Research Article

A New Microarray System to Detect Streptococcus pneumoniae Serotypes

Yuka Tomita,¹ Akira Okamoto,² Keiko Yamada,² Testuya Yagi,³ Yoshinori Hasegawa,⁴ and Michio Ohta²

¹ Department of Infectious Disease, Nagoya University Hospital, 65 Tsurumai-cho, Showa-ku, Nagoya 466-8550, Japan

² Department of Bacteriology, Nagoya University Graduate School of Medicine, 65 Tsurumai-cho, Showa-ku, Nagoya 466-8550, Japan

³ Center of National University Hospital for Infection Control, Nagoya University Hospital, Nagoya 466-8550, Japan

⁴ Department of Respiratory Medicine, Nagoya University Graduate School of Medicine, 65 Tsurumai-cho, Showa-ku, Nagoya 466-8550, Japan

Correspondence should be addressed to Yuka Tomita, yu-cat@med.nagoya-u.ac.jp

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Streptococcus pneumoniae, one of the most common gram-positive pathogens to colonize the human upper respiratory tract, is responsible for many severe infections, including meningitis and bacteremia. A 23-valent pneumococcal vaccine is available to protect against the 23 *S. pneumoniae* serotypes responsible for 90% of reported bacteremic infections. Unfortunately, current *S. pneumoniae* serotype testing requires a large panel of expensive antisera, assay results may be subjective, and serotype cross-reactions are common. For this study, we designed an oligonucleotide-based DNA microarray to identify glycosyltransferase gene sequences specific to each vaccine-related serotype. Out of 56 isolates representing different serotypes, only one isolate, representing serotype 23A, was not detected correctly as it could not be distinguished from serotype 23F. Our data suggest that the microarray provides a more cost-effective and reliable way of monitoring pneumococcal capsular types.

1. Introduction

Streptococcus pneumoniae is an important cause of bacteremia, community-acquired bacterial pneumonia, and meningitis, especially among young children and older adults [1-3]. Capsular polysaccharide is the primary S. pneumoniae virulence factor and encapsulated pneumococci are responsible for more diseases than unencapsulated strains [4]. After comparing the differences in capsular polysaccharides composition, S. pneumoniae can be divided into more than 90 serotypes [5] and the 23 serotypes responsible for 90% of disease cases [6] are represented in a 23-valent pneumococcal vaccine. Pneumococcal serogroup and serotype identification is currently performed by using large panels of expensive antisera by various methods, including the capsular swelling (Quellung) reaction, latex agglutination, and coagglutination. Cross-reactions between serotypes and discrepancies between methods can occur and some strains are nonserotypable. On the other hand,

molecular typing has the potential to improve discrimination and provide additional information. S. pneumoniae capsule production is predominantly controlled by capsular polysaccharide synthesis (cps) gene clusters [7, 8], which are responsible for each serotype-specific polysaccharide. The Sanger Institute has sequenced the cps gene clusters of 90 S. pneumoniae serotypes and predicted the general function of 1,973 of the 1,999 gene products [9, 10]. S. pneumoniae capsular polysaccharides represent a diverse group of polymers with distinct sugar compositions and linkages [10]. The key enzymes to link each serotype-specific sugar component are glycosyltransferases (GTs) [11], which transfer the sugar moiety from an activated nucleotide sugar to an acceptor to generate a serotype-specific capsular polysaccharide. After discovering that S. pneumoniae GT genes are highly variable and contain serotype- or serogroup-specific regions, we used GT sequences as probes in an oligonucleotide-based microarray to identify 23-valent pneumococcal vaccine and closely related S. pneumoniae serotypes. Our data suggest that the microarray provides a more cost-effective and reliable way of monitoring serotype distribution.

2. Materials and Methods

2.1. Bacterial Strains, Growth Conditions, Immunological Serotyping, and Genomic DNA Extraction. S. pneumoniae strains representing various serotypes were obtained from the American Type Culture Collection, the Statens Serum Institute, and clinical isolates (Table 1). Each strain was cultivated on brain-heart infusion broth (Eiken, Tokyo, Japan) supplemented with 0.3% yeast extract (Becton Dickinson, Boston, MA) (BHI-Y) for 24 h at 37°C in 5% CO₂. Conventional serotyping was performed for clinical isolates obtained in Japan by slide agglutination (Denka Seiken, Tokyo, Japan) or quellung reaction (Statens Serum Institute, Copenhagen, Denmark).

Genomic DNA was extracted using a Wizard Genomic DNA purification kit (Promega, Madison, WI).

2.2. DNA Array Preparation. Oligonucleotide probes were synthesized and spotted on a glass slide at Nihon Gaishi (Nagoya, Japan). The slide was stirred in a beaker filled with $2 \times SSC/0.2\%$ SDS for 15 min, transferred to a second beaker filled with $2 \times SSC/0.2\%$ SDS to incubate for 5 min at 95°C, rinsed three times with dH₂O, and centrifuged at 900 rpm for 3 min at 25°C in a horizontal microtiter plate rotor before being covered with a plastic seal.

2.3. Chromosomal DNA Labeling. 500 ng of genomic DNA was suspended in $21 \,\mu\text{L}$ dH₂O and $20 \,\mu\text{L}$ of $2.5 \times \text{Random}$ Primer Solution (Invitrogen, Carlsbad, CA), heated to 95°C for 5 min, and chilled on ice for 3 min. The DNA was labelled in a reaction including 5 µL of 10X dCTP Nucleotide Mix (Invitrogen, Carlsbad, CA), 5 µL Cy3 or Cy5-dCTP (GE Healthcare, Buckinghamshire, UK), and 1 µL of Exo-Klenow Fragment (Invitrogen, Carlsbad, CA). After a 2hour incubation at 37°C, 5 μ L of sodium acetate, 125 μ L of ethanol and 1 μ L of glycogen was added to 25 μ L of Cy3 and Cy5 labeled DNA, which was purified previously by QIAprep Spin Miniprep Kit (250) (Qiagen, Tokyo, Japan). Following a 30-minute incubation at -80° C in the dark, the probe mixture was centrifuged for 30 min at 14,000 rpm at 4°C. The supernatant was removed and the probe was air-dried for 5 min in the dark. The probe mixture was diluted in $70\,\mu\text{L}$ of the hybridization buffer (25% formamide, 0.1% SDS, 6 \times SSPE), incubated for 30 min at room temperature in the dark, heated for 8 min at 75°C, and incubated for 30 min at 42°C.

2.4. Probe Hybridization and Microarray Signal Detection. Prewarmed probe mixture was applied to the prepared microarray slide, placed in a hybridization chamber and incubated for 20 h at 42°C. After hybridization, the plastic seal was removed and the slide was washed with $1 \times$ SSC/0.1% SDS solution for 3 min, 0.05 × SSC for 3 min, and 95% ethanol for 90 s at room temperature. The washed microarray slide was dried by centrifugation and scanned

TABLE 1: Test strains.

Serotype	Strain designation	Serotype	Strain designation
1	ATCC6301 ^a	14	D59 ^b
2	ATCC6302 ^a	15F	ATCC6315 ^a
3	D36 ^b	15A	ATCC6330 ^a
4	JHK27 ^b	15B	ATCC10354 ^a
5	ATCC6305 ^a	15C	SSI15C/2 ^c
6A	MSC1943 ^b	17F	ATCC6317 ^a
6B	MSC1047 ^b	17A	SSI17A/2 ^c
7F	ATCC10351 ^a	18F	ATCC6318 ^a
7A	ATCC6307 ^a	18A	ATCC10344 ^a
7B	ATCC10348 ^a	18B	ATCC10355 ^a
7C	ATCC10350 ^a	18C	ATCC10356 ^a
8	ATCC6308 ^a	19F	D33 ^b
9A	ATCC8333 ^a	19A	D4 ^b
9V	KD10-11 ^b	19B	ATCC10358 ^a
9L	ATCC10349 ^a	19C	ATCC10359 ^a
9N	KD01-26 ^b	20	ATCC6320 ^a
10F	ATCC6310 ^a	22F	KD01-23 ^b
10A	ATCC8334 ^a	22A	ATCC10363 ^a
10B	SSI10B/2 ^c	23F	KD11-15 ^b
10C	SSI10C/2 ^c	23A	KD12-06 ^b
11F	ATCC6311 ^a	23B	ATCC10364 ^a
11A	SSI11A/2 ^c	33F	ATCC10370 ^a
11B	SSI11B/2 ^c	33A	ATCC8340 ^a
11C	ATCC10353 ^a	33B	ATCC10342 ^a
11D	SSI11D/1 ^c	33C	ATCC8339 ^a
12F	ATCC6312 ^a	33D	SSI33D/2 ^c
12A	SSI12A/5 ^c	44	SSI44/3 ^c
12B	SSI12B/1 ^c	46	SSI46/2 ^c

Explanatory notes: Serotypes represented in bold letter are those included in 23-valent pneumococcal vaccine.

^aAmerican Type Culture Collection.

^bClinical isolate obtained from Japan.

^cStatens Serum Institute.

using the DNA Microarray Scanner (Agilent, Santa Clara, CA).

2.5. Data Analysis. The signal and background intensities of each spot were quantified using GenePix Pro 6.0 software and the average was calculated with Microsoft Excel software.

3. Results

3.1. Target Gene Selection and Microarray Construction. In this study, we designed a DNA microarray to identify the 23 *S. pneumoniae* serotypes included in the 23-valent pneumococcal vaccine, using GT genes in *cps* locus. We compared the GT sequences of the 23-valent vaccine serotypes with other *S. pneumoniae* serotypes and found that these 23 serotypes were indistinguishable from 14 nonvaccine serotypes. Therefore, 37 serotypes, 23-valent vaccine serotypes and 14 closely related serotypes, were divided into 23 groups and each group had one to six GT genes in their *cps* locus TABLE 2: Twenty-three groups distinguished in this study and targeted glycosyltransferase genes.

