### Heliyon 8 (2022) e10273

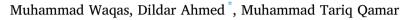
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# Heliyon

journal homepage: www.cell.com/heliyon

**Research article** 

# Surfactant-mediated extraction of capsaicin from *Capsicum annuum* L. fruit in various solvents



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#### ARTICLE INFO ABSTRACT Keywords: Capsaicin is a valuable compound found in Capsicum annuum. The present study aimed to explore the efficiency of Capsicum annuum different solvents and surfactants on its extraction by maceration. Ethyl acetate was found to be the best solvent Capsaicin followed by dichloromethane and acetone, respectively. Overall order of efficiency of the solvents used was this: Extraction ethyl acetate > dichloromethane > acetone > glycerol > acetonitrile > methanol > acetic acid > toluene. Surfactant-assisted Extractability of ethyl acetate for capsaicin remained unaffected by the surfactants. Tween-80 had very positive Solvent effect effect on the extraction efficiency of dichloromethane (DCM) and acetone. Kinetics of the extraction with the most efficient solvent ethyl acetate showed extraction of capsaicin to follow a pseudo-second order kinetic model. In conclusion, for extraction of capsaicin from green chili, ethyl acetate was the most powerful amongst the solvents used in the present work and tween-80 had a notable positive effect on the efficiency of DCM and acetone.

#### 1. Introduction

*Capsicum annuum* L. or chili is a well-known plant found through the world. Its fruit is commonly used as a spice and added in various types of cuisines and is also used in traditional medicine. The fruit of *Capsicum* species or hot peppers contains a group of compounds that give them their characteristic pungent flavour [1]. These compounds are called capsaicinoids. The major component of this group is capsaicin. Capsaicin and its related compounds are used in folk medicine for different purposes, such as to relieve pain and to treat arthritis [2, 3, 4]. The structures of some capsaicinoids are given in Table 1.

Chemically capsaicin is 8-methyl-*N*-vanillyl-6-nonenamide. In its pure form, it is a colorless, crystalline compound. It is hydrophobic and insoluble in water. The compound has been found to have remarkable property to relieve pain [5]. Apart from its use in food and medicine, capsaicin is also used in cosmetics [6]. In order to explore its possible applications to treat chronic diseases, the compound is also used in research in various fields of medicine, such as cancer [7, 8, 9]. The literature shows numerous studies on the medicinal activities of capsaicin and related compounds [10, 11, 12, 13]. Its analgesic and painkilling effect is well-known which has led to its use in pain relieving creams. Studies have also highlighted its role in treating metabolic disorders such as those related to obesity and diabetes. It has also found effective in treating some types of human cancers such as those affecting lungs, stomach, colon, and breast [13]. In fact, the literature shows an increasing number of studies focusing on the anti-cancer activities of capsaicin that has increased the value of this compound up to manifold [14, 15].

Generally, capsaicinoids obtained from hot red chilis contain 69% of capsaicin, 22% dihydrocapsaicin, 7% nordihydrocapsaicin, 1% homo-capsaicin and 1% homodihydracapsaicin [16, 17, 18].

In view of their importance as therapeutic agents, numerous studies report isolation and identification of capsaicinoids from chili and their applications [16, 19, 20]. For extraction of these compounds, several solvents and techniques have been evaluated.

In the recent years, surfactants-assisted extraction of bioactive compounds has gain attention [21, 22, 23, 24]. Surfactants are amphiphilic compounds and have a hydrophilic and a lipophilic part. In aqueous media, the surfactant molecules form micelles when the concentration of the surfactant is higher than its CMC (critical micelle concentration). They form micelles by orienting themselves in a way that their hydrophobic parts make the inner core while the hydrophilic parts make the micellar surface. Surfactants assist the extraction of bioactive compounds from the plant cells by playing role in cleaving the cell membrane and cell wall and by increasing the solubility of otherwise less soluble compounds in aqueous medium [25]. The factors affecting their effectiveness for extraction of chemical compounds from biomasses include nature of surfactants depending upon the nature of target molecules. For this purpose, the surfactants commonly used include cetyltrimethylammonium bromide (CTAB), sodium dodecyl sulfate (SDS) and tween-80.

