



Review

Geochemistry and the Origin of Life: From Extraterrestrial Processes, Chemical Evolution on Earth, Fossilized Life's Records, to Natures of the Extant Life

Satoru Nakashima ^{1,2,*}, Yoko Kebukawa ^{1,3}, Norio Kitadai ⁴, Motoko Igisu ⁵ and Natsuki Matsuoka ²

- Department of Earth and Space Science, Osaka University, Toyonaka, Osaka 560-0043, Japan
- Undergraduate School of Physics, Osaka University, Toyonaka, Osaka 560-0043, Japan; nmatsuoka@ess.sci.osaka-u.ac.jp
- ³ Faculty of Engineering, Yokohama National University, 79-5 Tokiwadai, Hodogaya-ku, Yokohama 240-8501, Japan; kebukawa@ynu.ac.jp
- ⁴ Earth-Life Science Institute, Tokyo Institute of Technology, 2-12-1, Ookayama, Meguro-ku, Tokyo 152-8550, Japan; nkitadai@elsi.jp
- Department of Subsurface Geobiological Analysis and Research, Japan Agency for Marine-Earth Science and Technology (JAMSTEC), Kanagawa 237-0061, Japan; igisu@jamstec.go.jp
- * Correspondence: satoru@ess.sci.osaka-u.ac.jp; Tel.: +81-6-6850-5799

Received: 10 August 2018; Accepted: 17 September 2018; Published: 20 September 2018



Abstract: In 2001, the first author (S.N.) led the publication of a book entitled "Geochemistry and the origin of life" in collaboration with Dr. Andre Brack aiming to figure out geo- and astro-chemical processes essential for the emergence of life. Since then, a great number of research progress has been achieved in the relevant topics from our group and others, ranging from the extraterrestrial inputs of life's building blocks, the chemical evolution on Earth with the aid of mineral catalysts, to the fossilized records of ancient microorganisms. Here, in addition to summarizing these findings for the origin and early evolution of life, we propose a new hypothesis for the generation and co-evolution of photosynthesis with the redox and photochemical conditions on the Earth's surface. Besides these bottom-up approaches, we introduce an experimental study on the role of water molecules in the life's function, focusing on the transition from live, dormant, and dead states through dehydration/hydration. Further spectroscopic studies on the hydrogen bonding behaviors of water molecules in living cells will provide important clues to solve the complex nature of life.

Keywords: building blocks; biopolymers; polymerization; extraterrestrial inputs; mineral surfaces; metabolism; photosynthesis; water; hydrogen bonding (9: 3-10)

1. Introduction

Life is generally characterized by the following three functions [1]: (1) metabolism: the ability to capture energy and material resources, staying away from thermodynamic equilibrium, (2) replication: the ability to process and transmit heritable information to progeny, and (3) compartmentalization: the ability to keep its components together and distinguish itself from the environment. These functions are operated by biopolymers such as proteins, DNAs, RNAs, and phospholipids (Figure 1). Proteins are made of amino acids linked together by peptide bonds. DNAs and RNAs are made of nucleotides (composed of (deoxy)ribose and nucleobases) bound by phosphodiester linkages. Phospholipids are made of two fatty acids esterified to a glycerol phosphate molecule.

These vital components were generally assumed to be synthesized abiotically, accumulated somewhere, condensed into polymers, interacted mutually, and eventually evolved into a self-sustaining system on the primitive Earth [2–5]. A considerable number of experimental and theoretical simulations have been made to explore favorable conditions for the respective prebiotic steps. However, geo- and astro-chemical processes essential for the origin of life are still not fully clear.

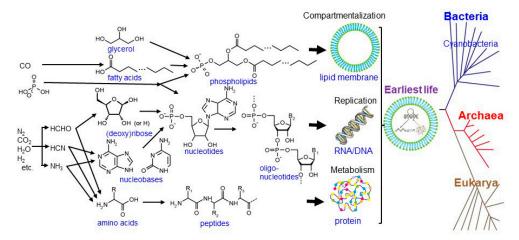


Figure 1. Structures, components, and abiotic synthetic pathways of bio-macromolecules operating in accordance with the three fundamental functions of life (compartmentalization, replication, and metabolism), modified from Kitadai and Maruyama [6].

To tackle these questions from Earth and planetary science perspectives, Nakashima and colleagues organized an international symposium, "Geochemistry and the origin of life," hosted by the Interactive Research Center of Science at the Tokyo Institute of Technology on 30 March 2001. International experts in biophysics, biochemistry, geochemistry, geology, and paleontology discussed various topics leading to a publication of a book, "Geochemistry and the origin of life," edited by Nakashima, Maruyama, Brack, and Windley [5]. This book provided our important starting points for future studies.

Here, we overview the research progress of this field since the book's publication, with a special focus on the achievements from our group. First, we summarize the findings for the formation and survival of life's building blocks in extraterrestrial environments, the chemical evolution of life in deep-sea hydrothermal systems, and the nature of ancient microorganisms inferred from fossilized biomolecules such as lipids. A new hypothesis on the origin and early evolution of photosynthesis is then proposed based on the changes in the redox and photochemical conditions through the Earth's history. Besides these bottom-up approaches, the extant life's activities with water are discussed highlighting the transition from live, dormant, and dead states for life with anhydrobiosis (life without water).

2. Origins of the Building Blocks of Life

2.1. Organics in the Cosmic Environments and Their Delivery to Earth

Since the primitive Earth around 4.6 billion years ago had abundant asteroid impacts, it is considered to have been covered with a "magma ocean". Therefore, neither life nor its constituents could be present at least on the Earth's surface. After the decrease of asteroid collisions, leading to the cooling of the Earth's surface and the formation of water oceans, life is considered to have appeared around 3.8 billion years ago [7]. Most of the raw materials for life (building blocks) are then expected to have been delivered from cosmic environments, although some of them could have been synthesized inside the Earth by the processes mentioned below.

Organic compounds detected in cosmic environments are mainly small molecules. Some of them are considered to be adsorbed or included in an amorphous ice coating on the surface of small silicate grains (Figure 2) (e.g., [8,9]). Greenberg [9] proposed these small micrometer-sized grains as cometary dust particles that have a silicate mineral core, organic mantle, and an amorphous ice crust ("Greenberg particles") (Figure 2), although their real structures have not yet been discovered. Ultraviolet and cosmic ray radiation are considered to induce reactions and transformations of some compounds leading to the formation of complex organic compounds including amino acids [10–12]. It should be noted that some submicron-sized organic nanoglobules have been observed in carbonaceous chondrites [13]. The large hydrogen isotope D/H ratios for the organic globules have been considered to indicate their pre-solar origin, at least for their raw materials [14]. They are associated with hydrous phyllosilicates but do not have silicate cores [13]. Therefore, they do not likely correspond to "Greenberg particles".

Beyond the snow line of the protoplanetary disk, these icy dust grains accreted into planetesimals (Figure 2). The interior of some of these planetesimals might have become warm by radioactive decay heat and others (e.g., [15]). If the temperature increased, ice can be melted to liquid water containing some organic components and aqueous alteration occurred in the planetesimals' interior (0–150 $^{\circ}$ C, for up to \approx 10 million years) [16–18]. More heating over water evaporation could have resulted in further thermal metamorphism without water [19] (Figure 2).

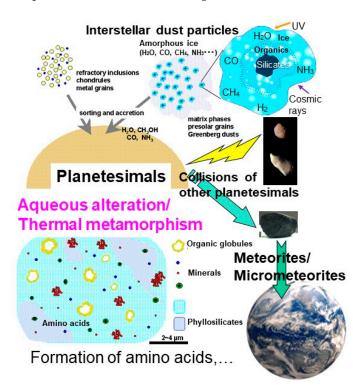


Figure 2. Schematic processes of evolution of organic molecules from interstellar icy dust particles, their accumulation to planetesimals, and aqueous alteration/thermal metamorphism inside the planetesimals leading to the formation of life's building blocks such as amino acids. They could have been delivered in meteorites and micrometeorites to Earth by collisions with other planetesimals.

To understand the nature and origins of organic matter in these interstellar or planetesimal interior environments, extensive studies on analyses of organic compounds in meteorites, micrometeorites, interplanetary dust particles (IDPs), and cometary dust particles and laboratory experiments have been conducted. Nucleic acid bases and sugar-related compounds have been detected in carbonaceous chondrites [20–22]. Various amino acids have also been detected from a wide variety of chondrite groups including thermally metamorphosed chondrites ([23] and references therein). Primitive

chondrites with some low temperature aqueous alteration contains larger amounts of amino acids compared to thermally metamorphosed chondrites ([23] and references therein).

In order to examine the formation processes of these building blocks in extraterrestrial environments, numerous experimental studies have been conducted, in particular simulating interstellar ice chemistry and in the presence of liquid water simulating aqueous alteration in planetesimals. UV photolysis and cosmic ray bombardment of interstellar ices could produce amino acids [10–12], nucleobases ([24] and references therein), as well as refractory organic matter (e.g., [25]). Fischer-Tropsch type (FTT) reactions could also produce amino acids and macromolecular organic matter from high-temperature gases (CO, H₂, and NH₃) with the presence of mineral catalysts in the solar nebula and/or planetesimals [26,27]. In the presence of liquid water in planetesimals, amino acids could be formed by Strecker reactions from aldehydes, NH3, and HCN, but Strecker reactions can only produce α -amino acids (e.g., [28]). Cody et al. [29] found that complex organic solids resembling insoluble organic matter (IOM) in primitive carbonaceous chondrites were produced from formaldehyde and water starting from a formose reaction (well-known sugar formation reaction [30]) by further heating. Kebukawa et al. [31] added ammonia in this system, resulting in the formation of macromolecules with heterocyclic N and the enhancement of yields of organic solids. Kebukawa and Cody [32] determined the first order reaction rates forming these organic solids with and without ammonia. Kebukawa et al. [33] identified several amino acids in these reaction products with ammonia with relative abundances similar to those extracted from carbonaceous chondrites. These data can be considered as experimental proofs of reaction processes forming amino acids, sugars, and complex insoluble organics during aqueous alteration in planetesimals.

Some of these organics can be eventually delivered to Earth surviving radiogenic heating in planetesimals, impacts, and atmospheric entries, and used as raw materials for life after their delivery to Earth as meteorites and IDPs. Kebukawa et al. [34] studied the thermal stabilities of chondritic organics and determined decreased rates of aliphatic CHs by using in situ heating infrared micro-spectroscopy. They evaluated preservation of organics based on the above kinetic experimental data with a thermal history of an asteroid (9 km radius). For asteroids with a radius of less than 2 km, some organics were found to survive on the surface.

