; Cabibbo, M.; Conigliaro, A.; Barreca, M.M.; Musumeci, T.; Giammona, G.; Cavallaro, G. Rapamycin-Loaded Polymeric Nanoparticles as an Advanced Formulation for Macrophage Targeting in Atherosclerosis. *Pharmaceutics* **2021**, *13*, 503. [[CrossRef](#)]
17. Negishi, T.; Koizumi, F.; Uchino, H.; Kuroda, J.; Kawaguchi, T.; Naito, S.; Matsumura, Y. NK105, a paclitaxel-incorporating micellar nanoparticle, is a more potent radiosensitising agent compared to free paclitaxel. *Br. J. Cancer* **2006**, *95*, 601–606. [[CrossRef](#)] [[PubMed](#)]
18. Hamaguchi, T.; Matsumura, Y.; Suzuki, M.; Shimizu, K.; Goda, R.; Nakamura, I.; Nakatomi, I.; Yokoyama, M.; Kataoka, K.; Kakizoe, T. NK105, a paclitaxel-incorporating micellar nanoparticle formulation, can extend in vivo antitumour activity and reduce the neurotoxicity of paclitaxel. *Br. J. Cancer* **2005**, *92*, 1240–1246. [[CrossRef](#)]
19. Mukai, H.; Kato, K.; Esaki, T.; Ohsumi, S.; Hozomi, Y.; Matsubara, N.; Hamaguchi, T.; Matsumura, Y.; Goda, R.; Hirai, T.; et al. Phase I study of NK105, a nanomicellar paclitaxel formulation, administered on a weekly schedule in patients with solid tumors. *Investig. New Drugs* **2016**, *34*, 750–759. [[CrossRef](#)]
20. Fujiwara, Y.; Mukai, H.; Saeki, T.; Ro, J.; Lin, Y.-C.; Nagai, S.E.; Lee, K.S.; Watanabe, J.; Ohtani, S.; Kim, S.B.; et al. A multi-national, randomised, open-label, parallel, phase III non-inferiority study comparing NK105 and paclitaxel in metastatic or recurrent breast cancer patients. *Br. J. Cancer* **2019**, *120*, 475–480. [[CrossRef](#)] [[PubMed](#)]

21. Matsumura, Y.; Hamaguchi, T.; Ura, T.; Muro, K.; Yamada, Y.; Shimada, Y.; Shirao, K.; Okusaka, T.; Ueno, H.; Ikeda, M.; et al. Phase I clinical trial and pharmacokinetic evaluation of NK911, a micelle-encapsulated doxorubicin. *Br. J. Cancer* **2004**, *91*, 1775–1781. [[CrossRef](#)]
22. Harada, M.; Bobe, I.; Saito, H.; Shibata, N.; Tanaka, R.; Hayashi, T.; Kato, Y. Improved anti-tumor activity of stabilized anthracycline polymeric micelle formulation, NC-6300. *Cancer Sci.* **2011**, *102*, 192–199. [[CrossRef](#)]
23. Takahashi, A.; Yamamoto, Y.; Yasunaga, M.; Koga, Y.; Kuroda, J.-i.; Takigahira, M.; Harada, M.; Saito, H.; Hayashi, T.; Kato, Y.; et al. NC-6300, an epirubicin-incorporating micelle, extends the antitumor effect and reduces the cardiotoxicity of epirubicin. *Cancer Sci.* **2013**, *104*, 920–925. [[CrossRef](#)]
24. Wei, M.; Gao, Y.; Li, X.; Serpe, M.J. Stimuli-responsive polymers and their applications. *Polym. Chem.* **2017**, *8*, 127–143. [[CrossRef](#)]
25. Adelnia, H.; Tran, H.D.N.; Little, P.J.; Blakey, I.; Ta, H.T. Poly(aspartic acid) in Biomedical Applications: From Polymerization, Modification, Properties, Degradation, and Biocompatibility to Applications. *ACS Biomater. Sci. Eng.* **2021**, *7*, 2083–2105. [[CrossRef](#)]
26. Jalalvandi, E.; Shavandi, A. Polysuccinimide and its derivatives: Degradable and water soluble polymers (review). *Eur. Polym. J.* **2018**, *109*, 43–54. [[CrossRef](#)]
27. Gu, X.; Wang, J.; Liu, X.; Zhao, D.; Wang, Y.; Gao, H.; Wu, G. Temperature-responsive drug delivery systems based on polyaspartamides with isopropylamine pendant groups. *Soft Matter* **2013**, *9*, 7267–7273. [[CrossRef](#)]
