

CASE REPORT

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A novel *TAB2* nonsense mutation (p.S149X) causing autosomal dominant congenital heart defects: a case report of a Chinese family

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Abstract

Background: *TAB2* is an activator of MAP 3 K7/TAK1, which is required for the IL-1 induced signal pathway. Microdeletions encompassing *TAB2* have been detected in various patients with congenital heart defects (CHD), indicating that haploinsufficiency of *TAB2* causes CHD. To date, seven variants within *TAB2* were reported associated with CHD, only two of them are nonsense mutations.

Case presentation: Here we describe a three-generation Chinese family that included five CHD patients with heart valvular defects, such as mitral or tricuspid valves prolapse or regurgitation, and aortic valve stenosis or regurgitation. Our proband was a pregnant woman presenting with mitral, tricuspid, and aortic defects; her first child experienced sudden cardiac death at the age of 2 years. Whole-exome sequencing of the proband revealed a novel nonsense variant in *TAB2* (c.C446G, p.S149X), which results in the elimination of the majority of C-terminal amino acids of *TAB2*, including the critical TAK1-binding domain. The variant was identified in five affected patients but not in the eight unaffected family members using Sanger sequencing and was classified as “pathogenic” according to the latest recommendation on sequence variants laid out by the American College of Medical Genetics and Genomics and the Association for Molecular Pathology.

Conclusion: We described a family with CHD caused by a novel *TAB2* nonsense mutation. Our study broadens the mutation spectrum of *TAB2*; to the best of our knowledge, this is the first report of a pathogenic mutation within *TAB2* in a Chinese population.

Keywords: Congenital heart defects, Valvular anomalies, *TAB2*, Whole-exome sequencing

Background

TAB2 is a gene located on chromosome 6q25.1 [OMIM *605101] and encodes the TGF- β -activated kinase 1/ MAP 3 K7 binding protein 2 (*TAB2*). As an adapter protein linking TGF- β -activated kinase 1 (TAK1/MAP 3 K7) and TNF receptor-associated factor 6 (TRAF6), *TAB2* plays an essential role in the activation of JNK/NF- κ B signaling induced by IL-1 [1]. Haploinsufficiency of *TAB2* has been linked to congenital heart defects (CHD)

via mapping of the smallest overlapping region of various 6q25.1 microdeletions in different patients with CHD [2]. Further investigations have shown that *TAB2* is expressed in embryonic cardiac tissues of both humans and zebrafish. Knocking down *TAB2* in zebrafish embryos resulted in delayed epiboly progression, convergent extension defects during gastrulation (at approximately 12 h postfertilization), and severe heart failure 36–48 h postfertilization [2]. These findings indicate that *TAB2* dysfunction causes CHD. Additionally, *TAB2* mutations have also been found in patients with frontometaphyseal dysplasia (FMD), with apparently different phenotypes from CHD [3, 4].

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According to the latest Human Gene Mutation Database (HGMD; <http://www.hgmd.cf.ac.uk/ac/index.php>), 21 deletions encompassing *TAB2* and seven disease-causing variants within the gene have been associated with CHD; only two of them are nonsense mutations. Here we examined a three-generation Chinese family including eight patients with CHD (Fig. 1a; Table 1). Heart valvular defects were detected in most affected patients by standard echocardiography. A novel heterozygous variant (c.C446G) in exon5 of *TAB2* was detected in the proband by Whole-Exome Sequencing (WES) and identified in all the surviving affected family members using Sanger sequencing (Fig. 1b; Table 1). This nonsense variant creates a premature stop codon at the 149th residue of *TAB2* (p.S149X) and removes the majority of amino acids from the protein, including the TAK1-binding domain (TAK1 BD) (Fig. 2) [1]. This variant is classified as “pathogenic” as per the latest recommendation on sequence variants interpretation laid out by the American College of Medical Genetics and Genomics (ACMG) and the Association for Molecular Pathology (AMP) [5]. Our study expands the variant diversity of *TAB2* and provide more clinical symptoms of CHD patients caused by a *TAB2* mutation in a Chinese population.