2.2. Required Conditions for Prebiotic Chemistry

Besides the extraterrestrial inputs summarized in Section 3, various terrestrial geochemical events have been proposed to have produced life's building blocks as summarized by Kitadai and Maruyama [6]. Although laboratory simulations of aquatic environments on land have reported a number of great successes [35,36], demonstrated organic reactions typically start from activated carbon compounds (e.g., formaldehyde, hydrogen cyanide) that are incompatible with biologically and geologically inferred early life's metabolism strategy [37–39].

Deep-sea hydrothermal systems associated with serpentinization of ultramafic rocks have been suggested to be among the most plausible settings for life to originate from both biological and geological perspectives [40]. However, experimental verification about what geochemical processes drove chemical evolution remains insufficient [41].

Recent in situ electrochemical survey of the Okinawa Trough hydrothermal fields discovered spontaneous and widespread electricity generation in deep-sea black smoker chimneys [42]. Field and laboratory investigations have verified that electrons are catalytically provided by the oxidation of reductive chemicals (e.g., H_2S and H_2) at the hydrothermal fluid-mineral interface and are transported toward the outer chimney surface via electrically conductive sulfide rocks across the redox gap between the fluids and seawater [43–45]. Considering the ubiquity of sulfide deposits in the present-day and early ocean hydrothermal environments [46,47], together with the ever-existing redox disequilibrium between the Earth's surface and the interior [48], the geoelectrochemical systems would have been distributed extensively on the seafloor through the Earth's history. Kitadai et al. [49] demonstrated efficient CO_2 electroreduction to carbon monoxide (CO) on some metal sulfides (for example, CdS and

Life **2018**, 8, 39 5 of 21

Ag₂S) simulating early ocean hydrothermal vent environments. The reaction conditions favorable for the CO production were consistent with that assumed in Wächtershäuser's abiotic organic synthesis starting from CO [50–52]. Therefore, the early ocean alkaline hydrothermal systems were suggested to have favored the prebiotic CO_2 fixation, and for the subsequent evolution of primordial metabolism toward the origin of life [49].

The sulfide-promoted efficient CO₂-to-CO electroreduction requires electric potentials less than -0.8 V (versus the standard hydrogen electrode; SHE) [49,53]. Occurrence of this reaction in terrestrial hydrothermal environments would be difficult because of a low boiling temperature of water (e.g., 100 °C at 1 bar) that restricts geoelectrochemically generable potential range. However, if more efficient mineral catalysts allow the electrochemical CO₂ conversions to reactive C1 chemicals under potentials available on land (e.g., -0.7 V versus SHE in the Cedars spring systems [54]), chemical evolution starting from CO₂ could also have been realized in terrestrial geothermal fields. In contrast to deep-sea vents, such on-land settings likely enabled the involvement of solar light as an energy source [55] and diverse organic compounds that were produced through atmospheric/photochemical reactions [56] or delivered by extraterrestrial materials [57]. In a pond near the geothermal field, water evaporation possibly led to the concentration of key precursors and their condensation into life's building blocks [58]. Taking these advantages into account, a formamide-based volcanic origin-of-life scenario has been proposed by Saladino and coworkers [59]. It should be noted, however, that life's origin in water-poor environments must overcome "the water paradox": water is essential for all biological functions although its presence thwarts many proposed prebiotic reaction schemes [60]. Further experimental studies are needed to determine which of the two systems (deep-sea vs on-land) offered the best environmental situation to realize a smooth and uninterrupted transition from ancient geochemistry to biochemistry.

2.3. Roles of Mineral Surfaces

It has long been suggested that mineral surfaces have played a crucial role in the chemical evolution of life. The proposed functions beneficial to life's origin include the protection, selection, and concentration of key monomers from dilute aqueous solutions [61–63], the activation of the adsorbed monomers' polymerization to biopolymers [51,64–74], and the promotion of the emergence of biological functions such as replication, metabolism, and compartmentalization [4,75–77].

Biomolecules' polymerization in water is thermodynamically unfavorable (e.g., 17.1 kJ per one mole of peptide bond formation at 25 °C and 1 bar [78]) (Figure 3). Nakashima and Shiota [79] suggested that this difficulty can be overcome by coupling the dehydration–polymerization of monomers with the hydration of minerals when the overall reaction is thermodynamically downhill (Figure 3). This possibility was tested by Kitadai et al. [80] in a dry heating experiment of glycine with several anhydrous salts (MgSO₄, SrCl₂, BaCl₂, and Li₂SO₄) at 140 °C for up to 20 days. They showed that glycine polymerization is promoted by the salt hydration through the hydration–dehydration interactions, and that the salt having lower hydration Gibbs energy (easier to hydrate) exhibits a greater promotion effect. This type of coupling could also occur in water on crystalline oxide minerals with low hydration $\Delta_r G^{\circ}$ (e.g., anorthite (CaAl₂Si₂O₃) and forsterite (Mg₂SiO₄)).

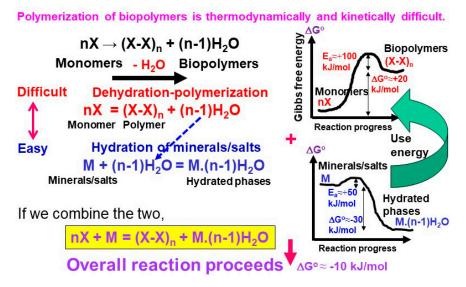


Figure 3. Schematic illustration of thermodynamic and kinetic difficulty for polymerization of biopolymers. Hydration of minerals/salts would enable these polymerization processes (modified from Nakashima and Shiota [79]).

Another problem regarding biomolecules' polymerization is their sluggish reaction rates due to the presence of high activation energy barriers (Figure 3). To examine surface-catalyzed polymerization and underlying molecular mechanism, Kitadai et al. [81] compared various oxide minerals as the catalysts for glycine polymerization. Titanium oxides (rutile and anatase) were found to provide a very effective reaction site, where around 25% of glycine converted to 2- to 6-mers after 10 days' heating at 80 °C, while only 0.01% of glycine dimerized in the absence of catalyst. Alumina (γ and α types), iron oxides (magnetite and hematite), and silica (quartz and amorphous silica) also facilitated the reaction with the degrees in this order. These observations suggest that polymerization activation arises from deprotonation of the –NH₃+ group to the nucleophilic –NH₂ one, and from the withdrawal of electron density from the carboxyl carbon to the surface metal ion (Figure 4). The surface structure and crystallinity of oxide minerals also influence the reactivity, but are not the primary factors. In addition, forsterite intermediately assisted glycine polymerization, possibly because of the hydration–dehydration interaction [81], but the basicity of the solid sample used by Kitadai et al. [81] is a more likely cause [82].

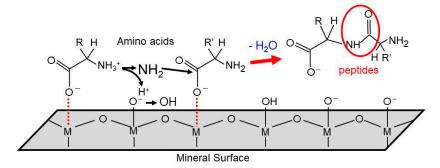


Figure 4. Schematic surface activation mechanisms of glycine polymerization on oxide minerals, modified from Kitadai et al. [81] with copyright permission from the journal.

Minerals can promote biomolecules' polymerization even in the presence of water because polymers tend to adsorb on the surface more tightly than the corresponding monomers, leading to the shift of monomer–polymer equilibria toward the polymer sides. Kitadai et al. [83] evaluated this thermodynamic effect using lysine and amorphous silica as a model combination. By combining experimentally determined adsorption constants of lysine and its dimer on amorphous silica with

thermodynamic data of these organic compounds in water reported in the literature [78], it was predicted that a silica surface favors lysine dimerization, particularly at alkaline pH (pH \approx 9) and lower ionic strength (1mM NaCl). In those conditions, around 50 times larger equilibrium dimer concentration was expected assuming infiltrating water in sandstone with a typical pore diameter (4 μm or the pore surface-to-volume ratio of 1000 m 2 L $^{-1}$) compared with that without silica. Importantly, the amounts of adsorption and the thermodynamically attainable dimer concentration depended greatly on the aqueous condition (e.g., pH, ionic strength). Thus, once lysine dimerization occurs in a favorable condition, changes in water chemistry can release most of the adsorbed dimers to the solution, thereby keeping the surface available for newly adsorbed species.

The thermodynamic computation as a function of environmental parameters introduced above is performable only for the lysine–amorphous silica system because of a lack of adsorption constants for polymers. However, the surface affinities for ion adsorption of oxide minerals are theoretically predictable [84,85]. Amino acid adsorptions on oxide minerals are controlled mostly by electrostatic interaction [86,87]. Therefore, future experimental characterizations of some representative acidic, basic, and neutral amino acids (e.g., lysine, aspartate, and alanine) and their corresponding peptides by the methodology reported by Kitadai et al. [83] will enable evaluation of favorable geological settings for peptide bond formation on the primitive Earth.

Recently, amorphous silica was found to stabilize ribose [88] and promote its condensation with phosphate and adenine into adenosine monophosphate under a dry condition [89]. Nucleotides might also be synthetized in hydrothermal conditions [90].

3. Origin and Evolution of Photosynthesis: A New Hypothesis

The first life that appeared on Earth might have used metabolic pathways presented above in sea floor hydrothermal environments like present day hyperthermophiles. They might have lived in sulfide-rich hydrothermal vent environments with a primordial metabolic system that preceded the autotrophic CO₂ fixation such as the reductive acetyl-CoA pathway and the reductive tricarboxylic acid (rTCA) cycle [91,92].

With the evolution of Earth's interior forming a strong magnetic core, Earth is supposed to have become surrounded by a magnetic field shielding cosmic electromagnetic radiation around 3 billion years ago [7]. This might have enabled ancient microbes to survive at shallow sea environments. The microbes then started to use solar energy for metabolism which started photosynthesis.

The present-day photosynthesis operates with complex electron and proton transfer pathways by using many photosynthetic pigments, such as chlorophylls absorbing lights at 680 nm (P680) and at 700 nm (P700), and proteins such as cytochrome and ferredoxin to produce nicotinamide adenine dinucleotide phosphate (NADP) and adenosine triphosphate (ATP) (Figure 5) [93–96]. Recent genomic comparison of different photosynthetic microbes suggests that the photosynthetic systems I (PS I) and II (PS II) occurred separately and merged into oxygen generating systems in cyanobacteria and plants by means of endosymbiosis [95,96].

