

## TECHNICAL NOTE

# Diffusion-weighting Caused by Spoiler Gradients in the Fast Imaging with Steady-state Precession Sequence May Lead to Inaccurate $T_2$ Measurements in MR Fingerprinting

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Magnetic resonance fingerprinting (MRF) is a promising framework that allows the quantification of multiple magnetic resonance parameters with a single scan. MRF using fast imaging with steady-state precession (MRF-FISP) has robustness to off-resonance artifacts and has many applications in inhomogeneous fields. However, the spoiler gradient used in MRF-FISP is sensitive to diffusion motion, and may lead to quantification errors when the spoiler moment increases. In this study, we examined the effect of the diffusion weighting in MRF-FISP caused by spoiler gradients. The  $T_2$  relaxation times were greatly underestimated when large spoiler moments were used. The  $T_2$  underestimation was prominent for tissues with large values of  $T_2$  and diffusion coefficients. The  $T_2$  bias was almost independent of the apparent diffusion coefficient (ADC) and  $T_2$  values when the ADC map was measured and incorporated into the matching process. These results reveal that the  $T_2$  underestimation resulted from the diffusion weighting caused by the spoiler gradients.

**Keywords:** *diffusion weighting, magnetic resonance fingerprinting, magnetic resonance fingerprinting using fast imaging with steady-state precession, spoiler gradient,  $T_2$  underestimation*

## Introduction

Conventional clinical MRI scanners can detect disease pathologies, but in most cases, they are qualitative in nature. In contrast, quantitative MRI measurements could more directly reflect disease changes and be more helpful for disease detection and follow-up. These quantitative MRI assessments include  $T_1$  and  $T_2$  relaxation times,<sup>1</sup> proton density, diffusion<sup>2,3</sup> and perfusion<sup>4</sup> parameters, magnetic transfer effects,<sup>5</sup> and other tissue parameters. Despite many efforts, quantitative MRI measurements still face technical challenges, e.g., reproducibility and lengthy scan times to obtain multiple parameters.

Recently, a new framework for quantifying multiple tissue parameters in a single scan, called magnetic resonance fingerprinting (MRF), has been proposed.<sup>6</sup> MRF has the potential to overcome the limitations of conventional quantitative methods, enabling routine quantitative MRF in clinical and preclinical environments.

Magnetic resonance fingerprinting uses the responses of tissues or materials to repeated excitation and acquisition

schemes. MRF acquires a transient-state signal with pseudorandom acquisition parameters. The acquired signal is matched with a predetermined dictionary of possible signal evolutions calculated with appropriate tissue and system parameters, and the best-matched entry is used to assign multiple tissue properties simultaneously.

The acquisition schemes that are mostly used in the MRF framework are the balanced steady-state-based MRF (MRF-bSSFP)<sup>6</sup> and MRF using fast imaging with steady-state precession (MRF-FISP).<sup>7</sup> MRF-bSSFP has the advantage of high signal intensity, but quantification of relaxation parameters is affected by banding artifacts arising from the  $B_0$  inhomogeneity. In contrast, MRF-FISP uses the unbalanced gradient moment, which can alleviate the banding artifacts, and extends the application of MRF to inhomogeneous fields that are potentially challenging for MRF-bSSFP.

However, MRF-FISP can be affected by banding artifacts where there is a large off-resonance variation in an imaging volume, such as in high field strengths,<sup>8</sup> under insufficient shimming, and for wide FOV scanners. Moreover, it is not straightforward to suppress the banding artifacts in regions where sharp susceptibility transitions exist, such as nasal cavities and auditory canals.

In such cases, the use of much stronger spoiler gradients is a simple way to alleviate banding artifacts. However, gradient-spoiled imaging is particularly sensitive to diffusive motion, especially when the spoiler gradient precedes imaging.<sup>9–11</sup>

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Therefore, the increased gradient moment in MRF-FISP also causes increased sensitivity to diffusion and motion along the gradient direction. However, the diffusion-weighting effect has been neglected in the MRF-FISP framework and is not generally built into the MRF-FISP dictionaries that are used for pattern matching. The neglect of diffusion may lead to false estimates of relaxation times.

In this study, we examined the effect of diffusion weighting in MRF-FISP with spoiler moments, and we showed that the diffusion effect results in  $T_2$  underestimation, and that the  $T_2$  bias for tissues with large  $T_2$  and diffusion coefficients is nonnegligible even for moderate gradient moments.

## Methods

### Samples

For the phantom study, we used nine test tubes filled with water with  $\text{CuSO}_4$  (0.13, 0.4, and 2 g/l) to alter  $T_1$  and  $T_2$  and with glycerol (0, 12.5, and 25%) to alter the diffusion coefficient. For the reference, the  $T_1$ ,  $T_2$ , and apparent diffusion coefficient (ADC) maps were acquired using standard methods. The  $T_1$  map was acquired with inversion-recovery spin-echo sequences (inversion time = 20, 50, 100, 200, 400, 800, 1500, and 3000 ms; TE/TR = 10/8000 ms) and the  $T_2$  map was acquired with a multiple spin echo (MSE) sequence (interecho spacing = 10 ms, TR = 8000 ms). The ADC map was estimated using the standard Stejskal-Tanner method ( $b = 0, 35.1, 141, \text{ and } 316 \text{ s/mm}^2$ ).

### MRI systems

The phantoms were measured on a vertical bore 4.74T system (Oxford Instruments, UK; bore diameter = 88.3 mm) with shielded gradients, a home-built radio-frequency coil, and a digital MRI console<sup>12</sup> (DTRX6, MRTechnology, Japan). The field homogeneity was about 1 ppm over 20 mm in diameter of spherical volume (DSV).

For the *in vivo* study, MRF-FISP measurements were performed on a healthy C57BL/6JJcl mouse brain using a small-bore 1.5T system (bore diameter = 280 mm; JASTEC, Kobe, Japan) with shielded gradients. The field homogeneity was 50 ppm over 160 mm DSV.