https://doi.org/10.1016/j.heliyon.2022.e10273

Received 4 May 2022; Received in revised form 2 July 2022; Accepted 9 August 2022

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Capsaicinoids names	Structures
Capsaicin	
Dihydrocapsaicin	HO H H
Nordihydrocapsaicin	
Homodihydrocapsaicin	
Homocapsaicin	Hold Harrison
Nonivamide	

# Table 1. Capsaicin and related compounds (capsaicinoids).

There are many organic liquids that are used as solvents to extract chemical compounds from plants and other organisms. For extraction of capsaicin from chili, several studies have been conducted using various solvents. These include methanol-water, methanol-surfactant, methanolwater-surfactant, and water-surfactant solutions [26].

Quantification of capsaicin in the extracts were determined by UVvisible spectrophotometry method. This method has an obvious limitation since the extracts not only contained the desired compound capsaicin but also other phytochemicals extractable under the experimental conditions, which may contribute to the absorbance. However, the method is widely used as it is robust, convenient, and cost-effective and gives a reasonable estimate of the desired compound. The method has also been used for the quantification of capsaicin [19, 27].

With this background in view, the current work was designed with the objectives to evaluate the effectiveness of different solvents for extraction of capsaicin from chili and to investigate the effect of anionic, cationic and non-ionic surfactants on the extraction efficiency of the solvents with significant effectiveness.

#### 2. Materials and methods

### 2.1. Materials and reagents

The chemicals used in the current study were of analytical grade. CTAB, SDS, tween-80 was purchased from Sigma-Aldrich Chemicals (St. Louis, US). The solvents methanol, ethyl acetate and acetone, toluene, glycerol, acetonitrile, acetic acid and dichloromethane were obtained from Merck (Darmstadt, Germany). Standard capsaicin was purchased from Chem-Impex Int'l Inc. (Chicago, US).

#### 2.2. Collection of plant material

A sample of fresh chili (*Capsicum annuum* L.) fruit was purchased from a local market of Lahore, Pakistan. The fruit was green. It was dried in shade for 10 days at room temperature and ground to obtain a fine powder. This powder was used for extraction studies.

# 2.3. Extraction with different solvents

Several solvents were used for extraction, which included methanol, ethyl acetate, dichloromethane, acetic acid, toluene, glycerol, acetonitrile, and acetone. Based on the preliminary runs, a measured amount of the chili powder (1 g) was soaked in 50 mL of each solvent and the mixture was put on shaking for 6 h on an orbital shaker at room temperature. The mixture was filtered under gravity using Whatman filter paper no 1. In order to make it appropriate for UV-Visible spectrophotometric analysis, the filtrate was diluted with the same solvent (1 mL of the filtrate was diluted with 9 mL of the solvent). Each experiment was carried out three times.

# 2.4. Calculation of yield

The extraction yield of capsaicin was calculated by UV-visible spectrophotometry method according to the Lambert-Beer law. A standard curve of the standard capsaicin was drawn by measuring the absorbance of dilutions at 280 nm (the  $\lambda_{max}$  of capsaicin) and molar absorptivity was calculated. Absorbance of each extract was then recorded at 280 nm. The value of molar absorptivity was used to calculate the yield of capsaicin in the extract using equation of Lambert-Beer law.

#### 2.5. Extraction in the presence of surfactants

Three most effective solvents in the present study were ethyl acetate, dichloromethane, and acetone. Effect of surfactants on their extractability was evaluated. Three different surfactants were used (a) anionic (sodium dodecylsulfate or SDS), (b) non-ionic (tween-80) and (c) cationic (cetrimonium bromide or CTAB). A known amount of the chili powder (1 g) was mixed in a given amount (30 mL) of a given solvent and extraction was carried out in the presence or absence of a surfactant. The amount of a given surfactant added was a little more than its CMC. A summary of the parameters used are given in Table 2. Each experiment was carried out at least in triplicate.