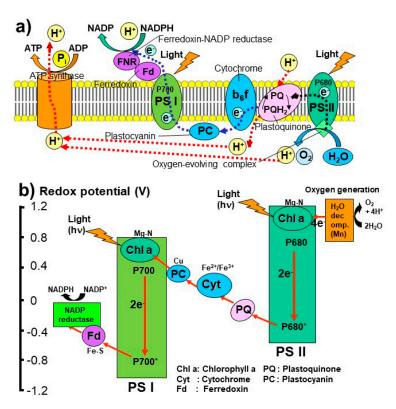


Figure 5. (a) Present day photosynthetic pathways in the lipid membrane and (b) redox potentials of the photosynthetic systems I (PSI and II (PSII), modified from Blankenship [95] and Martin et al. [96].

Sun lights arriving at the present Earth's surface is mostly in the 300 to 1300 nm wavelength range, while those at the depth of 20 m in sea water is narrower around 500 nm (Figure 6a) [95,97,98]. On the other hand, major photosynthetic pigments do not use lights around 500 nm but absorb 275 nm for ferredoxin, 420 and 550 nm for cytochrome, 430 and 670 nm for chlorophyll a, and 360, 580, and 780 nm for bacteriochlorophyll a (Figure 6b) [95,99].

Spectral characters of sunlight arriving at the Earth's surface and sea water around 3 billion years ago are not known but could be different from the present-day ones due to different compositions of the ancient atmosphere and ocean. Earth's atmosphere without oxygen (absorbing UV <240 nm) and ozone (absorbing UV <360 nm) [98] will permit UV light to reach Earth's surface and sea water.

Under these UV-rich oxygen-free environments around 3 billion years ago, the first photosynthetic molecules might have been absorbing UV light under reducing conditions. Therefore, ferredoxins with Fe-S centers with the lowest redox potential could have been the first photosynthetic molecule absorbing UV light around 275 nm ("proto-PS I") (Figures 5 and 6). This Fe-S containing protein could have been formed during the above "proto-metabolism" operating on the surface of metal sulfides.

With the further evolution of Earth's atmospheric and oceanic environments, wavelengths of sunlight might have shifted from UV to the visible region and the redox potential might have also increased. Cytochrome with Fe^{2+}/Fe^{3+} pairs could have been the second photosynthetic molecule ("proto-PS II"), absorbing light around 420 and 550 nm (Figures 5 and 6).

Chlorophylls might have been added later to these proto-PS I and II systems with further changes in atmospheric and oceanic conditions. They can work as antenna for absorbing mainly 680 and 700 nm in high (oxidizing) redox conditions leading to oxygen generating systems in cyanobacteria around 2.7 billion years ago.

This new hypothesis on the origin and evolution of photosynthesis proposed here based on geochemical redox and light environments is just a working one without mechanistic details. Several biologists [100–104] have proposed that the primitive photosynthetic reaction centers were porphyrin—FeS couples (PSI type) or quinone types [95]. Kritsky et al. pointed out that excited flavins,

which can photocatalyze reactions leading to the accumulation of free energy in the products, were available in the earliest stage of evolution [105]. Recent genomic and molecular structural comparison studies on various photosynthetic microbes suggested that at least either PS I or PS II systems might have started separately and then merged into oxygen generating systems in cyanobacteria and plants by means of endosymbiosis [95,96]. Because it is extremely difficult to test this hypothesis by detecting remnant photosynthetic molecules in the fossil record, this hypothesis cannot be confirmed easily. However, a new view based on geochemical environmental evolution of Earth should be considered in the origin and evolution of photosynthesis. In particular, ultraviolet-visible spectra arriving at the Earth's surface and sea water should be evaluated quantitatively by assuming atmospheric and oceanic compositions. We are currently working on the formation and degradation pathways and rates of photosynthetic pigments in present day plants in order to find clues for their evolutional pathways.

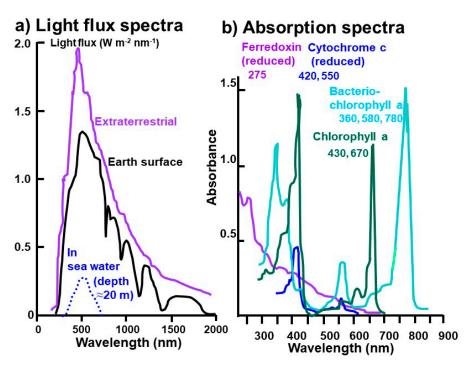


Figure 6. (a) Present day light flux spectra from the Sun, modified from Blankenship [95], Smith et al. [97], and Seinfeld and Pandis [98]. (b) Absorption spectra of photosynthetic pigments and proteins, modified from Blankenship [95] and Wang et al. [99]. Copyright permission was obtained from the journal, the book, and the author.

4. Fossilized Evidence of Life

The remnant of life in Archean (about 4–2.5 billion years ago) rocks has the potential for elucidating the origin of life on Earth. The presence of life on Earth has been inferred mainly from morphologically preserved microscopic fossils, stromatolites, molecular biomarker, and stable isotopic compositions in sedimentary rocks. Recently, the oldest putative microfossils were reported from \approx 3.8 billion-year-old ferruginous sedimentary rocks in Quebec, Canada [106]. In addition, the ¹³C-depleted graphite was found from \approx 3.95 billion-year-old metasedimentary rocks in Labrador, Canada, which provides the oldest evidence of life on Earth [38]. However, because of scarcity and the poor preservation of Archean rocks, interpretation of the ancient records for the presence of life is still controversial.

As an example, cell-like structures found in \approx 3.5 billion-year-old Apex chert (sedimentary rock composed of microcrystalline quartz) in Australia have been taken as the oldest microfossils [107]. However, Brasier et al. [108] questioned the biological origin of these structures and suggested that they can be formed by precipitation of organics from hydrothermal solutions. On the other hand, the chemical analyses of isolated organic matter have provided important information about the

physiological and phylogenetic characteristics of the source organic compounds (e.g., [109,110]). However, the bulk analysis of isolated organic matter generally involves the potential risk of post-depositional and experimental contamination of the organic matter (e.g., [110,111]).

In situ analysis have provided isotopic, elemental, and molecular signatures of individual microstructures in petrographic thin sections (e.g., [112–129]). Among these chemical signatures, molecular characteristics of microfossils have been obtained using spectroscopic techniques including Raman micro-spectroscopy, infrared (IR) spectroscopy, electron energy-loss spectroscopy (EELS), and X-ray absorption near-edge structure (XANES) spectroscopy. Raman micro-spectroscopy has been used to identify mineralogy of the samples. Especially for evaluating microfossils, Raman micro-spectroscopy has been used to clarify their carbonaceous composition; that is G (graphite) and D (disordered or defect) bands [114–118,124,125,128,130–132]. However, it should be noted that these G and D bands can also be observed for experimentally produced abiotic organic matter, and thus are not necessarily indicators for their biogenic origin [133]. These bands are also often used as an indicator for evaluating the maturation degree of carbonaceous matter in sedimentary rocks (e.g., [134,135]).

IR micro-spectroscopy revealed distributions of aliphatic C–H bonds corresponding to microfossil structures in Bitter Springs (0.83 billion year old) [120] and Gunflint cherts (1.9 billion year old) [121] (Figure 7a). The CH_3/CH_2 peak height ratios of microfossils, which correspond to relative length and/or branching of aliphatic chains of lipids [136], were found to be similar to those of modern cyanobacteria [121] (Figure 7b,c). Aliphatic C–H bonds have also been detected in microfossils and/or carbonaceous matter in Wumishan stromatolites (\approx 1.5 billion years) [137] and Doushantuo phosphorites and cherts (\approx 0.58 billion years) [137,138].

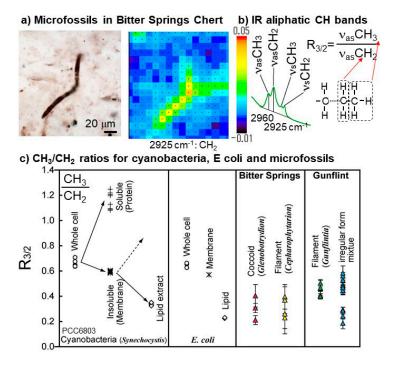


Figure 7. Infrared (IR) micro-spectroscopic analyses of microfossils. (a) Optical microscope image of a filamentous microfossil in Bitter Springs chert and IR map image for 2925 cm^{-1} peak height due to aliphatic CH_2 modified from Igisu et al. [120]. (b) Schematic images for determining the CH_3/CH_2 peak height ratio $(R_{3/2})$. (c) CH_3/CH_2 peak height ratio $(R_{3/2})$ of extant prokaryotes (cyanobacteria and E coli), and microfossils from Bitter Springs Group and Gunflint Formation modified from Igisu et al. [121] with copyright permission from the journal. Values for the whole cell, water-soluble (proteins), water-insoluble (membrane), and lipid extracts are also shown for cyanobacteria and *E. coli*.

EELS combined with transmission electron microscope (TEM) showed that organic matter from Apex chert possesses C=C of amorphous carbon [139]. XANES spectroscopy was applied to Apex and

Gunflint carbonaceous matter [140] and Gunflint microfossils [141]. They revealed the presence of COOH and aromatic COH components in Apex and Gunflint organic matter [140], and the presence of amide derived from protein in Gunflint microfossils [141].

These in situ methods detecting molecular information would be useful for identification and/or classification of microfossils. Although the functional groups such as C–H, C=C, COOH, and COH can be included in non-biogenic carbonaceous compounds, such as asphalt [60], their spatial distribution data by the in situ methods corresponding to microfossil structures would support their biogenicity. However, post-depositional degradation of microorganisms should be considered for comparison of the obtained data with extant microorganisms. Some experimental studies were performed in order to evaluate changes in the spectroscopic signatures during fossilization process [121,141,142]. It is important for a better understanding of the post-depositional degradation of microorganisms to accumulate information on the durability of organic components under various preservation conditions.

5. Roles of Water for Life through Studies on Anhydrobiosis

The presence of liquid water is essential for life's activities with functions of biomolecules. However, some microbes, plants, and insects can survive in the absence of water and this tolerance against anhydrous states is called "anhydrobiosis" (life without water) [143]. This anhydrobiosis can keep a sort of sleeping state (dormancy) during dehydrated states and can recover viable states upon rehydration. Recent biological studies suggested the following two important mechanisms for anhydrobiosis [143–145]: (1) protection of proteins against denaturation by means of surrounding sugars such as trehalose, and (2) fixation of cellular structures by changing lipid membranes into glassy states. Therefore, studies on transitions from live, dormant, and dead states through dehydration/hydration might provide clues for life's functions with water.