Case presentation

Clinical information

The study was approved by the institutional review board of Jiangxi maternal and child health hospital, Nanchang, China. All the enrolled subjects provided written informed consent. A family from the Jiangxi province, China, comprising 25 family members (11 males, 12 females, and 2 fetuses) across three generations (Fig. 1a) were included in the study. Our proband (II:5) was a 31-year-old female who was 15 weeks pregnant at her first visit to our hospital. She was slightly tachypneic and reported occasional fatigue since the age of 25 years old. She performed a transthoracic color echocardiogram examination at 30 years old and revealed mild aortic valve stenosis accompanied with mild mitral, tricuspid, and aortic regurgitation. She denied any surgical or pharmaceutical interventions. Her husband was 32 years old (II:6) and declared no cardiac symptoms or family history of heart diseases. Their first child (III:7) was a CHD girl with left and right ventricles dilatation. The girl then experienced sudden cardiac death (SCD) at 2 years of age. The second child (III:8) was detected with no heartbeat at 9 weeks of pregnancy using a Doppler fetal monitor, and subsequently, the pregnancy was

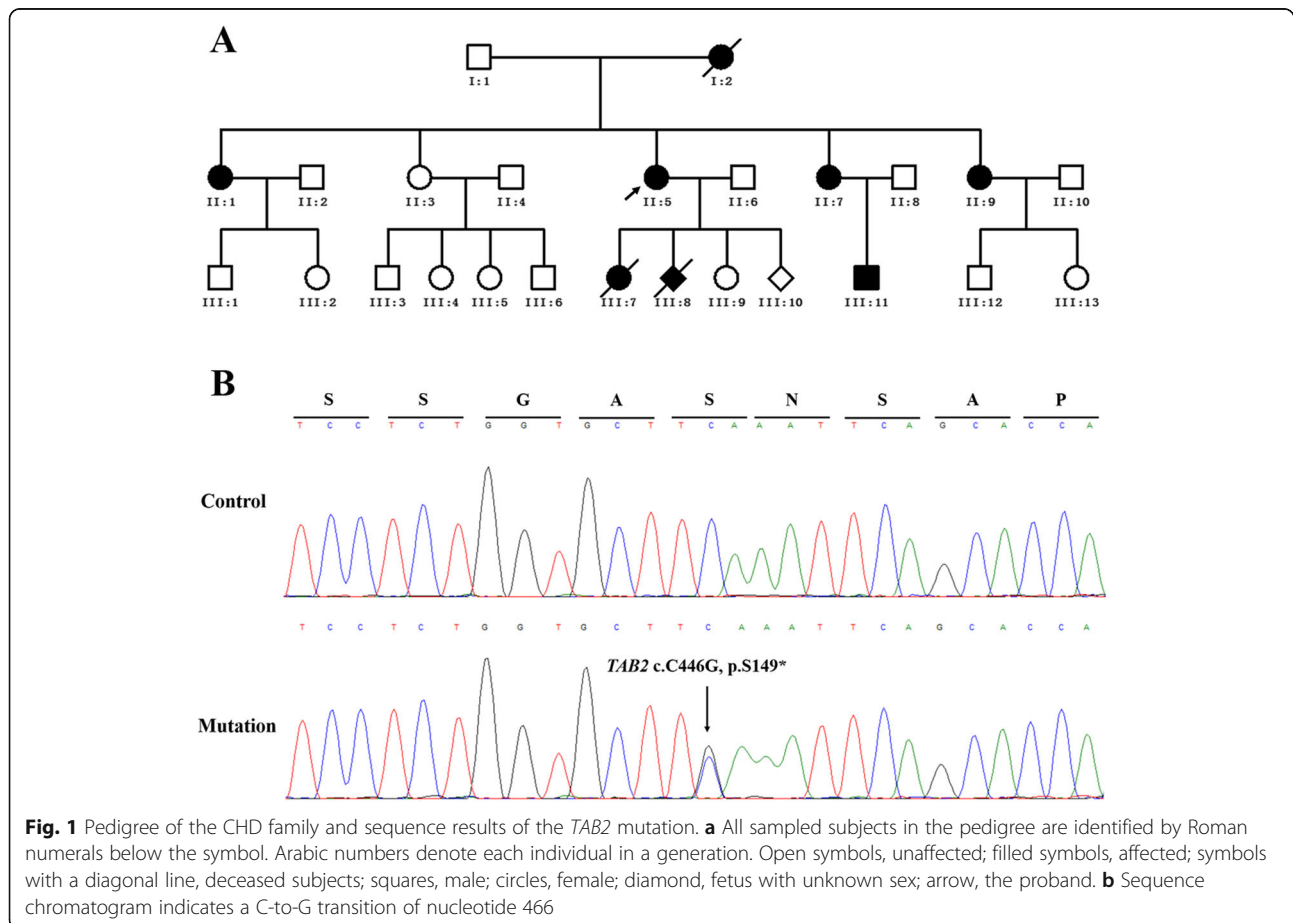


Table 1 Summary of the CHD family

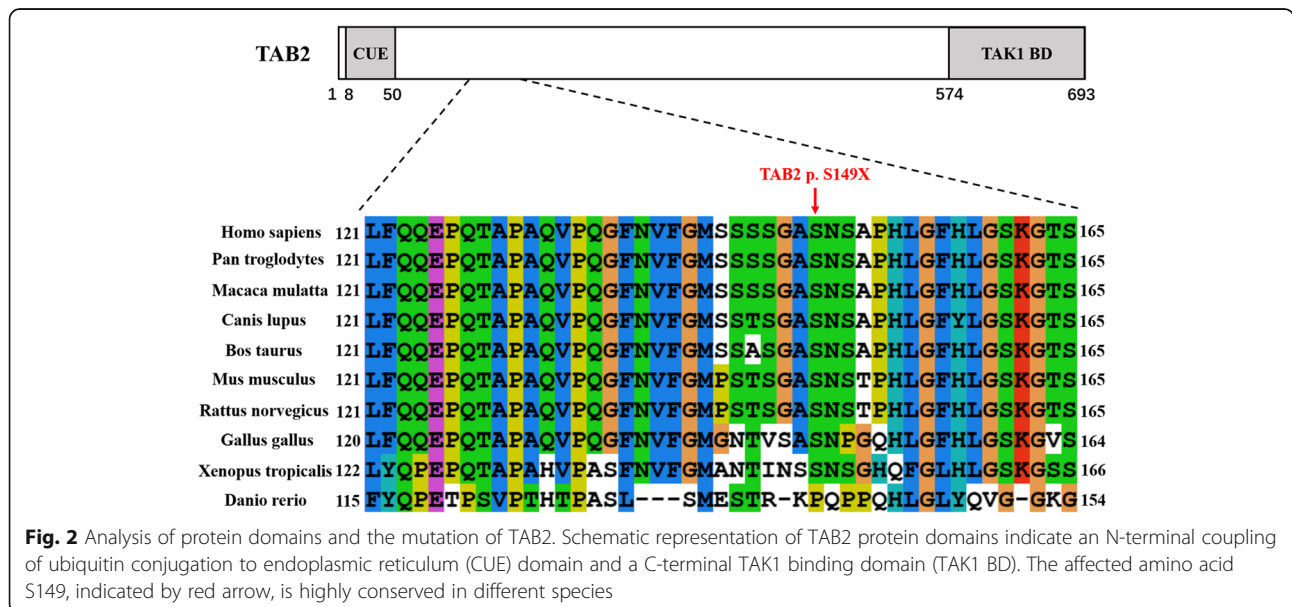
CHD Family members	CHD	Age at diagnosis	Heart defects according to transthoracic echocardiogram	TAB2	
				DNA	Protein
II:1	Yes	39y	atrial septal aneurysm, left coronary artery dilation, mild aortic regurgitation	c.C446G	p.S149X
II:3	No	36y	-	-	-
II:5	Yes	31y	mild mitral and tricuspid regurgitation, mild aortic valve stenosis with aortic regurgitation	c.C446G	p.S149X
II:6	No	32y	-	-	-
II:7	Yes	30y	atrial septal aneurysm, mild mitral valves prolapse with mitral regurgitation, mild pulmonic regurgitation	c.C446G	p.S149X
II:8	No	31y	-	-	-
II:9	Yes	29y	atrial septal aneurysm, left atrial and ventricular dilatation	c.C446G	p.S149X
III:3	No	12y	-	-	-
III:4	No	11y	-	-	-
III:5	No	3y	-	-	-
III:9	No	2y	-	-	-
III:10	High risk	19w pregnancy	-	c.C446G	p.S149X
III:11	Yes	3y	mild left ventricular and right atrial dilation, mild mitral valves prolapse with mitral regurgitation	c.C446G	p.S149X
III:13	No	1y	-	-	-