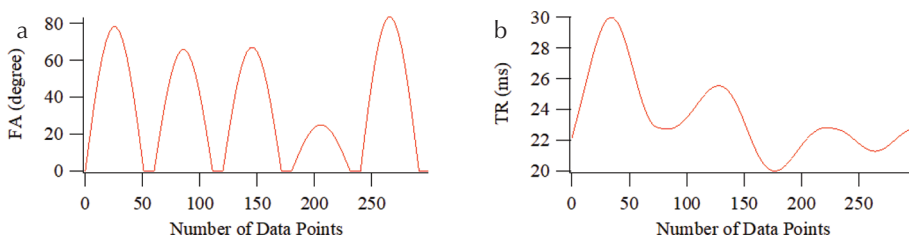
### MRF pulse sequences

The MRF-FISP acquisition was initiated with an adiabatic inversion pulse, which was followed by  $N$  successive FISP

acquisition periods with varying flip angle (FA) and TR.  $N = 300$  and TE was held constant (5 ms). FA was varied pseudo-randomly and sinusoidally ( $5\text{--}90^\circ$ ) and TR was varied using a Perlin noise pattern (20–30 ms) (Fig. 1). To avoid the potential imaging artifacts arising from hardware imperfections, Cartesian sampling<sup>13</sup> was used for the MRF acquisition. The acquired MRF signal evolution profile was matched to an MRF dictionary. To correct the  $B_1$  inhomogeneity, the  $B_1$  dimension was added to the MRF dictionaries, and a  $B_1$  map measured with a double-angle method was incorporated into the dictionary-matching process. Vector-based inner product comparisons were used as a matching algorithm. To examine the effect of diffusion weighting in MRF-FISP, the moment of the spoiler gradient was varied from  $2\pi$  to  $32\pi$  per slice thickness. Here, we set the slice thickness to be 2.5 mm, and the moment ranged from 9.5 to 150 mT/m $\times$ ms.

Conventionally, the MRF-FISP dictionary is generated without considering the diffusion effect. To consider this effect, we generated two types of MRF-FISP dictionaries; one without the diffusion effect (dictionary D $-$ ) and one with the diffusion effect (dictionary D $+$ ). The MRF dictionaries D $-$  with a range of  $T_1$  (20–1500 ms),  $T_2$  (10–1200 ms),  $B_1$  (0.6–1.2) were created using an extended phase graph (EPG) algorithm.<sup>14</sup> The MRF-FISP dictionary D $+$  with the additional dimension of the diffusion coefficient  $D$  ( $0\text{--}2.6 \times 10^{-3} \text{ mm}^2/\text{s}$ ) was also generated using EPG with isotropic diffusion operators,  $\tilde{D}_L(k, t) = \exp\left(-\frac{1}{T_D}k^2t\right)$  and  $\tilde{D}_T(k, t) = \exp\left(-\frac{1}{T_D}\left(k^2 + k + \frac{1}{3}\right)t\right)$ , where  $\tilde{D}_L$  and  $\tilde{D}_T$  are the diffusion operators for the longitudinal and transverse configuration states, respectively,  $k$  is the Fourier space coordinate, and  $T_D = \frac{1}{D(\gamma G \tau)^2}$  where  $\gamma G \tau$  is the zeroth moment of the spoiler gradient.

For the phantom study, the slice thickness was 2.5 mm, and the FOV was  $25.6 \times 25.6 \text{ mm}^2$ . The whole TR was 12 s, and the measurement time was 26 min for MRF-FISP [number of excitations (NEX) = 1]. For the *in vivo* study, the slice thickness was 2.5 mm, and the FOV was  $32 \times 32 \text{ mm}^2$ . The whole TR was 12 s, and the measurement time was 26 min. The matrix size was  $128 \times 128$  for all the measurements.



**Fig. 1** Patterns of (a) flip angle (FA) and (b) TR used for the magnetic resonance fingerprinting with steady-state precession (MRF-FISP) sequence.