Several studies indicated usefulness of infrared (IR) spectroscopy for characterizing molecular changes of lipids and proteins [144,146]. However, IR spectroscopic signatures of transitions from dormancy to live states and from live states to dormancy or death have not been elucidated.

Yeast (*Saccharomyces cerevisiae*) is one of the unicellular eukaryotic microbes employed in fabrication of breads. Dried yeast cells are known to be in dormancy and can return back to activities upon rehydration. We are now studying these transition processes among dormancy, viable states, and death by means of an original in situ IR microspectroscopy system with controls of relative humidity (RH) and temperature (T) (Figure 8). RH in a plastic container with a CaF_2 window was controlled by flowing a mixture of dry and wet air. By changing the flow rates of the dry and wet air with flowmeters to be the total flow rate at 1 L min⁻¹, the mixed air introduced to the plastic cell can be controlled from about 2 to 80% RH [147]. Dry yeast powders were placed on an Al plate on a Peltier cooling–heating stage (-20 to 120 °C) in the RH/T control cell (Figure 8). IR transflection (transmission–reflection) spectra were measured every 1 minute during RH/T changes.

Aggregated dry yeast cells dispersed with one drop of pure water placed on an Al plate showed the following IR absorption bands (Figure 9a): a broad band at 3700–3000 cm⁻¹ due to OH and NH stretching of water molecules and proteins (amides A and B); a group of several bands at 3000–2770 cm⁻¹ due to CH stretching of lipids; two bands at 1720–1480 cm⁻¹ due to C=O stretching (amide I), and CNH (amide II) and HOH bending of proteins and water molecules; and a broad band at 1170–1000 cm⁻¹ due to C=O stretching of sugars.

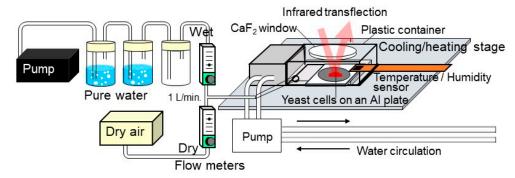


Figure 8. In situ IR micro-spectroscopy system with controls of relative humidity (RH) and temperature (T) for studying changes of yeast cells during dehydration/rehydration and/or heating/cooling.

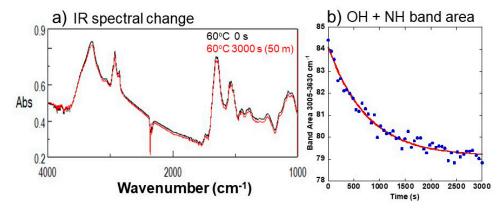


Figure 9. (a) IR spectral changes with time of the yeast cells heated at $60 \,^{\circ}$ C for 0 and $3000 \, \text{s}$. (b) Changes with time for the first $3000 \, \text{s}$ in the IR band area at $3005 - 3630 \, \text{cm}^{-1}$ (OH + NH) of the yeast cells heated at $60 \,^{\circ}$ C. A fitting exponential curve is also shown.

Our preliminary heating experiments of the yeast cells at 60 °C at ambient room RH condition showed IR spectral changes. In the first stage (0 to 3000 s), OH+NH (Figure 9b), amides I+II+H₂O, and CH band areas decreased exponentially. In the later second stage, OH+NH and amides I+II+H₂O band areas increased gradually, while the CH band area continued to decrease gradually. Changes with time for the first 3000 s in band areas at 3005–3630 cm⁻¹ (OH + NH)(Figure 9b), at 1480–1720 cm⁻¹ (amides I + II + H₂O), and at 2775–3005 cm⁻¹ (aliphatic CH) could be fitted by exponential equations to determine the first order reaction rate constant k_1 to be 1.3×10^{-3} s⁻¹ for the OH + NH band area, 0.93×10^{-3} s⁻¹ for the amides I + II + H₂O band area, and 0.87×10^{-3} s⁻¹ for the aliphatic CH band area. These reaction rate constants are all on the order of 1×10^{-3} s⁻¹, showing similar decay constants. Since the 3005–3630 cm⁻¹ band area corresponds to stretching vibrations of OH in water and NH in proteins, and that at 1480–1720 cm⁻¹ to peptide bonds in proteins (amides I, II) with a contribution of bending vibration of H₂O molecules, these band area decreases indicate a loss of water associated with protein changes.

Since yeast cells are considered to lose viability at 60 °C, the above decreases in water, proteins, and lipids can be corresponding to a loss of life's functions possibly due to a loss of hydrogen bonding. Although further detailed studies are required, the present first results indicate the usefulness of the new method for elucidating transitions among dormancy, live, and dead states, leading to a better understanding of the life's function with water.

The life's functions are mostly maintained by hydrogen bonding: (1) secondary and higher order structures of proteins are formed by hydrogen bonding mainly among peptide bonds, (2) information transfer is operated by base pairing among DNAs and RNAs by hydrogen bonding, (3) lipid membranes separating cellular compartments have polar head groups facing to liquid water by hydrogen bonding, and (4) liquid water molecules containing all these biopolymers are hydrogen

bonded to each other. Therefore, hydrogen bonds are considered to be essential for life's activities [148] and further studies on the nature of hydrogen bonds among and within biopolymers and water molecules are necessary.

6. Summary and Future Perspectives

(1) Extraterrestrial inputs

Primitive meteorites contain a wide variety of organic compounds, including potential building blocks of life, such as amino acids, nucleobases, and sugar-related compounds. These organic compounds would have been formed and evolved in various environments, e.g., icy dust in molecular clouds and planetesimals. Some of these would have been delivered to the early Earth within meteorites, comets, and IDPs. Despite the numerous works on meteorite analyses and experimental studies, the origins and evolutions of organic compounds are not fully understood. Future sample return missions such as Hayabusa2 and OSIRIS-REx would provide more clues for origin, evolution, distribution, and delivery to Earth of prebiotic molecules in space environments.

(2) Prebiotic chemistry

Laboratory experiments have demonstrated the syntheses of almost all of life's building blocks and their polymers under simulated prebiotic conditions. However, how these compounds spontaneously assembled to generate proto-biological functions (e.g., metabolism, replication) on the Hadean Earth remains unclear. Geochemically plausible CO_2 fixation systems and their developments in response to the acquisition or synthesis of simple organic molecules are the next important topics to be explored for constraining environments and mechanisms for life to originate on Earth and other Earth-type planets.

(3) First metabolism and photosynthesis

The "proto-metabolism" might have initiated in the geoelectrochemical environments in deep-sea alkaline hydrothermal systems through abiotic CO₂ reduction and fixation into carboxylic acids (e.g., TCA intermediates) on metal sulfides.

The photosynthesis then started in shallow sea water. Although the origin and evolution of complex photosynthetic systems are still under debate among biologists, geochemical environments, such as redox and light conditions, should be considered. The first photosynthetic molecule could have been ferredoxins with Fe-S centers, which could have been formed by the above proto-metabolism, absorbing UV light around 275 nm ("proto-PS I") under UV-rich oxygen-free Earth environments around 3 billion years ago. Cytochrome with Fe²⁺/Fe³⁺ pairs could have been the second photosynthetic molecule ("proto-PS II"), absorbing light around 420 and 550 nm. Chlorophylls absorbing mainly 680 and 700 nm at high (oxidizing) redox conditions could have been added later to these proto-PS I and II systems, leading to oxygen generating systems in cyanobacteria around 2.7 billion years ago.

(4) Fossil records

Geological records provide direct evidence of the presence of life on Earth in four main ways: microfossils, stromatolites, molecular biomarkers, and stable isotope ratios. However, interpretation of the ancient records of the presence of life is still under debate because Archean rocks are rare on Earth, and poorly preserved. In situ analyses of individual microstructures reveal their elemental, isotopic, and molecular compositions, and enable us to examine spatial relationships between individual morphological structures and the surrounding minerals. Further careful descriptions of such microscopic chemical and morphological information are necessary for better understanding of their biogenicity.

(5) Water, hydrogen bonds, and life

Studies on transitions from live, dormant, and dead states through dehydration/hydration might provide clues for life's functions with water. Our preliminary heating experiments of the yeast cells at $60\,^{\circ}\text{C}$ at an ambient room RH condition by using an original in situ IR microspectroscopy system with controls of relative humidity (RH) and temperature (T) showed decreases in water, proteins, and lipids. They can correspond to a loss of life's functions, possibly due to a loss of hydrogen bonding.

Further studies on the nature of hydrogen bonds among and within biopolymers and water molecules are necessary.

Since the publication of the book entitled "Geochemistry and the origin of life" in 2001, the first author (S.N.) tried with colleagues to conduct the following four research targets: (1) material proofs of building blocks of life, mainly from extraterrestrial materials; (2) experimental evidence of formation processes of building blocks and polymers, both in extraterrestrial and terrestrial environments; (3) roles of mineral surfaces on the chemical evolution; and (4) molecular signatures of ancient life's fossil records. These studies could constrain some favorable environments for origin and evolution of life to be (a) aqueous alteration in planetesimals, (b) seafloor hydrothermal environments for polymerization and proto-metabolism, and (c) some mineral surfaces. Although most of the analytical and experimental studies in the literature showed inventories and showcases of building blocks and polymers, our group has provided thermodynamic and kinetic constraints with some logical background on the chemical evolution to life.

These studies suggest that mineral–water–organic interfaces are often playing key roles in the origin and evolution of life. Therefore, further interdisciplinary researches linking cosmochemistry, geochemistry, mineralogy, organic chemistry, electrochemistry, and photochemistry to investigate quantitatively mineral–water–organic interactions are required. They are not only important in the origin and evolution of life in the past, but also in quantitative prediction of Earth's resources and environments toward our future.

Author Contributions: Conceptualization, S.N.; Methodology, S.N., Y.K., N.K., M.I., and N.M.; Software, N.D.; Validation, Y.K., N.K., M.I., and N.M.; Formal Analysis, S.N.; Investigation, S.N., Y.K., N.K., M.I., and N.M.; Resources, S.N.; Data Curation, S.N., Y.K., N.K., M.I., and N.M.; Writing-Original Draft Preparation, S.N.; Writing-Review & Editing, S.N., Y.K., N.K., M.I., and N.M.; Visualization, S.N. and N.K.; Supervision, S.N.; Project Administration, S.N., Y.K., N.K., and M.I.; Funding Acquisition, S.N., Y.K., N.K., and M.I.