28. Vega-Chacon, J.; Piazza, R.D.; Costa Marques, R.F.; Elaissari, A.; Jafellici, M. The influence of pH, hydrolysis and degree of substitution on the temperature-sensitive properties of polyaspartamides. *Polym. Int.* **2019**, *68*, 88–93. [[CrossRef](#)]
29. Liu, N.; Li, B.; Gong, C.; Liu, Y.; Wang, Y.; Wu, G. A pH- and thermo-responsive poly(amino acid)-based drug delivery system. *Colloids Surf. B Biointerfaces* **2015**, *136*, 562–569. [[CrossRef](#)]
30. Moon, J.R.; Park, Y.H.; Kim, J.-H. Synthesis and characterization of novel thermo- and pH-responsive copolymers based on amphiphilic polyaspartamides. *J. Appl. Polym. Sci.* **2009**, *111*, 998–1004. [[CrossRef](#)]
31. Moon, J.; Kim, J.-H. Preparation of Biodegradable Thermo-responsive Polyaspartamides with N-Isopropylamine Pendant Groups (I). *Bull. Korean Chem. Soc.* **2006**, *27*, 1981–1984. [[CrossRef](#)]
32. Tachibana, Y.; Kurisawa, M.; Uyama, H.; Kakuchi, T.; Kobayashi, S. Biodegradable thermoresponsive poly(amino acid)s. *Chem. Commun.* **2003**, 106–107. [[CrossRef](#)] [[PubMed](#)]
33. Zhang, G.; Zhou, J.; Wang, P. A novel cross-linkable, polyaspartamide derivative-containing cinnamoyl groups with temperature and pH dual stimuli-responsiveness. *Iran. Polym. J.* **2019**, *28*, 157–166. [[CrossRef](#)]
34. Ma, Y.; Zhang, G.; Li, L.; Yu, H.; Liu, J.; Wang, C.; Chu, Y.; Zhuo, R.; Jiang, X. Temperature and pH Dual-Sensitive Polyaspartamide Derivatives for Antitumor Drug Delivery. *J. Polym. Sci. Part A Polym. Chem.* **2016**, *54*, 879–888. [[CrossRef](#)]
35. Du, X.; Jiang, Y.; Zhuo, R.; Jiang, X. Thermosensitive and Photocleavable Polyaspartamide Derivatives for Drug Delivery. *J. Polym. Sci. Part A Polym. Chem.* **2016**, *54*, 2855–2863. [[CrossRef](#)]
36. Hsu, S.-P.; Chu, I.-M.; Yang, J.-D. Thermo- and pH-Responsive Polymersomes of Poly( $\alpha,\beta$ -N-substituted-DL-aspartamide)s. *J. Appl. Polym. Sci.* **2012**, *125*, 133–144. [[CrossRef](#)]
37. Ma, Y.; Jiang, X.; Zhuo, R. Biodegradable and thermosensitive polyaspartamide derivatives bearing aromatic structures. *Mater. Lett.* **2014**, *121*, 78–80. [[CrossRef](#)]
38. Zhang, G.; Zhang, Y.; Chu, Y.; Ma, Y.; Zhuo, R.; Jiang, X. Facile synthesis of thermosensitive functional polyaspartamide derivatives by click chemistry. *J. Polym. Sci. Part A Polym. Chem.* **2015**, *53*, 1296–1303. [[CrossRef](#)]
39. Czaderna-Lekka, A.; Kozanecki, M.; Matusiak, M.; Kadlubowski, S. Phase transitions of poly(oligo(ethylene glycol) methyl ether methacrylate)-water systems. *Polymer* **2021**, *212*, 12. [[CrossRef](#)]
40. Zuppari, F.; Malinconico, M.; D'Agosto, F.; D'Ayala, G.G.; Cerruti, P. Well-Defined Thermo-Responsive Copolymers Based on Oligo(Ethylene Glycol) Methacrylate and Pentafluorostyrene for the Removal of Organic Dyes from Water. *Nanomaterials* **2020**, *10*, 1779. [[CrossRef](#)]
41. Zhou, C.P.; Shi, Z.W.; Xu, F.; Ling, Y.; Tang, H.Y. Preparation and properties of thermo- and pH-responsive polypeptide bearing OEG and aldehyde pendants. *Colloid Polym. Sci.* **2020**, *298*, 1293–1302. [[CrossRef](#)]