#### 2.6. Study of kinetics of capsaicin extraction

A known amount of ground dried fresh chili powder (0.1 g) was soaked in 10 mL ethyl acetate solvent and was allowed to shake for 10, 20, 30, 60, 120, 240, 360, 480 and 1440 min on a shaker. The mixture was filtered under gravity using Whatman filter paper no 1. Then, 1 mL of the filtrate was diluted with 3 mL methanol for the measurement of absorbance at 280 nm. Extraction yield of capsaicin was calculated using Lambert-Beer law. Each experiment was carried out in triplicate.

# 2.7. Statistical analysis

All the determinations were made at least three times, and statistical mean was calculated with standard deviation using Microsoft Excel 365 (Las Vegas, NV, US).

### 3. Results and discussion

Capsaicin, the pungent principle in chili, is a valuable natural product because of having medicinal and analytical applications. It is highly valuable to extract this bioactive substance in high yield. Although the literature reports several studies to this effect, the yield is still a problem. The present study, therefore, was planned to explore methods to extract

Table 2. A summary of the parameters used for surfactant-assisted extraction of capsaicin from ground dried fresh green fruit of *Capsicum annuum* 

Solvent	Solvent to solid ratio (mL/g)	SDS (g)	CTAB (g)	Tween-80 (mL)
(1) Acetone	30	0.714	0.086	0.21
(2) Ethyl acetate				
(3) Dichloromethane				

The absorbance was measured at 280 nm to calculate the yield of capsaicin using Lambert-Beer law; shaking time: 6 h.

Table 3. Extraction yield of capsaicin from fresh green chili powder in different solvents.								
Solvent	Ethyl acetate	DCM	Acetone	Glycerol	Acetonitrile	Methanol	Acetic acid	Toluene
Yield (mg/10 g)	$73.97 \pm 2.25$	$35.98 \pm 4.36$	$\textbf{28.8} \pm \textbf{2.92}$	$17.78\pm0.20$	$7.87 \pm 1.82$	$7.37\pm3.24$	$4.08\pm0.94$	$2.77\pm2.03$

 Table 4. Effect of surfactants on extraction of capsaicin from fresh green chili

 fruit powder in different solvents.

Solvent and Surfactant	Yield in different solvents (mg/10 g)				
	Ethyl acetate	Dichloromethane	Acetone		
Solvent	$\textbf{73.97} \pm \textbf{2.255}$	$35.98 \pm 4.362$	$28.8\pm2.916$		
Solvent + SDS	$\textbf{72.29} \pm \textbf{2.941}$	$46.53 \pm 11.079$	$43.7\pm 6.689$		
Solvent + CTAB	$\textbf{72.62} \pm \textbf{2.649}$	$35.57 \pm 1.302$	$\textbf{46.7} \pm \textbf{13.837}$		
Solvent + Tween-80	$\textbf{72.74} \pm \textbf{0.641}$	$71.20\pm1.273$	$61.6\pm4.348$		

capsaicin from *C. annuum* in high yield. The objective was to assess the effect of three different surfactants on the extraction of capsaicin using a variety of solvents. The surfactants used included SDS, CTAB, and Tween-80. The SDS (sodium dodecylsulfate) is an anionic, CTAB (cetyl-trimethylammonium bromide) is cationic while Tween-80 is a neutral surfactant.

# 3.1. Extraction of capsaicin in different solvents

Capsaicin was extracted from dried powdered green chili using different solvents and in the presence of three different surfactants. The quantification of capsaicin in the extracts were done according to UV-visible spectrophotometry method by measuring absorbance at the  $\lambda$ max of capsaicin. The results are shown in Table 3.