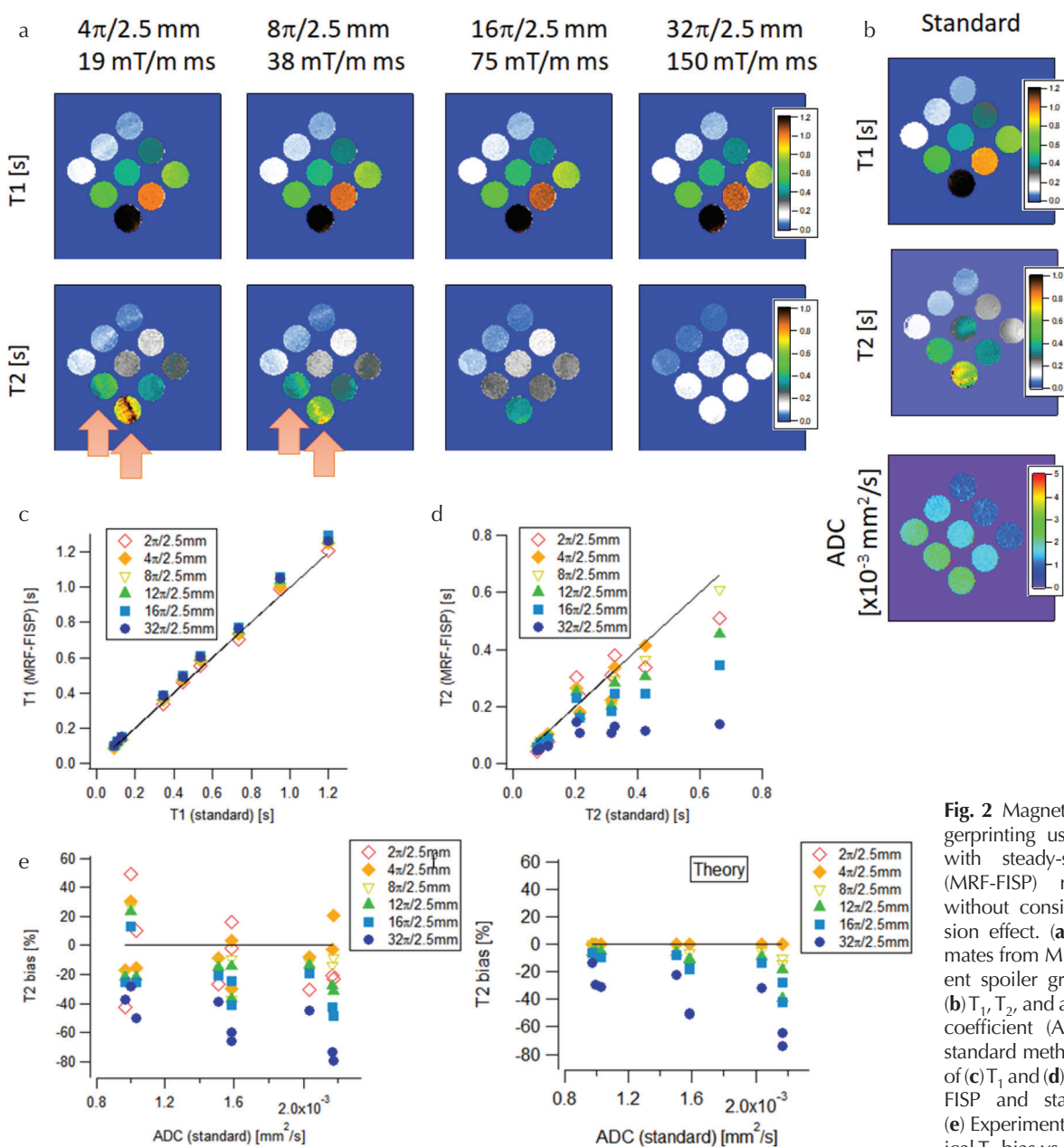
## Results

### Magnetic resonance fingerprinting using fast imaging with steady-state precession

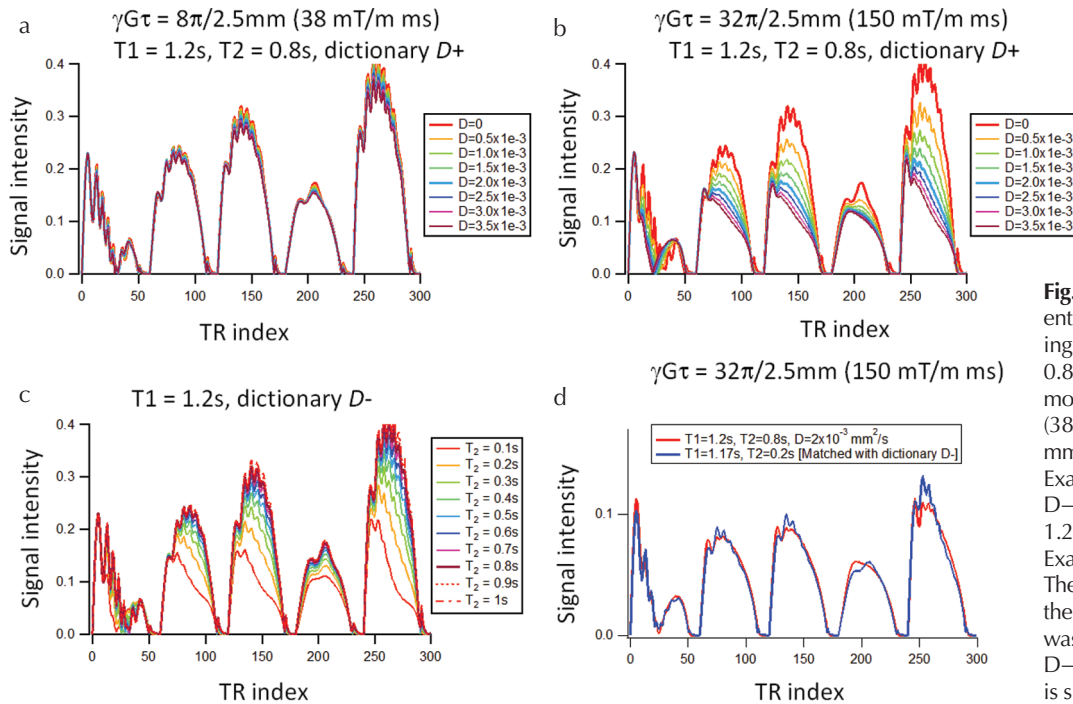
Figure 2 shows  $T_1$  and  $T_2$  estimates from MRF-FISP using dictionary  $D^-$  (excluding diffusion). At small spoiler moments, both the  $T_1$  and  $T_2$  maps suffered from banding artifacts (indicated by arrows in the figure) and showed large variations in the estimated values in the region where the banding appeared. As the moment of the spoiler gradient increased, the banding artifacts became less clear, and the  $T_1$  and  $T_2$  values became almost constant in each phantom. The  $T_1$  estimates were almost independent of the moment, and showed a high correlation with the  $T_1$  standards (Fig. 2c).

However, the  $T_2$  estimates significantly decreased as the moment increased (Fig. 2d), and deviated from the  $T_2$  standards. The  $T_2$  bias, which was defined as the difference between the  $T_2$  estimates and the  $T_2$  standards, became severe (increased negatively) as the moment increased (Fig. 2e). For large moments, the  $T_2$  bias increased negatively as the ADC increased. For small moments, the  $T_2$  bias had large variations because of the banding artifact.

Figure 3a and 3b show an example of MRF-FISP signal entries from dictionary  $D^+$  (including diffusion). The signals were almost independent of  $D$  for the moderate moment ( $38 \text{ mT/m} \times \text{ms}$ ). However, they decreased significantly with  $D$  for the large moment ( $150 \text{ mT/m} \times \text{ms}$ ). Similar behavior is observed in the signal entries of dictionary  $D^-$  (Fig. 3c); the



**Fig. 2** Magnetic resonance fingerprinting using fast imaging with steady-state precession (MRF-FISP) results obtained without considering the diffusion effect. (a)  $T_1$  and  $T_2$  estimates from MRF-FISP for different spoiler gradient moments. (b)  $T_1$ ,  $T_2$ , and apparent diffusion coefficient (ADC) maps from standard methods. Comparison of (c)  $T_1$  and (d)  $T_2$  between MRF-FISP and standard methods. (e) Experimental and (f) theoretical  $T_2$  bias vs ADC.



**Fig. 3** (a and b) Example of entries in dictionary  $D^+$  including diffusion for  $T_1 = 1.2 \text{ s}$ ,  $T_2 = 0.8 \text{ s}$ , and varying  $D$ . The moment was (a)  $8\pi/2.5 \text{ mm}$  ( $38 \text{ mT/m} \times \text{ms}$ ) and (b)  $32\pi/2.5 \text{ mm}$  ( $150 \text{ mT/m} \times \text{ms}$ ). (c) Example of entries in dictionary  $D^-$  excluding diffusion for  $T_1 = 1.2 \text{ s}$  and varying  $D$ . (d) Example of  $T_2$  underestimation. The signal evolution including the diffusion effect (red line) was matched with dictionary  $D^-$  and the best-matched entry is shown as a blue line.

signal intensity increased as  $T_2$  increased. As shown in Fig. 3d, the signal evolutions for large  $D$  values in  $D^+$  (Fig. 3b) were similar to those for small  $T_2$  values in  $D^-$  (Fig. 3c). This characteristic would lead to  $T_2$  underestimation when dictionary  $D^-$  was used for matching.