Funding: N.K. is supported by the Astrobiology Center Program of National Institutes of Natural Sciences (NINS) (Grant Number AB292004). Y.K. is supported by the Astrobiology Center of National Institutes of Natural Sciences (grant number AB291005 and AB301020) and Japan Society for the Promotion of Science KAKENHI (grant number JP17H02991, JP17H06458 and JP18K03722).

Acknowledgments: The authors are grateful to Andre Brack for giving us an opportunity of this review. We thank Kiyoshi Nakazawa, Shigenori Maruyama and Kensei Kobayashi for their supports and discussion.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

- 1. Zubay, G. Origins of Life on the Earth and in the Cosmos, 2nd ed.; Academic Press: San Diego, CA, USA, 2000; 564p.
- 2. Brack, A. Introduction. In *The Molecular Origin of Life. Assembling Pieces of the Puzzle*; Brack, A., Ed.; Cambridge University Press: Cambridge, UK, 1998; pp. 1–10.
- 3. Maurel, M.C. *La Naissance de la Vie et L'evolution Prebiotique a L'evolution Biologique*; Diderot-Multimedia-EDL: Paris, France, 1999; 176p.
- 4. Lahav, N. Biogenesis. Theories of Life's Origin; Oxford University Press: Oxford, UK, 1999; 349p.
- 5. Nakashima, S.; Maruyama, S.; Brack, A.; Windley, B.F. *Geochemistry and the Origin of Life*; Universal Academy Press: Tokyo, Japan, 2001; 355p.
- 6. Kitadai, N.; Maruyama, S. Origins of building blocks of life: A review. *Geosci. Front.* **2018**, *9*, 1117–1153. [CrossRef]
- 7. Maruyama, S.; Isozaki, Y.; Nakashima, S.; Windley, B.F. History of the Earth and life. In *Geochemistry and the Origin of Life*; Nakashima, S., Maruyama, S., Brack, A., Windley, B.F., Eds.; Universal Academy Press: Tokyo, Japan, 2001; pp. 285–328.
- 8. Herbst, E.; van Dishoeck, E.F. Complex organic interstellar molecules. *Annu. Rev. Astron. Astrophys.* **2009**, 47, 427–480. [CrossRef]
- 9. Greenberg, J.M. Making a comet nucleus. Astron. Astrophys. 1998, 330, 375–380.

10. Kasamatsu, T.; Kaneko, T.; Saito, T.; Kobayashi, K. Formation of organic compounds in simulated interstellar media with high energy particles. *Bull. Chem. Soc. Jpn.* **1997**, *70*, 1021–1026. [CrossRef]

- 11. Bernstein, M.P.; Dworkin, J.P.; Sandford, S.A.; Cooper, G.W.; Allamandola, L.J. Racemic amino acids from the ultraviolet photolysis of interstellar ice analogues. *Nature* **2002**, *416*, 401–403. [CrossRef] [PubMed]
- 12. Muñoz Caro, G.; Meierhenrich, U.; Schutte, W.; Barbier, B.; Arcones Segovia, A.; Rosenbauer, H.; Thiemann, W.-P.; Brack, A.; Greenberg, J. Amino acids from ultraviolet irradiation of interstellar ice analogues. *Nature* 2002, 416, 403–406. [CrossRef] [PubMed]
- 13. Nakamura, K.; Zolensky, M.E.; Tomita, S.; Nakashima, S.; Tomeoka, K. Hollow organic globules in the Tagish Lake meteorite as possible products of primitive organic reactions. *Int. J. Astrobiol.* **2002**, *1*, 179–189. [CrossRef]
- 14. Nakamura-Messenger, K.; Messenger, S.; Keller, L.P.; Clemett, S.J.; Zolensky, M.E. Organic globules in the tagish lake meteorite: Remnants of the protosolar disk. *Science* **2006**, *314*, 1439–1442. [CrossRef] [PubMed]
- 15. MacPherson, G.J.; Davis, A.M.; Zinner, E.K. The distribution of Al-26 in the early solar-system—A reappraisal. *Meteoritics* **1995**, *30*, 365–386. [CrossRef]
- 16. Zolensky, M.E.; Bourcier, W.L.; Gooding, J.L. Aqueous alteration on the hydrous asteroids—Results of EQ3/6 computer-simulations. *Icarus* **1989**, *78*, 411–425. [CrossRef]
- 17. Brearley, A.J. The action of water. In *Meteorites and the Early Solar System II*; Lauretta, D.S., McSween, J.H.Y., Eds.; University of Arizona Press: Tucson, AZ, USA, 2006; pp. 587–624.
- 18. Fujiya, W.; Sugiura, N.; Hotta, H.; Ichimura, K.; Sano, Y. Evidence for the late formation of hydrous asteroids from young meteoritic carbonates. *Nat. Commun.* **2012**, *3*, 627. [CrossRef] [PubMed]
- 19. Huss, G.R.; Rubin, A.E.; Grossman, J.N. Thermal metamorphism in chondrites. In *Meteorites and the Early Solar System II*; Lauretta, D.S., McSween, J.H.Y., Eds.; University of Arizona Press: Tucson, AZ, USA, 2006; pp. 567–586.
- 20. Cooper, G.; Kimmich, N.; Belisle, W.; Sarinana, J.; Brabham, K.; Garrel, L. Carbonaceous meteorites as a source of sugar-related organic compounds for the early Earth. *Nature* **2001**, *414*, 879–883. [CrossRef] [PubMed]
- 21. Callahan, M.P.; Smith, K.E.; Cleaves, H.J., II; Ruzicka, J.; Stern, J.C.; Glavin, D.P.; House, C.H.; Dworkin, J.P. Carbonaceous meteorites contain a wide range of extraterrestrial nucleobases. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 13995–13998. [CrossRef] [PubMed]
- 22. Martins, Z.; Botta, O.; Fogel, M.L.; Sephton, M.A.; Glavin, D.P.; Watson, J.S.; Dworkin, J.P.; Schwartz, A.W.; Ehrenfreund, P. Extraterrestrial nucleobases in the Murchison meteorite. *Earth Planet. Sci. Lett.* **2008**, 270, 130–136. [CrossRef]
- 23. Elsila, J.E.; Aponte, J.C.; Blackmond, D.G.; Burton, A.S.; Dworkin, J.P.; Glavin, D.P. Meteoritic amino acids: Diversity in compositions reflects parent body histories. *ACS Cent. Sci.* **2016**, 2, 370–379. [CrossRef] [PubMed]
- 24. Sandford, S.A.; Bera, P.P.; Lee, T.J.; Materese, C.K.; Nuevo, M. Photosynthesis and photo-stability of nucleic acids in prebiotic extraterrestrial environments. In *Photoinduced Phenomena in Nucleic Acids II: DNA Fragments and Phenomenological Aspects*; Barbatti, M., Borin, A.C., Ullrich, S., Eds.; Springer: Berlin, Germany, 2015; Volume 356, pp. 123–164.
- 25. Greenberg, J.M.; Li, A.; Mendozagomez, C.X.; Schutte, W.A.; Gerakines, P.A.; Degroot, M. Approaching the interstellar grain organic refractory component. *Astrophys. J.* **1995**, *455*, L177–L180. [CrossRef]
- 26. Nuth, J.A., III; Johnson, N.M.; Manning, S. A self-perpetuating catalyst for the production of complex organic molecules in protostellar nebulae. *Astrophys. J. Lett.* **2008**, *673*, L225–L228. [CrossRef]
- 27. Pizzarello, S. Catalytic syntheses of amino acids and their significance for nebular and planetary chemistry. *Meteorit. Planet. Sci.* **2012**, *47*, 1291–1296. [CrossRef]
- 28. Pizzarello, S.; Cooper, G.W.; Flynn, G.J. The nature and distribution of the organic material in carbonaceous chondrites and interplanetary dust particles. In *Meteorites and the Early Solar System II*; Lauretta, D.S., McSween, J.H.Y., Eds.; University of Arizona Press: Tucson, AZ, USA, 2006; pp. 625–651.
- 29. Cody, G.D.; Heying, E.; Alexander, C.M.O.; Nittler, L.R.; Kilcoyne, A.L.D.; Sandford, S.A.; Stroud, R.M. Establishing a molecular relationship between chondritic and cometary organic solids. *Proc. Natl. Acad. Sci. USA* **2011**, *108*, 19171–19176. [CrossRef] [PubMed]
- 30. Butlerow, A. Formation synthétique d'une substance sucrée. C. R. Acad. Sci. 1861, 53, 145–147.

31. Kebukawa, Y.; Kilcoyne, A.L.D.; Cody, G.D. Exploring the potential formation of organic solids in chondrites and comets through polymerization of interstellar formaldehyde. *Astrophys. J.* **2013**, 771, 19. [CrossRef]