Group name		Targeted G	T genes in cps locus	(probe number ^a)		
1	wchB (1, 2, 3)	wchD (4, 5, 6)				
2	wchF (7, 8, 9)	wchG (10, 11, 12)	wchH (13, 14, 15)	wchI (16, 17, 18)		
3	wchE (19, 20, 21)					
4	wciJ (22, 23, 24)	wciK (25, 26, 27)	wciL (28, 29, 30)			
5	wciJ (31, 23, 24)	whaC (32, 33, 34)	whaD (35, 36, 37)			
6A/6B	wciN (38, 39, 40)	wciP (41, 42, 43)				
7F/7A	wchF (44, 45, 46)	wcwA (47, 48, 49)	wcwF (50, 51, 52)	wcwG (53, 54, 55)	wcwH (56, 57, 58)	
8	wciR (59,60,61)	wciR (62, 63, 64)	wciS (65, 66, 67)	wciT (68, 69, 70)		
9A/9V	wchO (71,72,73,74)	wcjA (75, 76, 77)	wcjB (78, 84, 85)	WcjC (81, 82, 83)		
9L/9N	wchO (71, 72, 73, 74)	wcjA (75, 76, 77)	wcjB (78, 79, 80)	wcjC (81, 82, 83)		
10A	wciB (86, 87, 8)	wcrC (89, 90, 91)	wcrD (92, 93, 94)	wciF (95, 96, 97)	wcrG (98, 99, 100)	
11A/11D	wchK (101, 102, 103)	wcyK (104, 105, 106)	wcrL (107, 108, 109)			
12F/12A/ 12B/44/46	wciJ (110, 111, 112)	wcxB (113, 114, 115)	wcxD (116, 117, 118)	wcxE (119, 120, 121)	wcxF (122, 123, 124)	
14	wchK (125, 126, 127)	wchL (128, 129, 130, 131)	wchM (132, 133, 1334)	wchN (135, 136, 137)		
15B/15C	wchK (138, 139, 125)	wchL (128, 140, 141, 131)	wchM (142, 143)	wchN (135)		
17F	wchF (144, 145, 146)	abp1 (147, 148, 149)	wciP (150, 151, 152)	wcrV (153, 154, 155)		
18B/18C	wchF (156, 157)	wciU (158, 159, 160)	wciV (161, 162, 163)	wciW (164, 165, 166)		
19F	wchO (167, 72, 168, 169)	wchQ (171, 172, 173)				
19A	wchO (71, 170, 73, 74)	wchQ (171, 172, 173)				
20	wciB (174, 175, 176)	whaJ (177, 178, 179)	wciL (180, 181, 182)	wcwK (183, 184, 185)	wciD (186, 187, 188)	whaF (189, 190, 191)
22F/22A	wchF (7, 8, 192, 193)	wcwA (48, 49, 194)	wcwV (195, 196, 197)	whaB (198, 199, 200)		
23F	wchF (144, 156, 145, 193, 201)	wchV (202, 203, 204)	wchW (205, 206, 207)			
33F/33A/ 37	wciB (208, 209, 210)	wciC (211, 212, 213)	wciD (214, 215, 216)	wciE (217, 218, 219)	wciF (220, 221, 222)	

Explanatory notes: ^aProbes containing 60-bp oligonucleotides were designed and named as 1, 2, 3 etc from Group 1. The name of each GT gene (*wchB* etc) was derived from the Sanger Institute.

(Table 2). The 60-bp oligonucleotide probes contained the variable middle region of each open reading frame and were designed from published sequences at the Sanger Institute (http://www.sanger.ac.uk/Projects/S_pneumoniae/CPS/) and Genbank websites. In most cases, the designed probes were gene specific, although some probes included sequences from more than one gene. Each serotype group was identified using 3 to 18 probes (Table 2) and a total of 222 probes

were designed to target 23 groups (Table 3). 26 positive control probes were designed to hybridize *S. pneumoniae* housekeeping genes and 16S rDNA. In addition, 26 negative control probes were designed to detect housekeeping genes of other bacterial respiratory pathogens, including *Klebsiella pneumoniae*, *Staphylococcus aureus*, *Legionella pneumophila*, *Chlamydophila pneumoniae*, *Mycoplasma pneumoniae*, *Pseudomonas aeruginosa*, and *Streptococcus pyogenes*. A schematic

TABLE 3: Oligonucleotide probes used in this study.

Spot identifier	Targeted GT gene	Specificity	Probe sequence $(5'-3')$
1	wchB	Serotype1	ATAAGATTATTGAGAAAATATAGACCGGATGTAGTCTTGACATATACC- GTGAAACCAAAT
2	wchB	Serotype1	TTTATTGGTAGGATATTAAAAGAAAAAGGTATAGATACTTATCTGGCTGCT- GCCCAAATT
3	wchB	Serotype1	GAAAATGAAAAACGAAAAGAGATGGGACTTCAAGGGAGAATGTATATA- GAGCAATATTTT
4	wchD	Serotype1	TTATTGAAGGAATGATTGATAGCGACTTAATAGTTGTTCGTATTCCGTCTA- TAATTGGAT
5	wchD	Serotype1	GCCATAGATTTGTATTGGAAGCAATGAAGAGATTAGAAATACAAGGTA- TTTTGTTGGATT
6	wchD	Serotype1	AGCGATTGCGGGATCTATTATAGATTTTATTAGTATGGATAAGGAAAA- GATGGTGATAAA
7	wchF	Serotype2, 21, 22F, 22A,23B, 32F, 32A	TTTGTTGAGAAATTAACAGAATATCAAAAAGATGGTAACATCCAATACTAT- GTTGCCTGC
8	wchF	Serotype2, 21, 22F, 22A, 27, 32F, 32A	CTAAAAAAGACTTTGTTCTCATTACAAATGTGGAACAGAATAAGTTTTACGATCAGTTGC
9	wchF	Serotype2	TTATTGAAGCAGTGGAGCAATTTGATGAGAACGCCATTTCTGAACTAG- ATAAAAAATCTA
10	wchG	Serotype2	GCAATACCAAGAAAAATACCCTAAAAAAATTAAGGTTATCACAGATTCCTC- TGTTATAGG
11	wchG	Serotype2	TAGAAGTTTAAACAATCTGTTAGATTTGAATAGTAATGCAGTAGCTATGCA- TGATTGGTG
12	wchG	Serotype2	TTATCAGAATTCTCTAAGTAATGAGGAGACAGATATTATTCGTGAATTTAT- CAGCATTCC
13	wchH	Serotype2	TAGAAACCAGACAATTTTTTATCGGATAAAAGCTTCTTTGGGGAATACTCT- AAAAAACG
14	wchH	Serotype2	CGTATTCCAGAAAAGTTACCTGATACCTATAATGTGTTGATTAATCCTGAA- AGAGAAAAA
15	wchH	Serotype2	CTTTGTTGGAACTCTCAAATGGTCAGAATACTATAGTTGTAGAAGAGTTAT- CAGAAATAT
16	wchI	Serotype2	CATTTTACCAGAACATGGAAATGTGGAAGATGAGCTTGTAAACAAAGG- AATTAAATACTA
17	wchI	Serotype2	TGATTTAGTTAGAGCGATAGCTAATCTTCCTGAGAGATATAAACAGATGTT- TAAAGTTGA
18	wchI	Serotype2	TACAAAAGAGATAATTTCTACAGGAGAAACAGGATATCTGTATGAACC- AGGAGATTATAT
19	wchE	Serotype3	TATAAGTCCTACAGTTGTAGTGTAAGTGATGAGAAGTTATTTAGTTCTGTA- ATTATCCCT
20	wchE	Serotype3	ACTTTAAAAAAAGGCTATAAAACTGTTATGCAGGATACTTCTGTTGTGTAT- ACAGATGCT
21	wchE	Serotype3	ACTGCAATTGTTTATACAGCTTCATGGTGGGAAATTATTTTATATGTTCTT- TTGGGAATG
22	wciJ	Serotype4, 45	GATTGTTGAATTATTTTAGCTTTGCAATTAGTTCTACTTTAGGAGTTTTAT- TGGGGAGGT
23	wciJ	Serotype4, 5, 45	GCCACAATATGCAGAAGATCTTTTTATCCCTGATGAATCTATAGTTAATAA- AGAAAGTGT
24	wciJ	Serotype4, 5, 45	CCTTCTATAAAAAATCAGATGCTATGTTAGTTTCTTTAATAGGAGACTCGA- TAGTTTCTC
25	wciK	Serotype4	GGTTCAGAAACAATTGGTGAAAAATTCTTTAATGAATATCGTTTCTTCAGA- CGGCTATAA
26	wciK	Serotype4	TCGATTTCAGTTGAATTTTATAGGTACTAATGCAGGAGAATTAAGGGAATT- TTGTCAAGA

Spot identifier	Targeted GT gene	Specificity	Probe sequence $(5'-3')$
27	wciK	Serotype4	GTGAAGATACTTATATGGAAAAAGTGTCAATAGAGAATGGTTTTGGTT- TCGTTTTACCTA
28	wciL	Serotype4	AAAAGCCTCTACATCAGTTTCTCTCTCTTGCTAGAATAATAAAGAAAG
29	wciL	Serotype4	AGAACTCATTTTAATCAAACCAAATGTTATTTTACTCCTAGTTGGTAATGG- TGAGGATGA
30	wciL	Serotype4	AAAAACATTAGTTACTTACCTATCAACGAAGAGTCTGTGTTGCTATGGAAA- GATAAAGTA
31	wciJ	Serotype5	TTACATAGGATATTAAATTATTTTAGTTTTGCTATCAGTTCCTCGATAGGG- GTTCTACTG
32	whaC	Serotype5	TTTCTGACTCTCACAAGTATGATGGATTGGTATTACCAAAGAAAAATACAG- TTCGCAATT
33	whaC	Serotype5	TATATCCCGAACCTCAACTTTTGAACCTTTTAACGAGAAATATCATATCCG- TCAGATTAT
34	whaC	Serotype5	GAAGACTAAACTTCAGCGTGAATTGAAACTAGAAGAAGCACGCTATAA- AGGAAATAGATT
35	whaD	Serotype5	AAGACGGCAGTACGCTATTTCTGTTGATGGTATAATAAATCATAGTAATAT- CTCACTTAA
36	whaD	Serotype5	GTTTTCACAAGATATAGTATTCGAAAATCTGAGAAAAATCTGCTTTTTGTG- GGACAGTTT
37	whaD	Serotype5	TCTAATATACATATAATTCCTTTTCTAGAAAAAACTGATATCCTAGAGTTG- ATGCGGGTG
38	wciN	Serotype6A, 6B, 33D	AATAGATTATCAAAACAATTTGCGCAGAGAGAAATTAATT
39	wciN	Serotype6A, 6B, 33D	TTACAAGGAGATTTAGGGGTTTTAAATGCAGTTTTATATAACTCATTTGGT- GTACTTCCT
40	wciN	Serotype6A, 6B	GCAAGAAGGCAGTAATGTTGCACATATAGACCAATTTAAAAAATACTA- TGAAGGTAGTTA
41	wciP	Serotype6A, 6B	GGACACTTTTTATTAGGGATGATGGATCAAAAGATAAAACAATAGAAG- TAATACAGAGGT
42	wciP	Serotype6A, 6B	CAGGTTTTAATCATGCATTGCTAGAGATGGTTCCTTCAGTTGATATTGATA- AAGATTATT
43	wciP	Serotype6A, 6B	CTTACACATGCTGGGGTATATAATCAAACTCTTTATATGCTAAAAAAAGCT- TCTGGAAAA
44	wchF	Serotype7F, 7A	ATACAATACTATGTTGCTTGTATGCGTGAAAATTCAGCTAAATCTGGCATC- ATGGATGAT
45	wchF	Serotype7F, 7A	AAAAATATATCCAAGAGGATTATAAGCAGTACCAACCAAAGACCACCT- ATATTGCCTATG
46	wchF	Serotype7F, 7A, 23B	TTGTTACAGGATACTGGTTTTGATAAAGATCCTAGGGTTAAATTTGTT- GGGACTGTCTAT
47	wcwA	Serotype7F, 7A	AAGTGCATCTTTCCAACGTCAAAAAGAATTCTTTTCGTTGGAAAGTTATAT- TCGGAATTT
48	wcwA	Serotype7F, 7A, 21, 22F, 22A	GTGTGTTGGATTATCCGATTTTACGTAGAAAATACTTTAATCCTAAGGGGA- TTTTAGAAT
49	wcwA	Serotype7F, 7A, 21, 22F, 22A	CTCACAAATCAAACGAATTGACTATTATGAGCATACGACTGAGCTTTATAA- TATGTTTGA
50	wcwF	Serotype7F, 7A	AAATATGAAGTTATTCTAGTAAATGATGGCAGTACAGATGCTTCACCCAAT- ATTTGTGAA
51	wcwF	Serotype7F, 7A	TATTTTATTGGGAATGATGCGGCTATTACCAAACAGTGGTCTGAAAAA- AAAATTAGTGAT
52	wcwF	Serotype7F, 7A	ATGAAATTGTATGAAGAAAATCAGGAAGACACTCAACTTTTAGGTTGATA- CTTGCAGAA