42. Cheng, Y.; He, C.; Xiao, C.; Ding, J.; Zhuang, X.; Chen, X. Versatile synthesis of temperature-sensitive polypeptides by click grafting of oligo(ethylene glycol). *Polym. Chem.* **2011**, *2*, 2627–2634. [[CrossRef](#)]
43. Zhang, G.; Bao, C.; Yi, H. Investigation of the Temperature Responsive Behaviors of Novel Polyaspartamide Derivatives Bearing Alkyl Ether-type Pendants. *Arab. J. Chem.* **2021**, *14*, 103287. [[CrossRef](#)]
44. Lagant, P.; Vergoten, G.; Loucheux, C.; Fleury, G. Study of Poly(L-aspartic acid). I. Laser Raman Spectrometry. *Polym. J.* **1979**, *11*, 345–351. [[CrossRef](#)]
45. Németh, C.; Gyarmati, B.; Abdullin, T.; László, K.; Szilágyi, A. Poly(aspartic acid) with adjustable pH-dependent solubility. *Acta Biomater.* **2017**, *49*, 486–494. [[CrossRef](#)] [[PubMed](#)]
46. Kim, J.H.; Oh, Y.T.; Lee, K.S.; Yun, J.M.; Park, B.T.; Oh, K.T. Development of a pH-sensitive polymer using poly(aspartic acid-graft-imidazole)-block-poly(ethylene glycol) for acidic pH targeting systems. *Macromol. Res.* **2011**, *19*, 453–460. [[CrossRef](#)]
47. Chu, Y.; Yu, H.; Ma, Y.; Zhang, Y.; Chen, W.; Zhang, G.; Wei, H.; Zhang, X.; Zhuo, R.; Jiang, X. Synthesis and characterization of biodegradable pH and reduction dual-sensitive polymeric micelles for doxorubicin delivery. *J. Polym. Sci. Part A Polym. Chem.* **2014**, *52*, 1771–1780. [[CrossRef](#)]



48. Gong, C.; Shan, M.; Li, B.; Wu, G. A pH and redox dual stimuli-responsive poly(amino acid) derivative for controlled drug release. *Colloids Surf. B Biointerfaces* **2016**, *146*, 396–405. [[CrossRef](#)]
49. Kim, J.; Kim, J.-H.; Kim, D. pH-Sensitive Amphiphilic Biodegradable Graft Co-Polymer Aggregates Based on Polyaspartamide for Intracellular Delivery. *J. Biomater. Sci. Polym. Ed.* **2012**, *23*, 1255–1269. [[CrossRef](#)]
50. Wang, L.; Luan, J.; Du, L.; Li, L.; Liu, J.; Liu, Z.; Zhuo, R. Polyaspartamide-based multi-responsive micelle with sheddable shell for antitumor drug delivery. *RSC Adv.* **2016**, *6*, 111161–111169. [[CrossRef](#)]
51. Seo, K.; Chung, S.W.; Byun, Y.; Kim, D. Paclitaxel loaded nano-aggregates based on pH sensitive polyaspartamide amphiphilic graft copolymers. *Int. J. Pharm.* **2012**, *424*, 26–32. [[CrossRef](#)]
52. Lin, W.; Kim, D. pH-Sensitive Micelles with Cross-Linked Cores Formed from Polyaspartamide Derivatives for Drug Delivery. *Langmuir* **2011**, *27*, 12090–12097. [[CrossRef](#)]
53. Seo, K.; Kim, D. pH-dependent hemolysis of biocompatible imidazole-grafted polyaspartamide derivatives. *Acta Biomater.* **2010**, *6*, 2157–2164. [[CrossRef](#)] [[PubMed](#)]
54. Ngoc Bich, T.; Kim, J.Y.; Kim, Y.-C.; Kim, Y.J.; Kim, J.-H. CO<sub>2</sub>-responsive swelling behavior and metal-ion adsorption properties in novel histamine-conjugated polyaspartamide hydrogel. *J. Appl. Polym. Sci.* **2016**, *133*, 43305. [[CrossRef](#)]
55. Moon, J.R.; Jeon, Y.S.; Zrinyi, M.; Kim, J.-H. pH-Responsive PEGylated nanoparticles based on amphiphilic polyaspartamide: Preparation, physicochemical characterization and in vitro evaluation. *Polym. Int.* **2013**, *62*, 1218–1224. [[CrossRef](#)]
56. Gu, X.; Wang, J.; Wang, Y.; Wang, Y.; Gao, H.; Wu, G. Layer-by-layer assembled polyaspartamide nanocapsules for pH-responsive protein delivery. *Colloids Surf. B Biointerfaces* **2013**, *108*, 205–211. [[CrossRef](#)] [[PubMed](#)]