To estimate the theoretical  $T_2$  bias caused by the diffusion effect, the signal evolution in dictionary  $D^+$  was matched with that in dictionary  $D^-$  and the resulting bias between the true  $T_2$  value and the best-matched  $T_2$  value was calculated. Figure 2f shows the theoretical  $T_2$  bias for the nine phantoms. It was negligibly small for the small moment, while it became severe for large moments and large ADC values. These features agreed with those observed in the experimental results (Fig. 2e).

To validate experimentally that the  $T_2$  underestimation originates from the diffusion effect, the ADC map was also measured and incorporated into the matching processes with dictionary  $D^+$ . Figure 4 shows the resulting  $T_1$  and  $T_2$  estimates. Both estimates agreed with their standard values for different gradient moments (Fig. 4c and 4d), and the  $T_2$  bias was almost independent of the ADC value (Fig. 4e). These results support the claim that the measured  $T_2$  underestimation originates from the diffusion weighting caused by the spoiler gradient.

Figure 5 shows the theoretical  $T_2$  bias for different gradient moments calculated without including diffusion. At a fixed moment, the  $T_2$  underestimation became larger as the theoretical  $T_2$  and  $D$  values became larger. The  $T_2$  bias was almost independent of the true  $T_1$ . Figures 5c and d show the  $T_2$  bias estimated for  $T_1$ ,  $T_2$ , and  $D$  values of typical tissues<sup>15–28</sup> (Table 1). For tissues with relatively short  $T_2$  and small  $D$  values (e.g., white matter and gray matter), the  $T_2$

bias was not severe even with the large moments, but those with long  $T_2$  and large  $D$  values showed severe  $T_2$  bias even with the small moment. For example, the  $T_2$  bias for cerebrospinal fluid (CSF) ( $T_1 = 5000 \text{ ms}$ ,  $T_2 = 2100 \text{ ms}$ , and  $D = 3.2 \times 10^{-3} \text{ mm}^2/\text{s}$ ) reached to  $-26\%$  even for the moment of  $19 \text{ mT/m} \times \text{ms}$  ( $4\pi/2.5 \text{ mm}$ ).

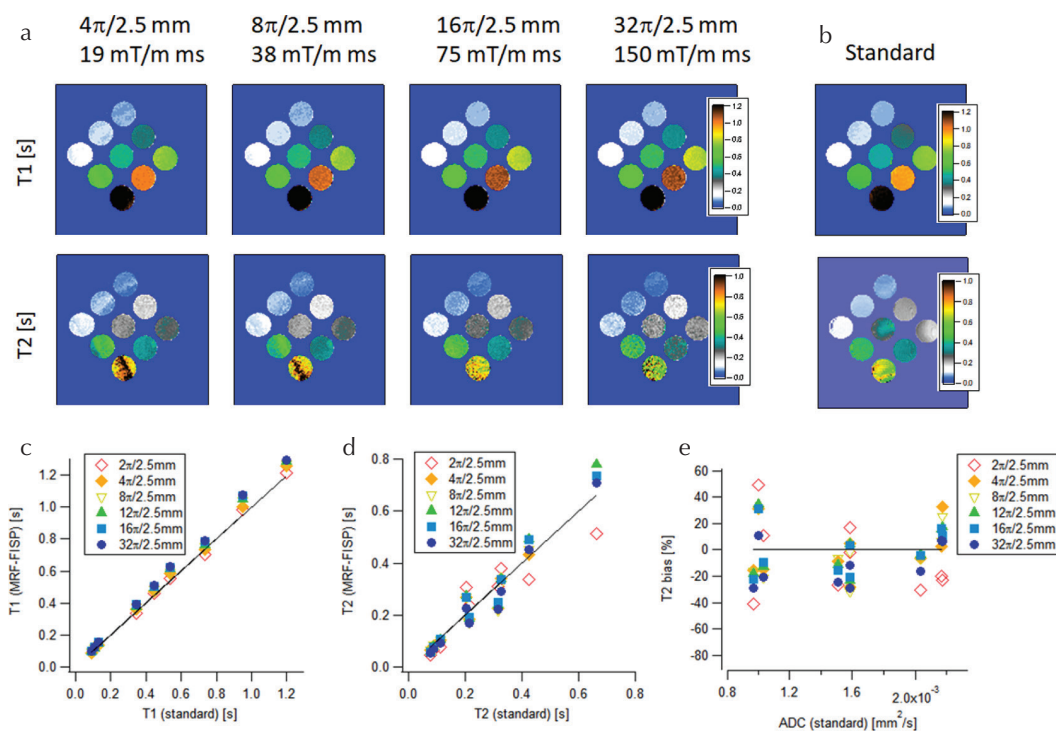
### In vivo study

Figure 6 shows the MRF-FISP and MSE results for the *in vivo* mouse brain.  $T_2$  estimates from MRF-FISP excluding diffusion showed a reduced  $T_2$  value for the larger moment, especially in CSF and in the region outside the brain.