CHD Congenital heart defect, y Years old, w Weeks; "-", no defect or mutation detected

terminated. The third child (III:9) was a 2-year-old girl with no symptoms of cardiac disorders. She was not diagnosed with any heart abnormalities using echocardiography.

The proband's other family members were interviewed, and cardiac symptoms, such as tachypnea, shortness of breath, and fatigue, were reported. Four females and one male were identified with clinical symptoms (Fig. 1a). The proband's mother (I:2) died in her late

fifties due to sudden cardiac arrhythmia. Evident atrial septal aneurysms were detected in the proband's sisters (II:1, II:7, and II:9) using echocardiography. Left coronary artery dilation was screened in the patient II:1 by echocardiography and then identified using coronary arteriography. Other cardiac anomalies, such as mild mitral valves prolapse with mitral and pulmonic regurgitation, and left atrial and ventricular dilatation, were found in II:7 and II:9, respectively. III:11 was a 3-year-old boy who was



diagnosed with mild left ventricular and right atrial dilation as well as mild mitral valves prolapse with mitral regurgitation using echocardiography. Transabdominal fetal echocardiography was performed on the proband's fetus (III:10) at 15 weeks of pregnancy, and no cardiac abnormality was detected. The heart defects in the individuals enrolled in this study are summarized in Table 1.

Mutation detection

An autosomal dominant inheritance pattern was suggested on the basis of vertical transmission of CHD in the family. Peripheral blood was obtained from 13 family members (II:1, II:3, II:5, II:6, II:7, II:8, II:9, III:3, III:4, III:5, III:9, III:11, and III:13). Genomic DNA was extracted from the peripheral blood lymphocytes using the QIAamp DNA blood mini kit (Qiagen).

To determine the causative mutation in the family, WES of the proband (II:5) was performed as previously described with minor modifications [6]. Two micrograms of the genomic DNA from the proband II:5 was used for human whole-exome analysis with paired-end-sequencing at 100× resolution. Libraries were constructed using the SureSelect Human All ExonV7 kit (Agilent Technologies, USA) and sequenced on the Illumina HiSeq 2500 platform (Illumina, San Diego, CA), as per the manufacturer's instructions. The reads were aligned to the human reference genome (University of California Santa Cruz, UCSC hg19) using SOAPaligner. Single-nucleotide polymorphism (SNP) and indel (insertion or deletion) identification was performed using SAMtools and/or the Genome Analysis Toolkit (GATK), and SNPs with a read depth > 4 and quality > 20 were used for subsequent analyses. SNPs and indels were annotated using SeattleSeq annotation. Known polymorphisms in the dbSNP (<https://www.ncbi.nlm.nih.gov/snp/>) (minor allele frequency, > 0.01) and 1000 genomes (<https://www.ncbi.nlm.nih.gov/variation/tools/1000genomes/>) (genotype frequency, > 0.005) databases as well as synonymous single-nucleotide variants and variants not located in exonic or splicing regions were excluded. Pathogenicity of the obtained variants was predicted using Polyphen-2 (<http://genetics.bwh.harvard.edu/pph2/index.shtml>) [7], SIFT (<https://sift.bii.a-star.edu.sg/>) [8], and MutationTaster (<http://www.mutationtaster.org/>) [9].