As Table 3 shows, ethyl acetate was found to be the most effective solvent to extract capsaicin from the powder of dried green chili followed by dichloromethane and acetone, respectively. Ethyl acetate provided maximum yield (73.97 mg out of 10 g sample). On the other hand, dichloromethane provided 35.98 mg out of 10 g sample. Acetone also proved to be a good solvent after dichloromethane and provided 28.88 mg out of 10 g sample. The extraction efficiency of ethyl acetate was, therefore, remarkably higher than the other solvents used in the study. Capsaicin and other capsaicinoids are phenolic compounds and the

efficiency of ethyl acetate to extract phenolic has been reported in many studies [28]. The importance of the study stems from the need to discover the most efficient ways to extract capsaicin and related compounds as well as the other bioactive compounds found in chili such as phenolics and carotenoids [29, 30]. Since one of the most important factors affecting extraction of chemical constituents of a plant material is choice of solvents, the findings of the present work are important for future studies. Ethyl acetate has been found not only a very efficient extracting medium for capsaicin, but it is also considered an environmentally friendly or green solvent [31, 32, 33]. The efficacy of ethyl acetate in extracting phenolics has been attributed to its moderately polar nature [34]. An ethyl acetate extract of chili, therefore, should be considered as safe product for industrial applications such as its use in pain killer creams for topical applications [35, 36].

#### 3.2. Effect of surfactants on the extractability of solvents

Effect of surfactants on the extraction of capsaicin from chili material was investigated and the results are shown in Table 4.

As Table 4 shows, the surfactants had little effect on the extraction efficiency of ethyl acetate. However, all the surfactants had a pronounced impact on the efficiency of acetone to extract capsaicin, and Tween-80 was most effective followed by CTAB. Tween-80 had very positive effect on the extraction efficiency of dichloromethane, the trend supported other studies that have shown non-ionic surfactants to be more effective than the ionic ones [23]. SDS too had a fairly good positive effect. However, CTAB had almost no effect. Since Tween-80 is known to be a nontoxic surface acting compound, it can be used for extracting valuable bioactive compounds for nutraceutical or therapeutic purposes without any concern about the safety [21]. The efficacy of tween-80 in extracting phenolic compounds has also been demonstrated by other studies [37]. It has been suggested that the surfactant binds itself with phenolics through hydrogen bonding and thereby assisted in their extraction [38].

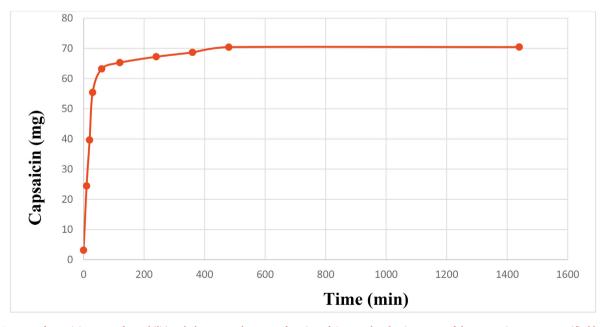


Figure 1. Amount of capsaicin extract from chili in ethyl acetate solvent as a function of time; each value is a mean of three experiments as quantified by measuring absorbance of the extracts at 280 nm.

### 3.3. Study of rate kinetics for capsaicin extraction

Amount of capsaicin extract from chili as a function of time was determined for kinetics study. The results are shown in Figure 1.

The kinetic study of the extraction with the most efficient solvent ethyl acetate exhibited interesting results, according to which the rate of extraction of capsaicin with ethyl acetate appeared to follow the second order kinetic model. Similar trend has been found in extraction of polyphenols in other studies [39, 40]. Notably, during the first hour of the extraction, there was a rapid extraction of capsaicin after the extraction slowed down and attained a steady state. The study, therefore, suggests that most capsaicin is extractable in the first 60 min of the process.

As capsaicin is a valuable bioactive compound with various actual and potential medicinal applications, exploring efficient methods for its extraction from chili is important [13, 29]. Numerous studies have been carried out to this end, but the extraction yield is still a problem [36]. The present work, therefore, is a contribution to the ongoing work on this subject paving way for further in-depth studies. The efficacy of ethyl acetate as extracting medium for capsaicin has been demonstrated with the finding that most of extraction occurs within the first hour of the process.