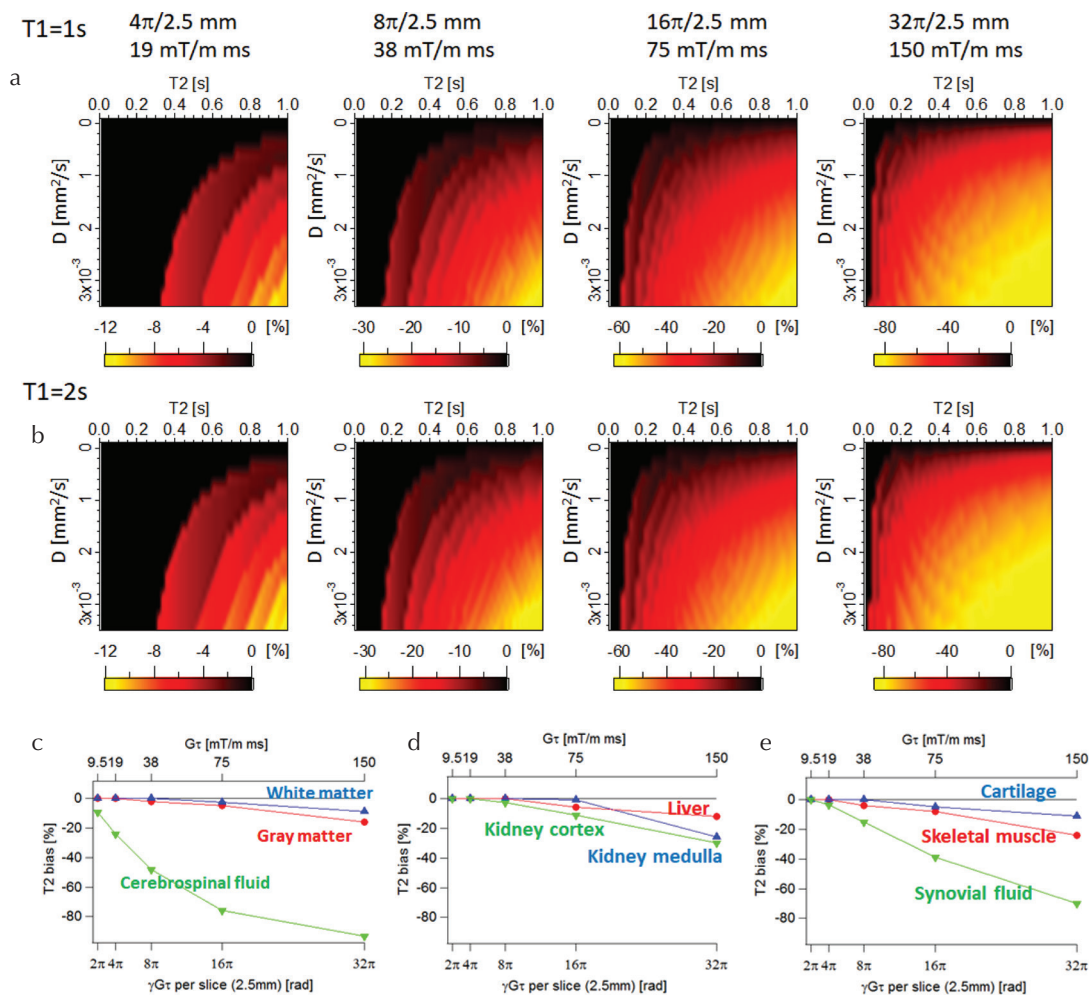
### Discussion

We experimentally and theoretically validated the diffusion-weighting effect caused by spoiler gradients in MRF-FISP, which has been neglected in the MRF-FISP framework. In the phantom experiment, the  $T_2$  estimates from MRF-FISP excluding diffusion depended on the gradient moment, and decreased as the moment increased. The  $T_2$  bias negatively increased as  $T_2$  and ADC increased, agreeing with the theoretical results. The phantom results of MRF-FISP with the additional ADC map did not show the  $T_2$  underestimation (Fig. 4). These results reveal that the  $T_2$  underestimation resulted from the diffusion weighting caused by the spoiler gradients.

The diffusion weighting in MRF-FISP can be understood by considering signal pathways in the Fourier transformation space.<sup>9,10</sup> For a FISP sequence in which TR and FA are fixed, echoes are formed through free induction decay (FID) and different refocusing pathways, which include those



**Fig. 4** Magnetic resonance fingerprinting using fast imaging with steady-state precession (MRF-FISP) results obtained while considering the diffusion effect. (a)  $T_1$  and  $T_2$  estimates from MRF-FISP for different spoiler gradient moments. (b)  $T_1$  and  $T_2$  maps from standard methods. Comparison of (c)  $T_1$  and (d)  $T_2$  between MRF-FISP and standard methods. (e)  $T_2$  bias vs apparent diffusion coefficient (ADC).

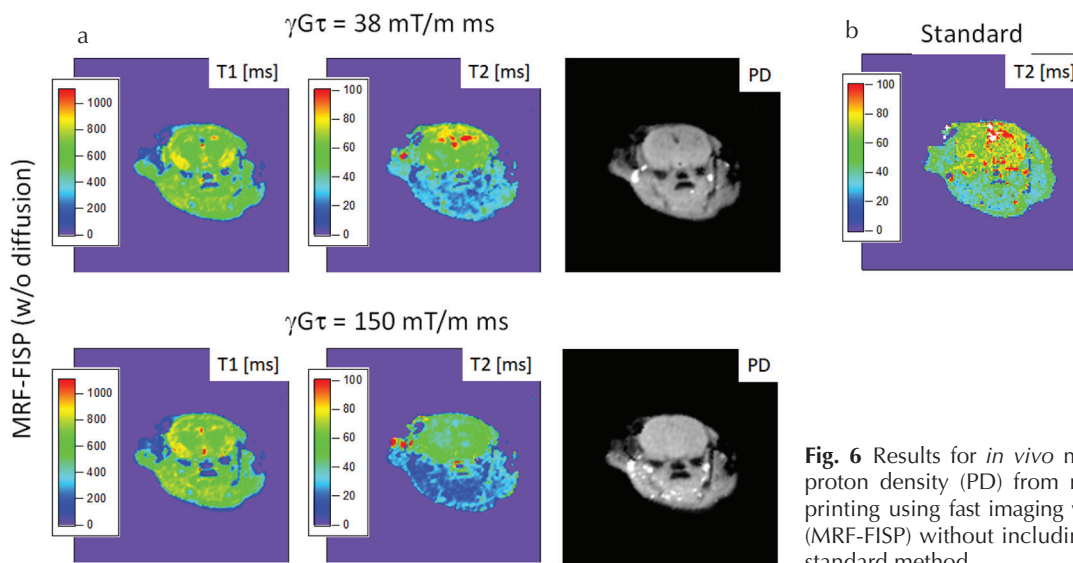


**Fig. 5** Theoretical  $T_2$  bias calculated without including diffusion.  $T_2$  bias maps for (a)  $T_1 = 1$  s and (b)  $T_2 = 2$  s. (c-e)  $T_2$  bias for typical tissues as a function of spoiler moment.

