

REVIEW

Open Access



How chronic inflammation fuels carcinogenesis as an environmental epimutagen

Yu-Yu Liu¹ , Yui Ohashi¹ and Toshikazu Ushijima^{1*}

*Correspondence:

Toshikazu Ushijima
tushijima142@hoshi.ac.jp
¹Department of Epigenomics,
Institute for Advanced Life Sciences,
Hoshi University, Tokyo
142-8501, Japan

Abstract

Chronic inflammation, induced by environmental and intrinsic factors, frequently leads to carcinogenesis. In inflammation-associated cancers, such as gastric, colon, and cervical cancers, an important role of epigenetic alterations has been implicated. Such epigenetic alterations include aberrant DNA methylation and histone modifications, some of which can permanently alter cellular characteristics and predispose cells to malignant transformation. Even in normal-appearing tissues, high levels of aberrant DNA methylation can be present, with these levels correlating to future cancer risk. The mechanisms of how chronic inflammation induces aberrant DNA methylation involve the repression of DNA methylation erasers, TET enzymes, and the increased activity of DNMTs. Aberrant DNA methylation in normal tissues can serve as a biomarker for cancer risk in the stomach and uterus. Beyond chronic inflammation, factors such as folate, vitamin B₁₂, and vitamin C deficiency can affect DNA methylation through metabolic pathways. Additionally, environmental chemicals, such as arsenic and NNK, and microbial metabolites, have been implicated in inducing epigenetic alterations. Further research is warranted on epigenetic alterations due to environmental factors, and the factors themselves, namely epimutagens.

Keywords Chronic inflammation, Carcinogenicity, DNA methylation, Epimutagen

1 Introduction

Chronic inflammation arises from environmental and intrinsic factors, such as persistent infections (e.g., *Helicobacter pylori* [*H. pylori*], human papilloma virus [HPV], and hepatitis viruses), autoimmune disorders, long-term exposure to environmental toxins (e.g., tobacco smoke), metabolic disorders, tissue damage, and chronic stress. Chronic inflammation is well known to be associated with cancer development, traditionally thought to be driven by inducing mutations [1–3]. However, growing evidence suggests a great impact of epigenetic alterations induced by chronic inflammation on carcinogenesis [4–7]. At a molecular level, chronic inflammation can induce aberrant DNA methylation, particularly genes pre-marked with specific histone modifications, via activation of the NF- κ B pathway and production of nitric oxide [8–10]. Aberrant DNA methylation



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

can disrupt gene regulation without affecting the DNA sequence itself, leading to long-lasting cellular changes [11–13].

The causal role of epigenetic alterations in cancer development has been shown by multiple lines of evidence. First, global DNA hypomethylation promoted tumor formation in mice [14, 15], and suppression of aberrant DNA methylation by DNA demethylating agents repressed tumorigenesis in mouse models [16–19]. Second, tumor suppressor genes mutated in human cancers are also silenced by aberrant promoter DNA methylation during precancerous stages, with this silencing maintained in tumors [20–22]. Third, the therapeutic efficacy of DNA demethylating agents and histone modification inhibitors has been shown in human hematological tumors and multiple animal models of human solid and hematological tumors, providing strong support for the causal role of epigenetic alterations in the development of these tumors [23–29].

Notably, a substantial amount of epigenetic alterations can accumulate in non-cancer cells, enabling cells to bypass normal growth checkpoints and ultimately predisposing them to malignant transformation [30, 31]. For example, while *BRAF* mutations typically induce cellular senescence in normal cells, the accumulation of epigenetic alterations in non-cancer cells can circumvent this process, allowing continued cell proliferation despite the presence of a *BRAF* mutation, ultimately leading to colorectal tumor formation [32]. Additionally, global hypomethylation can also occur even in non-cancer tissues, and can lead to chromosomal instability [15].

This review examines the role of chronic inflammation as an epigenetic mutagen (epimutagen), with a particular focus on stomach, colon, and uterine cervical cancers where chronic inflammation is heavily involved, and discusses potential applications of the findings. We will also explore substances that act as epimutagens and modulators of epigenetic processes.

2 Mutations and epigenetic alterations in cancers

Some cancers are associated with defined mutagens, such as tobacco smoking for squamous cell lung cancers (SCCs) and ultraviolet radiation exposure for melanomas. These cancers often present with frequent driver gene mutations, such as *PIK3CA* mutations in SCCs and *BRAF* mutations in melanomas, along with high mutation burdens [33]. Other cancers, such as gastric and hepatocellular cancers, are associated with chronic inflammation and display less frequent driver gene mutations [34, 35], suggesting the importance of epigenetic alterations in these cancers. For example, although driver genes such as *TP53* and *CDHI* are identifiable in some gastric cancers, a substantial portion lack sufficient numbers of driver gene mutations [36]. These tumors often involve epigenetic alterations, such as the silencing of *CDKN2A*, *CDHI*, and *MLHI* through DNA hypermethylation. This shows that some cancer types are heavily influenced by mutations, while others are more by epigenetic alterations. Furthermore, it suggests that even within the same cancer type, the relative importance of mutations and epigenetic alterations depends on the individual.

3 Mutations and epigenetic alterations in normal-appearing tissues and their association with cancer risk

Both mutations and epigenetic alterations are now known to be present even in normal-appearing tissues, and are likely to play a crucial role in predisposing cells to malignancies. Regarding mutations, their presence in normal-appearing tissues was recently demonstrated in various tissues by the use of small amounts of samples, which increases the relative proportion of a clone, and/or high-accuracy sequencing [37]. In contrast with mutations, aberrant DNA methylation can be present at high levels even in normal-appearing tissues, and its accumulation in tissues exposed to chronic inflammation has been known for more than two decades [38–40]. The high levels of aberrant DNA methylation enabled us to show that its accumulation levels can be correlated with cancer risk [41, 42]. The predisposed status of a normal-appearing tissue due to accumulation of epigenetic alterations is termed as an epigenetic field for cancerization [30].