Spot identifier	Targeted GT gene	Specificity	Probe sequence (5'-3')
53	wcwG	Serotype7F, 7A	AAAACGATTACCCGGATTTTTATTCCATAATTGGTGGTTAGAAGAATGGTC- TAGAAAATT
54	wcwG	Serotype7F, 7A	GGTGCAGATAAAGGAAGATTGCCAAAATTAAAAAGCTTAGCTAAGCAG- ATAGTTTTAAAT
55	wcwG	Serotype7F, 7A	ATAAAAAGGGACAGGATGGTCTAACCCTTAGAGCAATGGAATCCATTT- TTTATAAAAAAA
56	wcwH	Serotype7F, 7A	GGAACAGAGTTACTAAGAATTGTAAAATCAAATCAATTGTAGGCAATA- TACGTGGCAAAA
57	wcwH	Serotype7F, 7A	ATTTGCTAAATCCTATAGAGAAACGAAACCCATTTCATCTAGGTATGTTAT- ATCATGAAG
58	wcwH	Serotype7F, 7A	TATTTGAATATGCAATTGATGGCGAGAATGCACTTTTATCTCCGATAAAAGATAGTGTTA
59	wciO	Serotype8	AACTAATGAAGCTTGAACCGATTATGAGACAACAAGACAGCTATTTAA- TCACAGAATATA
60	wciO	Serotype8	AAATCACTTTATATACTGTTAAGAATACGCCCAAAAGTAGTTATCTGTACA- GGTGTTCTT
61	wciO	Serotype8	TTTATATTGAATCTTTTGCAAAAGTGACCACTCCTACTTTAACAGGTAGAA- TACTATACC
62	wciR	Serotype8	AAAATAGATCAACTTATTGAATTAGAAGTGATAAAGGAAGAGGTGTTT- GCTCAGATTGGA
63	wciR	Serotype8	TAAAGCTTTGAAATTAAGAAAAAAAATTATAGCGGTTCCACGATTAGA- GCAGTTTGGAGA
64	wciR	Serotype8	ATGCTTTGATATAGAGCAGTTAGGAACTGTTTATCAAAAAGCTCAGACTTT- TACAACAAA
65	wciS	Serotype8	TTTATTGATGGCTCTCTTGTAACACGGTTAACCTATTCTAGTTATGCTCTT- CTTAAATTT
66	wciS	Serotype8	AACAACTTTCTTTTTTAGGAAGGATGGGCAAAAGAAAGGAGCCTAT- GATTTAATAGAT
67	wciS	Serotype8	AGGATAATGGCTGGTTAATTCAACCGGGTGATATTTCTCAGTTATCTAATATTATTTTAG
68	wciT	Serotype8	ATGGAATGAGGATAATTTTGATTTATCAGATTCACAATTTGCGAAGTCTGC- ATATGAATC
69	wciT	Serotype8	GGTGCAATATTATGAGCAAGCAAGTTTTGATATCAATCATTTGGTAACTGT- CAATACAAT
70	wciT	Serotype8	AATAATTGATGGATTAGCAATTTATCCAGATGATTACTTTTGTGGTTATGA- TCAGGAGGT
71	wchO	Serotype9A, 9V, 9L, 9N, 19A	ATTAACGATGAAAGAAACAGTGGATGCTGTTGAACAGTATGTTTTAAA- GAAGCATCCTTT
72	wchO	Serotype9A, 9V, 9L, 9N, 19F, 19B, 19C	TTTTGATGTATTATCAGGACACATTAAACGAGCTCCATTATGGATGCAAAA- ATTGAATCT
73	wchO	Serotype9A, 9V, 9L, 9N, 19A	GAAAGAATATATTATCCAATCATTCATGGATAATGGAATTAATGCTGTGTT- TATGGGGGT
74	wchO	Serotype9A, 9V, 9L, 9N, 19A	GAGTAGCGGGTATTGATTTGATGCAATGTCTTTTAGAGTTGTCAAATA- AAAAAGGATATT
75	wcjA	Serotype9A, 9V, 9L, 9N	AACAGGTGGACTATGGGAGAGCAAACTTTTATCAAAAGGAGTTCAACA- TCATAAAATTTT
76	wcjA	Serotype9A, 9V, 9L, 9N	TTAAAAAAAGCGTATTGTGTAGCTGTGGGTAAAGCGGTTAATGATAAT- TTGAAACATGAT
77	wcjA	Serotype9A, 9V, 9L, 9N	GTTGTTGAATGTATCAATAGTTTTGATTACTTAGTGTCATCATCTTTATAT- GAGGGGTTG

Spot identifier	Targeted GT gene	Specificity	Probe sequence $(5'-3')$
78	wcjB	Serotype9A, 9V, 9L, 9N	GAAAAGGCTAATTTAGAAAATGAACTAATTGTTTCGTTTACAACAATTCCA- AGCCGTCTT
79	wcjB	Serotype9L, 9N	AGTTGTTCTAGTTGATGACGATATCATTTATCCTCGAAATACTATAAAGAA- ACTGATTGC
80	wcjB	Serotype9L, 9N	CAATCCTGAGGAGAGTTTGGTATATTTGAATACCGTATATGATAACAA- CAATGATAAATG
81	wcjC	Serotype9A, 9V, 9L, 9N	AAATTTCTAGCTGAACAACTTGTAAAAGAAGGACATGAGGTATTTGCA- TACTCTGATGAT
82	wcjC	Serotype9A, 9V, 9L, 9N	TTATCAATAAAGGATTTATTAACCCATCTTCTCAAAAATGTATGGCCATTG- AAAATGCTG
83	wcjC	Serotype9A, 9V, 9L, 9N	CGAAAGTGATCCTAGAATACAATATTTAGGCTTTCAAGATACAAAAAACCT- CTATGAAAC
84	wcjB	Serotype9A, 9V	ATTGTAATTTTGGTTGATGATGACACTGTCTATTCATCGAATACCATCGAA- AAGTTAGTT
85	wcjB	Serotype9A, 9V	ACCCTGAAGAGAGTTTGGTGTATCTGAATGCTATATATGATAATAATGATAATAATGATAGGTGTA
86	wciB	Serotype10F, 10A, 10B, 10C, 47A	ATCAAGGTAATCATATCTCACACCTCAATCCTTATTATTGTGAATTGACAG- GATTATACT
87	wciB	Serotype10F, 10A, 10B, 10C, 31, 47A	TTTAGATGTAACGCGAGAAATTATAAAAGAGGTTTCGCCAGAATATTT- AGCAACATTTGA
88	wciB	Serotype10F, 10A, 10B, 10C, 47A	TGAATTTATTTGAGAAGGGCAAATCCTTCTTGAAAGCCAAGTATTTCG- GAAAAAATATG
89	wcrC	Serotype10A, 10C, 34, 35F, 43, 47F, 47A	GTTGCTGTATCTTTGGCAAACGAACTTACAAAAAGTATGAAGTTCATTTG- ATTGGAATT
90	wcrC	Serotype10A, 10C	TAACTGTTGGTCGTTTTGATTATCAAAAAGGATATGATTATCTTATCCAAG- TCGCGAAAA
91	wcrC	Serotype10A, 10C	TGGTTATCTGATAGATTGTTATGATACCGATAAGATGAGTGAG
92	wcrD	Serotype10A, 10B	GGATATGGTTCTTACGGATTTTACAGAACAACATGTTTATAACAATACTAC- TGTTCGAAA
93	wcrD	Serotype10A, 10B	ACCTATAGAACATCTATCCTAGTCGACAATAGAATTCGTTTAAGTGAAAAGACGTTTTAT
94	wcrD	Serotype10A, 10B	TATCAGAAGAATTATACAGACAAATTGAGCAGAGTTCTTATGAGTATATCC- CTACGAAAA
95	wciF	Serotype10A, 10B	AAGCATCATCAGATTGGATTTTCTTTCTAGATCCAGATGATTATTTGGAAG- ATTATACTC
96	wciF	Serotype10A, 10B	GGATAAAATTGTGATTAGTCCACTTGAAACATATAACTATTACCGTAGAGA- AGGTAGTAT
97	wciF	Serotype10A, 10B	AGGCTGACTCTGGTTTAACAGATTTTTCGAAAGATCGAAACCTATTAA- AAGTTGATTTTA
98	wcrG	Serotype10A, 10B	CTCTGTTGGATTATAAGGAACATGATATTTTTATTATTGTAGGCAGCAAAG- TTAATGTGG
99	wcrG	Serotype10A, 10B	GCTAGAAATATTCAGAACAAATATGTTCGTAAATTTGTAGCATATTACCGT- AAGCTAGAG
100	wcrG	Serotype10A, 10B	GCATCTAACTGGGTATCTATTAATCAGGATTTAGTTAGAATAATACTAGAA-GAAGAAA
101	wchK	Serotype11A, 11B, 11C, 11D,	GATAGATTAAAAGGTGAGGGATTTATTCAGGATGATGTTTTTATTCAGACT- GGTTTTTCA

14, 15F, 15A

TABLE 3: Continued.