57. Moon, J.R.; Kim, M.W.; Kim, D.; Jeong, J.H.; Kim, J.-H. Synthesis and self-assembly behavior of novel polyaspartamide derivatives for anti-tumor drug delivery. *Colloid Polym. Sci.* **2011**, *289*, 63–71. [[CrossRef](#)]
58. Moon, J.R.; Kim, J.-H. Biodegradable stimuli-responsive hydrogels based on amphiphilic polyaspartamides with tertiary amine pendent groups. *Polym. Int.* **2010**, *59*, 630–636. [[CrossRef](#)]
59. Huang, X.; Jiang, X.; Zhuo, R. Microwave-assisted solid-phase synthesis of pH-responsive polyaspartamide derivatives. *Carbohydr. Polym.* **2012**, *89*, 788–794. [[CrossRef](#)] [[PubMed](#)]
60. Le, T.T.; Kim, D. Folate-PEG/Hyd-curcumin/C18-g-PSI micelles for site specific delivery of curcumin to colon cancer cells via Wnt/beta-catenin signaling pathway. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2019**, *101*, 464–471. [[CrossRef](#)]
61. Lu, C.; Wang, X.; Wu, G.; Wang, J.; Wang, Y.; Gao, H.; Ma, J. An injectable and biodegradable hydrogel based on poly(alpha,beta-aspartic acid) derivatives for localized drug delivery. *J. Biomed. Mater. Res. Part A* **2014**, *102*, 628–638. [[CrossRef](#)]
62. Lim, C.; Cho, E.-B.; Kim, D. pH-triggered intracellular release of doxorubicin from polyaspartamide-encapsulated mesoporous silica nanoparticles. *Korean J. Chem. Eng.* **2019**, *36*, 166–172. [[CrossRef](#)]
63. Yu, H.; Sun, J.; Zhang, Y.; Zhang, G.; Chu, Y.; Zhuo, R.; Jiang, X. pH- and beta-Cyclodextrin-Responsive Micelles Based on Polyaspartamide Derivatives as Drug Carrier. *J. Polym. Sci. Part A Polym. Chem.* **2015**, *53*, 1387–1395. [[CrossRef](#)]
64. Sonawane, S.J.; Kalthapure, R.S.; Govender, T. Hydrazone linkages in pH responsive drug delivery systems. *Eur. J. Pharm. Sci.* **2017**, *99*, 45–65. [[CrossRef](#)]
65. Liu, N.; Han, J.M.; Zhang, X.C.; Yang, Y.; Liu, Y.; Wang, Y.M.; Wu, G.L. pH-responsive zwitterionic polypeptide as a platform for anti-tumor drug delivery. *Colloids Surf. B Biointerfaces* **2016**, *145*, 401–409. [[CrossRef](#)]
66. Lim, C.W.; Kim, D. Bone targeting nano-aggregates prepared from self-assembled polyaspartamide graft copolymers for pH sensitive DOX delivery. *Biomater. Sci.* **2021**, *9*, 1660–1667. [[CrossRef](#)]
67. Van Tran Thi, T.; Lim, C.W.; Park, J.H.; Ahn, C.-H.; Kim, D. Self-assembled nanoaggregates based on polyaspartamide graft copolymers for pH-controlled release of doxorubicin. *J. Mater. Chem. B* **2015**, *3*, 2978–2985. [[CrossRef](#)]
68. Nguyen, M.P.; Thuy, V.T.; Kim, D. Integration of iron oxide nanoparticles and polyaspartamide biopolymer for MRI image contrast enhancement and an efficient drug-delivery system in cancer therapy. *Nanotechnology* **2020**, *31*, 11. [[CrossRef](#)]
69. Jang, J.; Cha, C. Multivalent Polyaspartamide Cross-Linker for Engineering Cell-Responsive Hydrogels with Degradation Behavior and Tunable Physical Properties. *Biomacromolecules* **2018**, *19*, 691–700. [[CrossRef](#)] [[PubMed](#)]