The relative importance of genetic and epigenetic alterations in cancer risk was estimated by measuring point mutations and aberrant DNA methylation in normal-appearing tissues of healthy individuals (low risk), individuals exposed to risk factors (intermediate risk), and patients with esophageal or gastric cancers (high risk) [43]. In esophageal mucosa, both point mutations and aberrant DNA methylation increased as cancer risk increased. In gastric mucosa, point mutations increased from low to intermediate risk, but not from intermediate to high risk. In contrast, aberrant DNA methylation increased along with cancer risk. This showed that epigenetic alterations in normal-appearing tissues play a more important role than genetic alterations in cancer risk assessment, depending on the cancer type and its etiology (Fig. 1).

4 Molecular mechanisms for induction of epigenetic alterations by chronic inflammation

In the stomach, *H. pylori* infection was strongly associated with high levels of aberrant DNA methylation [39]. In the stomach of Mongolian gerbils, even when *H. pylori* was present in large amounts, suppressing inflammatory responses with cyclosporin A almost completely inhibited methylation induction, highlighting the critical role of

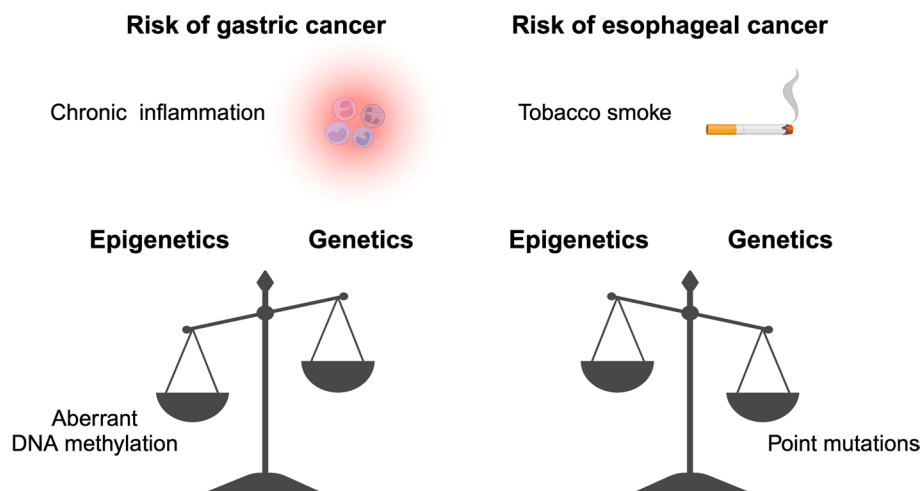


Fig. 1 Balance between genetic and epigenetic alterations in carcinogenesis. Certain types of cancer driven by chronic inflammation primarily involve epigenetic alterations, while other types of cancer driven by mutagens primarily involve mutations.

inflammation in driving epigenetic alterations [10]. In addition, while *H. pylori* infection potently induced severe DNA methylation, dietary salt and alcohol did not [44]. These agents caused repeated acute inflammation, which is characterized by prolonged neutrophil infiltration, while *H. pylori* infection caused chronic inflammation characterized by macrophage and lymphocyte infiltration. The lack of aberrant DNA methylation induction despite sustained cell proliferation in the repeated acute inflammation suggests that signals from macrophages or lymphocytes are required for induction of aberrant DNA methylation.

In *H. pylori* infection-induced chronic inflammation, the infiltration of macrophages stimulates the secretion of pro-inflammatory cytokines such as TNF- α , IL-1 β , and IL-6 [44–47]. TNF- α and IL-1 β activate the NF- κ B pathway, which turned out to be a major driver of inflammation-induced aberrant DNA methylation. NF- κ B translocates to the nucleus upon activation, acting as a transcription factor that induces genes involved in cell survival, proliferation, and inflammation [48]. Importantly, activated NF- κ B upregulates *MIR20A*, *MIR26B*, and *MIR29C* that target ten-eleven translocation (TET) enzymes, and down-regulates *TET* expression [8]. Since TET enzymes mediate DNA demethylation, their down-regulation can contribute to the induction of aberrant DNA methylation.

Another hint was massive production of reactive oxygen and nitrogen species (ROS/RNS) by immune cells to fight pathogens during inflammation [49, 50]. Particularly nitric oxide (NO), produced by nitric oxide synthase 2 (*NOS2*), was reported to enhance the activity of DNA methyltransferases (DNMTs) [51] and to inhibit the activity of TET enzymes [52]. Although the impact of NO only was minimal, a combination of NF- κ B-mediated down-regulation of TET enzymes and NO-induced enhancement of DNMT activity synergistically drove widespread induction of aberrant DNA methylation across the genome [8] (Fig. 2).

Histone modifications are also altered by chronic inflammation, especially by NO. NO-mediated *S*-nitrosylation inhibits activity of histone deacetylases (HDACs), increasing histone or protein acetylation [53, 54]. In addition, NO is reported to inhibit the activity of lysine-specific histone demethylase KDM3A, and potentially other Jumonji-C demethylases, which in turn leads to an increase of KDM3A expression. NO also reduces the iron cofactor of demethylases, and thereby increases global levels of H3K9me2 [55].

5 Clinical applications of epigenetic alterations associated with chronic inflammation

In the stomach, aberrant DNA methylation is accumulated in *H. pylori*-infected gastric mucosa [56–58]. Importantly, the methylation level in gastric mucosa decreases after *H. pylori* eradication [56–58]. Since DNA methylation of a CpG island does not disappear without therapeutic drugs, this phenomenon suggests that the aberrant DNA methylation occurs mainly in differentiated epithelial cells. Most stem cells in gastric mucosa remain unmethylated, and the overall methylation level decreases when differentiated cells with aberrant DNA methylation are replaced by cells newly produced from unaffected stem cells after *H. pylori* eradication. Our recent organoid and single-cell analysis on gastric stem cells and differentiated cells supported this model (Takeuchi et al., manuscript in preparation).