Spot identifier	Targeted GT gene	Specificity	Probe sequence $(5'-3')$
102	wchK	Serotype11A, 11D	TTCTTATGATGAGATGAATCGCTATATAGATGAAGCAAATATTATCATTAC- ACATGGCGG
103	wchK	Serotype11F, 11A, 11D	CGAAGGGTATGAATTATCTCTGATTAATGATATAAGCGAATTGCAGTA- TAGTTTAAAGCA
104	wcyK	Serotype11F, 11A, 11D	CATCATAAATTAGATAAACTACTACGACCGATATTTTATCGAGTTTATACT- CAGGCATGT
105	wcyK	Serotype11F, 11A, 11D	CTAGGTCATGTTGGACGTTTTAATACTCAAAAAATCAATGTTTTCTAGTG- TCTCTAATG
106	wcyK	Serotype11F, 11A, 11D	GGTCAATTTGATGATATGAAATCTTTTGTGTCATCAATGGATATAATGTTG- CTTCCAAGT
107	wcrL	Serotype11F, 11A, 11D	TGTATTGAAAATCAAAATCAATTTGTGCAGGATGCATATAGAGATAAA- GCATGGGCTTTT
108	wcrL	Serotype11F, 11A, 11D	GTTATGTCCTGAATTAAATACACCTGTATTTAAACGTCTTGGTTATACTTA- TTCTGACTG
109	wcrL	Serotype11F, 11A, 11D	CAGATAAAACATTCTCTATTCATCATTATAGTGCTTCTTGGACTTCCTTAA- GAAATCAGA
110	wciJ	Serotype12F, 12A, 12B, 44, 46	GGAAATATATGCTGATTATCGTAAGAGAAAAAAAAGAAGAGAGAG
111	wciJ	Serotype12F, 12A, 12B, 44, 46	ACTTAACTTTTGCTGGAAATATTGGAAAAGCTCAGAATTTAGAGACTATTT- TGAAAGCAG
112	wciJ	Serotype12F, 12A, 12B, 44, 46	AATGTTGATCAGTTAGTGAGAAATATTCGTAAGTTCTGTTTGCTTTCTGTA- GAGGAAAGA
113	wcxB	Serotype12F, 12A, 12B, 44, 46	ATGTTCCGAAACAATTTCAACAGTATGCAGTGAAAATTGGTACAAAGT- CTGATATTCGTT
114	wcxB	Serotype12F, 12A, 12B, 44, 46	AAAAGAATATCCAGTGAAAGTAATTCATAATGGTATTGATACTACTGTCTT- TCAACCGAG
115	wcxB	Serotype12F, 12A, 12B, 44, 46	TAGAAAGTGCTAAACTTTATGGTCTCGTTTGTCAGGATAGAAACGTAG- CTTCTATTTTAT
116	wcxD	Serotype12F, 12A, 12B, 44, 46	GAAGAAGAATTTTTTTAAAGTTAGTGGAGCTTTGCGAAAAGTGTTGAA- AAAACAGCAGTT
117	wcxD	Serotype12F, 12A, 12B, 44, 46	TGTCAGCTCTCTTCTAAAAAAATTATCAGTGTTGGATCTTTAGTACGACAA- AAAGGTTTT
118	wcxD	Serotype12F, 12A, 12B, 44, 46	GATAGAGAAAAATTAGAGGAGAAAGTCAGGGAATACCAATTAGAAGGC- TTTATAAATTTG
119	wcxE	Serotype12F, 12A, 12B, 44, 46	ATAAAATCCCCGATAATCTTACCCAATTTTTTGGACGAGAAAATATAGAAG- AGAGAGATA
120	wcxE	Serotype12F, 12A, 12B, 44, 46	TATAAAACCTTGATTACTCCCATTTTGATAAAAGAACAGATACCAATTATT- CGGACGCAA
121	wcxE	Serotype12F, 12A, 12B, 44, 46	AGGTAGCAGATTTTGCTTTATTTCCTAAACAATGTAGTTTAAGTTTTATG- ATGCACAGG
122	wcxF	Serotype12F, 12A, 12B, 44, 46	AAGTTACAATGAGAAATATAATCATGATGAAAATTACGGTCGTTAGTTGTGA- CCATAAGGA

Spot identifier	Targeted GT gene	Specificity	Probe sequence (5'-3')
123	wcxF	Serotype12F, 12A, 12B, 44, 46	TGATTGTTTTTTTGGACGTATCAACAAAAATAAAGGTATCAAAGAACTGC- TTGAAGCCT
124	wcxF	Serotype12F, 12A, 12B, 44, 46	GAAATGCTCTTCGGTTATTACTTCTAATAGAGATAGAGGAGCCTATTTTC- TATTGAAAA
125	wchK	Serotype13, 14, 15B, 15C	CTTATGAAAAAATGAATCAATTGATTAAGGAATCAGATATTATCATTACCC- ATGGCGGTC
126	wchK	Serotype14	TAAAAATCCAATAATTGTTCCGCGGCTAAAAAAATTTGGTGAGCATGTAAA- TGATCACCA
127	wchK	Serotype14	AGGACAAACATTTTGAAACTTATTTGAATAACGAGAGATTTAATGTACGTTTCAATGTGG
128	wchL	Serotype14, 15B, 15C	TTGTGTTGATAGTGCCTTAAAGCAAAATTTAGAATCTCTTGAAGTGATTTT- GGTGAATGA
129	wchL	Serotype14, 15F, 15A	AAAAATTCTTGAACAGTATGGTGATAATCCCCAAGTGATGATTTTCCATCA- AGTGAACAT
130	wchL	Serotype14	GCTAAGTTATTTCTTCGTAGAAGAATTGAGGAAAACAATATTGCTTTTTCGACTGAAATG
131	wchL	Serotype14, 15F, 15A, 15B, 15C	TCCTAAAATTGAGGAGAACTACTACAAGCAACATATGGATTTTAGATTTTA- TCTTGCTAG
132	wchM	Serotype14	AATAGAAAGTATTTTGAATCAAACGTATGATAACCTTGAGGTTCTATTAGT- CGATGATGG
133	wchM	Serotype14	AATAGAAAGTATTTTGAATCAAACGTATGATAACCTTGAGGTTCTATTAGT- CGATGATGG
134	wchM	Serotype14	CAGTATTGTAACTGGATTGTTACAATAACTGTTAGTCATTACAATGTTTTG- AATGTAGCC
135	wchN	Serotype14, 15F, 15A, 15B, 15C	CAAAAAAATGATATGAACATTTCGAATAAAGTTTGGATTTGTTGGTTTCAG- GGCGAAGAA
136	wchN	Serotype14	TATGCGAGAAAACTACTCTGGGAGTATTGGCGTAGAAAAAATAGTTTA- TGCAATTATTTT
137	wchN	Serotype14	GAGTTAAATAATCAATTTTCAGAAAAAAGGTGGGAACAGCTAAAACAG- ATATCGGTGTTT
138	wchK	Serotype13, 15B, 15C	GATGAAGTATTTATTCAAATAGGATATTCCAGTTATATTCCGAAATATTGT- GAGTGGGAA
139	wchK	Serotype15B, 15C	GCATGTGAATGACCATCAGCTTCAATTCGTAAAACTGACGAAAGAAA
140	wchL	Serotype15B, 15C	AGAAATTTTGAACCAGTACGACAGGAATTCAAGGGTTAAGATTTTTCA- TCAGCTTAATAA
141	wchL	Serotype15F, 15A, 15B, 15C	GAAGAAAATAATATTACTTTTTCGACTGAGATGTCACTAGGTGAAGATATG- TCATTTGTG
142	wchM	Serotype15F, 15A, 15B, 15C	GAAAGTATTTTGAATCAGACTTATCAAAATATCGAGATTTTATTGGTTGAT- GACGGAAGC
143	wchM	Serotype15F, 15A, 15B, 15C	GTACTGCAATTGGATTGTTACAGCGACTACCAATCATAGTAAGATTTTAAA- TCCTAATTT
144	wchF	Serotype7B, 16F, 17F, 18F, 18A, 18B, 18C, 23F, 23A, 24F, 24A, 24B, 28F, 28A, 40, 48	GAAACTTTTGTTGAAAAATTAACAGCCTTCCAACAAGATAAGGCTATCCAA TATTATGTG

Spot identifier	Targeted GT gene	Specificity	Probe sequence $(5'-3')$
145	wchF	Serotype16F, 17F, 18F, 18A, 18B, 18C, 23F, 24F, 24A, 24B, 28F, 28A, 48	AAGGTCTTATGGTCAAACATGCAGCTCTTTTAGTGTGTGATAGTAAGA- ATATTGAAAAAT
146	wchF	Serotype16F, 17F, 18F, 18B, 18C, 23A	TTCGTTACTTGAAGCATTAGCATCCACAAAGTTAAACTTACTACTCGATGT- TGGTTTTAA
147	abp1	Serotype17F, 24F, 24A, 24B, 48	GCCAGTCATTATCTATACCCTTGAAAAATTTCAAAATCATCCAGAAATTGA- TGAAATCTG
148	abp1	Serotype17F, 24F, 24A, 24B, 48	ACACAAACTCCTCATGTTTACCATCTTGATAATATTCTATCGCTTCATGAA- AAAGCATTA
149	abp1	Serotype17F, 24F, 24A, 24B, 48	TTATTTCTCTCTTGGAACAGAGAAAAACTTGAAAATTACGACTGTAGAAGA- TCTCGATAT
150	wciP	Serotype17F	GAAGAAAAAGATAGACGGATTAAATTGATTGAAAAACATATCGGAATAT- CATGGAGCCTAT
151	wciP	Serotype17F	GTATACCAATCCTATCTCAACTTTTATGGCTCATAAGGTTTATGGATGTAA- TACGTTATT
152	wciP	Serotype17F	ATCTTAAAACGTATCTCGAAAATTGATGAATTAGCTAAAGATCATGCCTTGACTTACAAG
153	wcrV	Serotype17F	TCGACAGATAGTAGCAAACAGATAATTAACGAGTATCTTAATGCAGAC- AGTAGATTTAAA
154	wcrV	Serotype17F	CATGCAAAACTTAAGTTGTTCTGTCAGAATTTTAAGTTAGTGAGGAAA- CAGATTTTTAGG
155	wcrV	Serotype17F	CGATTTAATCTACTAAAAAATAACGGAGGAATGTGGGTTGACTCCACT- ATATATTTTACT
156	wchF	Serotype7B, 16F, 17F, 18F, 18A, 18B, 18C, 23F, 23A, 24F, 24A, 24B, 28F, 28A, 40, 48	TATAGCGTATGATATCGCTGCAATTAACAGAGCTATTGAAATTGCCAA- AGAAAATAAGGA
157	wchF	Serotype16F, 18F, 18B, 18C	TATAATCAGCTATTAGCAAGTACTGGATTTGATAAAGATCCACGAGTG- AAATTTGTTGGA
158	wciU	Serotype16F, 18F, 18A, 18B, 18C, 28F, 28A	AGAAAAAGTACAACCCGACATTATACATATTCACTCGTTTATGGGATTGCA- TAAAGAATT
159	wciU	Serotype16F, 18F, 18B, 18C, 28F, 28A	TCATCATCAGAGATTGACAACTGCAAATAATAAAATTAGAGTTGCTTATAT- TGGTCCAGA
160	wciU	Serotype18A, 18B, 18C	GACAAGGAAGATTTGTTGGCTAAAATCATCAATAATCAGTTGAAGAAA- ATTCCGCTTAAA
161	wciV	Serotype18A, 18B, 18C	AAATACATAACCTTTGTAGATTCAGATGACTATGTTTCTCTAGATATGCTG- CAAACTCTA
162	wciV	Serotype18F, 18A, 18B, 18C	AGAAGATGCTATTTTTCAAATTGATTGTTTAAAATTAGCAACATCTGCCCT- TGTTATCCC
163	wciV	Serotype18F, 18B, 18C	ACCCAATATCAAAATCAGTATTACGTCATTATCCAATCCATCGTTTACCTT- TTACTAAAC

TABLE 3: Continued.