**Table 1**  $T_1$ ,  $T_2$ , and  $D$  values used for simulation. The numbers in square brackets show reference numbers

Tissue	$T_1$ (ms)	$T_2$ (ms)	$D$ ( $\times 10^{-3}$ mm <sup>2</sup> /s)
Gray matter	2000* <sup>1</sup> (16–19)	90* <sup>1</sup> (16–19)	0.83 (24)
White matter	700* <sup>1</sup> (16–19)	70* <sup>1</sup> (16–19)	0.64 (24)
CSF	5000* <sup>1</sup> (16–19)	2100* <sup>1</sup> (16–19)	3.2 (24)
Liver	809* <sup>2</sup> (20)	34* <sup>2</sup> (20)	1.24 (25)
Kidney medulla	1545* <sup>2</sup> (20)	81* <sup>2</sup> (20)	2.21 (26)
Kidney cortex	1142* <sup>2</sup> (20)	76* <sup>2</sup> (20)	2.26 (26)
Skeletal muscle	1017* <sup>2</sup> (21,22)	50* <sup>2</sup> (21,22)	2.2 (27)
Cartilage	1015.6* <sup>2</sup> (23)	39.1* <sup>2</sup> (23)	1.088 (28)
Synovial fluid	2564.7* <sup>2</sup> (23)	652.9* <sup>2</sup> (23)	1.84 (29)

\*1 Measured at 1.5T, \*2 Measured at 3T; CSF, cerebral spinal fluid.



**Fig. 6** Results for *in vivo* mouse brain. (a)  $T_1$ ,  $T_2$ , and proton density (PD) from magnetic resonance fingerprinting using fast imaging with steady-state precession (MRF-FISP) without including diffusion. (b)  $T_2$  from the standard method.

from configuration states with higher-order  $k$ . These higher-order states are affected by diffusion in a stronger way and contribute strongly to signal attenuation within each TR. Similarly, for an MRF-FISP in which pseudo-steady states were achieved by slightly varying TR and FA, higher-order states contribute to signal attenuation, leading to the underestimation of  $T_2$  when diffusion is not considered.

It should be noted that the calculation results indicate that the  $T_2$  bias is large for tissues with large  $T_2$  and  $D$  values. For example, a large  $T_2$  bias of CSF was observed even for moderate moments ( $G\tau = 19$  mT/m  $\times$  ms). This moment value was also used in the original MRF-FISP study,<sup>7</sup> and was of a similar order of magnitude to those found in previous studies.<sup>29,30</sup> The same tendency was observed for different acquisition parameters  $N$ , TR, and FA (Fig. 7). Although there are high degrees of freedom in designing an MRF-FISP and the  $T_2$  bias may depend on the acquisition scheme to varying degrees, care should be taken when tissues with large  $T_2$  and  $D$  values are estimated using MRF-FISP.  $T_2$  underestimation is especially pronounced when a

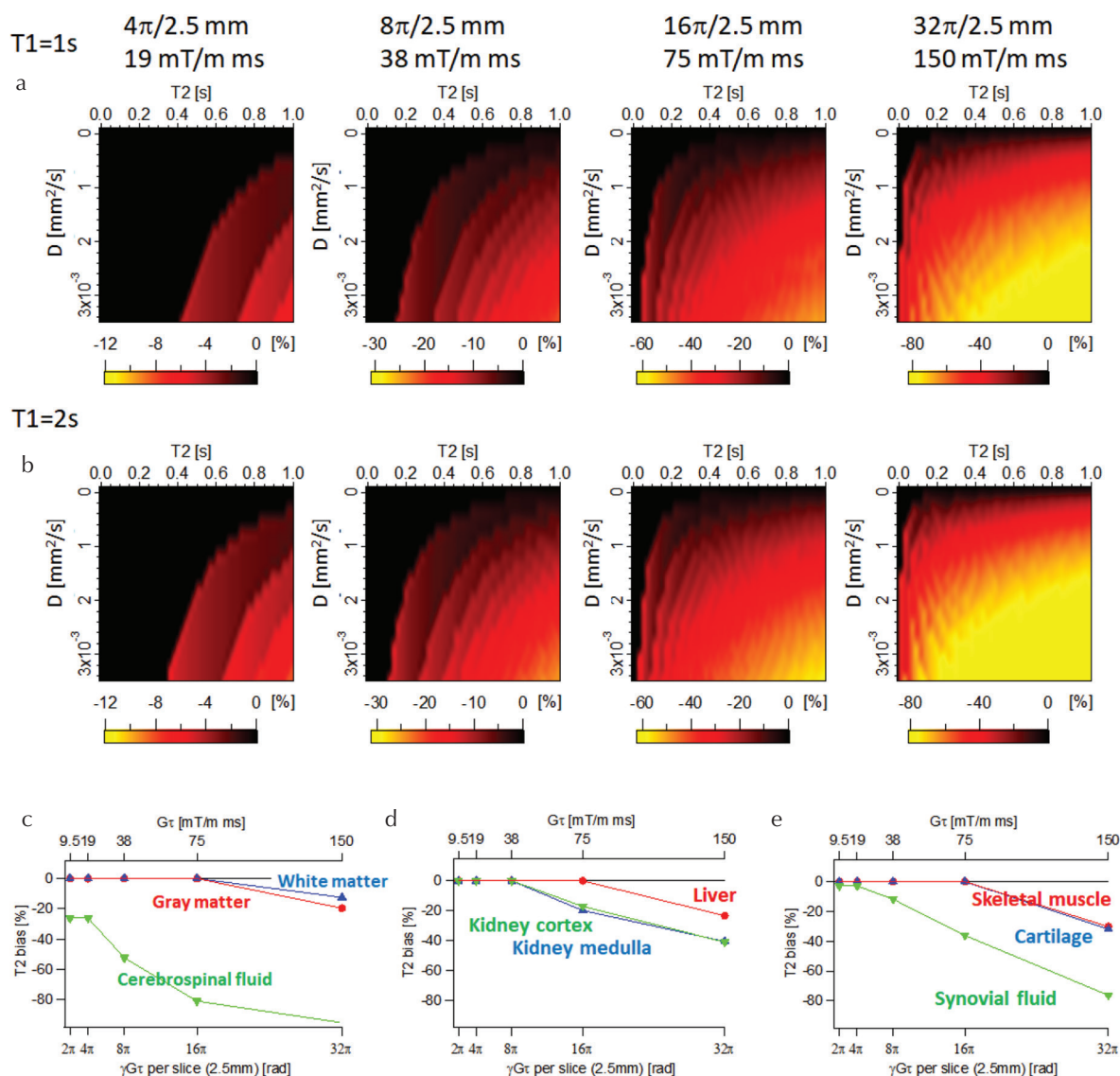
much stronger spoiler gradient is required to alleviate strong banding artifacts caused by large field inhomogeneities. For example, when the moment exceeds 75 mT/m  $\times$  ms, most tissues are affected by diffusion, resulting in significant  $T_2$  underestimation.