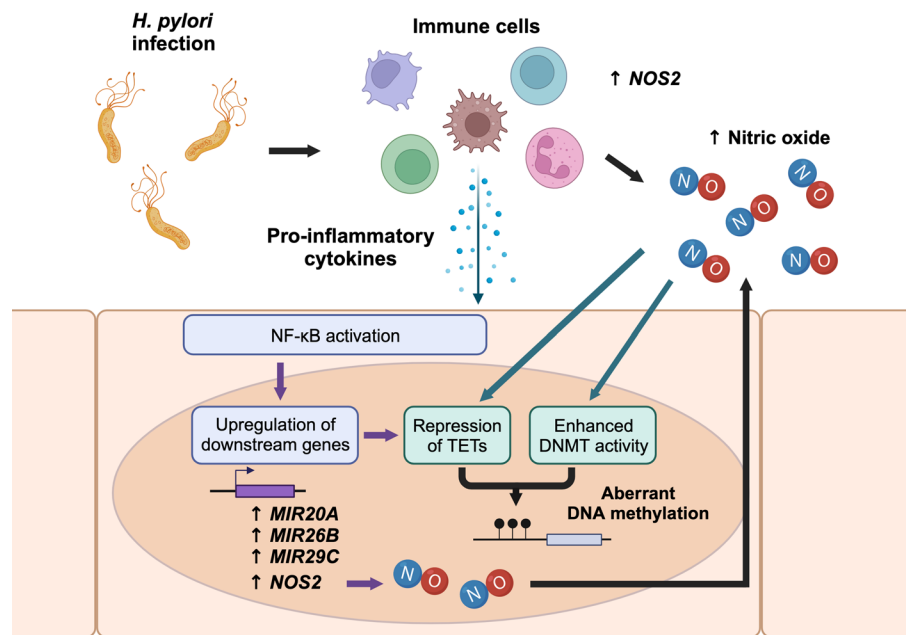


Fig. 2 Induction mechanism of aberrant DNA methylation by chronic inflammation triggered by *H. pylori* infection. Proinflammatory cytokines, such as IL-1 β and TNF- α , activate the NF- κ B pathway, repressing TETs and upregulating downstream genes, including *NOS2*. NO from inflammatory and epithelial cells increases DNMT activity and inhibits TET expression. Simultaneous occurrence of TET repression and increased DNMT activity potentially drive aberrant DNA methylation induction in gastric epithelial cells.

The methylation is widespread and involves numerous genomic loci [59], including promoter CpG island hypermethylation that silences tumor suppressor genes such as *CDKN2A* [58], *RUNX3* [50], *CDH1* (E-cadherin) [60], and *MLH1* [61]. Additionally, global hypomethylation is observed and is considered to contribute to chromosomal instability [15, 62]. These hyper- and hypomethylations in gastric stem cells produce an epigenetic field for cancerization [30] and closely overlap with precancerous conditions such as atrophic gastritis and intestinal metaplasia [22, 63].

The severity of an epigenetic field can be measured by the DNA methylation level of a marker gene, and the methylation level is associated with cancer risk, namely with a pathway to cancer or that to longevity after *H. pylori* eradication [41]. To bring this finding into cancer risk diagnosis, a multi-center prospective study demonstrated that assessing the accumulation of aberrant DNA methylation at the *MIR124-3* gene in the gastric mucosa of gastric cancer patients cured by endoscopic treatment can predict the risk of metachronous gastric cancer, with a multivariate-adjusted hazard ratio of 2.3 [64]. The methylation level of *MIR124-3* reflects the genome-wide burden of epimutations, supporting its use as a surrogate marker for cancer risk [41]. Additionally, another multi-center prospective study demonstrated that measuring DNA methylation levels of *RIMS1* can stratify gastric cancer risk among healthy individuals after *H. pylori* eradication [65] (Fig. 3).

In colonic and intestinal mucosa, aberrant DNA methylation is accumulated in patients with inflammatory bowel diseases (IBD), namely ulcerative colitis (UC) and Crohn's disease (CD). Tumor suppressor genes including *CDKN2A* [66], *CDH1* [67], *RASSF1A* and *p14/ARF* [68] undergo hypermethylation, while global hypomethylation further contributes to genomic instability [15, 69]. These epigenetic alterations occur not