Spot identifier	Targeted GT gene	Specificity	Probe sequence $(5'-3')$
164	wciW	Serotype18F, 18A, 18B, 18C	AAGTGCAACTTGAAGATAGGGCCTACAGAATACTAAAAAAGAAATACG- GTTCTTTAATTT
165	wciW	Serotype18F, 18A, 18B, 18C	TGGATTGACTCAACAGTGTATTGTACAGGAATTACTACCATAGAGACA- ATTGAAAAAAAT
166	wciW	Serotype18F, 18A, 18B, 18C	TACGAACGCAACACCACATATAATGGTTGATGAATTAAATAATGTTTTTC- AAAGGAACG
167	wchO	Serotype19F, 19B, 19C	ATAGATAGTGTAGAACAATATGTATTAGAAAAAAGACCACTACACTTG- ATGGGGGTGAAT
168	wchO	Serotype19F, 19B, 19C	GCTCAAAGTATTTAAGAGAGATTATCCAAATTTGATAGTTATTGGACACAGAAATGGCTA
169	wchO	Serotype19F, 19A, 19B, 19C	AATTTAGAGTGGTTATTCCGTGTAGCTAATGAGCCTAAACGTCTCTTTAAA-CGTTATTTT
170	wchO	Serotype19A	GAGTTGCTGGAATAGACTTGATGAAACATTTACTAGAGTTGTCTAATG- AAAAAGGATACT
171	wchQ	Serotype19F, 19A, 19B, 19C	ATCAGATTTAGAAATTGATGTTTTGATTAACCATGAAAATGCTGGTTTTGC- TCGTGGAAA
172	wchQ	Serotype19F, 19A, 19B, 19C	ATCAGTAGACTATAGAAAACAGGTAGAAAACCCAATTCTTCATGGTTCTTT- TATTGTATA
173	wchQ	Serotype19F, 19A	GGATACAAGAGAATTTATACACCTAAAATTAGAGTTTTGCACCATCAAAAT-GTTGCAACT
174	wciB	Serotype20	ATACTGGGGAAAACATTTCCCAGTTAAACCCTTATTACTGTGAATTAACAGGTTTATATT
175	wciB	Serotype20	AAAAAGGAAATATTATATTGAAACTCTATGTTCTCATTATGCACACACGCT-AGATGCTAG
176	wciB	Serotype20	AATGGCTGTTTCCGATTTTAGATTGTATGTTTGATCAGATTAATCTTTCAG-AGTTAACTG
177	whaJ	Serotype20	TTTCTCAAAAATTAGCGACCGAAAAACTCAAATATACGAGTCTTGAAATCAGATAAAGGAA
178	whaJ	Serotype20	GATTGATGAGTACGGTTTGAAGTTTAATACGAATTTGAGAGTTTCAGA- AGATAGTGATTT
179	whaJ	Serotype20	CTATGTTTTTTGAGCCTATACAAAATCTATCTGTATCTAGTGTTAGCAATT- TATCGCTAG
180	wciL	Serotype20	GATACGTTATTATTGGGAAATGTATAGATTCTTCAAAGAATATGCATCTGA- TTATCAGGC
181	wciL	Serotype20	TATACATTAGACAATAAATTTGTGCTAGGTCATGTAGGACGTTTGCATTTT-CAGAAGAAT
182	wciL	Serotype20	GACACTACTCTCAGAAGAAGGTGTACCAAAGGAAGTAAAAATCAATGA- TAATACTTTTTT
183	wcwK	Serotype20	AAACAAGATATAGAGATATGGATTTGTTTCAATATTGGTTTCGAGCGG- TAGAAAAACATG
184	wcwK	Serotype20	AATCTATTTAGCATTTTTTATTCAGGGATTATTGGTTATCATGATGCTCAT- GTCGCTATG
185	wcwK	Serotype20	GTGAATATGTGCCTCTGGCTTATTCAGGTAAAATTGAATCTATTATTCACAAAACAAAAGA
186	wciD	Serotype20	TGGCTCAGAAACTGGAAAAAGAGTATTCTGGCATAGTTAGT
187	wciD	Serotype20	CATAAAAATTGATGAGAATATGTTCTACGTTGACATGGAGTATATTGTTTT- TCCAACTCC
188	wciD	Serotype20	GAGACAATTGCTAGATGTGTTACTATTATGACAAATGTTTGTCTATCAATG- GAAGATACT

TABLE 3: Continued.

TABLE 3: Continued.

Spot identifier	Targeted GT gene	Specificity	Probe sequence (5'–3')
189	whaF	Serotype20	ACTTTAATACAAAAAACTGAATTTCCTAAATTTATCTGGACTATGTGGTGG- CAAGGAGAA
190	whaF	Serotype20	ATTTGGTTAGATTCAACGATGTATGTCCATCCAGATTTCCCTATTGAAATA- TTAGAAAGA
191	whaF	Serotype20	AGGGAAATAATAAAAAGTATCCCTAGATATTCTAGTCAAGAAGACATCTTT- TGGTTGAGA
192	wchF	Serotype22F, 22A	ACTTATATCGCCTATGGAACAGATACAAGCAAGTCTATTTTAAAACCTGAT- GACGAAAAA
193	wchF	Serotype18F, 22F, 22A, 23F	ATCGCTTTTAGAAGCTCTTGCTTCAACAAAGCTTAATTTACTGCTAGATGT- TGGCTTTAA
194	wcwA	Serotype22F, 22A	TAAGAAGACAAGGAGAATCGTTTTCTTTGGAATCTTATATCCGTAGTTTCT- CAGAATTAT
195	wcwV	Serotype22F, 22A	GAAAAACGGGGAAAAAATTAAAGTATTTTGGAGAAGGGGAATAAGATT- ATTTAGAAGTGG
196	wcwV	Serotype22F, 22A	GGAGAATAAGCAAAATATTCTTTATGTAGGCTCACTATCAAAAAGAAAAAA CACAGCTCA
197	wcwV	Serotype22F, 22A	ACCTTATTTAAAGAACTCTCAGCTTCAATTTATTTACCCATCATCACAACT- ATTTGTGCT
198	whaB	Serotype22F, 22A	TGGCAGTATAGAAAGGGTAGAAGCCTTATTTGCAAATAATGACGAGAT- AGTTATAATAAA
199	whaB	Serotype22F, 22A	CATCATCAAAGTCCTGTTGTTGAGAAGATCAATTCTATATCTAAGGCAAAT- AAAGAACTT
200	whaB	Serotype22F, 22A	TTATTTTACATGGGAGTTGTGTAATTTTTTCACCATTATATGTTTCAGAGG- AGGAGTTTG
201	wchF	Serotype23F, 23A	CCATTTACTGGAAGAAAGATAATCTTCATGAGATTATTGAAACGAGTG- AACAAAAAACAC
202	wchV	Serotype23F, 23A, 23B	CCTCATTTTTGTTGACAGTGATGATTTTGTCTCTCAAGATATGGTATCTTA- TTTAGTATC
203	wchV	Serotype23F, 23A	GGCCAAGATATTTAAAAGAGAGTTGTTTGATGATATAAGATTTCCTGT- AGGTAAGCTATT
204	wchV	Serotype23F, 23A	TTTTGGAGATTACGAACACAATTATTAATCACTATGGTGATAATTTACGCG- TGTATACTG
205	wchW	Serotype23F, 23A	ATTTGAAACAAAATTATCAAATAAACTTGGCCTACAAAAATCTTTGCATGG- AAAGGGTGG
206	wchW	Serotype23F, 23A	CGGGGGGATATTATACAAAAGAGTATAAACAACTATTCAGTTCGGTAG- TAGAAAATATTA
207	wchW	Serotype23F, 23A	CCTATAGAGTAAATCTCCATCAATTTTTAATAAACGAGATCTCAGATGCTA- CAGTAAGAT
208	wciB	Serotype33F, 33A, 34, 35A, 35B, 35C, 37, 41F, 41A, 42	TTGGTTTTATCGGTGATAATACTGGCGATAATATATCCTCTCTAAATCCAT- ATTATTGTG
209	wciB	Serotype33F, 33A, 34, 35A, 35B, 35C, 37, 41F	ATAGTTCCAAAGAAGCGAAAGTATTATATTGAAACTCTTTATTCACATTAT- GCCCATACC
210	wciB	Serotype33F, 33A, 34, 35A, 35B, 35C, 37, 41F	AACTATTAGATGATTATTTACCGTGGCTTTTTTCTATTCTGGATACTATGT- ACGAACAGA
211	wciC	Serotype33F, 33A, 37	CAAATTTTAATATCTGATACAGATGTTTATTATTTTACTCCAGCTGGTTCA- GTAGCTGGT
212	wciC	Serotype33F, 33A, 37	TTACGAAATTTTATTAGAAGTTGCTAAGAAGATGGTGGGGGATGAGAA- ATATCACTTTTA
213	wciC	Serotype33F, 33A, 37	GTTTTACCATCGTATTATAAAGATGAAACTTTACCTATCAGTATGTTAGAA-GCAATGGCA