- 32. Kebukawa, Y.; Cody, G.D. A kinetic study of the formation of organic solids from formaldehyde: Implications for the origin of extraterrestrial organic solids in primitive solar system objects. *Icarus* **2015**, 248, 412–423. [CrossRef]
- 33. Kebukawa, Y.; Chan, Q.H.S.; Tachibana, S.; Kobayashi, K.; Zolensky, M.E. One-pot synthesis of amino acid precursors with insoluble organic matter in planetesimals with aqueous activity. *Sci. Adv.* **2017**, 3, e1602093. [CrossRef] [PubMed]
- 34. Kebukawa, Y.; Nakashima, S.; Zolensky, M.E. Kinetics of organic matter degradation in the Murchison meteorite for the evaluation of parentbody temperature history. *Meteorit. Planet. Sci.* **2010**, *45*, 99–113. [CrossRef]
- 35. Powner, M.W.; Gerland, B.; Sutherland, J.D. Synthesis of activated pyrimidine ribonucleotides in prebiotically plausible conditions. *Nature* **2009**, 459, 239–242. [CrossRef] [PubMed]
- 36. Patel, B.H.; Percivalle, C.; Ritson, D.J.; Duffy, C.D.; Sutherland, J.D. Common origins of RNA, protein and lipids precursors in a cyanosulfidic protometabolism. *Nat. Chem.* **2015**, *7*, 301–307. [CrossRef] [PubMed]
- 37. Weiss, M.C.; Sousa, F.L.; Mrnjavac, N.; Neukirchen, S.; Roettger, M.; Nelson-Sathi, S.; Martin, W.F. The physiology and habitat of the last universal common ancestor. *Nat. Microbiol.* **2016**, *1*, 16116. [CrossRef] [PubMed]
- 38. Tashiro, T.; Ishida, A.; Hori, M.; Igisu, M.; Koike, M.; Méjean, P.; Takahata, N.; Sano, Y.; Komiya, T. Early trace of life from 3.95 Ga sedimentary rocks in Labrador, Canada. *Nature* **2017**, *549*, 516–518. [CrossRef] [PubMed]
- 39. Nunoura, T.; Chikaraishi, Y.; Izaki, R.; Suwa, T.; Sato, T.; Harada, T.; Mori, K.; Kato, Y.; Miyazaki, M.; Shimamura, S.; et al. A primordial and reversible TCA cycle in a facultatively chemolithoautotrophic thermophile. *Science* 2018, 359, 559–563. [CrossRef] [PubMed]
- 40. Russell, M.J.; Hall, A.J.; Martin, W. Serpentinization as a source of energy at the origin of life. *Geobiology* **2010**, *8*, 355–371. [CrossRef] [PubMed]
- 41. McCollom, T.M. Abiotic methane formation during experimental serpentinization of olivine. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 13965–13970. [CrossRef] [PubMed]
- 42. Yamamoto, M.; Nakamura, R.; Kasaya, T.; Kumagai, H.; Suzuki, K.; Takai, K. Spontaneous and widespread electricity generation in natural deep-sea hydrothermal fields. *Angew. Chem. Int. Ed.* **2017**, *56*, 5725–5728. [CrossRef] [PubMed]
- 43. Nakamura, R.; Takashima, T.; Kato, S.; Takai, K.; Yamamoto, M.; Hashimoto, K. Electrical current generation across a black smoker chimney. *Angew. Chem. Int. Ed.* **2010**, 49, 7692–7694. [CrossRef] [PubMed]
- 44. Ang, R.; Khan, A.U.; Tsujii, N.; Takai, K.; Nakamura, R.; Mori, T. Thermoelectricity generation and electron–magnon scattering in a natural chalcopyrite mineral from a deep-sea hydrothermal vent. *Angew. Chem. Int. Ed.* **2015**, *54*, 12909–12913. [CrossRef] [PubMed]
- 45. Yamamoto, M.; Nakamura, R.; Oguri, K.; Kawagucci, S.; Suzuki, K.; Hashimoto, K.; Takai, K. Generation of electricity and illumination by an environmental fuel cell in deep-sea hydrothermal vents. *Angew. Chem. Int. Ed.* **2013**, 52, 10758–10761. [CrossRef] [PubMed]
- 46. Russell, M.J.; Hall, A.J. The emergence of life from monosulphide bubbles at a submarine hydrothermal redox and pH front. *J. Geol. Soc. Lond.* **1997**, *154*, 377–402. [CrossRef]
- 47. Maslennikov, V.V.; Maslennikova, S.P.; Large, R.R.; Danyushevsky, L.V.; Herrington, R.J.; Ayupova, N.R.; Zaykov, V.V.; Lein, A.Y.; Tseluyko, A.S.; Melekestseva, I.Y.; et al. Chimneys in Paleozoic massive sulfide mounds of the Urals VMS deposits: Mineral and trace element comparison with modern black, grey, white and clear smokers. *Ore Geol. Rev.* 2017, 85, 64–106. [CrossRef]
- 48. Russell, M.J.; Barge, L.M.; Bhartia, R.; Bocanegra, D.; Bracheer, P.J.; Branscomb, E.; Kidd, R.; McGlynn, S.; Meier, D.H.; Nitschke, W.; et al. The drive to life on wet and icy worlds. *Astrobiology* **2014**, *14*, 308–343. [CrossRef] [PubMed]
- 49. Kitadai, N.; Nakamura, R.; Yamamoto, M.; Takai, K.; Li, Y.; Yamaguchi, A.; Gilbert, A.; Ueno, Y.; Yoshida, N.; Oono, Y. Geoelectrochemical CO production: Implications of the autotrophic origin of life. *Sci. Adv.* **2018**, *4*, eaao7265. [CrossRef] [PubMed]
- 50. Huber, C.; Wächtershäuser, G. Activated acetic acid by carbon fixation on (Fe,Ni)S under primordial conditions. *Science* **1997**, 276, 245–247. [CrossRef] [PubMed]

51. Huber, C.; Wächtershäuser, G. Peptides by activation of amino acids with CO on (Ni,Fe)S surfaces: Implications for the origin of life. *Science* **1998**, *281*, 670–672. [CrossRef] [PubMed]

- 52. Huber, C.; Wächtershäuser, G. α-hydroxy and α-amino acids under possible Hadean, volcanic origin-of-life conditions. *Science* **2006**, *314*, 630–632. [CrossRef] [PubMed]
- 53. Yamagushi, A.; Yamamoto, M.; Takai, K.; Ishii, T.; Hashimoto, K.; Nakamura, R. Electrochemical CO₂ reduction by Ni-containing iron sulfides; How is CO₂ electrochemically reduced at bisulfide-bearing deep-sea hydrothermal precipitates? *Electrochim. Acta* **2014**, *141*, 311–318. [CrossRef]
- 54. Morrill, P.L.; Kuenen, J.G.; Johnson, O.J.; Suzuki, S.; Rietze, A.; Sessions, A.L.; Fogel, M.L.; Nealson, K.H. Geochemistry and geobiology of a present-day serpentinization site in California: The Cedars. *Geochim. Cosmochim. Acta* 2013, 109, 222–240. [CrossRef]
- 55. Mulkidjaniana, A.Y.; Bychkovc, A.Y.; Dibrova, D.V.; Galperine, M.Y.; Koonin, E.V. Origin of first cells at terrestrial, anoxic geothermal fields. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, E821–E830. [CrossRef] [PubMed]
- 56. Fenus, M.; Michalčíková, R.; Shestivská, V.; Sponer, J.; Sponer, J.E.; Civiš, S. High-energy chemistry of formamide: A simpler way for nucleobase formation. *J. Phys. Chem. A* **2014**, *118*, 719–736.
- 57. Saladino, R.; Carotaa, E.; Bottaa, G.; Kapralovb, M.; Timoshenkob, G.N.; Rozanovb, A.Y.; Krasavinb, E.; Di Mauro, E. Meteorite-catalyzed syntheses of nucleosides and of other prebiotic compounds from formamide under proton irradiation. *Proc. Natl. Acad. Sci. USA* **2015**, *112*, E2746–E2755. [CrossRef] [PubMed]
- 58. Saladino, R.; Botta, G.; Mattia Bizzarri, B.; Di Mauro, E.; Garcia Ruiz, J.M. A Global scale scenario for prebiotic chemistry: Silica-based self-assembled mineral structures and formamide. *Biochemistry* **2016**, *55*, 2806–2811. [CrossRef] [PubMed]
- 59. Sponer, J.E.; Sponer, J.; Novkov, O.; Brabec, V.; Sedo, O.; Zdrhal, Z.; Costanzo, G.; Pino, S.; Saladino, R.; Di Mauro, E. Emergence of the first catalytic oligonucleotides in a formamide-based origin scenario. *Chem. Eur. J.* **2016**, 22, 3572–3586. [CrossRef] [PubMed]
- 60. Benner, S.A. Paradoxes in the origin of life. Orig. Life Evol. Biosph. 2014, 44, 339–343. [CrossRef] [PubMed]
- 61. Brack, A.; Barbier, B.; Boillot, F.; Chabin, A. Extraterrestrial organic molecules and the origin of life. In *Geochemistry and the Origin of Life*; Nakashima, S., Maruyama, S., Brack, A., Windley, B.F., Eds.; Universal Academy Press: Tokyo, Japan, 2001; pp. 17–38.
- 62. Maurel, M.C. Functional capabilities of biomolecules at the beginnings of life. —RNA selectivity and adaptability—. In *Geochemistry and the Origin of Life*; Nakashima, S., Maruyama, S., Brack, A., Windley, B.F., Eds.; Universal Academy Press: Tokyo, Japan, 2001; pp. 107–118.
- 63. Hazen, R.M.; Filley, T.R.; Goodfriend, G.A. Selective adsorption of L- and D-amino acids on calcite: Implications for biochemical homochirality. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 5487–5490. [CrossRef] [PubMed]
- 64. Bernal, J.D. The physical basis of life. Proc. Phys. Soc. 1949, 62, 537–558. [CrossRef]
- 65. Bujdak, J.; Faybikova, K.; Eder, A.; Yongyai, Y.; Rode, B. Peptide-chain elongation—A possible role of montmorillonite in prebiotic synthesis of protein precursors. *Orig. Life Evol. Biosph.* **1995**, 25, 431–441. [CrossRef] [PubMed]
- 66. Ferris, J.P.; Hill, A.J.; Liu, R.; Orgel, L. Synthesis of long prebiotic oligomers on mineral surfaces. *Nature* **1996**, 381, 59–61. [CrossRef] [PubMed]
- 67. Bujdak, J.; Rode, B.M. The effect of smectite composition on the catalysis of peptide bond formation. *J. Mol. Evol.* **1996**, 43, 326–333. [CrossRef] [PubMed]
- 68. Bujdak, J.; Rode, B. Silica, alumina, and clay-catalyzed alanine peptide bond formation. *J. Mol. Evol.* **1997**, 45, 457–466. [CrossRef]
- 69. Bujdak, J.; Rode, B. Silica, alumina and clay catalyzed peptide bond formation: Enhanced efficiency of alumina catalyst. *Orig. Life Evol. Biosph.* **1999**, 29, 451–461. [CrossRef] [PubMed]
- 70. Ferris, J.P. Catalyzed RNA synthesis for the RNA world. In *The Molecular Origin of Life. Assembling Pieces of the Puzzle*; Brack, A., Ed.; Cambridge University Press: Cambridge, UK, 1998; pp. 255–268.
- 71. Orgel, L.E. Polymerization on the rocks: Theoretical introduction. *Orig. Life Evol. Biosph.* **1998**, *28*, 227–234. [CrossRef] [PubMed]
- 72. Hill, A.R.; Böhler, C.; Orgel, L. Polymerization on the rocks: Negatively charged *α*-amino acids. *Orig. Life Evol. Biosph.* **1998**, *28*, 235–243. [CrossRef] [PubMed]

73. Liu, R.; Orgel, L. Polymerization on the rocks: *β*-amino acids and arginine. *Orig. Life Evol. Biosph.* **1998**, *28*, 245–257. [CrossRef] [PubMed]