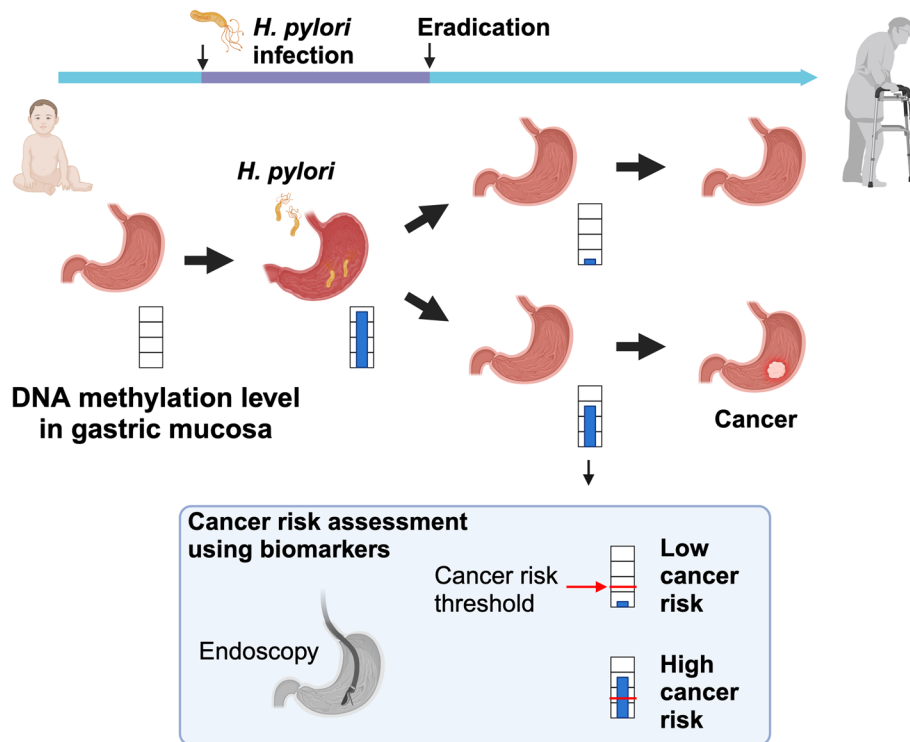


Fig. 3 Aberrant DNA methylation accumulates in the gastric mucosa following *H. pylori* infection during childhood and its eradication in adulthood. *H. pylori* infection potentially induces aberrant DNA methylation of many genes. After eradication, these methylation changes can either decrease or persist. The decrease in methylation levels suggests that differentiated cells are more susceptible to methylation induction, and that the DNA methylation level after *H. pylori* eradication is correlated with epigenomic damage in stem cells and thus with gastric cancer risk (see the text).

only in epithelial cells but also in immune cells. For example, depletion of *UHRF1*, a key component of maintenance of DNA methylation, in mice with UC led to severe pathology due to excessive TNF- α production through its hypomethylation in macrophages [70]. Moreover, the histone methyltransferase G9a was found to mediate H3K9me2 deposition in T cells, which restricted Th17 and Treg differentiation by limiting chromatin accessibility and responsiveness to TGF- β , driving dysregulated immune responses and colitis progression [71].

Promising biomarkers for cancer risk in IBD have been identified, each with unique diagnostic and prognostic utility. For CD, *FHIT* hypermethylation was associated with the presence of CD and its progression [72]. Combined *TGF β 1* and *IL-6* hypermethylation in whole peripheral blood samples showed promise as a non-invasive diagnostic tool for pediatric CD [73]. For UC, *CXCL5* hypermethylation in whole peripheral blood was reported to serve as a less invasive diagnostic marker, helping to distinguish UC from other inflammatory conditions [74]. As a practical and non-invasive approach for early cancer detection, altered methylation levels of *BMP3* and *VAV3* in stool samples, relative to *ZDHHC1*, have also been reported [75]. Lastly, hypermethylation of *ITGA4* and *TFPI2* has been identified as risk markers for colitis-associated cancer, enabling cancer risk assessment and preventive strategies in high-risk individuals [76]. Collectively, these epigenetic biomarkers are likely to contribute to an advancement in the assessment of cancer risks.

In the uterine cervix, persistent infection with high-risk HPV types 16 and 18, along with its resultant chronic inflammation, is a major driver of cervical carcinogenesis. As in the stomach and colon, chronic inflammation due to HPV infection is known to alter cellular DNA methylation patterns [77]. Importantly the accumulation of aberrant DNA methylation in cervical epithelial cells has been associated with an increased risk of cervical cancer [78, 79]. A recent prospective study demonstrated the effectiveness of DNA methylation biomarkers, particularly those targeting HPV-associated and host genes, in cervical cancer screening, highlighting their potential as a cancer risk biomarker [42].

These findings in the stomach, colon, and uterine cervix confirm the broad utility of DNA methylation as a reliable tool for early detection and risk assessment in chronic inflammation-associated cancers.

6 Other environmental factors

Environmental factors such as nutritional deficiencies can significantly impact cancer risk through epigenetic alterations in tissues with or even without chronic inflammation. Folate and vitamin B₁₂ are involved in the synthesis of S-adenosylmethionine (SAM), a universal methyl donor required for DNA methylation. Folate supplementation was found to reduce the expression of pro-inflammatory cytokines and inhibit NF- κ B activation after liposaccharide (LPS) induction [80], showing the potential for correcting inflammation-induced epigenetic alterations. On the other hand, supplementation of vitamin C, a cofactor for oxidative DNA demethylation by TET enzymes, can boost TET activity [81]. Importantly, a clinical study in patients with TET2 germline mutations showed that vitamin C supplementation decreased aberrant DNA methylation in blood cells [82]. These findings emphasize the promising potential of dietary and pharmacological interventions to target epigenetic alterations for therapeutic and preventive purposes.

Other environmental chemicals, such as arsenic and NNK (a tobacco-specific nitrosamine), are also reported as epimutagens that can disrupt normal epigenetic regulation. Chronic and low-level arsenic exposure was associated with DNA hypomethylation and aberrant gene expressions [83]. These alterations are accompanied by reduced DNMT activity, attributed to arsenic's direct binding and its subsequent degradation of DNMT through the ubiquitin-proteasome pathway [84]. Such epigenetic alterations are reported to activate oncogenes and drive carcinogenesis in tissues, including the skin, bladder and lungs [85]. Similarly, NNK was reported to induce DNMT1 nuclear localization and hypermethylation of key genes involved in cell cycle regulation and DNA repair, thus further driving cancer development in smokers [86, 87]. These findings highlight the profound role of environmental factors in driving epigenetic alterations, emphasizing the need to understand their broader implications in carcinogenesis and potential strategies to mitigate their effects.