70. Sanjoh, M.; Hiki, S.; Lee, Y.; Oba, M.; Miyata, K.; Ishii, T.; Kataoka, K. pDNA/poly(L-lysine) Polyplexes Functionalized with a pH-Sensitive Charge-Conversional Poly(aspartamide) Derivative for Controlled Gene Delivery to Human Umbilical Vein Endothelial Cells. *Macromol. Rapid Commun.* **2010**, *31*, 1181–1186. [[CrossRef](#)] [[PubMed](#)]
71. Maeda, Y.; Pittella, F.; Nomoto, T.; Takemoto, H.; Nishiyama, N.; Miyata, K.; Kataoka, K. Fine-Tuning of Charge-Conversion Polymer Structure for Efficient Endosomal Escape of siRNA-Loaded Calcium Phosphate Hybrid Micelles. *Macromol. Rapid Commun.* **2014**, *35*, 1211–1215. [[CrossRef](#)]
72. Cao, L.; Xiao, Y.; Lu, W.; Liu, S.; Gan, L.; Yu, J.; Huang, J. Nanomicelle drug with acid-triggered doxorubicin release and enhanced cellular uptake ability based on mPEG-graft-poly(N-(2-aminoethyl)-L-aspartamide)-hexahydrophthalic acid copolymers. *J. Biomater. Appl.* **2018**, *32*, 826–838. [[CrossRef](#)] [[PubMed](#)]
73. Hill, M.R.; MacKrell, E.J.; Forsthoefel, C.P.; Jensen, S.P.; Chen, M.; Moore, G.A.; He, Z.L.; Sumerlin, B.S. Biodegradable and pH-Responsive Nanoparticles Designed for Site-Specific Delivery in Agriculture. *Biomacromolecules* **2015**, *16*, 1276–1282. [[CrossRef](#)] [[PubMed](#)]
74. Zhang, Y.; Xiao, Y.; Huang, Y.; He, Y.; Xu, Y.; Lu, W.; Yu, J. Poly(ethylene glycol) shell-sheddable TAT-modified core cross-linked nano-micelles: TAT-enhanced cellular uptake and lysosomal pH-triggered doxorubicin release. *Colloids Surf. B Biointerfaces* **2020**, *188*, 110772. [[CrossRef](#)] [[PubMed](#)]

75. Liu, J.; Xu, Y.; Yang, Q.; Li, C.; Hennink, W.E.; Zhuo, R.; Jiang, X. Reduction biodegradable brushed PDMAEMA derivatives synthesized by atom transfer radical polymerization and click chemistry for gene delivery. *Acta Biomater.* **2013**, *9*, 7758–7766. [[CrossRef](#)]
76. Zhang, G.; Liu, J.; Yang, Q.; Zhuo, R.; Jiang, X. Disulfide-Containing Brushed Polyethylenimine Derivative Synthesized by Click Chemistry for Nonviral Gene Delivery. *Bioconjug. Chem.* **2012**, *23*, 1290–1299. [[CrossRef](#)]
77. Zhang, Y.; Jiang, Q.; Bi, B.; Xu, L.; Liu, J.; Zhuo, R.; Jiang, X. A bioreducible supramolecular nanoparticle gene delivery system based on cyclodextrin-conjugated polyaspartamide and adamantyl-terminated polyethylenimine. *J. Mater. Chem. B* **2018**, *6*, 797–808. [[CrossRef](#)]
78. Park, C.W.; Yang, H.-M.; Woo, M.-A.; Lee, K.S.; Kim, J.-D. Completely disintegrable redox-responsive poly(amino acid) nanogels for intracellular drug delivery. *J. Ind. Eng. Chem.* **2017**, *45*, 182–188. [[CrossRef](#)]
79. Birhan, Y.S.; Darge, H.F.; Hanurry, E.Y.; Andrgie, A.T.; Mekonnen, T.W.; Chou, H.-Y.; Lai, J.-Y.; Tsai, H.-C. Fabrication of Core Crosslinked Polymeric Micelles as Nanocarriers for Doxorubicin Delivery: Self-Assembly, In Situ Diselenide Metathesis and Redox-Responsive Drug Release. *Pharmaceutics* **2020**, *12*, 580. [[CrossRef](#)]
80. Gyarmati, B.; Vajna, B.; Nemethy, A.; Laszlo, K.; Szilágyi, A. Redox- and pH-Responsive Cysteamine-Modified Poly(aspartic acid) Showing a Reversible SolGel Transition. *Macromol. Biosci.* **2013**, *13*, 633–640. [[CrossRef](#)]
81. Krisch, E.; Gyarmati, B.; Barczikai, D.; Lapeyre, V.; Szilágyi, B.Á.; Ravaine, V.; Szilágyi, A. Poly(aspartic acid) hydrogels showing reversible volume change upon redox stimulus. *Eur. Polym. J.* **2018**, *105*, 459–468. [[CrossRef](#)]