Spot identifier	Targeted GT gene	Specificity	Probe sequence (5'–3')
214	wciD	Serotype33F, 33A, 37	AATAGCAAGACAATTCGAGAGAGAGAATATGAGGGAATTGTTAGAGTTAT- AAGTAAGGAAAA
215	wciD	Serotype33F, 33A, 37	TGCAAGAGAACAATATTCGGCTGTCTGAAAAAATGTTCTATGTAGATA- TGGAATATATTG
216	wciD	Serotype33F, 33A, 37	GCATAAACAAGTGATCTATCATTTGGTTGATTTTTATAATCAAATGAGATC- TAGCGCTGT
217	wciE	Serotype33F, 33A, 37	GCCAATTTTTAAAATCCTATAACTTTAAAGAGGTATCGCACAAGGAGA- TAGAACAAAGAA
218	wciE	Serotype33F, 33A, 37	CTGAATTATTTAAAAAAAGATTTTTATACTATTCGAGCAAAGACACATGAGAGAGA
219	wciE	Serotype33F, 33A, 37	TAAAGTCGAAGAAAATAATCAGGAGTTGTTCTTTTTGGCAGACAATTTTTC- TAACCAGTA
220	wciF	Serotype33F, 33A, 37	CTGGGGAAATATGTGATGAATATGGGAAACTGTATGATAATATTCATG- TTTTCCATAAGA
221	wciF	Serotype33F, 33A, 37	CAGAGCGTTTGTTGAATATTAAAACAGTTGCTCATACCGATTTGCCTATAT- ATCATTATT
222	wciF	Serotype33F, 33A, 37	AAGGAATTGTTAGCAGCCTTAAATGCTAAAAGAGTAATTGGCTCCTTTATT- TTGAGTAAT
223	16S	Streptococcus pneumoniae	TATTGGAAACGATAGCTAATACCGCATAAGAGTAGATGTTGCATGACA- TTTGCTTAAAAG
224	16S	Streptococcus pneumoniae	ATAAGTCTGAAGTTAAAGGCTGTGGCTTAACCATAGTAGGCTTTGGAA- ACTGTTTAACTT
225	aroE	Streptococcus pneumoniae	ATTTCAAAAACGGTGTTTCAAAGGGTTGGTATATGATATCTGCAACTA- AGAGAGTTTCTG
226	aroE	Streptococcus pneumoniae	AGCAGGGTCATCTTTTTACCTGAAATTGTAAAAGAAGGCAAGCACTTA- AAAAATCCCTTG
227	aroE	Streptococcus pneumoniae	TCAAAGGCTCTATTGTGGATGAAGGGAGAAATAGAATGCTTAATAGGA- TTGGCAACAACT
228	ddl	Streptococcus pneumoniae	TATTGAGCTCGTTGAGAAAAATCTCTCCCTTATCTGTATAGAAGAAATCGC- AACGAGATA
229	ddl	Streptococcus pneumoniae	ATGTTTGACGGCTTAGTGAAGACTGGATAAGCCAATTTTTCTTCCACTTCA- GCGATTTTA
230	ddl	Streptococcus pneumoniae	CATGACTAAATTCCTGTGTTTTGATAAAGTCACCTGACTGA
231	gdhA	Streptococcus pneumoniae	TGAATTCCTCCAAGCTGTTGAAGAATTTTTCAACACTTTGGAACCTGTATT- TGAAAAACA
232	gdhA	Streptococcus pneumoniae	TAAACCAAGGGATTTTGAAATTCCTCGGATTTGAACAAATCTTTAAAAACG- TCTTGACTG
233	gdhA	Streptococcus pneumoniae	TATACTGAAGAAATGCTCAAAGCTAACGGTAACAGCTTTGCTGGTAAG- AAAGTGGTTATT
234	gdhA	Streptococcus pneumoniae	ATGGACGTCTCAAAGACATCATGACCAACATCTTTAACACAGCTAAAA- CAACTTCAGAAA
235	glcK	Streptococcus pneumoniae	AAATGGTCAATCAAGACCAACATTTTGGATGAGGGAAGTCATATCGTT- GATGATATGATT
236	glcK	Streptococcus pneumoniae	AACAAAAGATTGAAAAAGCTTTGGGCATTCCATTTTTCATCGATAATGATG- CCAACGTAG
237	glcK	Streptococcus pneumoniae	AGAATTCCTTCTACAAGGTGTTCAAAAAGTTTACGATGAAAATAGTTTCCC-ACAAGTACG
238	spi	Streptococcus pneumoniae	AGAAGGTATTCTCCTTCTGGAACAGTAAAGCTAAAGTTGGTGTTGTAG- TTGACATCAACT
239	spi	Streptococcus pneumoniae	TGATATAGTCTGCTAGATAAGGCTCGTCCGTTTCTTTGTCATTGATGTAGA- GTTTATCAT
240	spi	Streptococcus pneumoniae	AACATTGCTCCAAAAAAAGATACGGCTCAAAGCTAGTAATGACAGAAT- CAGGAGGAATAA
241	tktA	Streptococcus pneumoniae	ACAATTTTTACAAGATTTTCTACAGTAAAGCCATATTCTGCCAATACTTTT- GGTGCTGGG

TABLE 3: Continued.

Spot identifier	Targeted GT gene	Specificity	Probe sequence (5'-3')
242	tktA	Streptococcus pneumoniae	CAATCAAGATGGTATCAAAGTCGGCTGCATTTTCATATACAACATAAG- CACCTTTAGCAA
243	tktA	Streptococcus pneumoniae	TTCAAGATTGTTCCCATTGCAAATTCACGAACACCAAACTGAATGTTACGA TTCAAGCGA
244	tktA	Streptococcus pneumoniae	AACATGTTCTTTGAAATCAGCATATACTTGTTCTGGAATTTCAAATGGTTC- GTAGTCCCA
245	tktA	Streptococcus pneumoniae	TCTCCACAGATAACGTAAGTATAGTGGTCAAAGATATTGTAGCCTTCA- CGGTTATATTTG
246	xpt	Streptococcus pneumoniae	AGATTCCTTTTTAACCCACCAAGTTGACTTTAGCTTGATGCGAGAGAT- TGGTAAGGTTTT
247	xpt	Streptococcus pneumoniae	ATGATTTTCGCCAAAAAAGCTAAGAACATCACCATGAACGAAGGCATC- TTAACTGCTCAA
248	xpt	Streptococcus pneumoniae	TTTGATTATCGACGATTTCCTTGCTAATGGCCAAGCTGCTAAAGGCTTGAT- TCAAATCAT
249	KP _ gapA	Klebsiella pneumoniae	GACGTTGTTGCTGAAGCAACCGGTATCTTCCTGACCGACGAAACCGCT- CGTAAACACATC
250	KP_rpoB	Klebsiella pneumoniae	AACGGTGTGGTTACTGACGAAATTCACTACCTGTCTGCTATCGAAGAA- GGCAACTACGTT
251	KP_mdh	Klebsiella pneumoniae	TGTACGATAAAAACAAACTGTTCGGCGTTACCACGCTGGACATCATCC- GTTCCAATACCT
252	KP_pgi	Klebsiella pneumoniae	CCTGGCCTTTGGTAAATCCCGCGAAGTGGTTGAGCAGGAATATCGCGA- TCAGGGTAAAGA
253	SA_arcC	Staphylococcus aureus	TGATAGGCTATTGGTTGGAAACTGAAATCAATCGCATTTTAACTGAAA- TGAATAGTGATA
254	SA_aroE	Staphylococcus aureus	AAGTTTTGATTGGTCATTAGTTCCTGGTTATATTGTTGCTCAAATGTTAGG- TGCAATTGT
255	SA_glpF	Staphylococcus aureus	TAAGAATTACTTTGCCAACTTTTTAAGTGAGATTATCGGAACAATGGCATT- AACTTTAGG
256	SA_gmk	Staphylococcus aureus	CGTAGATTACTTTTTTAAAACTAGGGATGCGTTTGAAGCTTTAATCAAAGA- TGACCAATT
257	LP_acnF	Legionella pneumophila	CGAAAAAAGGGGTTGTTGGTAAATTTGTTGAATTTTATGGTCCTGGA- CTTAATGATTTA
258	LP_mompS	Legionella pneumophila	TCAATGTGAACTGGTATCATTTTGATAACGACAGTGATCACTGGTTTGATT- TTGCTAACT
259	CP_groES	Chlamydophila pneumoniae	TTCTTTACCTGCGTTACTTGCAATTTGCTTTAATGGAGCTGTTAATGCTTT- TAGAATAAT
260	CP_gyrA	Chlamydophila pneumoniae	GTTTGGTGGCTAAAAATAAGAAGCCGGCATTATCAAAATTCTTAATATTCAATATAGCTG
261	CP_gyrB	Chlamydophila pneumoniae	CCAAGACCTTTATACCTCTGAATTTCTATGCCTTTTCTTCCAAGATTTTTA- AGATAGTTA
262	CP_dnaA	Chlamydophila pneumoniae	CCTGCTGCTTCTAAAACATCTTTTAAAAGAGTTTTCACATCATCTTCATAT- AGTAATTGG
263	CP_accA	Chlamydophila pneumoniae	TGATAACAGTATCGATAATGCCAAATTGTTTTAAGTTTTCTCCATGCATTT- TCAACATGG
264	CP_dnaK	Chlamydophila pneumoniae	TCAAAAAACAAGAAGGCATTGATCTTAGCAAAGATAATATGGCCTTAC- AAAGACTTAAAG
265	MP_gyrB	Mycoplasma pneumoniae	AGGAACCTTTATTTGAGGACATTATCTTTGGTGAAAAAACCGATACTGTTAAATCAGTTA
266	MP_gyrA	Mycoplasma pneumoniae	ACAAGATCAAATTGACAAAATTCGTCAGGAATTAGCACAATCAGCAAT- TAAAAACATCTC
267	MP_dnaJ	Mycoplasma pneumoniae	TTGCGCAAGCTCAAGGAATTTATTAAACCTAATCAAGAGGTAAAACAATAT- TTAAACGCA

Spot identifier	Targeted GT gene	Specificity	Probe sequence (5'-3')
268	MP_lgt	Mycoplasma pneumoniae	TGGGATTGCCTTTGGCATCTTAATGTTTGTCTTGAAGTTAATTTACTTTTA- CAAGATTCA
269	MP_fus	Mycoplasma pneumoniae	TAAGCTTCCGTGAAACCTTCAATAAAGAAAGTGAAGTTGAGGGTAAAT- ACATTAAACAAT
270	MP_lspA	Mycoplasma pneumoniae	TTTGAAGAACTGAATTAAAAAGGAGAGGAACAGACCAATAAAACTAAA- GGTAATGCAACA
271	PA_trpE	Pseudomonas aeruginosa	TCACCGAAAAAATGGTGATCGAACGTTACTCCAACGTCATGCACATCG- TGTCCAACGTCA
272	PA_nuoD	Pseudomonas aeruginosa	GATCATGATGGCGGAGTTCTTCCGTATCCTGAACCACCTGCTGTACCT- GGGCACCTATAT
273	SP_gki	Streptococcus pyogenes	ATTCAGCCATCAAAGCAGCTATTGACAATGGTGAAGGTGTTACCAGTA- AAGACATTTTCA
274	SP_xpt	Streptococcus pyogenes	ATCGCTGGTAAATTCCTATCTAAAGAAGACAAGGTTTTGATTATTGATGAC- TTTTTAGCT

TABLE 3: Continued.

diagram of the probe positions on the microarray is shown in Figure 1(a).