The microbiome is also an environmental factor influencing host cells, and plays a crucial role in shaping the epigenome and directing health and disease (Fig. 4), a topic that has gained significant attention in recent studies [88]. Its composition is profoundly shaped by environmental factors, mainly dietary components. In a healthy state, the gut microbiome produces short-chain fatty acids (SCFAs) such as acetate, propionate, and butyrate, which regulate host cell energy metabolism by serving as substrates for acetyl-CoA production and driving the tricarboxylic acid (TCA) cycle, which prevents

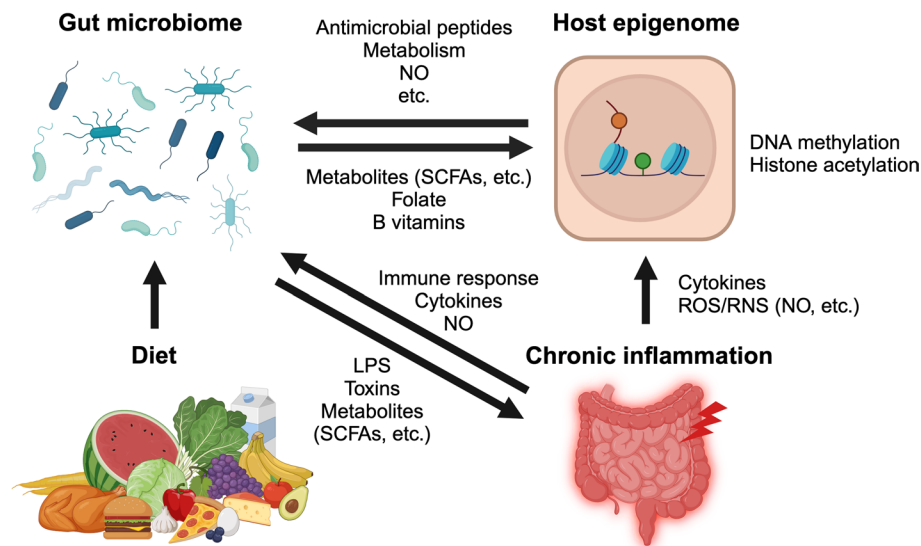


Fig. 4 The microbiome influences the host epigenomes through its metabolites and metabolism of vitamin B₁₂ and folate. The gene expression changes, in turn, affect the microbiome composition through secreted factors such as NO. Additionally, chronic inflammation can further alter the gut microbiome composition and host epigenomes.

autophagy [89]. SCFAs also inhibit HDACs [90], modulate gene expression, supporting barrier function and reducing immune responses in the colon [89, 91]. Additionally, microbial metabolism of vitamin B₁₂ and folate affects DNA methylation in host cells [90]. In contrast, under disease conditions, chronic inflammation significantly impacts microbial diversity and functionality, partly through NO overproduction [92], leading to widespread epigenetic alterations in host cells as discussed in previous sections. These mechanisms highlight the complex crosstalk between diet, the microbiome, and the host epigenome in healthy and diseased states, which can either enhance homeostasis or contribute to disease.

7 Outlook

Chronic inflammation, induced by environmental and intrinsic factors, clearly drives carcinogenesis by promoting epigenetic alterations, acting as an epimutagen. In addition, folate, vitamin B₁₂, and vitamin C are modulators of epigenetic processes with mechanistic support. Other chemicals, however, require more robust analysis to understand their underlying mechanisms. The interplay between the gut microbiome and epigenetic alterations is a growing field, with potential therapeutic implications for modulating microbiota to prevent or reverse harmful epigenetic alterations in inflammatory conditions. However, the extreme complexity of this field calls for further research. Moreover, factors such as age and genetic background are likely to influence epigenetic alterations induced by chronic inflammation and chemical substances, warranting more refined and larger scale studies.

Application of epigenetic alterations in normal tissues to cancer risk diagnosis is now approaching clinical use in the uterus and stomach [42, 64]. In other tissues, such as the liver and colon, similar approaches are likely to be successful [93, 94]. For therapeutic purposes, aberrant DNA methylation in tumors is now used as a target [95, 96]. To advance cancer prevention, early detection, and risk stratification, further investigation

is needed to clarify how specific environmental exposures drive epigenetic alterations in normal tissues.

8 Conclusions

Chronic inflammation driven by environmental factors such as infection plays a critical role in carcinogenesis, and induction of epigenetic alterations is the major mechanism involved. Such alterations are present even in normal tissues and are involved in field cancerization in certain cancer types. Emerging biomarkers derived from inflammation-induced aberrant DNA methylation offer promising avenues for cancer risk diagnosis. Targeting these epigenetic alterations, alongside preventive strategies such as *H. pylori* eradication or vitamin supplementation, may help reduce cancer risks associated with chronic inflammation. Advancing this field will require further understanding of environmental and biological factors driving epigenetic alterations.

Acknowledgements

The figures were created using Biorender, and the original source can be accessed at: <https://BioRender.com/h12m159>.

Author contributions

YY Liu, Y Ohashi, and T Ushijima wrote the manuscript and created the figures. All authors gave final approval to the submitted and published versions.

Funding

This work was supported by a Grant-in-Aid for Scientific Research from JSPS under Grant Number JP20K16347 (YY. L.) and AMED-CREST under Grant Number JP20gm1310006 (T. U.).

Data availability

No datasets were generated or analysed during the current study.

Declarations

Competing interests

We declare a conflict of interest with Sysmex company, which is developing an epigenetic gastric cancer risk diagnosis. We declare a conflict of interest with Sysmex company, which is developing an epigenetic gastric cancer risk diagnosis.

