

Microinflammation in the pathogenesis of diabetic nephropathy

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ABSTRACT

Diabetic nephropathy is the leading cause of end-stage renal failure in developed countries. Furthermore, diabetic nephropathy is related to the risk of cardiovascular diseases and an increase in mortality of diabetic patients. Several factors are involved in the development of nephropathy, including glomerular hyperfiltration, oxidative stress, accumulation of advanced glycation end-products, activation of protein kinase C, acceleration of the polyol pathway and over-expression of transforming growth factor- β . Recently, accumulated data have emphasized the critical roles of chronic low-grade inflammation, 'microinflammation', in the pathogenesis of diabetic nephropathy, suggesting that microinflammation is a common mechanism in the development of diabetic vascular complications. Expression of cell adhesion molecules, chemokines and pro-inflammatory cytokines are increased in the renal tissues of diabetic patients and animals. Deficiency of pro-inflammatory molecules results in amelioration of renal injuries after induction of diabetes in mice. Plasma and urinary levels of cytokines, chemokines and cell adhesion molecules, are elevated and correlated with albuminuria. Several kinds of drugs that have anti-inflammatory actions as their pleiotropic effects showed renoprotective effects on diabetic animals. Modulation of the inflammatory process prevents renal insufficiency in diabetic animal models, suggesting that microinflammation is one of the promising therapeutic targets for diabetic nephropathy, as well as for cardiovascular diseases. (*J Diabetes Invest*, doi: 10.1111/jdi.12050, 2013)

KEY WORDS: Diabetic nephropathy, Inflammation, Microinflammation

INTRODUCTION

Diabetic nephropathy (DN) is the leading cause of end-stage renal failure in developed countries. Moreover, DN is related to the risk of cardiovascular diseases and an increase in mortality of diabetic patients. However, the established therapeutic strategies based on strict control of blood glucose level and blood pressure, and blockade of the renin-angiotensin system cannot prevent the progression of DN completely.

Several factors are involved in the development of DN, including genetic factors, glomerular hyperfiltration¹, oxidative stress², accumulation of advanced glycation end-products (AGEs)³, acceleration of the polyol pathway, activation of protein kinase C⁴, overexpression of transforming growth factor- β (TGF β), followed by increase of extracellular matrices⁵. Recently, accumulated data have emphasized the critical role of the inflammatory process in the pathogenesis of DN. Many kinds of pro-inflammatory molecules, including adhesion molecules, chemokines and cytokines, have been known to play roles in the development of diabetic nephropathy⁶. These pro-inflammatory molecules might be new therapeutic targets for DN, as well as for other inflammatory diseases.

The present review will focus on the role of microinflammation in the pathogenesis of DN as a common pathway of development of diabetic vascular complications.

MICROINFLAMMATION

It is well known that the inflammatory process is involved in the pathogenesis of atherosclerosis⁷. Activated macrophages play critical roles for the migration and proliferation of smooth muscle cells in the intima, and the rupture of plaque resulting in an acute coronary event. Inflammatory cells mainly composed of macrophages are also seen in the glomeruli and interstitium of patients with DN, suggesting that the inflammatory process is also involved in the development of DN^{8,9}. Inflammation is characterized by infiltration of inflammatory cells, increased expression of adhesion molecules, chemokines and pro-inflammatory cytokines, and elevation of serum C-reactive protein (CRP) level. These features are also seen in DN and atherosclerosis although they are quite mild as compared with classic inflammatory diseases, such as rheumatoid arthritis. Therefore, the low-grade inflammation that occurs in atherosclerosis and DN is termed 'microinflammation' to distinguish it from classic inflammation.

ADHESION MOLECULES

Infiltration of leukocytes into inflammatory lesions is mediated by adhesion to endothelial cells and transmigration from vascular lumen to inflammatory sites. Adhesion molecules are

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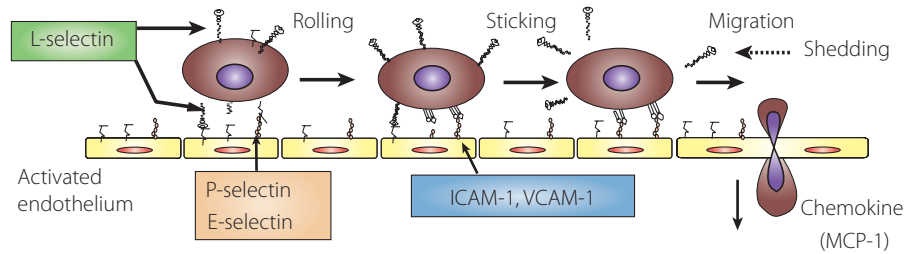


Figure 1 | Mechanism of macrophage infiltration into the inflammatory lesion. Leukocyte infiltration is mediated by the adhesion molecules and chemokines. Selectin molecules mediate the leukocyte rolling on the vascular endothelial cells. Intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1) promote firm attachment of leukocytes and endothelial cells. Monocyte chemoattractant protein-1 (MCP-1) induces the migration of leukocytes from vascular lumen into subendothelium.

expressed on the cell surface, and mediate cell–cell binding and cell–matrix attachment. Leukocyte adhesion to vascular endothelial cells is promoted by adhesion molecules expressed on leukocytes and endothelial cells (Figure 1). Selectin molecules mediate the leukocyte rolling along with endothelial cells at the first step of leukocyte infiltration into inflammatory lesions. At the second step, tight adhesion of leukocytes to the endothelium is mediated by intercellular adhesion molecule-1 (ICAM-1) and vascular cell adhesion molecule-1 (VCAM-1)¹⁰.