- 74. Rode, B.M.; Son, H.L.; Suwannachot, Y. The combination of self-induced peptide formation reaction and clay catalysis: A way to higher peptides under primitive Earth conditions. *Orig. Life Evol. Biosph.* 1999, 29, 273–286. [CrossRef] [PubMed]
- 75. Cairns-Smith, A.G. *Genetic Takeover and the Mineral Origin of Life*; Cambridge University Press: Cambridge, UK, 1982; 488p.
- 76. Wächtershäuser, G. Before enzymes and templates: Theory of surface metabolism. *Microbiol. Rev.* **1988**, *52*, 452–484. [PubMed]
- 77. Wächtershäuser, G. Origin of life in an iron-sulfur world. In *The Molecular Origin of Life. Assembling Pieces of the Puzzle*; Brack, A., Ed.; Cambridge University Press: Cambridge, UK, 1998; pp. 206–218.
- 78. Kitadai, N. Thermodynamic prediction of glycine polymerization as a function of temperature and pH consistent with experimentally obtained results. *J. Mol. Evol.* **2014**, *78*, 171–187. [CrossRef] [PubMed]
- 79. Nakashima, S.; Shiota, D. Organic-inorganic interactions and the origin and evolution of life. In *Geochemistry and the Origin of Life*; Nakashima, S., Maruyama, S., Brack, A., Windley, B.F., Eds.; Universal Academy Press: Tokyo, Japan, 2001; pp. 135–177.
- 80. Kitadai, N.; Yokoyama, T.; Nakashima, S. Hydration-dehydration interactions between glycine and anhydrous salts: Implications for chemical evolution of life. *Geochim. Cosmochim. Acta* **2011**, 75, 6285–6299. [CrossRef]
- 81. Kitadai, N.; Oonishi, H.; Umemoto, K.; Usui, T.; Fukushi, K.; Nakashima, S. Glycine polymerization on oxide minerals. *Orig. Life Evol. Biosph.* **2017**, 47, 123–143. [CrossRef] [PubMed]
- 82. Sakata, K.; Kitadai, N.; Yokoyama, T. Effects of pH and temperature on dimerization rate of glycine: Evaluation of favorable environmental conditions for chemical evolution of life. *Geochim. Cosmochim. Acta* **2010**, *74*, 6841–6851. [CrossRef]
- 83. Kitadai, N.; Nishiuchi, K.; Nishii, A.; Fukushi, K. Amorphous silica-promoted lysine dimerization: A thermodynamic prediction. *Orig. Life Evol. Biosph.* **2018**, *48*, 23–34. [CrossRef] [PubMed]
- 84. Sverjensky, D.A. Prediction of surface charge on oxides in salt solutions: Revisions for 1:1 (M⁺L⁻) electrolytes. *Geochim. Cosmochim. Acta* **2005**, *69*, 225–257. [CrossRef]
- 85. Kitadai, N.; Nishiuchi, K.; Tanaka, M. A comprehensive predictive model for sulfate adsorption on oxide minerals. *Geochim. Cosmochim. Acta* **2018**, 238, 150–168. [CrossRef]
- 86. Kitadai, N.; Yokoyama, T.; Nakashima, S. ATR-IR spectroscopic study of L-lysine adsorption on amorphous silica. *J. Colloid Interface Sci.* **2009**, 329, 31–37. [CrossRef] [PubMed]
- 87. Kitadai, N.; Yokoyama, T.; Nakashima, S. In situ ATR-IR investigation of L-lysine adsorption on montmorillonite. *J. Colloid Interface Sci.* **2009**, *338*, 395–401. [CrossRef] [PubMed]
- 88. Akouche, M.; Jaber, M.; Zins, E.-L.; Maurel, M.-C.; Lambert, J.-F.; Georgelin, T. Thermal behavior of d-ribose adsorbed on silica: Effect of inorganic salt coadsorption and significance for prebiotic chemistry. *Chem. Eur. J.* **2016**, 22, 15834–15846. [CrossRef] [PubMed]
- 89. Akouche, M.; Jaber, M.; Maurel, M.-C.; Lambert, J.-F.; Georgelin, T. Phosphoribosyl pyrophosphate: A molecular vestige of the origin of life on minerals. *Angew. Chem. Int. Ed.* **2017**, *56*, 1–5. [CrossRef] [PubMed]
- 90. Pérez-Villa, A.; Saitta, M.; Georgelin, T.; Lambert, J.F.; Guyot, F.; Maurel, M.C.; Pietrucci, F. Synthesis of RNA nucleotides in plausible prebiotic conditions from ab initio computer simulations. *J. Phys. Chem. Lett.* **2018**, *9*, 4981–4987. [CrossRef] [PubMed]
- 91. Wächtershäuser, G. 1990 Evolution of the first metabolic cycles. *Proc. Natl. Acad. Sci. USA* **1990**, 87, 200–204. [CrossRef] [PubMed]
- 92. Aono, M.; Kitadai, N.; Oono, Y. A principled approach to the origin problem. *Orig. Life Evol. Biosph.* **2015**, 45, 327–338. [CrossRef] [PubMed]
- 93. Blankenship, R.E. Early evolution of photosynthesis. *Plant Physiol.* **2010**, *154*, 434–438. [CrossRef] [PubMed]
- 94. Hohmann-Marriott, M.F.; Blankenship, R.E. Evolution of photosynthesis. *Annu. Rev. Plant Biol.* **2011**, *62*, 515–548. [CrossRef] [PubMed]
- 95. Blankenship, R.E. *Molecular Mechanisms of Photosynthesis*, 2nd ed.; John Wiley & Sons: Oxford, UK, 2014; 296p.

96. Martin, W.F.; Bryant, D.A.; Beatty, J.T. A physiological perspective on the origin and evolution of photosynthesis. *FEMS Microbiol. Rev.* **2018**, 42, 205–231. [CrossRef] [PubMed]

- 97. Smith, R.C.; Prezelin, B.B.; Baker, K.S.; Bidigare, R.R.; Boucher, N.P.; Coley, T.; Karentz, D.; MacIntyre, S.; Matlick, H.A.; Menzies, D.; et al. Ozone depletion: Ultraviolet radiation and phytoplankton biology in Antarctic waters. *Science* **1992**, *255*, 952–959. [CrossRef] [PubMed]
- 98. Seinfeld, J.H.; Pandis, S.N. *Atmospheric Chemistry and Physics—From Air Pollution to Climate Change*, 2nd ed.; John Wiley and Sons: Hoboken, NJ, USA, 2006; 1203p.
- 99. Wang, S.; Huang, H.; Kahnt, J.; Thauer, R.K. A Reversible Electron-Bifurcating Ferredoxin- and NAD-Dependent [FeFe]-Hydrogenase (HydABC) in *Moorella thermoacetica*. *J. Bacteriol.* **2013**, 195, 1267–1275. [CrossRef] [PubMed]
- 100. Granick, S. Evolution of heme and chlorophyll. In *Evolving Genes and Proteins*; Bryson, V., Vogel, H.J., Eds.; Academic Press: New York, NY, USA, 1965; pp. 67–88.
- 101. Olson, J.M.; Pierson, B.K. Evolution of reaction centers in photosynthetic prokaryotes. *Int. Rev. Cytol.* **1987**, *108*, 209–248. [PubMed]
- 102. Mauzerall, D. Light, iron, Sam Granick and the origin of life. *Photosynth. Res.* **1992**, *33*, 167–175. [CrossRef] [PubMed]
- 103. Olson, J.M. Early evolution of chlorophyll-based photosystems. *Chemtracts Biochem. Mol. Biol.* **1999**, 12, 468–482.
- 104. Olson, J.M. Photosynthesis in the Archean era. Photosynth. Res. 2006, 88, 109–117. [CrossRef] [PubMed]
- 105. Kritsky, M.S.; Telegina, T.A.; Vechtomova, Y.L.; Buglak, A.A. Why flavins are not competitors of chlorophyll in the evolution of biological converters of solar energy. *Int. J. Mol. Sci.* **2013**, *14*, 575–593. [CrossRef] [PubMed]
- 106. Dodd, M.S.; Papineau, D.; Grenne, T.; Slack, J.F.; Rittner, M.; Pirajno, F.; O'Neil, J.; Little, C.T.S. Evidence for early life in Earth's oldest hydrothermal vent precipitates. *Nature* **2017**, *543*, 60–64. [CrossRef] [PubMed]
- 107. Schopf, J.W. Microfossils of the early Archean Apex chert: New evidence of the antiquity of life. *Science* **1993**, 260, 640–646. [CrossRef] [PubMed]
- 108. Brasier, M.D.; Green, O.R.; Jephocoat, A.P.; Kleppe, A.K.; Van Kranendonk, M.J.; Lindsay, J.F.; Steele, A.; Grassineau, N.V. Questioning the evidence for Earth's oldest fossils. *Nature* **2002**, *416*, 76–81. [CrossRef] [PubMed]
- 109. Hayes, J.M. Global methanotrophy at the Archean-Proterozoic transition. In *Early life on Earth*; Schopf, J.W., Ed.; Columbia University Press: New York, NY, USA, 1994.
- 110. Brocks, J.J.; Logan, G.A.; Buick, R.; Summons, R.E. Archean molecular fossils and the early rise of eukaryotes. *Science* **1999**, *285*, 1033–1036. [CrossRef] [PubMed]
- 111. Rasmmusen, B.; Fletcher, I.R.; Brocks, J.J.; Kilburn, M.R. Reassessing the first appearance of eukaryotes and cyanobacteria. *Nature* **2008**, *455*, 1101–1104. [CrossRef] [PubMed]
- 112. House, C.H.; Schopf, J.W.; McKeegan, K.D.; Coath, C.D.; Harrison, T.M.; Stetter, K.O. Carbon isotopic composition of individual Precambrian microfossils. *Geology* **2000**, *28*, 707–710. [CrossRef]
- 113. House, C.H.; Oehler, D.Z.; Sugitani, K.; Mimura, K. Carbon isotopic analyses of ca. 3.0 Ga microstructures imply planktonic autotrophs inhabited Earth's early oceans. *Geology* **2013**, *41*, 651–654. [CrossRef]
- 114. Kudryavtsev, A.B.; Schopf, J.W.; Agresti, D.G.; Wdowiak, T.J. In situ laser-Raman imagery of Precambrian microscopic fossils. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 823–826. [CrossRef] [PubMed]
- 115. Ueno, Y.; Isozaki, Y.; Yurimoto, H.; Maruyama, S. Carbon isotopic signatures of individual Archean microfossils (?) from Western Australia. *Int. Geol. Rev.* **2001**, *43*, 196–212. [CrossRef]
- 116. Schopf, J.W.; Kudryavtsev, A.B.; Agresti, D.G.; Wdowiak, T.J.; Czaja, A.D. Laser-Raman imagery of Earth's earliest fossils. *Nature* **2002**, *416*, 73–76. [CrossRef] [PubMed]
- 117. Schopf, J.W.; Kudryavtsev, A.B.; Agresti, D.G.; Czaja, A.D.; Wdowiak, T.J. Raman imagery: A new approach to assess the geochemical maturity and biogenicity of permineralized Precambrian fossils. *Astrobiology* **2005**, *5*, 333–371. [CrossRef] [PubMed]
- 118. Schopf, J.W.; Kitajima, K.; Spicuzza, M.J.; Kudryavtsev, A.B.; Valley, J.W. SIMS analyses of the oldest known assemblage of microfossils document their taxon-correlated carbon isotope compositions. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 53–58. [CrossRef] [PubMed]
- 119. Oehler, D.Z.; Robert, F.; Mostefaoui, S.; Meibom, A.; Selo, M.; McKay, D.S. Chemical mapping of Proterozoic organic matter at submicron spatial resolution. *Astrobiology* **2006**, *6*, 838–850. [CrossRef] [PubMed]