Received: 2 January 2025 / Accepted: 10 June 2025

Published online: 18 June 2025

References

1. Waldum H, Fossmark R. Inflammation and digestive Cancer. *Int J Mol Sci.* 2023;24(17):13503.
2. Liu W, et al. Cancer Evo-Dev: a theory of inflammation-induced oncogenesis. *Front Immunol.* 2021;12:768098.
3. Reuter S, et al. Oxidative stress, inflammation, and cancer: how are they linked? *Free Radic Biol Med.* 2010;49(11):1603–16.
4. Rokavec M, Oner MG, Hermeking H. Inflammation-induced epigenetic switches in cancer. *Cell Mol Life Sci.* 2016;73(1):23–39.
5. Maiuri AR, O'Hagan HM. Interplay between inflammation and epigenetic changes in cancer. *Prog Mol Biol Transl Sci.* 2016;144:69–117.
6. Yang ZH, Dang YQ, Ji G. Role of epigenetics in transformation of inflammation into colorectal cancer. *World J Gastroenterol.* 2019;25(23):2863–77.
7. Padmanabhan N, Ushijima T, Tan P. How to stomach an epigenetic insult: the gastric cancer epigenome. *Nat Rev Gastroenterol Hepatol.* 2017;14(8):467–78.
8. Takeshima H, et al. TET repression and increased DNMT activity synergistically induce aberrant DNA methylation. *J Clin Invest.* 2020;130(10):5370–9.
9. Abu-Remaileh M, et al. Chronic inflammation induces a novel epigenetic program that is conserved in intestinal adenomas and in colorectal cancer. *Cancer Res.* 2015;75(10):2120–30.
10. Niwa T, et al. Inflammatory processes triggered by *Helicobacter pylori* infection cause aberrant DNA methylation in gastric epithelial cells. *Cancer Res.* 2010;70(4):1430–40.
11. Sherif ZA, Ogunwobi OO, Ransom HW. Mechanisms and technologies in cancer epigenetics. *Front Oncol.* 2024;14:1513654.
12. Issa JP, Kantarjian HM. Targeting DNA methylation. *Clin Cancer Res.* 2009;15(12):3938–46.
13. Jones PA, Taylor SM. Cellular differentiation, cytidine analogs and DNA methylation. *Cell.* 1980;20(1):85–93.
14. Gaudet F, et al. Induction of tumors in mice by genomic hypomethylation. *Science.* 2003;300(5618):489–92.
15. Chen RZ, et al. DNA hypomethylation leads to elevated mutation rates. *Nature.* 1998;395(6697):89–93.
16. Lantry LE, et al. 5-Aza-2'-deoxycytidine is chemopreventive in a 4-(methyl-nitrosamino)-1-(3-pyridyl)-1-butanone-induced primary mouse lung tumor model. *Carcinogenesis.* 1999;20(2):343–6.