ICAM-1 AND VCAM-1

ICAM-1 is an adhesion molecule of the immunoglobulin-superfamily and binds to $\beta 2$ integrins, such as lymphocyte function-associated antigen-1 (LFA-1) and macrophage-1 antigen (Mac-1). There are several studies that have shown the increased expression of adhesion molecules in patients with diabetic nephropathy. Upregulation of ICAM-1 occurs in response to several kinds of stimuli, including pro-inflammatory cytokines^{10,11}, shear stress¹², oxidative stress, protein kinase C activation and AGEs¹³.

We have shown that ICAM-1 is upregulated in the glomeruli and interstitium of diabetic kidney¹⁴. Increased expression of ICAM-1 has been shown in several models of DN¹⁵ (Figure 2). Furthermore, we showed that the blockade of macrophage infiltration using anti-ICAM-1 antibody ameliorated renal injury

and infiltration of macrophage in the glomeruli in streptozotocin-induced diabetic rats¹⁴. Furthermore, urinary albumin excretion (UAE), renal tissue injuries and inflammation are prevented in ICAM-1 knockout (KO) mice after induction of diabetes by streptozotocin¹⁶. Interestingly, UAE was not changed between ICAM-1 KO mice and wild-type mice at 4 weeks after induction of diabetes, but significantly decreased in ICAM-1 KO mice rather than in wild-type mice at 12 and 24 weeks. Similar findings are noted in ICAM-1 deficient *db/db* mice¹⁷. Plasma levels of ICAM-1 are increased in patients with DN¹⁸. Interestingly, Lin *et al.*¹⁹ reported that an elevated baseline plasma level of ICAM-1 is associated with an increasing rate of UAE and the onset of microalbuminuria in the patients with type 1 diabetes who participated in the Diabetes Control and Complications Trial. These findings suggest that the inflammatory axis of ICAM-1 activation to macrophage infiltration plays a pivotal role in the development of diabetic nephropathy (Figure 3).

VCAM-1 is also expressed on endothelial cells, and promotes the adhesion between leukocytes and endothelial cells. VCAM-1 is shown to be increased on endothelial cells and infiltrating cells in the renal interstitium in the diabetic animal model²⁰. Circulating VCAM-1 level is increased and is correlated with albuminuria in patients with type 2 diabetes²¹. In addition, it has been shown that high plasma concentrations of soluble VCAM-1 is a risk factor for death²².

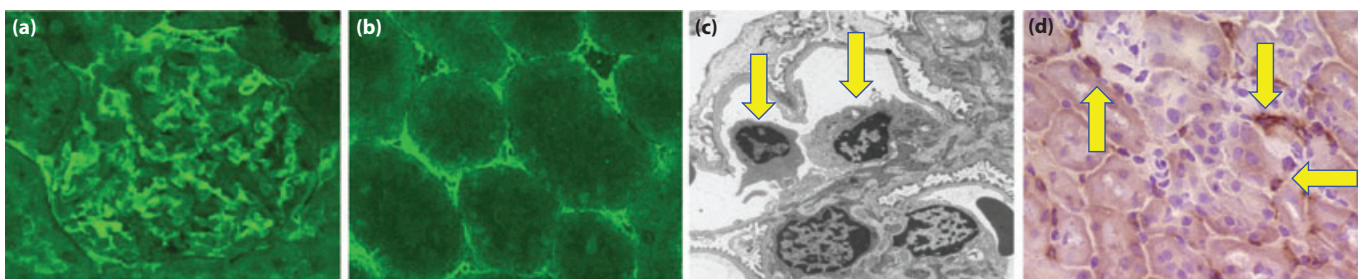


Figure 2 | Expression of intercellular adhesion molecule-1 (ICAM-1) in the (a) glomerulus and (b) interstitium. ICAM-1 is expressed along endothelial cells in the glomerulus and interstitium. Macrophages are infiltrated in the (c) glomerulus and (d) interstitium.

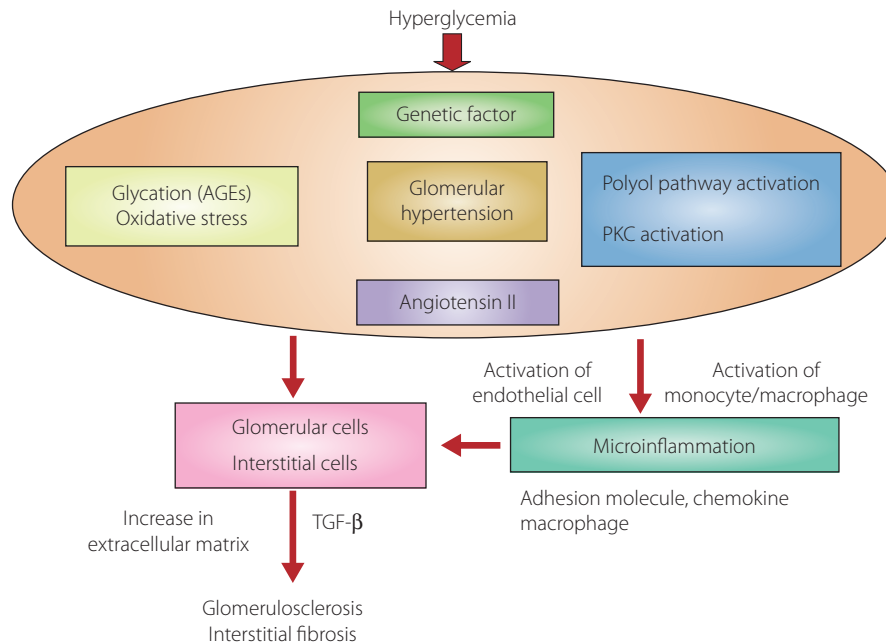


Figure 3 | Pathogenesis of diabetic nephropathy. AGEs, advanced glycation end-products; PKC, protein kinase C; TGF- β , transforming growth factor- β .