82. Krisch, E.; Messenger, L.; Gyarmati, B.; Ravaine, V.; Szilágyi, A. Redox- and pH-Responsive Nanogels Based on Thiolated Poly(aspartic acid). *Macromol. Mater. Eng.* **2016**, *301*, 260–266. [[CrossRef](#)]
83. Szilágyi, B.Á.; Gyarmati, B.; Horvát, G.; Laki, Á.; Budai-Szűcs, M.; Csányi, E.; Sandri, G.; Bonferoni, M.C.; Szilágyi, A. The effect of thiol content on the gelation and mucoadhesion of thiolated poly(aspartic acid). *Polym. Int.* **2017**, *66*, 1538–1545. [[CrossRef](#)]
84. Park, C.W.; Yang, H.-M.; Lee, K.S.; Kim, J.-D. Disulfide and  $\beta$ -sheet stabilized poly(amino acid) nanovesicles for intracellular drug delivery. *J. Ind. Eng. Chem.* **2017**, *56*, 277–284. [[CrossRef](#)]
85. Gyarmati, B.; Mammadova, A.; Barczikai, D.; Stankovits, G.; Misra, A.; Alavijeh, M.S.; Varga, Z.; László, K.; Szilágyi, A. Side group ratio as a novel means to tune the hydrolytic degradation of thiolated and disulfide cross-linked polyaspartamides. *Polym. Degrad. Stab.* **2021**, *188*, 109577. [[CrossRef](#)]
86. Budai-Szűcs, M.; Kiss, E.L.; Szilágyi, B.Á.; Szilágyi, A.; Gyarmati, B.; Berkó, S.; Kovács, A.; Horvát, G.; Aigner, Z.; Soós, J.; et al. Mucoadhesive Cyclodextrin-Modified Thiolated Poly(aspartic acid) as a Potential Ophthalmic Drug Delivery System. *Polymers* **2018**, *10*, 199. [[CrossRef](#)] [[PubMed](#)]
87. Wang, K.; Chen, S.; Zhang, W. A New Family of Thermo-, pH-, and CO<sub>2</sub>-Responsive Homopolymers of Poly[Oligo(ethylene glycol) (N-dialkylamino) methacrylate]s. *Macromolecules* **2017**, *50*, 4686–4698. [[CrossRef](#)]
88. Eskandari, P.; Roghani-Mamaqani, H.; Salami-Kalajahi, M.; Abousalman-Rezvani, Z. Modification of cellulose nanocrystal with dual temperature- and CO<sub>2</sub>-responsive block copolymers for ion adsorption applications. *J. Mol. Liq.* **2020**, *310*, 113234. [[CrossRef](#)]
89. Furusho, Y.; Endo, T. Reversible capture and release of carbon dioxide by binary system of polyamidine and polyethylene glycol. *Polym. Bull.* **2017**, *74*, 1207–1219. [[CrossRef](#)]
90. Schattling, P.; Pollmann, I.; Theato, P. Synthesis of CO<sub>2</sub>-responsive polymers by post-polymerization modification. *React. Funct. Polym.* **2014**, *75*, 16–21. [[CrossRef](#)]
91. Bich Ngoc, T.; Bui, Q.T.; Jeon, Y.S.; Park, H.S.; Kim, J.-H. Preparation and characterization of CO<sub>2</sub>-responsive poly(amino acid) derivatives with guanidine group. *Polym. Bull.* **2015**, *72*, 2605–2620. [[CrossRef](#)]
92. Wei, J.; Sun, J.; Yang, X.; Ji, S.; Wei, Y.; Li, Z. Self-crosslinking assemblies with tunable nanostructures from photoresponsive polypeptoid-based block copolymers. *Polym. Chem.* **2020**, *11*, 337–343. [[CrossRef](#)]
93. Jia, F.; Wang, Y.; Wang, H.; Jin, Q.; Cai, T.; Chen, Y.; Ji, J. Light cross-linkable and pH de-cross-linkable drug nanocarriers for intracellular drug delivery. *Polym. Chem.* **2015**, *6*, 2069–2075. [[CrossRef](#)]
94. Park, J.H.; Moon, J.R.; Hong, K.H.; Kim, J.-H. Photo-crosslinked polyaspartamide hybrid gel containing thermo-responsive Pluronic triblock copolymer. *J. Polym. Res.* **2011**, *18*, 273–278. [[CrossRef](#)]