17. Hagiwara H, et al. 5-Aza-2'-deoxycytidine suppresses human renal carcinoma cell growth in a xenograft model via up-regulation of the connexin 32 gene. *Br J Pharmacol*. 2008;153(7):1373–81.
18. Hsieh HH, et al. Epigenetic modifications as novel therapeutic strategies of Cancer chemoprevention by phytochemicals. *Pharm Res*. 2025;42(1):69–78.
19. McCabe MT, et al. Inhibition of DNA methyltransferase activity prevents tumorigenesis in a mouse model of prostate cancer. *Cancer Res*. 2006;66(1):385–92.
20. Kanai Y, Hirohashi S. Alterations of DNA methylation associated with abnormalities of DNA methyltransferases in human cancers during transition from a precancerous to a malignant state. *Carcinogenesis*. 2007;28(12):2434–42.
21. Gao W, et al. Variable DNA methylation patterns associated with progression of disease in hepatocellular carcinomas. *Carcinogenesis*. 2008;29(10):1901–10.
22. Takeuchi C, et al. Precancerous nature of intestinal metaplasia with increased chance of conversion and accelerated DNA methylation. *Gut*. 2024;73(2):255–67.
23. Yu X, et al. Cancer epigenetics: from laboratory studies and clinical trials to precision medicine. *Cell Death Discov*. 2024;10(1):28.
24. Perner F, et al. Therapeutic targeting of chromatin alterations in leukemia and solid tumors. *Int J Cancer*. 2025;1–27.
25. Geissler F, et al. The role of aberrant DNA methylation in cancer initiation and clinical impacts. *Ther Adv Med Oncol*. 2024;16:17588359231220511.
26. Chovanec M, et al. Incorporating DNA methyltransferase inhibitors (DNMTis) in the treatment of genitourinary malignancies: A systematic review. *Target Oncol*. 2018;13(1):49–60.
27. Shen N, et al. Inactivation of receptor tyrosine kinases reverts aberrant DNA methylation in acute myeloid leukemia. *Clin Cancer Res*. 2017;23(20):6254–66.
28. Huisman C, et al. Re-expression of selected epigenetically silenced candidate tumor suppressor genes in cervical Cancer by TET2-directed demethylation. *Mol Ther*. 2016;24(3):536–47.
29. Song Y, Zhang C. Hydralazine inhibits human cervical cancer cell growth in vitro in association with APC demethylation and re-expression. *Cancer Chemother Pharmacol*. 2009;63(4):605–13.
30. Ushijima T. Epigenetic field for cancerization. *J Biochem Mol Biol*. 2007;40(2):142–50.
31. Besselink N, et al. The genome-wide mutational consequences of DNA hypomethylation. *Sci Rep*. 2023;13(1):6874.
32. Ushijima T, Suzuki H. The origin of CIMP, at last. *Cancer Cell*. 2019;35(2):165–7.
33. Vogelstein B, et al. Cancer Genome Landscapes. *Sci*. 2013;339(6127):1546–58.
34. Consortium IT. Pan-cancer analysis of whole genomes. *Nature*. 2020;578(7793):82–93.
35. Sinkala M. Mutational landscape of cancer-driver genes across human cancers. *Sci Rep*. 2023;13(1):12742.
36. Liu Y, et al. Comparative molecular analysis of Gastrointestinal adenocarcinomas. *Cancer Cell*. 2018;33(4):721–e7358.
37. Kakiuchi N, Ogawa S. Clonal expansion in non-cancer tissues. *Nat Rev Cancer*. 2021;21(4):239–56.
38. García-Manero G, et al. Aberrant DNA methylation in pediatric patients with acute lymphocytic leukemia. *Cancer*. 2003;97(3):695–702.
39. Maekita T, et al. High levels of aberrant DNA methylation in *Helicobacter pylori*-infected gastric mucosae and its possible association with gastric cancer risk. *Clin Cancer Res*. 2006;12(3 Pt 1):989–95.
40. Zhang N, et al. Insights into the role of nucleotide methylation in metabolic-associated fatty liver disease. *Front Immunol*. 2023;14:1148722.
41. Irie T, et al. The methylation level of a single cancer risk marker gene reflects methylation burden in gastric mucosa. *Gastric Cancer*. 2023;26(5):667–76.
42. Schreiberhuber L, et al. Cervical cancer screening using DNA methylation triage in a real-world population. *Nat Med*. 2024;30(8):2251–7.
43. Yamashita S, et al. Genetic and epigenetic alterations in normal tissues have differential impacts on cancer risk among tissues. *Proc Natl Acad Sci U S A*. 2018;115(6):1328–33.
44. Hur K, et al. Insufficient role of cell proliferation in aberrant DNA methylation induction and involvement of specific types of inflammation. *Carcinogenesis*. 2011;32(1):35–41.
45. Basso D, et al. *Helicobacter pylori* infection enhances mucosal interleukin-1 beta, interleukin-6, and the soluble receptor of interleukin-2. *Int J Clin Lab Res*. 1996;26(3):207–10.
46. Fan XG, et al. Increased gastric production of interleukin-8 and tumour necrosis factor in patients with *Helicobacter pylori* infection. *J Clin Pathol*. 1995;48(2):133–6.
47. Yamaoka Y, et al. Induction of various cytokines and development of severe mucosal inflammation by CagA gene positive *Helicobacter pylori* strains. *Gut*. 1997;41(4):442–51.
48. Wu Y, Zhou BP. TNF-alpha/NF-kappaB/Snail pathway in cancer cell migration and invasion. *Br J Cancer*. 2010;102(4):639–44.
49. Kay J, et al. Inflammation-induced DNA damage, mutations and cancer. *DNA Repair (Amst)*. 2019;83:102673.
50. Katayama Y, Takahashi M, Kuwayama H. *Helicobacter pylori* causes runx3 gene methylation and its loss of expression in gastric epithelial cells, which is mediated by nitric oxide produced by macrophages. *Biochem Biophys Res Commun*. 2009;388(3):496–500.
51. Hmadcha A, et al. Methylation-dependent gene Silencing induced by Interleukin 1beta via nitric oxide production. *J Exp Med*. 1999;190(11):1595–604.
52. Thomas D, et al. Nitric oxide inhibits ten-eleven translocation DNA demethylases to regulate 5mC and 5hmC across the genome. *Res Sq*. 2024.
53. Okuda K, Ito A, Uehara T. Regulation of histone deacetylase 6 activity via S-Nitrosylation. *Biol Pharm Bull*. 2015;38(9):1434–7.
54. Colussi C, et al. HDAC2 Blockade by nitric oxide and histone deacetylase inhibitors reveals a common target in Duchenne muscular dystrophy treatment. *Proc Natl Acad Sci U S A*. 2008;105(49):19183–7.
55. Hickok JR, et al. Nitric oxide modifies global histone methylation by inhibiting Jumonji C domain-containing demethylases. *J Biol Chem*. 2013;288(22):16004–15.
56. Nakajima T, et al. Persistence of a component of DNA methylation in gastric mucosae after *Helicobacter pylori* eradication. *J Gastroenterol*. 2010;45(1):37–44.
57. Ushijima T, Nakajima T, Maekita T. DNA methylation as a marker for the past and future. *J Gastroenterol*. 2006;41(5):401–7.