Selectins and Selectin Ligands

The selectin family is composed of L-, E- and P-selectin, which promote leukocyte rolling along with vascular endothelial cells in the inflammatory sites²³. E-selectin is expressed on activated endothelial cells and mediates leukocyte rolling on endothelial cells. Expression of E-selectin is induced by pro-inflammatory cytokines, such as interleukin-1 (IL-1) and tumor necrosis factor- α (TNF- α). Expression of E-selectin is upregulated in the peritubular capillaries and is correlated with the number of infiltrating macrophages in the interstitium of patients with diabetic nephropathy²⁴. It was also reported that plasma levels of E-selectin are positively correlated with albuminuria and cardiovascular disease in patients with type 1 diabetes²⁵. L-selectin is constitutively expressed on leukocytes, and interacts with its ligands distributed on endothelial cells. We previously reported that sulfatide is a major L-selectin-binding molecule in the kidney, and that the interaction between L-selectin and sulfatide plays a critical role in monocyte infiltration into the kidney interstitium; however, it is unknown whether this binding pathway is involved in pathogenesis of DN²⁶.

Macrophage Scavenger Receptor-A

Macrophage scavenger receptor-A (SR-A) is a multifunctional receptor expressed on macrophages. A number of studies have established the important roles of SR-A in the pathogenesis of atherosclerosis. SR-A is involved in foam cell formation, activation of macrophages, and adhesion of macrophages to atherosclerotic lesions^{27,28}. We induced diabetes in SR-A KO and wild-type mice by streptozotocin and found that UAE, and

renal tissue injuries, were markedly diminished in diabetic SR-A KO mice²⁹. Interestingly, macrophage infiltration and gene expression of pro-inflammatory molecules in the kidneys was dramatically decreased in diabetic SR-A KO mice compared with diabetic wild-type mice. Furthermore, anti-SR-A antibody blocked the attachment of monocytes to type IV collagen substratum, but not to endothelial cells, showing that SR-A promotes macrophage migration into diabetic kidneys by enhancement of the attachment to renal extracellular matrices.

CHEMOKINES

Interaction of chemokines and their receptors promotes the recruitment of inflammatory cells into inflammatory sites. Recent studies showed that several kinds of chemokines, including C-C motif chemokine 2 (CCL2, monocyte chemoattractant protein-1), C-X3-C motif chemokine 1 (CX3CL1, fractalkine) and C-C motif chemokine 5 (CCL5, RANTES), play important roles in the pathogenesis of DN³⁰.

CCL2 plays a key role in the migration of monocytes into the kidney³¹. Increased expression of CCL2 occurs in the tubulointerstitial lesions of patients with diabetic nephropathy³². Urinary excretion of CCL2 is increased in type 2 diabetic patients, and the urinary level of CCL2 correlates with the clinical stage of DN³². CCL2 is produced by renal resident cells, as well as from inflammatory cells³³. It has been reported that high glucose concentrations, AGEs, protein kinase C, oxidative stress and angiotensin II might contribute to the upregulation of CCL2 in the diabetic kidney³⁴. CCL2 deficiency reduced renal macrophage infiltration and the progression of diabetic

renal injury in a streptozotocin-induced diabetes model³⁵. In addition, C-C chemokine receptor type 2 (CCR2) KO mice also showed a reduction of macrophage infiltration and fibrosis in the kidney in diabetic model³⁶.

CX3CL1 also promotes the migration of mononuclear cells and also induces adhesion between cells that express its receptor, CX3CR1³⁷. CX3CL1 and CX3CR1 are upregulated in the kidneys of diabetic animals³⁸. High glucose levels, AGEs and cytokine activation have been reported to upregulate CX3CR1 in diabetic kidneys³⁹. Interaction between CX3CL1 and CX3CR1 might enhance the infiltration of monocytes into the interstitium and interstitial injury.

CCL5 (RANTES) is expressed in mesangial cells and tubular epithelial cells⁴⁰. CCL5 is also upregulated by angiotensin II, and pro-inflammatory cytokine. Expression of CCL5 is increased in the interstitium of the kidney in patients with DN⁴¹.

PRO-INFLAMMATORY CYTOKINES

Pro-inflammatory cytokine are increased in inflammatory lesions, and contribute to accelerating and maintaining chronic inflammation in various kinds of inflammatory diseases. These pro-inflammatory cytokines, such as TNF- α and IL-1, have been reported to participate in the pathogenesis of DN⁴².

TNF- α induces expression of a variety of effector molecules, such as cytokines and adhesion molecules, apoptosis and necrosis, through the receptors. TNF- α is considered to play a pivotal role in the pathogenesis of diabetic nephropathy through cytotoxicity, apoptosis, necrosis and increased endothelial cell permeability⁴³⁻⁴⁷. Expression of TNF- α is increased in glomerular and proximal tubular epithelial cells, and is correlated with UAE⁴⁸⁻⁵¹. Serum and urinary concentrations of TNF- α are elevated and related to disease progression in patients with DN⁵². We also reported that serum levels of TNF- α were independently associated with UAE in type 2 diabetic patients⁵³.

IL-1 is a major pro-inflammatory cytokine that plays a central role in the mechanism of acute and chronic inflammation. IL-1 is known to be increased in the kidney of animal models of DN⁵⁴. In addition, IL-1 stimulates synthesis of expression of prostaglandin E2, suggesting that IL-1 might be related to the change of glomerular hemodynamics⁵⁵.