Approximately 99.71% of the sequencing reads were mapped to the human genome hg19, with mean 186.76× sequencing depth. A heterozygous variant in *TAB2* (c.C446G, NM_015093.5), which results in a premature stop codon with early termination of protein translation at residue 149 (p.S149X), passed the filtering criteria. Using Sanger sequencing, this variant was detected in all the affected family members (II:1, II:5, II:7, II:9, and III:11); it was absent in the unaffected individuals (II:3, II:6, II:8, III:3, III:4, III:5, III:9, and III:13) (Fig. 1b). The

variant, with no record in 1000 genomes, the Exome Variant Server (EVS; <http://evs.gs.washington.edu/EVS/>), the Genome Aggregation Database (gnomAD; <http://gnomad.broadinstitute.org/>), and the HGMD databases, was not detected in our 531 control cohorts using high-resolution melting analysis (SsoFast EvaGreen, Bio-Rad). *TAB2* protein sequences (from *Danio rerio* to *Homo sapiens*) were obtained from the protein database of the National Center for Biotechnology Information (NCBI, <https://www.ncbi.nlm.nih.gov/>). Alignments of *TAB2* protein family members using cluster 2.0 software [10] revealed that the affected amino acid is evolutionarily conserved (Fig. 2). According to the latest recommendation by ACMG and AMP on sequence variants interpretation, the c.C446G variant in *TAB2* would be classified as “pathogenic”, having met the requirements of fulfilling one very strong (PVS1), one moderate (PM2) and three supporting (PP1, PP3 and PP4) criteria (Fig. 3) [5].

A prenatal molecular genetic diagnosis for the fetus (III:10) was recommended for evaluating the risk for CHD. With complete informed consent of the proband (II:5) and her husband (II:6), a prenatal diagnosis of the fetus (III:10) was performed at 19 weeks of pregnancy. The amniocytes was obtained by amniocentesis, and fetal genomic DNA (III:10) was extracted using the QIAamp DNA mini kit (Qiagen). A heterozygous *TAB2* c. C446G variant was detected, indicating that the fetus had a high risk for CHD.

Discussion and conclusions

TAB2 is a 693-amino acid protein and plays an important role in the IL-1 signaling pathway and cardiac development. In the present study, a nonsense mutation in *TAB2* was identified in five living patients with cardiac defects in a Chinese family. Most patients in the family presented mild clinical symptoms, such as slightly tachypnea and occasional fatigue, except for the proband's first girl child (III:7) who had left and right ventricular dilatation and suffered a SCD at 2 years old. The dilation of left ventricle is a hallmark of dilated cardiomyopathy (DCM) and may contribute to the unexpected death of patient III:7. DCM is defined by the presence of left ventricular dilatation and contractile dysfunction and serves as a leading cause of SCD, especially in childhood [11, 12]. Patients carrying *TAB2* mutations have been associated with DCM. A 7-month-old girl with *TAB2* c.1168delT mutation was diagnosed with DCM and conducted a heart transplant surgery at 9 months old, then died at 2.5 years old [13]. DCM was also detected in a 60-year-old brother and a 48-year-old sister who were both carrying a *TAB2* c.1398dupT mutation [14]. In addition, arrhythmia, another risk factor of SCD, had been reported in patients with *TAB2* mutations [2, 14].