#### 4. Conclusions

Capsaicin is a highly valuable compound found in chili peppers. It is, therefore, of great importance to discover efficient methods of its extraction. The realization led to the current study that demonstrated ethyl acetate to be a highly effective solvent for extracting capsaicin from chili. Out of the several solvents used, ethyl acetate gave the highest extraction yield with the efficiency order: ethyl acetate > dichloromethane > acetone > glycerol > acetonitrile > methanol > acetic acid > toluene. Moreover, the non-ionic surfactant tween-80 strongly enhanced the extracting ability of acetone and dichloromethane. Thus, based on the findings of the present study, new methods may be developed and optimized to obtain high yield of capsaicin from chili.

#### Declarations

#### Author contribution statement

Muhammad Waqas: Performed the experiments; Analyzed and interpreted the data; Wrote the paper. Dildar Ahmed: Conceived and designed the experiments; Analyzed and interpreted the data; Wrote the paper. Muhammad Tariq Qamar: Analyzed and interpreted the data; Contributed reagents, materials, analysis tools or data; Wrote the paper.

#### Funding statement

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

## Data availability statement

Data will be made available on request.

### Declaration of interest statement

The authors declare no conflict of interest.

#### Additional information

No additional information is available for this paper.

# Acknowledgements

The authors are grateful to the Department of Chemistry, FCCU, for providing lab facilities for this work.