3.2. Evaluation of the Microarray. A total of 274 oligonucleotide probes were used in this microarray, including positive and negative controls and GT gene-specific probes. The microarray probes were tested using 36 pneumococcal isolates from 23 vaccine-associated serotypes and 19 additional pneumococcal isolates belonging to other serotypes (Table 1). Figure 1(b) shows the examples of scanned pictures of 6 strains representing different serotypes. Examples of the same serotype were tested repeatedly and shown to have an identical signal pattern, for example, 5 times for serotype 3 (data not shown). Of 23 strains representing 23-valent vaccine serotype, 18 strains hybridized to all the specific set of probes, and four strains hybridized to almost all the specific set of probes (Table 4). The strain representing serotype 22F may actually belong to serotype group 22F/22A, since this sample failed to hybridize specifically to wchF and wcwA probes but hybridized to the rest of group 22F/22A specific probes. Of the 13 strains representing the 23 vaccinerelated serotypes, only 1 isolate (serotype 46), failed to hybridize to a specific probe while the other 12 strains hybridized perfectly. Of the 20 nonvaccine serotypes, 19 strains either hybridized partially to GT-specific probes or did not hybridize to any probes. One strain, representing serotype 23A, hybridized to most of the 23F-specific probe; thus, 23A may be indistinguishable from 23F using GT gene sequences.

4. Discussion

In order to develop a more effective *S. pneumoniae* vaccine, simple detection methods are required to serotype large numbers of clinical isolates. Conventional serotyping methods using large panels of antisera are labourious and require technical expertise. Our microarray method can determine serotype of a strain at one time and needs no expertise.

In addition, the microarray method described here has the potential to be automated. To our knowledge, our report describes the first microarray to utilize GT genes to predict serotype of any bacteria.

Several molecular typing methods have been developed based on serotype-specific sequences [12–21]. Wang et al. [21] described microarray method using *wzy* and *capA* genes. Our approach is different in that GT genes were selected as serotype-specific genes. Since GTs catalyze the transfer of the sugar moiety to an acceptor and generate a serotype-specific capsular polysaccharide, detecting GT genes can directly reflect polysaccharide structure. We discovered considerable variability within *S. pneumoniae* GT genes, which provides groundwork for future investigations into new *S. pneumoniae* capsular types. Our method using GT genes can not only discriminate serotypes but can give information of the capsular polysaccharide structure.

The DNA microarray described here accurately detects the majority of S. pneumoniae serotypes and serogroups included in the 23-valent vaccine and in the 7, 9, 11, 13-valent conjugate vaccines, which will permit serotype surveillance before and after vaccination. Since 1983, the 23-valent pneumococcal vaccine has been administered to persons in the United States aged >2 years with certain underlying medical conditions or aged >65 years. In 2000, the more effective PCV7, 7-valent pneumococcal conjugate vaccine, which protects against serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F was approved for administration [22]. As a result of PCV7, antibiotic-resistant invasive pneumococcal infections have decreased dramatically in young children and older persons [23]; however, an increase in disease associated with serotypes not included in the PCV7 vaccine, has been observed [24, 25]. To address serotype vaccine coverage, the Advisory Committee on Immunization Practices (ACIP) issued recommendations in February 2010 for a newly licensed 13-valent pneumococcal conjugate vaccine (PCV13), which contains the seven serotypes in PCV7 (4, 6B, 9V, 14, 18C, 19F, and 23F) and six additional serotypes (1, 3, 5, 6A, 7F, and 19A) [26]. Taken together, our DNA

84	67	80	66	51	43	50	42	27	19	26	18	77	76	61	57	60	56	37	31	36	24	13	6	12	5
85	65	70	64	47	41	46	40	25	17	22	16	75	74	59	55	49	54	35	23	34	29	11	4	10	3
69	63	68	62	45	38	44	38	21	15	20	14	72	85	48	53	58	52	33	29	32	28	9	2	8	1
155	149	154	148	130	122	129	121	104	98	103	97	158	157	140	133	139	128	116	110	115	109	92	86	91	71
153	147	151	143	127	120	126	119	102	96	101	95	156	146	138	125	137	134	114	108	113	107	90	83	89	82
151	142	150	141	124	118	123	117	100	94	99	93	145	144	136	132	135	131	112	106	111	105	88	81	87	78
Р	218	Р	217	200	194	199	193	177	171	176	169	Р	Р	212	206	211	205	189	183	188	182	166	160	165	159
Р	220	Р	219	202	196	201	195	179	173	178	172	Р	Р	214	208	213	207	191	185	190	184	73	162	170	161
Р	222	Р	221	204	198	203	197	181	175	180	174	Р	Р	216	210	215	209	192	187	7	186	168	164	167	163
N	N	N	N	N	N	N	N	Р	Р	Р	Р	N	N	N	N	N	N	N	N	N	N	Р	Р	Р	Р
E	E	Е	E	Е	E	N	Ν	N	Р	Ν	Р	N	E	E	E	E	Е	E	N	N	N	Р	Р	Р	Р

(a)



(b)

FIGURE 1: (a) Microarray oligonucleotide probes layout. Oligonucleotides 1 to 222 are provided in Tables 2 and 3. P represents *S. pneumoniae* housekeeping genes and 16S rDNA positive control oligonucleotides. N indicates negative control oligonucleotides designed from housekeeping genes of other bacterial species. E denotes empty spot. (b) Scanned microarray images of *S. pneumoniae* genomic DNA hybridized with 6 samples (serotype 3, 9V, 11A, 19F, 22F and 22A). The numbers correspond to the spot identifiers given in Tables 2 and 3, and Figure 1(a) P indicates positive spot.

	Saratura	Strain ID	Desitive probe	Microarray result			
	Selotype	Strain ID	Positive probe		Assined group		
	1	ATCC6301	1, 2, 3, 4, 5, 6	Perfectly matched	1		
	2	ATCC6302	7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18	Perfectly matched	2		
	3	D36	19, 20, 21	Perfectly matched	3		
	4	JHK27	22, 23, 24, 25, 26, 27, 28, 29, 30	Perfectly matched	4		
	5	ATCC6305	23, 24, 31, 32, 33, 34, 35, 36, 37	Perfectly matched	5		
23 serotypes	6B	MSC1047	38, 39, 41, 42, 43	1 probe of group 6A/6B did not hybridized	6A/6B		
1ncluded in 23-valent vaccine	7F	ATCC10351	44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58	Perfectly matched	7F/7A		
	8	ATCC6308	59, 60, 61, 62, 63, 64, 65, 66, 67, 68, 69, 70	Perfectly matched	8		
	9V	KD10-11	71, 72, 73, 74, 75, 76, 77, 78, 84, 85, 81, 82, 83	Perfectly matched	9A/9V		
	9N	KD01-26	71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83	Perfectly matched	9L/9N		
	10A	ATCC8334	86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100	Perfectly matched	10A		
	11A	SSI11A/2	101, 102, 103, 104, 105, 106, 107, 108, 109	Perfectly matched	11A/11D		
	12F	ATCC6312	11, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124	1 extra probe of group 2 hybridized	12F/12A/12B/44/46		
	14	D59	101, 125, 126, 127, 128, 129, 130, 131, 132, 133, 134, 135, 136, 137	Perfectly matched	14		
	15B	ATCC10354	125, 128, 131, 135, 138, 139, 140, 141, 142, 143	Perfectly matched	15B/15C		
	17F	ATCC6317	144, 145, 146, 147, 148, 149, 150, 151, 152, 153, 154, 155, 156	1 extra probe of group 18B/18C hybridized	17F		
	18C	ATCC10356	156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166	Perfectly matched	18B/18C		
	19F	D33	72, 167, 168,169, 171, 172, 173	Perfectly matched	19F		
	19A	D4	71, 73, 74, 169, 170, 171, 172, 173,	1 extra probe of group 19F hybridized	19A		

TABLE 4: Microarray results of each strain.

TABLE 4: 0	Continued.
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	Serotype	Strain ID	Positive probe ^a	Microarray result			
	ociotype	Strain 12	rostive probe		Assined group		
	20	ATCC6320	174, 175, 176, 177, 178, 179, 180, 181, 182, 183, 184, 185, 186, 187, 188, 189, 190, 191	Perfectly matched	20		
	22F	KD01-23	7, 8, 44, 195, 196, 197, 198, 199, 200	5 probes of group 22F/22A did not hybridized and 1 extra probe of group 7F/7A hybridized	22F/22A		
	23F	KD11-15	144, 145, 156, 193, 201, 202, 203, 204, 205, 206, 207	Perfectly matched	23F		
	33F	ATCC10370	208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222	Perfectly matched	33F/33A/37		
	6A	MSC1943	38, 39, 40, 41, 42, 43	Perfectly matched	6A/6B		
	7A	ATCC6307	44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58	Perfectly matched	7F/7A		
Other serotypes included in 23	9A	ATCC8333	71, 72, 73, 74, 75, 76, 77, 78, 84, 85, 81, 82, 83	Perfectly matched	9A/9V		
groups	9L	ATCC10349	71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83	Perfectly matched	9L/9N		
	11D	SSI11D/1	101, 102, 103, 104, 105, 106, 107, 108, 109	Perfectly matched	11A/11D		
	12A	SSI12A/5	110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 274	Perfectly matched	12F/12A/12B/44/46		
	12B	SSI12B/1	110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 274	Perfectly matched	12F/12A/12B/44/46		
	15C	SSI15C/2	125, 128, 131, 135, 138, 139, 140, 141, 142, 143	Perfectly matched	15B/15C		
	18B	ATCC10355	156, 157, 158, 159, 160, 161, 162, 163, 164, 165, 166	Perfectly matched	18B/18C		
	22A	ATCC10363	7, 48, 49, 192, 193, 194, 195, 196, 197, 198, 199, 200	Perfectly matched	22F/22A		
	33A	ATCC8340	208, 209, 210, 211, 212, 213, 214, 215, 216, 217, 218, 219, 220, 221, 222	Perfectly matched	33F/33A/37		