IL-18, which belongs to the IL-1 superfamily, is a pro-inflammatory cytokine secreted from mononuclear cells. Serum concentration of IL-18 is known to be a strong predictor of death in patients with cardiovascular diseases. IL-18 induces expression of pro-inflammatory cytokines, and adhesion molecules and apoptosis^{56,57}. In our study, serum and urinary IL-18 levels were significantly elevated in patients with type 2 diabetes as compared with control subjects⁵⁸. We found significant positive correlations between serum and urinary levels of IL-18 and UAE. Serum IL-18 levels were also correlated positively with carotid intima media thickness (IMT) and brachial-ankle pulse wave velocity (baPWV). Furthermore, serum and urinary IL-18 levels correlated positively with AER after 6 months and

changes in UAE during the follow-up period of 6 months, suggesting that serum levels of IL-18 might be a predictor of progression of diabetic nephropathy, as well as cardiovascular diseases⁵⁸. Araki *et al.*⁵⁹ also showed that elevated levels of IL-18 are a determinant of early renal dysfunction in patients with type 2 diabetes.

Suzuki *et al.*⁶⁰ showed that mesangial cells, tubular cells and infiltrating cells expressed IL-6 messenger ribonucleic acid in the renal tissues of patients with DN by *in situ* hybridization. Serum levels of IL-6 were substantially higher in patients with DN than in control patients without DN⁶¹.

THERAPEUTIC IMPLICATIONS

Microinflammation is a potential therapeutic target for DN, because recent growing evidence has shown that the inflammatory process underlies the pathogenesis of DN. To date, there has been no established drug that ameliorates diabetic nephropathy through anti-inflammatory actions in humans, although an increasing number of studies *in vitro* and *in vivo* have suggested the efficacy of anti-inflammatory agents on DN (Table 1).

The potential beneficial anti-inflammatory effects on DN by using immunosuppressive agents in both type 1 and type 2 diabetic animal models has been reported. Utimura *et al.*⁶² reported that treatment with mycophenolate mofetil, an immunosuppressive agent, had no effect on blood pressure, glomerular dynamics or blood glucose levels, but prevents albuminuria, glomerular macrophage infiltration and glomerulosclerosis in streptozotocin-induced diabetic rats. Similar findings have been reported in Zucker fatty rats⁶³. In contrast, we also examined the effects of methotrexate on streptozotocin-induced diabetic rats⁶⁴. The results showed that methotrexate decreased UAE, renal tissue injuries and inflammation in the kidney. These findings clearly support the fundamental concept that anti-inflammatory effects are beneficial for the treatment of DN,

Table 1 | Renoprotective and anti-inflammatory agents shown in the animal diabetic models

Statin	Ota <i>et al. Diabetologia</i> 2003 ⁶⁸ Usui <i>et al. Nephrol Dial Transplant</i> 2003 ⁶⁷
Thiazolidinedione	Ohga <i>et al. Am J Physiol, Renal Physiol</i> , 2007 ⁶⁹
Angiotensin II receptor antagonist	Lee <i>et al. J Am Soc Nephrol</i> 2004 ⁶⁵
Spironolactone	Han <i>et al. J Am Soc Nephrol</i> 2006 ⁶⁶
Immunosuppressant	Utimura <i>et al. Kidney Int</i> 2003 ⁶² Yozai <i>et al. J Am Soc Nephrol</i> 2005 ⁶⁴
Pentoxifyline	DiPetrillo <i>et al. Am J Nephrol</i> 2004 ⁷¹
Macrolide (erythromycin)	Tone <i>et al. Diabetologia</i> 2005 ⁸⁰
GLP-1 receptor agonist	Park <i>et al. J Am Soc Nephrol</i> 2007 ⁷⁷ Kodera <i>et al. Diabetologia</i> 2011 ⁷⁸ Hendarto <i>et al. Metabolism</i> 2012 ⁷⁹
Cholecystokinin	Miyamoto <i>et al. Diabetes</i> 2012 ⁷⁴

GLP-1, glucagon-like peptide-1.

although the adverse effects caused by immunosuppression have not been evaluated.

In contrast, several kinds of drugs that are used for diabetic patients, such as angiotensin converting enzyme (ACE) inhibitor, angiotensin II receptor antagonist (ARB), thiazolidinediones and statins, are known to have anti-inflammatory effects as their pleiotropic effects. The renoprotective effects of ARB and aldosterone receptor blocker are considered to be at least partly related to anti-inflammatory actions through inhibition of nuclear factor- κ B (NF- κ B)-dependent pathways^{65,66}.

Statin is known to exert anti-inflammatory effects through inhibition of small-G proteins and NF- κ B-dependent inflammatory pathway independent of cholesterol-lowering effects. We examined the renoprotective effects of statin using streptozotocin-induced diabetic rats. Cerivastatin showed the amelioration of UAE, glomerular infiltration of macrophage and activation of NF- κ B in the kidney without any change of serum cholesterol level⁶⁷. Okada also showed the preventive effects of cerivastatin on renal injuries in diabetic rats through an anti-inflammatory effect⁶⁸.

Thiazolidinediones are also known to have anti-inflammatory actions by stimulation of the peroxisome proliferator-activated receptor (PPAR- γ) in addition to the effects of improvement of insulin resistance. We administered pioglitazone to streptozotocin-induced diabetic rats and found that pioglitazone reduced albuminuria, intraglomerular infiltration of macrophages and activation of NF- κ B, suggesting that pioglitazone exerts renoprotective effects through anti-inflammatory effects independent of blood glucose-lowering effects⁶⁹.

TNF- α is a major pro-inflammatory cytokine, and is considered to play important roles in the development of diabetic

nephropathy. Thus, TNF- α might be a promising therapeutic target for the treatment of DN. Pentoxifylline, which is a xanthine derivative, modulates the expression of TNF- α and other pro-inflammatory cytokines, and attenuates cellular processes involved in the inflammatory response⁷⁰. Treatment with pentoxifylline reduces the expression of TNF- α , IL-1, and IL-6 in the kidney and UAE in diabetic animals⁷¹. This drug reduced proteinuria and serum levels of TNF- α in patients with diabetes mellitus^{72,73}.