120. Igisu, M.; Nakashima, S.; Ueno, Y.; Awramik, S.M.; Maruyama, S. In situ infrared microspectroscopy of ~850 million-year-old prokaryotic fossils. *Appl. Spectrosc.* **2006**, *60*, 1111–1120. [CrossRef] [PubMed]

- 121. Igisu, M.; Ueno, Y.; Shimojima, M.; Nakashima, S.; Awramik, S.M.; Ohta, H.; Maruyama, S. Micro-FTIR signature of bacterial lipids in Proterozoic microfossils. *Precambrian Res.* **2009**, *173*, 19–26. [CrossRef]
- 122. Van Zuilen, M.A.; Chaussidon, M.; Rollion-Bard, C.; Marty, B. Carbonaceous cherts of the Barberton Greenstone Belt, South Africa: Isotopic, chemical and structural characteristics of individual microstructures. *Geochim. Cosmochim. Acta* **2007**, 71, 655–669. [CrossRef]
- 123. Wacey, D.; Kilburn, M.R.; Saunders, M.; Cliff, J.; Brasier, M.D. Microfossils of sulphur-metabolizing cells in 3.4-billion-year-old rocks of Western Australia. *Nat. Geosci.* **2011**, *4*, 698–702. [CrossRef]
- 124. Wacey, D.; Battison, L.; Garwood, R.J.; Hickman-Lewis, K.; Brasier, M.D. Advanced analytical techniques for studying the morphology and chemistry of Proterozoic microfossils. *Geol. Soc. Lond. Spec. Publ.* **2016**, 448, 81–104. [CrossRef]
- 125. Lepot, K.; Willford, K.H.; Ushikubo, T.; Sugitani, K.; Mimura, K.; Spicuzza, M.J.; Valley, J.W. Texture-specific isotopic compositions in 3.4 Gyr old organic matter support selective preservation in cell like structures. *Geochim. Cosmochim. Acta* 2013, 112, 66–86. [CrossRef]
- 126. Williford, K.H.; Ushikubo, T.; Schopf, J.W.; Lepot, K.; Kitajima, K.; Valley, J.W. Preservation and detection of microstructural and taxonomic correlations in the carbon isotopic compositions of individual Precambrian microfossils. *Geochim. Cosmochim. Acta* 2013, 104, 165–182. [CrossRef]
- 127. Alleon, J.; Bernard, S.; Le Guillou, C.; Marin-Carbonne, J.; Pont, S.; Beyssac, O.; McKeegan, D.D.; Robert, F. Molecular preservation of 1.88 Ga Gunflint organic microfossils as a function of temperature and mineralogy. *Nat. Commun.* 2016, 7, 11977. [CrossRef] [PubMed]
- 128. Ueno, Y.; Isozaki, Y.; McNamara, K.J. Coccoid like microstructures in a 3.0 Ga chert from Western Australia. *Int. Geol. Rev.* **2006**, *48*, 78–88. [CrossRef]
- 129. Schopf, J.W.; Kudryavtsev, A.B. Confocal laser scanning microscopy and Raman imagery of ancient microscopic fossils. *Precambrian Res.* **2009**, *173*, 39–49. [CrossRef]
- 130. Chen, J.Y.; Schopf, J.W.; Bottjer, D.J.; Zhang, C.Y.; Kudryavtsev, A.B.; Tripathi, A.B.; Wang, X.Q.; Yang, Y.H.; Gao, X.; Yang, Y. Raman spectra of a Lower Cambrian ctenophore embryo from southwestern Shaanxi, China. *Proc. Natl. Acad. Sci. USA* **2007**, 104, 6289–6292. [CrossRef] [PubMed]
- 131. Sugitani, K.; Grey, K.; Allwood, A.; Nagaoka, T.; Mimura, K.; Minami, M.; Marshall, C.P.; Van Kranendonk, M.J.; Walter, M.R. Diverse microstructures from Archean chert from the Mount Gldsworthy-Mount Grant area, Pilbara Craton, Western Australia: Microfossils, dubiofossils, or pseudofossils? *Precambrian Res.* 2007, 158, 228–262. [CrossRef]
- 132. Qu, Y.; Engdahl, A.; Zhu, S.; Vajda, V.; McLoughlin, N. Ultrastructural heterogeneity of carbonaceous material in ancient cherts: Investigating biosignature origin and preservation. *Astrobiology* **2015**, *15*, 825–842. [CrossRef] [PubMed]
- 133. Pasteris, J.D.; Wopenka, B. Necessary, but not sufficient: Raman identification of disordered carbon as a signature of ancient life. *Astrobiology* **2003**, *3*, 727–738. [CrossRef] [PubMed]
- 134. Wopenka, B.; Pasteirs, J.D. Structural characterization of kerogens to granulite-facies graphite: Applicability of Raman microprobe spectroscopy. *Am. Mineral.* **1993**, *78*, 533–557.
- 135. Lahfid, A.; Beyssac, O.; Diville, E.; Negro, F.; Chopin, C.; Goffe, B. Evolution of the Raman spectrum of carbonaceous material in low-grade metasediments of the Glarus Alps (Switzerland). *Terra Nova* **2010**, 22, 354–360. [CrossRef]
- 136. Lin, R.; Ritz, G.P. Reflectance FT-IR microspectroscopy of fossil algae contained in organic-rich shales. *Appl. Spectrosc.* **1993**, 47, 265–271. [CrossRef]
- 137. Qu, Y.; Wang, J.; Xiao, S.; Whitehouse, M.; Engdahl, A.; Wang, G.; McLoughlin, N. Carbonaceous biosignatures of diverse chemotrophic microbial communities from chert nodules of the Ediacaran Doushantuo Formation. *Precambrian Res.* **2017**, 290, 184–196. [CrossRef]
- 138. Igisu, M.; Komiya, T.; Kawashima, M.; Nakashima, S.; Ueno, Y.; Han, J.; Shu, D.; Li, Y.; Maruyama, S.; Takai, K. FTIR microspectroscopy of Ediacaran phosphatized microfossils from the Doushantuo Formation, Weng'an, South China. *Gondwana Res.* **2014**, *25*, 1120–1138. [CrossRef]
- 139. De Gregorio, B.T.; Sharp, T.G. The structure and distribution of carbon in 3.5 Ga Apex chert: Implications for the biogenicity of earth's oldest putative microfossils. *Am. Mineral.* **2006**, *91*, 784–789. [CrossRef]

140. De Gregorio, B.T.; Sharp, T.G.; Flynn, G.J.; Wirick, S.; Hervig, R.L. Biogenic origin for earth's oldest putative microfossils. *Geology* **2009**, *37*, 631–634. [CrossRef]

- 141. Alleon, J.; Bernard, S.; Le Guillou, C.; Daval, D.; Skouri-Panet, F.; Pont, S.; Delbes, L.; Robert, F. Early entombment within silica minimizes the molecular degradation of microorganisms during advanced diagenesis. *Chem. Geol.* **2016**, *437*, 98–108. [CrossRef]
- 142. Igisu, M.; Yokoyama, T.; Nakashima, S.; Ueno, Y.; Shimojima, M.; Ohta, H.; Maruyama, S. Changes of aliphatic C–H bonds in cyanobacteria during experimental thermal maturation in the presence or absence of silica as evaluated by FTIR microspectroscopy. *Geobiology* **2018**, *16*, 412–428. [CrossRef] [PubMed]
- 143. Hoekstra, F.A.; Golovina, E.A.; Buitink, J. Mechanisms of plant desiccation tolerance. *Trends Plant Sci.* **2001**, *6*, 431–438. [CrossRef]
- 144. Crowe, J.H.; Crowe, L.M.; Carpenter, J.E.; Petrelski, S.; Hoekstra, F.A.; de Araujo, P.; Panek, A.D. Anhydrobiosis: Cellular adaptation to extreme dehydration. In *Handbook of Physiology, Section 13: Comparative Physiology*; Oxford University Press: Oxford, UK, 1977; pp. 1445–1477.
- 145. Cordone, L.; Cottone, G.; Cupane, A.; Emanuele, A.; Giuffrida, S.; Levantino, M. Proteins in saccharides matrices and the trehalose peculiarity: Biochemical and biophysical properties. *Curr. Org. Chem.* **2015**, *19*, 1684–1706. [CrossRef]
- 146. Wolkers, W.F.; Bochicchio, A.; Selvaggi, G.; Hoekstra, F.A. Fourier transform infrared microspectroscopy detects changes in protein secondary structure associated with desiccation tolerance in developing maize embryos. *Plant Physiol.* **1998**, *116*, 1169–1177. [CrossRef] [PubMed]
- 147. Kudo, S.; Ogawa, H.; Yamakita, E.; Watanabe, S.; Suzuki, T.; Nakashima, S. Adsorption of water to collagen as studied by infrared (IR) micro-Spectroscopy combined with relative humidity control system and quartz crystal microbalance. *Appl. Spectrosc.* **2017**, *71*, 1621–1632. [CrossRef] [PubMed]
- 148. Vladilo, G.; Hassanali, A. Hydrogen bonds and life in the universe. Life 2018, 8, 1. [CrossRef] [PubMed]



© 2018 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).