#### References

- T. Watanabe, T. Kawada, M. Yamamoto, K. Iwai, Capsaicin, a pungent principle of hot red pepper, evokes catecholamine secretion from the adrenal medulla of anesthetized rats, Biochem. Biophys. Res. Commun. 142 (1987) 259–264.
- [2] V. Govindarajan, D. Rajalakshmi, N. Chand, U.J. Salzer, Capsicum production, technology, chemistry, and quality. Part IV. Evaluation of quality, Crit. Rev. Food Sci. Nutr. 25 (1987) 185–282.
- [3] C. Caruntu, C. Negrei, M. Ilie Ghita, A. Caruntu, A. Ioana Bădărău, I.B. Ioan Buraga, D. Boda, A. Albu, D. Brăni Steanu, Capsaicin, a hot topic in skin pharmacology and physiology, FARMACIA 63 (2015) 487–491.
- [4] S. Basith, M. Cui, S. Hong, S. Choi, Harnessing the therapeutic potential of capsaicin and its analogues in pain and other diseases, Molecules 21 (2016) 966.
- [5] M. Hayman, P.C.A. Kam, Capsaicin: a review of its pharmacology and clinical applications, Curr. Anaesth. Crit. Care 19 (2008) 338–343.
- [6] I. Musfiroh, M. Mutakin, T. Angelina, M. Muchtaridi, Capsaicin level of various capsicum fruits, Int. J. Pharm. Pharmaceut. Sci. 5 (2013) 248–251.
- [7] A.M. Chapa-Oliver, L. Mejfa-Teniente, Capsaicin: from plants to a cancersuppressing agent, Molecules 21 (2016) 931.
- [8] A. Rasul, B.J. Zhao, J. Liu, B. Liu, J.X. Sun, J. Li, X.M. Li, Molecular mechanisms of casticin action: an update on its antitumor functions, Asian Pac. J. Cancer Prev. APJCP 15 (2014) 9049–9058.
- [9] Y.J. Choi, J.Y. Kim, S.B. Yoo, J.H. Lee, J.W. Jahng, Repeated oral administration of capsaicin increases anxiety-like behaviours with prolonged stress-response in rats, J. Biosci. 38 (2013) 561–571.
- [10] K. Bley, G. Boorman, B. Mohammad, D. McKenzie, S. Babbar, A comprehensive review of the carcinogenic and anticarcinogenic potential of capsaicin, Toxicol. Pathol. 40 (2012) 847–873.
- [11] K. Srinivasan, Biological activities of red pepper (*Capsicum annuum*) and its pungent principle capsaicin: a review, Crit. Rev. Food Sci. Nutr. 56 (2016) 1488–1500.
- [12] S.M. McConachie, R.A. Caputo, S.M. Wilhelm, P.B. Kale-Pradhan, Efficacy of capsaicin for the treatment of cannabinoid hyperemesis syndrome: a systematic review, Ann. Pharmacother. 53 (2019) 1145–1152.
- [13] F. Wang, Y. Xue, L. Fu, Y. Wang, M. He, L. Zhao, X. Liao, Extraction, purification, bioactivity and pharmacological effects of capsaicin: a review, Crit. Rev. Food Sci. Nutri. 62 (2022) 5322–5348.
- [14] L. Wu, S. Xu, X. Cheng, L. Zhang, Y. Wang, J. Wu, J. Bao, H. Yu, R. Lu, Capsaicin inhibits the stemness of anaplastic thyroid carcinoma cells by triggering autophagylysosome mediated OCT4A degradation, Phytother Res. 36 (2022) 938–950.
- [15] S. Kunjiappan, M. Sankaranarayanan, B.K. Kumar, P. Pavadai, E. Babkiewicz, P. Maszczyk, E. Glodkowska-Mrowka, S. Arunachalam, S.R.K. Pandian, V. Ravishankar, S. Baskararaj, S. Vellaichamy, L. Arulmani, T. Panneerselvam, Capsaicin-loaded solid lipid nanoparticles: design, biodistribution, in silico modeling and in vitro cytotoxicity evaluation, Nanotechnol 32 (2021), 095101.
- [16] A. Da Silva Antonio, L.S.M. Wiedemann, J.V.F. da Veiga, Food pungency: the evolution of methods for capsaicinoid analysis, Food Anal. Methods 12 (2019) 1327–1345.
- [17] A.S. Antonio, L.S.M. Wiedemann, J.V.F. Veiga, The genus: capsicum: a phytochemical review of bioactive secondary metabolites, RSC Adv 8 (2018) 25767–25784.
- [18] R. Alonso-Salces, E. Korta, A. Barranco, L. Berrueta, B. Gallo, F. Vicente, Pressurized liquid extraction for the determination of polyphenols in apple, J. Chromatogr. A 933 (2001) 37–43.
- [19] A. González-Zamora, E. Sierra-Campos, R. Pérez-Morales, C. Vázquez-Vázquez, M.A. Gallegos-Robles, J.D. López-Martínez, J.L. García-Hernández, Measurement of capsaicinoids in Chiltepin Hot Pepper: a comparison study between Spectrophotometric method and high-performance liquid chromatography analysis, J. Chem. 2015 (2015) 1–10.
- [20] M. Hussain, M.T. Qamar, D. Ahmed, Microwave-and ultrasound-assisted extraction of capsaicin from *Capsicum annuum* using deep eutectic solvents, Int. J. Veg. Sci. 28 (2021) 312–319.
- [21] F.A. Li, Y.W. Raza, X.Q. Wang, Xu, G.H. Chen, Optimization of surfactant-mediated, ultrasonic-assisted extraction of antioxidant polyphenols from rattan tea (*Ampelopsis grossedentata*) using response surface methodology, Phcog. Mag. 13 (2017) 446–453.
- [22] O. Čudina, K. Karljiković-Rajić, I. Ruvarac-Bugarčić, I. Janković, Interaction of hydrochlorothiazide with cationic surfactant micelles of cetyltrimethylammonium bromide, Colloids Surf. A Physicochem. Eng. Asp. 256 (2005) 225–232.
- [23] R. Hosseinzadeh, K. Khorsandi, S. Hemmaty, Study of the effect of surfactants on extraction and determination of polyphenolic compounds and antioxidant capacity of fruits extracts, PLoS One 8 (2013), e57353.
- [24] R. Hosseinzadeh, M. Gheshlagi, Interaction and micellar solubilization of diclofenac with cetyltrimethylammonium bromide: a spectrophotometric study, Collect. Czech Chem. Commun. 74 (2009) 503–513.
- [25] D.F. Cortés-Rojas, C.R.F. Souza, W.P. Oliveira, Surfactant mediated extraction of antioxidants from Syzygium aromaticum, Separ. Sci. Technol. 50 (2015) 207–213.
- [26] Y.Y. Dang, H. Zhang, Z.L. Xiu, Three-liquid-phase extraction and separation of capsanthin and capsaicin from Capsicum annum L, Czech J. Food Sci. 32 (2014) 109–114.