Recently, we have found that cholecystokinin (CCK) is expressed in the kidney and exerts renoprotective effects through its anti-inflammatory actions. Furthermore, administration of sulfated cholecystokinin octapeptide (CCK-8S) ameliorated albuminuria, podocyte loss, expression of pro-inflammatory genes and infiltration of macrophages in the kidneys of diabetic rats⁷⁴.

ANTI-INFLAMMATORY EFFECTS OF GLUCAGON-LIKE PEPTIDE-1 RECEPTOR AGONIST

Glucagon-like peptide-1 (GLP-1) is a gut incretin hormone that enhances glucose-dependent insulin secretion of pancreatic β -cells. Today, dipeptidylpeptidase-IV resistant long acting GLP-1 receptor agonists, exendin-4 and liraglutide, are available for treatment of type 2 diabetes. Previous reports have shown that GLP-1 receptor is expressed not only in the pancreas, but also in many organs including the kidney⁷⁵⁻⁷⁷. Park *et al.*⁷⁷ reported that long-term treatment of exendin-4 ameliorates diabetic nephropathy through improving metabolic anomalies in *db/db* mice. We have recently found that GLP-1 receptor was expressed on glomerular endothelial cells, and showed that exendin-4 directly acted on GLP-1 receptor and attenuated

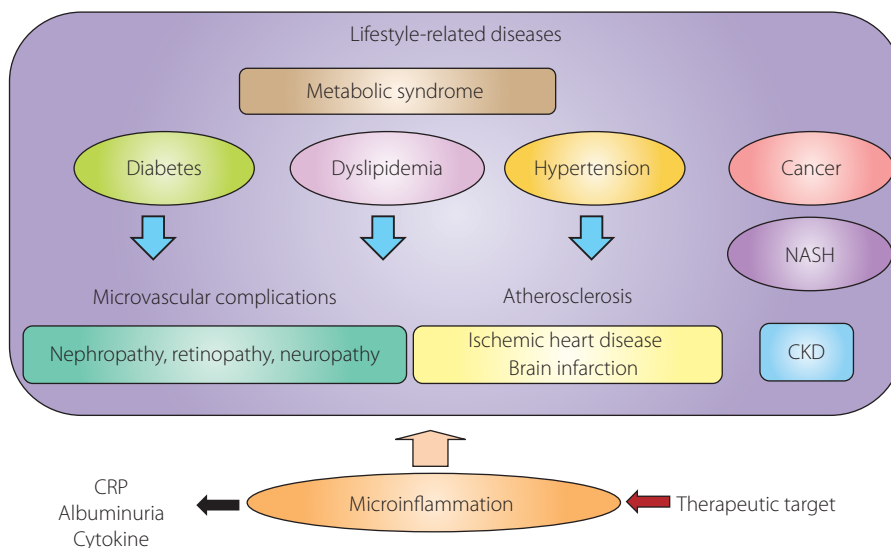


Figure 4 | Microinflammation and lifestyle-related diseases. Microinflammation is a common mechanism of the development of lifestyle-related diseases, including obesity-related insulin resistance, diabetic vascular complications, cardiovascular diseases, chronic kidney disease (CKD), non-alcoholic steatohepatitis (NASH) and some kinds of cancer. Albuminuria, C-reactive protein (CRP) and plasma levels of cytokines might be surrogate clinical markers for the microinflammation.

ICAM-1 expression on glomerular endothelial cells *in vitro*⁷⁸. Furthermore, exendin-4 ameliorated albuminuria, glomerular hyperfiltration, glomerular hypertrophy and mesangial matrix expansion in type 1 diabetic rats without changing blood pressure and bodyweight. Exendin-4 also prevented macrophage infiltration, expressions of ICAM-1 and type IV collagen, oxidative stress, and NF- κ B activation in kidney tissue⁷⁸. Furthermore, Hendaro *et al.*⁷⁹ recently reported that the GLP-1 analog, liraglutide, protects against oxidative stress and albuminuria in streptozotocin-induced diabetic rats through protein kinase A-mediated inhibition of renal nicotinamide adenine dinucleotide phosphate oxidases. These results show that GLP-1 receptor agonists might be beneficial for DN through anti-inflammatory and anti-oxidative actions independent of blood glucose-lowering effect.

CONCLUSION

There has been accumulating evidence showing that microinflammation is one of the key factors in the pathogenesis of diabetic nephropathy. Abnormalities of blood glucose, blood pressure or dyslipidemia trigger the activation of inflammatory pathways in the diabetic kidney followed by functional and structural renal injury. Microinflammation is a common pathological condition not only in diabetic vascular complications, but also in metabolic syndrome, non-alcoholic steatohepatitis and some kinds of cancer (Figure 4). These findings strongly suggest that microinflammation is a promising new therapeutic target for lifestyle-related diseases.

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We declare no conflicts of interest related to the manuscript.