In the conventional FISP, the diffusion sensitivity can be characterized by the diffusion time, which is defined as

$$T_D' = \frac{1}{D(\gamma G \cdot \text{TR})^2}.^{31}$$

The regime  $\text{TR} < T_D' < T_2$  with the flip

angle larger than the Ernst angle is the diffusion-sensitive regime in which the FID signal is highly sensitive to diffusion. Although FA and TR are varied, MRF-FISP would have the similar tendency, and thus  $T_D'$  would give a rough estimate of the diffusion sensitivity for different tissues. In MRF-FISP,  $\text{TR} < T_D'$  in most cases, and tissues with  $T_2 > T_D'$  would be strongly affected by diffusion. For TR = 12.5 ms and  $D = 2.0 \times 10^{-3}$  mm<sup>2</sup>/s,  $T_D' = 1.22$  s for the moderate moment of 38 mT/m  $\times$  ms, and most tissues would be unaffected by diffusion. As the moment increases,  $T_D'$



**Fig. 7** Theoretical  $T_2$  bias calculated without including diffusion using the same sequence parameters as used in Jiang et al.<sup>7</sup> ( $N = 1200$ ).  $T_2$  bias maps for (a)  $T_1 = 1$  s and (b)  $T_2 = 2$  s. (c–e)  $T_2$  bias for typical tissues as a function of spoiler moment.

decreases and the diffusion-sensitive regime becomes wider, and more tissues would be sensitive to diffusion. For the large moment of 150 mT/m × ms,  $T_D'$  reduces to 78.4 ms.

It can be expected that the diffusion weighting is also inherently present in MRF acquisitions using balanced gradients to varying degrees, because it is practically difficult to achieve fully balanced gradients because of eddy-current effects, concomitant fields, and other hardware imperfections resulting in gradient deviations.

The  $T_2$  bias could be alleviated by using dictionary D+ and incorporating the additionally measured ADC map into the matching process; we validated this approach with the phantom measurements. It should be noted that even using dictionary D+, if the ADC map was not incorporated, diffusion and  $T_2$  could not be accurately distinguished and estimated (data are not shown here). This is because the sequence

used was sensitive to both diffusion and  $T_2$ , and yielded similar signal evolutions for different  $T_2$  and  $D$  values. Similar problems are often seen in the MRF framework. For example, it has been shown that different pulse shapes used for slice selection produce different  $T_2$  values because the MRF sequences have similar sensitivities to  $B_1$  and  $T_2$ . Although MRF sequences have near-infinite possibilities of identifying magnetic resonance parameters of interest, pulse sequence components should be designed and implemented to impart differential sensitivity to the parameters of interest, and MRF sequence design and implementation will continue to be a significant open area of research to meet this requirement.

Regardless of whether the diffusion effect was included or not, the MRF-FISP results showed  $T_2$  variation originating from banding artifacts when the spoiler gradient was weak. One simple way of alleviating  $T_2$  variation is to increase the

strength of spoiler gradients, as was shown in Fig. 4. Another way is to use shorter TRs. In this study, we used relatively long TRs compared with the original studies, because of hardware limitations. Modern hardware could use shorter TRs and offer improved tolerance to field inhomogeneity. However, this approach would pose a limitation when the inhomogeneity is significantly large, such as in high field strengths and in an imaging volume where sharp susceptibility transitions exist. In such cases, the use of a stronger spoiler gradient is the practical solution to overcome the banding issues.

Another way of avoiding  $T_2$  bias in the MRF framework may be to use MRF based on a double-echo steady state (MRF-DESS<sup>32</sup>): to increase the diffusion sensitivity, the strength of the spoiler gradient is varied and both FID and echo signals are acquired. MRF-DESS is sensitive to both  $T_2$  and diffusion variations, and ADC values can also be estimated. MRF-DESS could be more efficient than MRF-FISP because it does not require the lengthy ADC measurements. Moreover, MRF-DESS uses a stronger spoiler gradient and hence is less susceptible to banding artifacts. However, MRF-DESS has several potential disadvantages compared with MRF-FISP. First, MRF-DESS requires high hardware performance to achieve fast switching of strong gradients with high fidelity within short TR intervals. Second, MRF-DESS tends to be less tolerant of systematic errors, because the echo signals are largely attenuated by the stronger spoiler gradients and the signal-to-noise ratios are reduced. Third, MRF-DESS has more complicated sensitivities to  $T_2$ ,  $D$ , and  $B_1$ , and has the increased similarity between different signal evolutions. Therefore, special care should be taken to design the MRF-DESS sequence to differentiate the  $T_2$  and  $D$  values. The protocol optimization<sup>33</sup> for DESS may serve to design the MRF-DESS sequence.

Our *in vivo* study has a limitation that we did not perform the ADC measurement because we needed to reduce the measurement time to keep the mouse alive. In the *in vivo* experiment,  $T_2$  estimates from MRF-FISP excluding diffusion were smaller in CSF and in the region outside the brain for the larger moment. This is probably because the diffusion coefficient was large in these regions, which could be confirmed by measuring the ADC map using a fast imaging method.

## Conclusion

Diffusion weighting caused by spoiler gradients in MRF-FISP was validated experimentally and theoretically. Without considering diffusion,  $T_2$  from MRF-FISP was greatly underestimated for large gradient moments. The  $T_2$  underestimation was more prominent for tissues with large  $T_2$  and  $D$  values. The  $T_2$  bias was almost independent of the ADC value when we incorporated the diffusion effect of the spoiler gradients into the dictionary and incorporated the ADC maps. These results reveal that the  $T_2$  underestimation resulted from the diffusion weighting caused by the spoiler gradients.

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## Conflicts of Interest

The authors or the authors' institutions have no conflicts of interest.