95. Cao, H.; Ma, X.Y.; Sun, S.H.; Su, H.J.; Tan, T.W. A new photocrosslinkable hydrogel based on a derivative of polyaspartic acid for the controlled release of ketoprofen. *Polym. Bull.* **2010**, *64*, 623–632. [[CrossRef](#)]
96. Ma, Y.; Jiang, X.; Zhuo, R. Biodegradable and thermosensitive micelles of amphiphilic polyaspartamide derivatives containing aromatic groups for drug delivery. *J. Polym. Sci. Part A Polym. Chem.* **2013**, *51*, 3917–3924. [[CrossRef](#)]
97. Zhang, G.; Jiang, X. Temperature Responsive Nanoparticles Based on PEGylated Polyaspartamide Derivatives for Drug Delivery. *Polymers* **2019**, *11*, 316. [[CrossRef](#)] [[PubMed](#)]
98. Dang, Q.D.; Park, J.H.; Bhang, S.H.; Kim, J.H. Synthesis and characterization of novel multi-hydroxy polyaspartamide derivative and its crosslinked hydrogels. *React. Funct. Polym.* **2020**, *147*, 9. [[CrossRef](#)]
99. Gong, C.; Lu, C.C.; Li, B.Q.; Shan, M.; Wu, G.L. Injectable dopamine-modified poly(alpha,beta-aspartic acid) nanocomposite hydrogel as bioadhesive drug delivery system. *J. Biomed. Mater. Res. Part A* **2017**, *105*, 1000–1008. [[CrossRef](#)]
100. Wang, B.; Jeon, Y.S.; Park, H.S.; Kim, J.-H. Self-healable mussel-mimetic nanocomposite hydrogel based on catechol-containing polyaspartamide and graphene oxide. *Mater. Sci. Eng. C Mater. Biol. Appl.* **2016**, *69*, 160–170. [[CrossRef](#)]
101. Nguyen, H.L.N.; Kim, J.H. Stimuli-sensitive complexation and the strongly adhesive antibacterial gel from biocompatible PolyAspAm(EA/EDA) and tannic acid. *Int. J. Polym. Mater. Polym. Biomater.* **2020**, *8*. [[CrossRef](#)]

102. Hladysh, S.; Oleshchuk, D.; Dvorakova, J.; Golunova, A.; Salek, P.; Panek, J.; Janouskova, O.; Kankova, D.; Pavlova, E.; Proks, V. Zwitterionic polyaspartamides based on L-lysine side-chain moieties: Synthesis, nonfouling properties and direct/indirect nanogel preparation. *Eur. Polym. J.* **2021**, *148*, 110347. [[CrossRef](#)]
103. Han, X.; Meng, X.; Wu, Z.; Wu, Z.; Qi, X. Dynamic imine bond cross-linked self-healing thermosensitive hydrogels for sustained anticancer therapy via intratumoral injection. *Mater. Sci. Eng. C* **2018**, *93*, 1064–1072. [[CrossRef](#)]
104. Kim, S.; Nishimoto, S.K.; Bumgardner, J.D.; Haggard, W.O.; Gaber, M.W.; Yang, Y. A chitosan/ $\beta$ -glycerophosphate thermo-sensitive gel for the delivery of ellagic acid for the treatment of brain cancer. *Biomaterials* **2010**, *31*, 4157–4166. [[CrossRef](#)]
105. Shriky, B.; Kelly, A.; Isreb, M.; Babenko, M.; Mahmoudi, N.; Rogers, S.; Shebanova, O.; Snow, T.; Gough, T. Pluronic F127 thermosensitive injectable smart hydrogels for controlled drug delivery system development. *J. Colloid Interface Sci.* **2020**, *565*, 119–130. [[CrossRef](#)]
106. Justus, C.; Dong, L.; Yang, L. Acidic tumor microenvironment and pH-sensing G protein-coupled receptors. *Front. Physiol.* **2013**, *4*, 354. [[CrossRef](#)] [[PubMed](#)]
107. Lee, R.J.; Wang, S.; Low, P.S. Measurement of endosome pH following folate receptor-mediated endocytosis. *Biochim. Biophys. Acta BBA Mol. Cell Res.* **1996**, *1312*, 237–242. [[CrossRef](#)]