REFERENCES

- Magee GM, Bilous RW, Cardwell CR, *et al.* Is hyperfiltration associated with the future risk of developing diabetic nephropathy? A meta-analysis. *Diabetologia* 2009; 52: 691–697.
- Zemin C, Mark EC. Pathogenesis of diabetic nephropathy. *J Diabetes Invest* 2011; 2: 243–247.
- Brownlee M, Cerami A, Vlassara H. Advanced glycosylation end products in tissue and the biochemical basis of diabetic complications. *N Engl J Med* 1988; 318: 1315–1321.
- Koya D, Jirousek MR, Lin YW. Characterization of protein kinase C beta isoform activation on the gene expression of transforming growth factor-beta, extracellular matrix components, and prostanoids in the glomeruli of diabetic rats. *J Clin Invest* 1997; 100: 115–126.
- Ziyadeh FN, Sharma K, Ericksen M, *et al.* Stimulation of collagen gene expression and protein synthesis in murine mesangial cells by high glucose is mediated by autocrine activation of transforming growth factor-beta. *J Clin Invest* 1994; 93: 536–542.
- Navarro-González JF, Mora-Fernández C, Muros de Fuentes M, *et al.* Inflammatory molecules and pathways in the pathogenesis of diabetic nephropathy. *Nat Rev Nephrol* 2011; 7: 327–340.
- Ross R. Atherosclerosis—an inflammatory disease. *N Engl J Med* 1999; 14: 115–126.
- Furuta T, Saito T, Ootaka T, *et al.* The role of macrophages in diabetic glomerulosclerosis. *Am J Kidney Dis* 1993; 21: 480–485.
- Shikata K, Makino H. Role of macrophages in the pathogenesis of diabetic nephropathy. *Contrib Nephrol* 2001; 134: 46–54.
- Staunton DE, Marlin SD, Stratowa C, *et al.* Primary structure of ICAM-1 demonstrates interaction between members of the immunoglobulin and integrin supergene families. *Cell* 1988; 52: 925–933.
- Sumagin R, Sarelius IH. TNF α activation of arterioles and venules alters distribution and levels of ICAM 1 and affects leukocyte-endothelial cell interactions. *Am J Physiol Heart Circ Physiol* 2006; 291: H2116–H2125.
- Sucosky P, Balachandran K, Elhammali A, *et al.* Altered shear stress stimulates upregulation of endothelial VCAM 1 and ICAM 1 in a BMP 4 and TGF β 1 dependent pathway. *Arterioscler Thromb Vasc Biol* 2009; 29: 254–260.
- Vlassara H, Fuh H, Donnelly T, *et al.* Advanced glycation endproducts promote adhesion molecule (VCAM-1, ICAM-1) expression and atheroma formation in normal rabbits. *Mol Med* 1995; 1: 447–456.
- Sugimoto H, Shikata K, Hirata K, *et al.* Increased expression of intercellular adhesion molecule-1 (ICAM-1) in diabetic rat glomeruli: glomerular hyperfiltration is a potential mechanism of ICAM-1 upregulation. *Diabetes*, 1997; 46: 2075–2081.
- Coimbra TM, Janssen U, Gröne HJ, *et al.* Early events leading to renal injury in obese Zucker (fatty) rats with type II diabetes. *Kidney Int* 2000; 57: 167–182.
- Okada S, Shikata K, Matsuda M, *et al.* Intercellular adhesion molecule-1-deficient mice are resistant against renal injury after induction of diabetes. *Diabetes* 2003; 52: 2586–2593.
- Chow FY, Nikolic-Paterson DJ, Ozols E, *et al.* Intercellular adhesion molecule-1 deficiency is protective against nephropathy in type 2 diabetic db/db mice. *J Am Soc Nephrol* 2005; 16: 1711–1722.
- Clausen P, Jacobsen P, Rossing K, *et al.* Plasma concentrations of VCAM-1 and ICAM-1 are elevated in patients with type 1 diabetes mellitus with microalbuminuria and overt nephropathy. *Diabet Med* 2000; 17: 644–649.
- Lin J, Glynn RJ, Rifai N, *et al.* Inflammation and progressive nephropathy in type 1 diabetes in the Diabetes Control and Complications Trial. *Diabetes Care* 2008; 31: 2338–2343.
- Ina K, Kitamura H, Okeda T, *et al.* Vascular cell adhesion molecule-1 expression in the renal interstitium of diabetic KKAY mice. *Diabetes Res Clin Pract* 1999; 44: 1–8.
- Rubio-Guerra AF, Vargas-Robles H, Lozano JJ, *et al.* Correlation between circulating adhesion molecule levels