#### M. Waqas et al.

- [27] K.G. Liljana, M. Viktorija, S.D. Marija, G. Rubin, I.J. Emilija, The effect of different methods of extractions of capsaicin on its content in the capsicum oleoresins, Scientific Works Volume Lx Food Sci. Eng. Technol. 60 (2013) 917–922.
- [28] I. Atique, D. Ahmed, M. Maqsood, W. Malik, Solvents for extraction of antidiabetic, iron chelating, and antioxidative properties from Bottle Gourd fruit, Int. J. Veg. Sci. 24 (2017) 212–226.
- [29] B.K. Saleh, A. Omer A, B. Teweldemedhin, Medicinal uses and health benefits of chili pepper (Capsicum spp.): a review, MOJ Food Process Technol 6 (2018) 325– 328.
- [30] S. Idrees, M.A. Hanif, M.A. Ayub, A. Hanif, T.M. Ansari, Medicinal Plants of South Asia, Elsevier, 2020, pp. 113–124. Chapter 9 – Chili Pepper.
- [31] S. Yewale, Z. Farash, S. Kulkarni, S. Palghadmal, N. Athawale, L. Sawant, S. Bhope, S. Padmanabhan, Effect of solvent polarity on extraction yield of total flavonoids with special emphasis to glabridin from *Glycyrrhiza glabra* roots, Fabad J. Pharm. Sci. 1 (2022) 1–12.
- [32] X. Zhang, Y. Shen, W. Prinyawiwatkul, Z. Xu, Volatile compounds in fresh-cut pineapple heated at different temperatures, J. Food Process. Preserv. 36 (2012) 567–573.
- [33] D. Pintać, T. Majkić, L. Torović, D. Orčić, I. Beara, N. Simin, N. Mimica–Dukić, M. Lesjak, Solvent selection for efficient extraction of bioactive compounds from grape pomace, Ind. Crop. Prod. 111 (2018) 379–390.

- [34] M.O. Gaylor, H.L. Juntunen, D. Hazelwood, P. Videau, Assessment of multiple solvents for extraction and direct GC–MS determination of the phytochemical inventory of sansevieria extrafoliar nectar droplets, J. Chromatogr. Sci. 56 (2018) 293–299.
- [35] P.F. White, Red-hot chili peppers: a spicy new approach to preventing postoperative pain, Anesth. Analg. 107 (2008) 6–8.
- [36] A. Azlan, S. Sultana, C.S. Huei, M. Razman, Antioxidant, anti-obesity, nutritional and other beneficial effects of different chili pepper: a review, Molecules 27 (2022) 898.
- [37] L. Skrypnik, A. Novikova, Response surface modeling and optimization of polyphenols extraction from apple pomace based on nonionic emulsifiers, Agronomy 10 (2020) 92.
- [38] C. Li, X. Meng, M. Tian, S. Li, Y. Tian, T. Wang, C. Zhao, A novel method to extract juglone from *Juglans mandshurica* waste branches using a water-in-oil microemulsion, Waste Biomass Valor 13 (2022) 1547–1563.
- [39] M. Cavdarova, D.P. Makris, Extraction kinetics of phenolics from carob (*Ceratonia siliqua L.*) kibbles using environmentally benign solvents, Waste Biomass Valor 5 (2014) 773–779.
- [40] O.R. Alara, N.H. Abdurahman, Kinetics studies on effects of extraction techniques on bioactive compounds from Vernonia cinerea leaf, J. Food Sci. Technol. 56 (2019) 580–588.