		Benign		Pathogenic			
		Strong	Supporting	Supporting	Moderate	Strong	Very Strong
Population Data	MAF frequency is too high for disorder OR observation in controls inconsistent with disease penetrance				Absent in 1000G, EVS and gnomAD PM2	Prevalence in affecteds statistically increased over controls PS4	
Computational & Predictive Data		Multiple lines of computational evidence suggest no impact on gene /gene product Missense in gene where only truncating cause disease	Multiple lines of computational evidence support a deleterious effect on the gene /gene product PP3		Novel missense change at an amino acid residue where a different pathogenic missense change has been seen before PM5 Protein length changing variant PM4	Same amino acid change as an established pathogenic variant PS1	Predicted null variant in a gene where LOF is a known mechanism of disease PVS1
Functional Data	Well-established functional studies show no deleterious effect		Missense in gene with low rate of benign missense variants and path. missense common PP2		Mutational hot spot or well-studied functional domain without benign variation PM1	Well-established functional studies show a deleterious effect PS3	
Segregation Data	Non-segregation with disease		Co-segregation with disease in multiple affected family members PP1	Increased segregation data →			
De novo Data					De novo (without paternity & maternity confirmed) PM6	De novo (paternity & maternity confirmed) PS2	
Allelic Data		Observed in <i>trans</i> with a dominant variant Observed in <i>cis</i> with a pathogenic variant			For recessive disorders, detected in <i>trans</i> with a pathogenic variant PM3		
Other Database		Reputable database = benign	Reputable database = pathogenic PP5				
Other Data		Found in case with an alternate cause	Patient's phenotype highly specific for gene PP4				

Fig. 3 Variant assessment of S149X in TAB2 according to the recommendations of the ACMG and the AMP. Criteria fulfilled by this variant are indicated in yellow. This figure has been adapted from Richards et al. [5]

The heart defects detected using transthoracic echocardiography in the affected members were heterogeneous and mainly involved the heart valves. The proband was diagnosed with mitral, tricuspid, and aortic valve regurgitation. Aortic, pulmonic, and mitral valve dysfunctions were observed in two of her sisters (II:1 and II:7) and one of her nephews (III:11), respectively. In addition, three of the proband's sisters (II:1, II:7, and II:9) had atrial septal aneurysm. Dilation in the coronary artery or heart chambers was detected in patients II:1, II:9, and III:11.

Mutations disrupting *TAB2* are associated with heart valvular defects. Several studies have reported that CHD patients with 6q24-q25 microdeletions, containing the *TAB2* locus, experience valve anomalies, such as valvular stenosis, mitral or aortic regurgitation, and atrial or ventricular septal defects [2, 14–17]. Similar valvular defects have also been detected in patients with missense, nonsense, and small insertion or deletion mutations within *TAB2*. A c.622 C > T (p.P208S) and a c.688 C > A (p.Q230K) mutation in *TAB2* were identified in a woman with aortic regurgitation and a man with bicuspid aortic valve, respectively [2]. WES of a

male child with polyvalvular syndrome revealed a c.1491 T > A nonsense mutation (p.Y497X) in *TAB2* [18]. Pulmonary artery aneurysm, moderate mitral regurgitation, and mild tricuspid regurgitation were discovered in a family with a c.1039 C > T nonsense (p.R347X) *TAB2* mutation [19]. A c.1398dupT mutation in *TAB2* was confirmed in a family with polyvalvular heart disease [14]. Recently, a girl with a de novo *TAB2* c.1168delT mutation was diagnosed with dilated cardiomyopathy at 7 months of age; she underwent a heart transplant after 2 months and died at 2.5 years of age [13].