- and albuminuria in type 2 diabetic hypertensive patients. *Kidney Blood Press Res* 2009; 32: 106–109.
22. Stehouwer CD, Gall MA, Twisk JW, *et al.* Increased urinary albumin excretion, endothelial dysfunction, and chronic low-grade inflammation in type 2 diabetes: progressive, interrelated, and independently associated with risk of death. *Diabetes*, 2002; 51: 1157–1165.
 23. Hara T, Ishida T, Cangara HM, *et al.* Endothelial cell-selective adhesion molecule regulates albuminuria in diabetic nephropathy. *Microvasc Res* 2009; 77: 348–355.
 24. Narumi S, Onozato ML, Tojo A, *et al.* Tissue-specific induction of E selectin in glomeruli is augmented following diabetes mellitus. *Nephron* 2001; 89: 161–171.
 25. Soedamah-Muthu SS, Chaturvedi N, Schalkwijk CG, *et al.* Soluble vascular cell adhesion molecule 1 and soluble E selectin are associated with micro- and macrovascular complications in type 1 diabetic patients. *J Diabetes Complications* 2006; 20: 188–195.
 26. Ogawa D, Shikata K, Honke K, *et al.* Cerebroside sulfotransferase deficiency ameliorates L-selectin-dependent monocyte infiltration in the kidney after ureteral obstruction. *J Biol Chem* 2004; 16: 2085–2090.
 27. Suzuki H, Kurihara Y, Takeya M, *et al.* A role for macrophage scavenger receptors in atherosclerosis and susceptibility to infection. *Nature* 1997; 386: 292–296.
 28. de Winther MP, van Dijk KW, Havekes LM, *et al.* Macrophage scavenger receptor class A: a multifunctional receptor in atherosclerosis. *Arterioscler Thromb Vasc Biol* 2000; 20: 290–297.
 29. Usui HK, Shikata K, Sasaki M. Macrophage scavenger receptor-a-deficient mice are resistant against diabetic nephropathy through amelioration of microinflammation. *Diabetes* 2007; 56: 363–372.
 30. Ruster C, Wolf G. The role of chemokines and chemokine receptors in diabetic nephropathy. *Front Biosci* 2008; 13: 944–955.
 31. Galkina E, Ley K. Leukocyte recruitment and vascular injury in diabetic nephropathy. *J Am Soc Nephrol* 2006; 17: 368–377.
 32. Wada T, Furuichi K, Sakai N, *et al.* Up-regulation of monocyte chemoattractant protein 1 in tubulointerstitial lesions of human diabetic nephropathy. *Kidney Int* 2000; 58: 1492–1498.
 33. Panzer U, Steinmetz OM, Stahl RA, *et al.* Kidney diseases and chemokines. *Curr Drug Targets* 2006; 7: 65–80.
 34. Amann B, Tinzmann R, Angelkort B. ACE inhibitors improve diabetic nephropathy through suppression of renal MCP 1. *Diabetes Care* 2003; 26: 2421–2425.
 35. Chow Y, Nikolic-Paterson DJ, Ozols E, *et al.* Monocyte chemoattractant protein 1 promotes the development of diabetic renal injury in streptozotocin-treated mice. *Kidney Int* 2006; 69: 73–80.
 36. Kitagawa K, Wada T, Furuichi K, *et al.* Blockade of CCR2 ameliorates progressive fibrosis in kidney. *Am J Pathol* 2004; 165: 237–246.
 37. Umehara H, Bloom ET, Okazaki T, *et al.* Fractalkine in vascular biology: from basic research to clinical disease. *Arterioscler Thromb Vasc Biol* 2004; 24: 34–40.
 38. Kikuchi Y, Ikee R, Hemmi N, *et al.* Fractalkine and its receptor, CX3CR1, upregulation in streptozotocin-induced diabetic kidneys. *Nephron Exp Nephrol* 2004; 97: e17–e25.
 39. Kikuchi Y, Imakiire T, Hyodo T, *et al.* Advanced glycation end-product induces fractalkine gene upregulation in normal rat glomeruli. *Nephrol Dial Transplant* 2005; 20: 2690–2696.
 40. Appay V, Rowland-Jones SL. RANTES: a versatile and controversial chemokine. *Trends Immunol* 2001; 22: 83–87.
 41. Mezzano S, Aros C, Droguett A, *et al.* NF-kappaB activation and overexpression of regulated genes in human diabetic nephropathy. *Nephrol Dial Transplant* 2004; 19: 2505–2512.
 42. Hasegawa G, Nakano K, Sawada M, *et al.* Possible role of tumor necrosis factor and interleukin 1 in the development of diabetic nephropathy. *Kidney Int* 1991; 40: 1007–1012.
 43. DiPetrillo K, Coutermarsh B, Gesek FA. Urinary tumor necrosis factor contributes to sodium retention and renal hypertrophy during diabetes. *Am J Physiol Renal Physiol* 2003; 284: F113–F121.
 44. Bertani T, Abbate M, Zoja C, *et al.* Tumor necrosis factor induces glomerular damage in rabbit. *Am J Pathol* 1989; 134: 419–430.
 45. Baud L, Perez J, Friedlander G, *et al.* Tumor necrosis factor stimulates prostaglandin production and cyclic AMP levels in rat cultured mesangial cells. *FEBS Lett* 1998; 239: 50–54.
 46. Laster SM, Wood JG, Gooding LR. Tumor necrosis factor can induce both apoptotic and necrotic forms of cell lysis. *J Immunol* 1938; 141: 2629–2634.
 47. McCarthy ET, Sharma R, Sharma M, *et al.* TNF α increases albumin permeability of isolated rat glomeruli through the generation of superoxide. *J Am Soc Nephrol* 1998; 9: 433–438.
 48. Sugimoto H, Shikata K, Wada J, *et al.* Advanced glycation end products cytokine nitric oxide sequence pathway in the development of diabetic nephropathy: aminoguanidine ameliorates the overexpression of tumour necrosis factor α and inducible oxide synthase in diabetic rat glomeruli. *Diabetologia* 1999; 42: 878–886.
 49. Navarro JF, Milena FJ, Mora C, *et al.* Renal pro-inflammatory cytokine gene expression in diabetic nephropathy: effect of angiotensin-converting enzyme inhibition and pentoxifylline administration. *Am J Nephrol* 2006; 26: 562–570.
 50. Navarro JF, Mora C. Role of inflammation in diabetic complications. *Nephrol Dial Transplant* 2005; 20: 2601–2604.
 51. Navarro-González JF, Mora-Fernández C. The role of inflammatory cytokines in diabetic nephropathy. *J Am Soc Nephrol* 2008; 19: 433–442.
 52. Navarro JF, Mora C, Macía M. Inflammatory parameters are independently associated with urinary albumin excretion in type 2 diabetes mellitus. *Am J Kidney Dis* 2003; 42: 53–61.
 53. Kajitani N, Shikata K, Nakamura A, *et al.* Microinflammation is a common risk factor for progression of nephropathy