108. Circu, M.L.; Aw, T.Y. Glutathione and modulation of cell apoptosis. *Biochim. Biophys. Acta BBA Mol. Cell Res.* **2012**, *1823*, 1767–1777. [[CrossRef](#)]
109. Mukai, H.; Kogawa, T.; Matsubara, N.; Naito, Y.; Sasaki, M.; Hosono, A. A first-in-human Phase 1 study of epirubicin-conjugated polymer micelles (K-912/NC-6300) in patients with advanced or recurrent solid tumors. *Investig. New Drugs* **2017**, *35*, 307–314. [[CrossRef](#)] [[PubMed](#)]
110. Chawla, S.P.; Goel, S.; Chow, W.; Braiteh, F.; Singh, A.S.; Olson, J.E.G.; Osada, A.; Bobe, I.; Riedel, R.F. A Phase 1b Dose Escalation Trial of NC-6300 (Nanoparticle Epirubicin) in Patients with Advanced Solid Tumors or Advanced, Metastatic, or Unresectable Soft-tissue Sarcoma. *Clin. Cancer Res.* **2020**, *26*, 4225–4232. [[CrossRef](#)] [[PubMed](#)]
111. Chang, S.; Wang, Y.F.; Zhang, T.Y.; Pu, X.H.; Zong, L.L.; Zhu, H.Y.; Zhao, L.L.; Feng, B. Redox-Responsive Disulfide Bond-Bridged mPEG-PBLA Prodrug Micelles for Enhanced Paclitaxel Biosafety and Antitumor Efficacy. *Front. Oncol.* **2019**, *9*, 11. [[CrossRef](#)] [[PubMed](#)]
112. Cai, M.; Li, B.; Lin, L.; Huang, J.; An, Y.; Huang, W.; Zhou, Z.; Wang, Y.; Shuai, X.; Zhu, K. A reduction and pH dual-sensitive nanodrug for targeted theranostics in hepatocellular carcinoma. *Biomater. Sci.* **2020**, *8*, 3485–3499. [[CrossRef](#)]
113. Tabata, K.; Abe, H.; Doi, Y. Microbial Degradation of Poly(aspartic acid) by Two Isolated Strains of *Pedobacter* sp. and *Sphingomonas* sp. *Biomacromolecules* **2000**, *1*, 157–161. [[CrossRef](#)]
114. Drobník, J.; Saudek, V.; Vlasák, J.; Kálal, J. Polyaspartamide—A potential drug carrier. *J. Polym. Sci. Polym. Symp.* **1979**, *66*, 65–74. [[CrossRef](#)]
115. Tabata, K.; Kajiyama, M.; Hiraishi, T.; Abe, H.; Yamato, I.; Doi, Y. Purification and Characterization of Poly(aspartic acid) Hydrolase from *Sphingomonas* sp. KT-1. *Biomacromolecules* **2001**, *2*, 1155–1160. [[CrossRef](#)]
116. Wei, J.; Xue, M.; Li, C.; Cao, H.; Tan, T. Effect of enzyme and mechanical stirring on the degradation of polyaspartic acid hydro-gel. *Prog. Nat. Sci. Mater. Int.* **2015**, *25*, 425–429. [[CrossRef](#)]
117. Nakato, T.; Yoshitake, M.; Matsubara, K.; Tomida, M.; Kakuchi, T. Relationships between Structure and Properties of Poly(aspartic acid)s. *Macromolecules* **1998**, *31*, 2107–2113. [[CrossRef](#)]
118. Alford, D.D.; Wheeler, A.P.; Pettigrew, C.A. Biodegradation of thermally synthesized polyaspartate. *J. Environ. Polym. Degrad.* **1994**, *2*, 225–236. [[CrossRef](#)]
119. Juriga, D.; Zrínyi, M. Biodegradation of Poly(aspartamide) Based Hydrogels. *Macromol. Symp.* **2019**, *385*, 1800194. [[CrossRef](#)]
120. Juriga, D.; Nagy, K.; Jedlovský-Hajdú, A.; Perczel-Kováč, K.; Chen, Y.M.; Varga, G.; Zrínyi, M. Biodegradation and Osteosarcoma Cell Cultivation on Poly(aspartic acid) Based Hydrogels. *ACS Appl. Mater. Interfaces* **2016**, *8*, 23463–23476. [[CrossRef](#)] [[PubMed](#)]
121. Lu, Y.; Chau, M.; Boyle, A.J.; Liu, P.; Niehoff, A.; Weinrich, D.; Reilly, R.M.; Winnik, M.A. Effect of Pendant Group Structure on the Hydrolytic Stability of Polyaspartamide Polymers under Physiological Conditions. *Biomacromolecules* **2012**, *13*, 1296–1306. [[CrossRef](#)] [[PubMed](#)]