## References

1. Cheng HL, Stikov N, Ghugre NR, Wright GA. Practical medical applications of quantitative MR relaxometry. *J Magn Reson Imaging* 2012; 36:805–824.
2. Koh DM, Collins DJ. Diffusion-weighted MRI in the body: applications and challenges in oncology. *AJR Am J Roentgenol.* 2007; 188:1622–1635.
3. Subhawong TK, Jacobs MA, Fayad LM. Insights into quantitative diffusion-weighted MRI for musculoskeletal tumor imaging. *AJR Am J Roentgenol* 2014; 203: 560–572.
4. Jackson A, O'Connor J, Thompson G, Mills S. Magnetic resonance perfusion imaging in neuro-oncology. *Cancer Imaging* 2008; 8:186–199.
5. Henkelman RM, Stanisz GJ, Graham SJ. Magnetization transfer in MRI: a review. *NMR Biomed.* 2001; 14:57–64.
6. Ma D, Gulani V, Seiberlich N, et al. Magnetic resonance fingerprinting. *Nature* 2013; 495:187–192.
7. Jiang Y, Ma D, Seiberlich N, Gulani V, Griswold MA. MR fingerprinting using fast imaging with steady state precession (FISP) with spiral readout. *Magn Reson Med* 2015; 74:1621–1631.
8. Terada Y. Initial implementation of magnetic resonance fingerprinting on a preclinical 14.1 T scanner. In *Proceedings of the 25th Annual Meeting of the ISMRM, Honolulu, Hawaii, USA, 2017*; 5220.
9. Wu E, Buxton R. Effect of diffusion on the steady-state magnetization with pulsed field gradients. *J Magn Reson* 1990; 90:243–253.
10. Buxton R. The diffusion sensitivity of fast steady-state free precession imaging. *Magn Reson Med* 1993; 29:235–243.
11. Hargreaves BA, Rapid gradient-echo imaging. *J Magn Reson Imaging* 2012; 36:1300–1313.
12. Hashimoto S, Kose K, Haishi T. Comparison of analog and digital transceiver systems for MR imaging. *Magn Reson Med Sci* 2014; 13:285–291.
13. Gao Y, Chen Y, Ma D, et al. Preclinical MR fingerprinting (MRF) at 7 T: effective quantitative imaging for rodent disease models. *NMR Biomed* 2015; 28:384–394.
14. Weigel M. Extended phase graphs: dephasing, RF pulses, and echoes - pure and simple. *J Magn Reson Imaging* 2015; 41:266–295.
15. Deoni SC, Peters TM, Rutt BK. High-resolution  $T_1$  and  $T_2$  mapping of the brain in a clinically acceptable time with DESPOT1 and DESPOT2. *Magn Reson Med* 2005; 53: 237–241.



16. Vymazal J, Righini A, Brooks RA, et al.  $T_1$  and  $T_2$  in the brain of healthy subjects, patients with Parkinson Disease, and patients with multiple system atrophy: relation to iron content. *Radiology* 1999; 211:489–495.
17. Whittall KP, MacKay AL, Graeb DA, Nugent RA, Li DK, Paty DW. In vivo measurement of  $T_2$  distributions and water contents in normal human brain. *Magn Reson Med* 1997; 37:34–43.
18. Poon CS, Henkelman RM. Practical  $T_2$  quantitation for clinical applications. *J Magn Reson Imaging* 1992; 2:541–553.
19. de Bazelaire CM, Duhamel GD, Rofsky NM, Alsop DC. MR imaging relaxation times of abdominal and pelvic tissues measured in vivo at 3.0 T: preliminary results. *Radiology* 2004; 230:652–659.
20. Chen Y, Lee GR, Aandal G, et al. Rapid volumetric  $T_1$  mapping of the abdomen using three-dimensional through-time spiral GRAPPA. *Magn Reson Med* 2016; 75:1457–1465.
21. Stanisz GJ, Odobina EE, Pun J, et al.  $T_1$ ,  $T_2$  relaxation and magnetization transfer in tissue at 3T. *Magn Reson Med* 2005; 54:507–512.
22. Jordan CD, Saranathan M, Bangerter NK, Hargreaves BA, Gold GE. Musculoskeletal MRI at 3.0 T and 7.0 T: a comparison of relaxation times and image contrast. *Eur J Radiol* 2013; 82:734–739.
23. Le Bihan D, Mangin JF, Poupon C, et al. Diffusion tensor imaging: concepts and applications. *J Magn Reson Imaging* 2001; 13:534–546.
24. Bruegel M, Holzapfel K, Gaa J, et al. Characterization of focal liver lesions by ADC measurements using a respiratory triggered diffusion-weighted single-shot echo-planar MR imaging technique. *Eur Radiol* 2008; 18:477–485.
25. Sulowska K, Palczewski P, Duda-Zysk A, et al. Diffusion-weighted MRI of kidneys in healthy volunteers and living kidney donors. *Clin Radiol* 2015; 70:1122–1127.
26. Heemskerk AM, Damon BM. Diffusion tensor MRI assessment of skeletal muscle architecture. *Curr Med Imaging Rev* 2007; 3:152–160.
27. Aoki T, Watanabe A, Nitta N, Numano T, Fukushi M, Niitsu M. Correlation between apparent diffusion coefficient and viscoelasticity of articular cartilage in a porcine model. *Skeletal Radiol* 2012; 41:1087–1092.
28. Barendregt AM, Nusman CM, Hemke R, et al. Feasibility of diffusion-weighted magnetic resonance imaging in patients with juvenile idiopathic arthritis on 1.0-T open-bore MRI. *Skeletal Radiol* 2015; 44:1805–1811.
29. Ma D, Pierre EY, Jiang Y, et al. Music-based magnetic resonance fingerprinting to improve patient comfort during MRI examinations. *Magn Reson Med* 2016; 75: 2303–2314.
30. Hamilton JI, Jiang Y, Chen Y, et al. MR fingerprinting for rapid quantification of myocardial  $T_1$ ,  $T_2$ , and proton spin density. *Magn Reson Med* 2017; 77:1446–1458.
31. Freed DE, Scheven UM, Zielinski LJ, Sen PN, Hürlimann MD. Steady-state free precession experiments and exact treatment of diffusion in a uniform gradient. *J Chem Phys* 2001; 115:4249.
32. Jiang Y, Ma D, Wright K, Seiberlich N, Gulani V, Griswold MA. Simultaneous  $T_1$ ,  $T_2$ , diffusion and proton density quantification with MR fingerprinting. In Proceedings of the 22th Annual Meeting of the ISMRM, Milan, Italy, 2014; 0028.
33. Gras V, Farrher E, Grinberg F, Shah NJ. Diffusion-weighted DESS protocol optimization for simultaneous mapping of the mean diffusivity, proton density and relaxation times at 3 Tesla. *Magn Reson Med* 2017; 78:130–141.