Apart from cardiac defects, *TAB2* mutations are associated with connective tissue disorders. The male child with CHD due to the *TAB2* p.Y497X mutation also had hypotonia, myopia, soft pale skin, joint hypermobility, and mild facial dysmorphism [18]. Similar clinical features were observed in the family with the *TAB2* c.1398dupT mutation [14]. However, no abnormality in the connective tissues was observed in the patients in our study. The family carrying the *TAB2* p.R347X nonsense mutation did not show any connective tissue disorders either [19]. These findings suggest clinical heterogeneity of extracardiac tissues in

patients with *TAB2* mutations. In addition, a c.1705 G > A mutation (p.E569K) and a c.1619 A > G mutation (p.Q540R) in *TAB2* were detected in patients with FMD, a progressive sclerosing skeletal dysplasia that affects the long bones and the skull. Different from the loss-of-function mutations involved in CHD, these *TAB2* mutations cause FMD through a gain-of-function mechanism [3, 4].

A detailed understanding of the loss-of-function mutations of *TAB2* that cause heart valvular defects remains unclear. The endothelial-to-mesenchymal transition (EndMT) process which endothelial cells migrate from the endocardial layer into the cardiac jelly and acquire mesenchymal characteristics to form the cardiac valves is essential in cardiac valvular development. This process is mainly regulated by the TGF- β signaling [20]. Disturbances in the TGF- β pathway undermine EndMT [21, 22] and cause valvular diseases, such as mitral valve degeneration and myxomatous atrioventricular valve diseases [23, 24]. *TAB2* plays an important role in the TGF- β pathway by phosphorylating the TAK1/MAP 3 K7 protein, which then activates NF- κ B and MAPK signaling [25]. *TAB2* mutations may cause valvular defects by disrupting the EndMT process controlled by the TGF- β pathway.

In summary, this study describes a three-generation Chinese family with a history of CHD. In this family, five living patients mainly presented with heart valvular defects. Whole-exome and direct sequencing were used to trace the genetic cause of the disease. We identified the first *TAB2* mutation (c.C446G, p.S149X) in a Chinese population. Molecular prenatal diagnosis was performed for the proband's fetus after the mutation was suggested to be pathogenic as per the latest recommendation on sequence variants interpretation laid out by the ACMG. The mutation may disturb cardiac valvular development by impeding the EndMT process regulated by the TGF- β pathway. Our study broadens the mutation spectrum of the *TAB2* gene and implies that *TAB2* plays a crucial role in the EndMT process.

Abbreviations

ACMG: The American College of Medical Genetics and Genomics; AMP: The Association for Molecular Pathology; CHD: Congenital heart diseases; DCM: Dilated cardiomyopathy; EndMT: Endothelial-to-mesenchymal transition; EVS: Exome variant server database; FMD: Frontometaphyseal dysplasia; GATK: Genome analysis toolkit; gnomAD: Genome aggregation database; HGMD: Human gene mutation database; indel: Insertion or deletion; NCBI: National Center for biotechnology information; SCD: Sudden cardiac death; SNP: Single-nucleotide polymorphism; TAK1 BD: TAK1-binding domain; WES: Whole-exome sequencing

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Authors' contributions

YL and JC designed the study. YL, JC, and YZ drafted the manuscript; JC and YZ performed the sequencing and the high-resolution melting analysis experiments; HY, LT, LP, and JX collected the clinical information; XW

performed the amniocentesis; KX, YY, and GC performed the DNA extraction and PCR. All authors have read and approved the final manuscript.

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Availability of data and materials

We did not use new software, databases, or applications/tools in the manuscript, all results and figures have already provided in the manuscript.

Ethics approval and consent to participate

All research was approved by the institutional review board of Jiangxi maternal and child health hospital, Nanchang, China. Written informed consents for the use of medical information and genetic analysis were obtained from all the enrolled subjects.

Consent for publication

The enrolled adults provided signed consents for the permission of using their clinical information and sequencing data for research purposes and/or scientific publications. For the participants under 16 years old, written informed consent was obtained from their parents. For the participant died at adult stage (I:2), written informed consent was obtained from her daughters.

Competing interests

The authors declare that they have no competing interests.

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