		Table 4	: Continued.					
	Serotype	Strain ID	Positive probe ^a	Microarray result				
	outotype		rounteproof	Assined group				
	44	SSI44/3	110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 274	Perfectly matched	12F/12A/12B/44/46			
	46	SSI46/2	110, 111, 112, 113, 114, 115, 116, 117, 119, 120, 121, 122, 123, 124, 274	1 probe of group 12F/12A/12B/44/46 did not hybridized	12F/12A/12B/44/46			
	7B	ATCC10348	143, 155	Partial hybridization	Not included in 23 group			
	7C	ATCC10350	none	None hybridization	Not included in 23 group			
	10F	ATCC6310	86, 87, 88	Partial hybridization	Not included in 23 group			
	10B	SSI10B/2	71, 72, 73, 74, 78, 79, 80, 81, 82, 83	Partial hybridization	Not included in 23 group			
	10C	SSI10C/2	71, 72, 73, 74, 75, 76, 77	Partial hybridization	Not included in 23 group			
Serotypes not included in 23	11F	ATCC6311	103, 104, 105, 106, 107, 108, 109	Partial hybridization	Not included in 23 group			
groups	11B	SSI11B/2	101	Partial hybridization	Not included in 23 group			
	11C	ATCC10353	101, 274	Partial hybridization	Not included in 23 group			
	15F	ATCC6315	101, 129, 131, 135, 141, 142, 143	Partial hybridization	Not included in 23 group			
	15A	ATCC6330	101, 129, 131, 135, 141, 142, 143	Partial hybridization	Not included in 23 group			
	17A	SSI17A/2	none	None hybridization	Not included in 23 group			
	18F	ATCC6318	144, 145, 156, 157, 158, 159, 162, 163, 164, 165, 166, 193	Partial hybridization	Not included in 23 group			
	18A	ATCC10344	144, 145, 156, 158, 160, 161, 162, 164, 165, 166	Partial hybridization	Not included in 23 group			
	19B	ATCC10358	72, 167, 168,169, 171, 172	Partial hybridization	Not included in 23 group			
	19C	ATCC10359	72,169,171,172	Partial hybridization	Not included in 23 group			
	23A	KD12-06	144, 146, 156, 201, 202, 203, 204, 205, 206, 207	1 probe of group 23F did not hybridized	23F			
	23B	ATCC10364	7, 46, 202,	Partial hybridization	Not included in 23 group			

none

none

49, 57

Explanatory notes: ^aThe numbers correspond to the spot identifiers given in Tables 2, 3, and Figure 1(a).

ATCC10342

ATCC8339

SSI33D/2

33B

33C

33D

Not included in 23

group

Not included in 23

group

Not included in 23

group

None

hybridization

None

hybridization

Partial

hybridization

microarray will be able to monitor serotype prevalence of all vaccine-related serotypes. However, in examining serotype replacement in vaccinated population a further study to distinguish more than 90 serotypes is required and is currently under investigation. Moreover, further study of the reproducibility of the microarray is needed.

5. Conclusion

We developed a *S. pneumoniae* DNA microarray that identifies GT gene polymorphisms to distinguish capsular types. We believe that our microarray system is more reliable and cost-effective and will help to survey the emergence of new *S. pneumoniae* serotype.

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References

- T. van der Poll and S. M. Opal, "Pathogenesis, treatment, and prevention of pneumococcal pneumonia," *The Lancet*, vol. 374, no. 9700, pp. 1543–1556, 2009.
- [2] K. McIntosh, "Community-acquired pneumonia in children," *New England Journal of Medicine*, vol. 346, no. 6, pp. 429–437, 2002.
- [3] CDC, "Preventing pneumococcal disease among infants and young children: recommendations of the Advisory Committee on Immunization Practices (ACIP)," *Morbidity and Mortality Weekly Report*, vol. 49, no. 6, pp. 1–35, 2000.
- [4] J. O. Kim and J. N. Weiser, "Association of intrastrain phase variation in quantity of capsular polysaccharide and teichoic acid with the virulence of *Streptococcus pneumoniae*," *Journal* of Infectious Diseases, vol. 177, no. 2, pp. 368–377, 1998.
- [5] J. Henrichsen, "Six newly recognized types of *Streptococcus pneumoniae*," *Journal of Clinical Microbiology*, vol. 33, no. 10, pp. 2759–2762, 1995.
- [6] J. O. Klein, "The epidemiology of pneumococcal disease in infants and children," *Reviews of Infectious Diseases*, vol. 3, no. 2, pp. 246–253, 1981.
- [7] J. Yother, "Capsule," in *The Pneumococcus*, E. I. Tuomanen, Ed., pp. 30–48, ASM Press, Washington, DC, USA, 2004.
- [8] E. García, D. Llull, R. Muñoz, M. Mollerach, and R. López, "Current trends in capsular polysaccharide biosynthesis of *Streptococcus pneumoniae*," *Research in Microbiology*, vol. 151, no. 6, pp. 429–435, 2000.
- [9] S. D. Bentley, D. M. Aanensen, A. Mavroidi et al., "Genetic analysis of the capsular biosynthetic locus from all 90 pneumococcal serotypes.," *PLoS Genetics*, vol. 2, no. 3, article e31, 2006.
- [10] J. E. G. Van Dam, A. Fleer, and H. Snippe, "Immunogenicity and immunochemistry of *Streptococcus pneumoniae* capsular polysaccharides," *Antonie van Leeuwenhoek*, vol. 58, no. 1, pp. 1–47, 1990.
- [11] D. M. Aanensen, A. Mavroidi, S. D. Bentley, P. R. Reeves, and B. G. Spratt, "Predicted functions and linkage specificities of the products of the *Streptococcus pneumoniae* capsular

biosynthetic loci," Journal of Bacteriology, vol. 189, no. 21, pp. 7856–7876, 2007.

- [12] D. A. Brito, M. Ramirez, and H. De Lencastre, "Serotyping Streptococcus pneumoniae by multiplex PCR," Journal of Clinical Microbiology, vol. 41, no. 6, pp. 2378–2384, 2003.
- [13] F. Kong, M. Brown, A. Sabananthan, X. Zeng, and G. L. Gilbert, "Multiplex PCR-based reverse line blot hybridization assay to identify 23 *Streptococcus pneumoniae* polysaccharide vaccine serotypes," *Journal of Clinical Microbiology*, vol. 44, no. 5, pp. 1887–1891, 2006.
- [14] F. Kong and G. L. Gilbert, "Using cpsA-cpsB sequence polymorphisms and serotype-/group-specific PCR to predict 51 Streptococcus pneumoniae capsular serotypes," Journal of Medical Microbiology, vol. 52, no. 12, pp. 1047–1058, 2003.
- [15] F. Kong, W. Wang, J. Tao et al., "A molecular-capsular-type prediction system for 90 *Streptococcus pneumoniae* serotypes using partial *cpsA-cpsB* sequencing and *wzy-* or *wzx-specific* PCR," *Journal of Medical Microbiology*, vol. 54, no. 4, pp. 351– 356, 2005.
- [16] E. R. Lawrence, C. A. Arias, B. Duke et al., "Evaluation of serotype prediction by *cpsA-cpsB* gene polymorphism in *Streptococcus pneumoniae*," *Journal of Clinical Microbiology*, vol. 38, no. 4, pp. 1319–1323, 2000.
- [17] E. R. Lawrence, D. B. Griffiths, S. A. Martin, R. C. George, and L. M. C. Hall, "Evaluation of semiautomated multiplex PCR assay for determination of *Streptococcus pneumoniae* serotypes and serogroups," *Journal of Clinical Microbiology*, vol. 41, no. 2, pp. 601–607, 2003.
- [18] R. Pai, R. E. Gertz, and B. Beall, "Sequential multiplex PCR approach for determining capsular serotypes of *Streptococcus pneumoniae* isolates," *Journal of Clinical Microbiology*, vol. 44, no. 1, pp. 124–131, 2006.
- [19] F. Zhou, F. Kong, Z. Tong, and G. L. Gilbert, "Identification of less-common *Streptococcus pneumoniae* serotypes by a multiplex PCR-based reverse line blot hybridization assay," *Journal of Clinical Microbiology*, vol. 45, no. 10, pp. 3411–3415, 2007.
- [20] S. L. Batt, B. M. Charalambous, T. D. McHugh, S. Martin, and S. H. Gillespie, "Novel PCR-restriction fragment length polymorphism method for determining serotypes or serogroups of *Streptococcus pneumoniae* isolates," *Journal of Clinical Microbiology*, vol. 43, no. 6, pp. 2656–2661, 2005.
- [21] Q. Wang, M. Wang, F. Kong et al., "Development of a DNA microarray to identify the *Streptococcus pneumoniae* serotypes contained in the 23-valent pneumococcal polysaccharide vaccine and closely related serotypes," *Journal of Microbiological Methods*, vol. 68, no. 1, pp. 128–136, 2007.
- [22] P. H. Mäkelä and J. C. Butler, "History of pneumococcal immunization," in *Pneumococcal Vaccines*, G. R. Siber, K. P. Klugman, and P. H. Mäkelä, Eds., chapter 1-2, pp. 19–29, ASM Press, Washington, DC, USA, 2006.
- [23] M. H. Kyaw, R. Lynfield, W. Schaffner et al., "Effect of introduction of the pneumococcal conjugate vaccine on drugresistant *Streptococcus pneumoniae*," *New England Journal of Medicine*, vol. 354, no. 14, pp. 1455–1463, 2006.
- [24] K. K. Hsu, J. E. Kellenberg, S. I. Pelton, D. S. Friedman, M. R. Moore, and H. T. Jordan, "Emergence of antimicrobial-resistant serotype 19A *Streptococcus pneumoniae*—Massachusetts, 2001–2006," *Morbidity and Mortality Weekly Report*, vol. 56, no. 41, pp. 1077–1080, 2007.
- [25] M. R. Jacobs, C. E. Good, S. Bajaksouzian, and A. R. Windau, "Emergence of *Streptococcus pneumoniae* serotypes 19A, 6C, and 22F and serogroup 15 in Cleveland, Ohio, in relation

to introduction of the protein-conjugated pneumococcal vaccine," *Clinical Infectious Diseases*, vol. 47, no. 11, pp. 1388–1395, 2008.

[26] CDC, "Licensure of a 13-valent pneumococcal conjugate vaccine (PCV13) and recommendations for use among children—advisory committee on immunization practices (ACIP), 2010," *Morbidity and Mortality Weekly Report*, vol. 59, no. 9, pp. 258–261, 2010.