58. Perri F, et al. Aberrant DNA methylation in non-neoplastic gastric mucosa of H. Pylori infected patients and effect of eradication. *Am J Gastroenterol*. 2007;102(7):1361–71.
59. Yamashita S, et al. Distinct DNA methylation targets by aging and chronic inflammation: a pilot study using gastric mucosa infected with *Helicobacter pylori*. *Clin Epigenetics*. 2019;11(1):191.
60. Huang FY, et al. *Helicobacter pylori* induces promoter methylation of E-cadherin via interleukin-1beta activation of nitric oxide production in gastric cancer cells. *Cancer*. 2012;118(20):4969–80.
61. Alvarez MC, et al. MGMT and MLH1 methylation in *Helicobacter pylori*-infected children and adults. *World J Gastroenterol*. 2013;19(20):3043–51.
62. Ushijima T, Clark SJ, Tan P. Mapping genomic and epigenomic evolution in cancer ecosystems. *Science*. 2021;373(6562):1474–9.
63. Correa P, Houghton J. Carcinogenesis of *Helicobacter pylori*. *Gastroenterology*. 2007;133(2):659–72.
64. Asada K, et al. Demonstration of the usefulness of epigenetic cancer risk prediction by a multicentre prospective cohort study. *Gut*. 2015;64(3):388–96.
65. Yamada H et al. Precision risk stratification of primary gastric cancer after eradication of H. pylori by a DNA methylation marker: a multicentre prospective study. *Gut*. 2025. <https://doi.org/10.1136/gutjnl-2025-335039>.
66. Hsieh CJ, et al. Hypermethylation of the p16INK4a promoter in colectomy specimens of patients with long-standing and extensive ulcerative colitis. *Cancer Res*. 1998;58(17):3942–5.
67. Azarschab P, et al. Epigenetic control of the E-cadherin gene (CDH1) by CpG methylation in colectomy samples of patients with ulcerative colitis. *Genes Chromosomes Cancer*. 2002;35(2):121–6.
68. Salama RH, et al. Interrelations of apoptotic and cellular senescence genes methylation in inflammatory bowel disease subtypes and colorectal carcinoma in Egyptians patients. *Appl Biochem Biotechnol*. 2019;189(1):330–43.
69. Gloria L, et al. DNA hypomethylation and proliferative activity are increased in the rectal mucosa of patients with long-standing ulcerative colitis. *Cancer*. 1996;78(11):2300–6.
70. Qi S, et al. Uhrf1-Mediated Tnf-alpha gene methylation controls Proinflammatory macrophages in experimental colitis resembling inflammatory bowel disease. *J Immunol*. 2019;203(11):3045–53.
71. Antignano F, et al. Methyltransferase G9A regulates T cell differentiation during murine intestinal inflammation. *J Clin Invest*. 2014;124(5):1945–55.
72. Kim TO, et al. Genome-wide analysis of the DNA methylation profile identifies the fragile histidine triad (FHIT) gene as a new promising biomarker of Crohn's disease. *J Clin Med*. 2020;9(5):1338.
73. Samarani S, et al. CpG methylation in TGFbeta1 and IL-6 genes as surrogate biomarkers for diagnosis of IBD in children. *Inflamm Bowel Dis*. 2020;26(10):1572–8.
74. Karatzas PS, et al. DNA methylation profile of genes involved in inflammation and autoimmunity in inflammatory bowel disease. *Med (Baltim)*. 2014;93(28):e309.
75. Kisiel JB, et al. Analysis of DNA methylation at specific loci in stool samples detects colorectal Cancer and High-Grade dysplasia in patients with inflammatory bowel disease. *Clin Gastroenterol Hepatol*. 2019;17(5):914–21. e5.
76. Gerecke C, et al. Hypermethylation of ITGA4, TFPI2 and VIMENTIN promoters is increased in inflamed colon tissue: putative risk markers for colitis-associated cancer. *J Cancer Res Clin Oncol*. 2015;141(12):2097–107.
77. Jimenez-Wences H, Peralta-Zaragoza O, Fernandez-Tilapa G. Human papilloma virus, DNA methylation and MicroRNA expression in cervical cancer (Review). *Oncol Rep*. 2014;31(6):2467–76.
78. Virmani AK, et al. Aberrant methylation during cervical carcinogenesis. *Clin Cancer Res*. 2001;7(3):584–9.
79. Widschwendter A, et al. Analysis of aberrant DNA methylation and human papillomavirus DNA in cervicovaginal specimens to detect invasive cervical cancer and its precursors. *Clin Cancer Res*. 2004;10(10):3396–400.
80. Cianciulli A, et al. Folic acid is able to polarize the inflammatory response in LPS activated microglia by regulating multiple signaling pathways. *Mediators Inflamm*. 2016;2016:p5240127.
81. Yue X, Rao A. TET family dioxygenases and the TET activator vitamin C in immune responses and cancer. *Blood*. 2020;136(12):1394–401.
82. Taira A, et al. Vitamin C boosts DNA demethylation in TET2 germline mutation carriers. *Clin Epigenetics*. 2023;15(1):7.
83. Zhao CQ, et al. Association of arsenic-induced malignant transformation with DNA hypomethylation and aberrant gene expression. *Proc Natl Acad Sci U S A*. 1997;94(20):10907–12.
84. Yan N, et al. Developmental arsenic exposure impairs cognition, directly targets DNMT3A, and reduces DNA methylation. *EMBO Rep*. 2022;23(6):e54147.
85. Reichard JF, Puga A. Effects of arsenic exposure on DNA methylation and epigenetic gene regulation. *Epigenomics*. 2010;2(1):87–104.
86. Lin RK, et al. The tobacco-specific carcinogen NNK induces DNA methyltransferase 1 accumulation and tumor suppressor gene hypermethylation in mice and lung cancer patients. *J Clin Invest*. 2010;120(2):521–32.
87. Damiani LA, et al. Carcinogen-induced gene promoter hypermethylation is mediated by DNMT1 and causal for transformation of immortalized bronchial epithelial cells. *Cancer Res*. 2008;68(21):9005–14.
88. Pepke ML, Hansen SB, Limborg MT. Unraveling host regulation of gut microbiota through the epigenome–microbiome axis. *Trends Microbiol*. 2024;32(12):1229–40.
89. Donohoe DR, et al. The Microbiome and butyrate regulate energy metabolism and autophagy in the mammalian colon. *Cell Metab*. 2011;13(5):517–26.
90. Woo V, Alenghat T. Epigenetic regulation by gut microbiota. *Gut Microbes*. 2022;14(1):2022407.
91. Kim M, et al. Microbial metabolites, short-chain fatty acids, restrain tissue bacterial load, chronic inflammation, and associated cancer in the colon of mice. *Eur J Immunol*. 2018;48(7):1235–47.
92. Leclerc M, et al. Nitric oxide impacts human gut microbiota diversity and functionalities. *mSystems*. 2021;6(5):e0055821.
93. Fu S, Debes JD, Boonstra A. DNA methylation markers in the detection of hepatocellular carcinoma. *Eur J Cancer*. 2023;191:112960.
94. Fatemi N, et al. DNA methylation biomarkers in colorectal cancer: clinical applications for precision medicine. *Int J Cancer*. 2022;151(12):2068–81.
95. Hattori N, et al. Combination of a synthetic retinoid and a DNA demethylating agent induced differentiation of neuroblastoma through retinoic acid signal reprogramming. *Br J Cancer*. 2021;125(12):1647–56.

96. Kurahashi Y, et al. Dual targeting of aberrant DNA and histone methylation synergistically suppresses tumor cell growth in ATL. *Blood Adv.* 2023;7(8):1545–59.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.