- and atherosclerosis in Japanese patients with type 2 diabetes *Diabetes Res Clin Pract* 2010; 88: 171–176.
54. Sassy-Prigent C, Heudes D, Mandet C, *et al.* Early glomerular macrophage recruitment in streptozotocin-induced diabetic rats. *Diabetes* 2000; 49: 466–475.
 55. Pfeilschifter J, Pignat W, Vosbeck K, *et al.* Interleukin 1 and tumor necrosis factor synergistically stimulate prostaglandin synthesis and phospholipase A2 release from rat renal mesangial cells. *Biochem Biophys Res Commun* 1989; 159: 385–394.
 56. Dai SM, Matsuno H, Nakamura H, *et al.* Interleukin 18 enhances monocyte tumor necrosis factor α and interleukin 1 β production induced by direct contact with T lymphocytes: implications in rheumatoid arthritis. *Arthritis Rheum* 2004; 50: 432–443.
 57. Mariño E, Cardier JE. Differential effects of IL 18 on endothelial cell apoptosis mediated by TNF α and Fas (CD59). *Cytokine* 2003; 22: 142–148.
 58. Nakamura A, Shikata K, Hiramatsu M, *et al.* Serum interleukin 18 levels are associated with nephropathy and atherosclerosis in Japanese patients with type 2 diabetes. *Diabetes Care* 2005; 28: 2890–2895.
 59. Araki S, Haneda M, Koya D, *et al.* Predictive impact of elevated serum level of IL 18 for early renal dysfunction in type 2 diabetes: an observational follow-up study. *Diabetologia* 2007; 50: 867–873.
 60. Suzuki D, Miyazaki M, Naka R, *et al.* *In situ* hybridization of interleukin 6 in diabetic nephropathy. *Diabetes* 1995; 44: 1233–1238.
 61. Sekizuka K, Tomino Y, Sei C, *et al.* Detection of serum IL 6 in patients with diabetic nephropathy. *Nephron* 1994; 68: 284–285.
 62. Utimura R, Fujihara CK, Mattar AL. Mycophenolate mofetil prevents the development of glomerular injury in experimental diabetes. *Kidney Int* 2003; 63: 209–216.
 63. Rodríguez-Iturbe B, Quiroz Y, Shahkarami A, *et al.* Mycophenolate mofetil ameliorates nephropathy in the obese Zucker rat. *Kidney Int* 2005; 68: 1041–1047.
 64. Yozai K, Shikata K, Sasaki M, *et al.* Methotrexate prevents renal injury in experimental diabetic rats via anti-inflammatory actions. *J Am Soc Nephrol* 2005; 16: 3326–3338.
 65. Lee FT, Cao Z, Long DM, *et al.* Interactions between angiotensin II and NF κ B dependent pathways in modulating macrophage infiltration in experimental diabetic nephropathy. *J Am Soc Nephrol* 2004; 15: 2139–2151.
 66. Han SY, Kim CH, Kim HS, *et al.* Spironolactone prevents diabetic nephropathy through an anti-inflammatory mechanism in type 2 diabetic rats. *J Am Soc Nephrol* 2006; 17: 1362–1372.
 67. Usui H, Shikata K, Matsuda M, *et al.* HMG-CoA reductase inhibitor ameliorates diabetic nephropathy by its pleiotropic effects in rats. *Nephrol Dial Transplant* 2003; 18: 265–272.
 68. Ota T, Takamura T, Ando H, *et al.* Preventive effect of cerivastatin on diabetic nephropathy through suppression of glomerular macrophage recruitment in a rat model. *Diabetologia* 2003; 46: 843–851.
 69. Ohga S, Shikata K, Yozai K, *et al.* Thiazolidinedione ameliorates renal injury in experimental diabetic rats through anti-inflammatory effects mediated by inhibition of NF-kappaB activation. *Am J Physiol Renal Physiol* 2007; 292: F1141–F1150.
 70. Han J, Thompson P, Beutler D. Dexamethasone and pentoxifylline inhibit endotoxin-induced cachectin/tumor necrosis factor synthesis at separate points in the signaling pathway. *J Exp Med* 1990; 172: 391–394.
 71. DiPetrillo K, Gesek FA. Pentoxifylline ameliorates renal tumor necrosis factor expression, sodium retention, and renal hypertrophy in diabetic rats. *Am J Nephrol* 2004; 24: 352–359.
 72. Guerrero-Romero F, Rodríguez-Morán M, Paniagua-Sierra JR, *et al.* Pentoxifylline reduces proteinuria in insulin-dependent and non insulin-dependent diabetic patients. *Clin Nephrol* 1995; 43: 116–121.
 73. Navarro JF, Mora C. Antiproteinuric effect of pentoxifylline in patients with diabetic nephropathy. *Diabetes Care* 1999; 22: 1006–1008.
 74. Miyamoto S, Shikata K, Miyasaka K, *et al.* Cholecystokinin plays a novel protective role in diabetic kidney through anti-inflammatory actions on macrophage: anti-inflammatory effect of cholecystokinin. *Diabetes* 2012; 61: 897–907.
 75. Bullock BP, Heller RS, Habener JF, *et al.* Tissue distribution of messenger ribonucleic acid encoding the rat glucagon-like peptide-1 receptor. *Endocrinology* 1996; 137: 2968–2978.
 76. Schlatter P, Beglinger C, Drewe J, *et al.* Glucagon-like peptide 1 receptor expression in primary porcine proximal tubular cells. *Regul Pept* 2007; 141: 120–128.
 77. Park CW, Kim HW, Ko SH, *et al.* Long-term treatment of glucagon-like peptide-1 analog exendin-4 ameliorates diabetic nephropathy through improving metabolic anomalies in db/db mice. *J Am Soc Nephrol* 2007; 18: 1227–1238.
 78. Kodera R, Shikata K, Kataoka HU, *et al.* Glucagon-like peptide-1 receptor agonist ameliorates renal injury through its anti-inflammatory action without lowering blood glucose level in a rat model of type 1 diabetes. *Diabetologia* 2011; 54: 965–978.
 79. Hendaro H, Inoguchi T, Maeda Y, *et al.* GLP-1 analog liraglutide protects against oxidative stress and albuminuria in streptozotocin-induced diabetic rats via protein kinase A-mediated inhibition of renal NAD(P)H oxidases. *Metabolism* 2012; 61: 1422–1434.
 80. Tone A, Shikata K, Sasaki M, *et al.* Erythromycin ameliorates renal injury via anti-inflammatory effects in experimental diabetic rats. *Diabetologia* 2005; 48: 2402–2411.