

Research Article

Value of Caffeic Acid Phenethyl Ester Pretreatment in Experimental Sepsis Model in Rats

Ozlem Alici,^{1,2} Havva Sahin Kavakli,^{1,3} Cemile Koca,^{1,4} Neriman Defne Altintas,^{1,5} Murat Aydin,^{1,6} and Suleyman Alici^{1,7}

¹Department of Infectious Diseases and Clinical Bacteriology and Department of Biochemistry, Faculty of Medicine, Fatih University, Ankara, Turkey

²Department of Infectious Diseases and Clinical Bacteriology, Fatih Sultan Mehmet Training and Research Hospital, 34746 Istanbul, Turkey

³Department of Emergency Medicine, Faculty of Medicine, Yıldırım Beyazıt University, Ankara, Turkey

⁴Department of Biochemistry, Faculty of Medicine, Yıldırım Beyazıt University, Ankara, Turkey

⁵Department of Internal Medicine, Faculty of Medicine, Ankara University, Ankara, Turkey

⁶Department of Biochemistry, Faculty of Medicine, Namik Kemal University, Tekirdağ, Turkey

⁷Department of Oncology, Faculty of Medicine, Bahcesehir University, Istanbul, Turkey

Correspondence should be addressed to Ozlem Alici; oalici@gmail.com

Received 23 December 2014; Revised 19 March 2015; Accepted 21 March 2015

Academic Editor: Nina Ivanovska

Copyright © 2015 Ozlem Alici et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background and Aim. The aim of this study was to determine the actions of caffeic acid phenethyl ester (CAPE) on the changes of endothelin-1 (ET-1) level, tumor necrosis factor- (TNF-) alpha, and oxidative stress parameters such as superoxide dismutase (SOD) activities and malondialdehyde (MDA) levels in experimental sepsis model in rats. **Materials and Methods.** Twenty-four rats were randomly divided into three experimental groups: sham (group 1), sepsis (group 2), and sepsis + CAPE (group 3), $n = 8$ each. CAPE was administered ($10 \mu\text{mol/kg}$) intraperitoneally to group 3 before sepsis induction. Serum ET-1, serum TNF-alpha, tissue SOD activity, and tissue MDA levels were measured in all groups. **Results.** Pretreatment with CAPE decreased ET-1, TNF-alpha, and MDA levels in sepsis induced rats. Additionally SOD activities were higher in rats pretreated with CAPE after sepsis induction. **Conclusion.** Our results demonstrate that CAPE may have a beneficial effect on ET and TNF-alpha levels and oxidative stress parameters induced by sepsis in experimental rat models. Therefore treatment with CAPE can be used to avoid devastating effects of sepsis.

1. Introduction

Sepsis is a complex process characterized by uncontrolled systemic inflammation that may eventually lead to multiorgan dysfunction and even death [1].

Endothelin-1 (ET-1) is the main isoform of the endothelin family, which have important roles in a wide range of diseases affecting the vascular system, kidney, heart, and lungs. Overexpression of ET-1 during sepsis, as a result of endothelial dysfunction, causes imbalance in local tissue perfusion [2, 3].

Excessive release of proinflammatory mediators which are induced by endotoxin like tumor necrosis factor-alpha

(TNF-alpha) and reactive oxygen species (ROS) causes systemic inflammatory response during sepsis [4]. TNF-alpha has important roles in the development of organ dysfunction related to sepsis. It induces organ damage through activation of neutrophils and endothelial cells as well as coagulation abnormalities in patients with sepsis. Based on this knowledge, inhibition of TNF- α might be critical for treating septic organ dysfunction [5, 6].

ROS have been associated with the onset, progression, and outcome of sepsis, both in experimental and in clinical studies [7, 8]. Excessive production of ROS during sepsis is the result of actions of leukocytes and may disturb

the antioxidative and oxidative balance resulting in organ damage leading to multiorgan failure [9]. Treatment options that may improve any of these parameters may be beneficial to treat sepsis.

Caffeic acid phenethyl ester (CAPE) is one of the major components of honeybee propolis and has been used in traditional medicine. It was found to be a potent free radical scavenger and antioxidant [10]. CAPE inhibits 5-lipoxygenase-catalysed oxygenation of linoleic acid and arachidonic acid in the micromolar concentration range [10]. It blocks production of reactive oxygen species (ROS) in human neutrophils and the xanthine/xanthine oxidase system at the concentration of 10 μ M [10]. It has been reported that CAPE is a potent anti-inflammatory and antioxidant agent and possesses cytostatic, antiviral, antibacterial, and antifungal properties [11].

The purpose of the current study was to evaluate the effects of CAPE on ET-1 levels, TNF-alpha, and oxidative stress in a rat sepsis model.

2. Materials and Methods

2.1. Study Design. Animal experiment was performed in accordance with the National Institute of Health guidelines for animal research and was approved by the Animal Research Ethics Committee of Fatih University School of Medicine, Istanbul. 24 adult male Wistar rats (weight range, 250 to 270 g) were used for the experiment. All animals had access to commercial standard diet and water ad libitum throughout the study. The rats were divided randomly into three groups of eight animals each: sham group (group 1), sepsis group (group 2), and sepsis group treated with CAPE (group 3).

Sham group (group 1) received only 1 mL intraperitoneal (i.p.) injection of 0.9% saline solution. Sepsis in group 2 and group 3 was induced by intraperitoneal (i.p.) injection of 2×10^{10} CFU of *Escherichia coli* ATCC 25922. *E. coli* ATCC 25922 was grown in brain-heart infusion broth. In the logarithmic phase of the growth, the suspension was centrifuged at 1000 g for 15 minutes, the supernatant was discarded, and the bacteria were resuspended and diluted in sterile saline. The rats received an i.p. inoculum of 1 mL of saline containing 2×10^{10} CFU of *E. coli* ATCC 25922 [12]. Six hours after bacterial challenge, group 2 received isotonic sodium chloride solution and group 3 received CAPE (10 μ mol/kg) by intraperitoneal injection. The CAPE was synthesized by standard method of Grunberger to prepare 25 micromol/mL of CAPE solution [13].

2.2. Sample Collection. 24 hours after bacterial challenge, all animals were sacrificed using ketamine and cardiac puncture. Blood samples were drawn from vena cava inferior; liver tissue samples were immediately removed and stored at -80°C for the determination of tissue associated malondialdehyde (MDA) levels and superoxide dismutase (SOD) activities, as the parameters of oxidative stress. Blood samples were collected for the determination of endothelin and TNF-alpha levels, centrifuged at 3000 g for 10 minutes, and stored at -80°C .

2.3. Quantitative Determination of Serum Endothelin Levels. Rat big ET-1 levels were measured using commercially available enzyme-linked immunosorbent assay (ELISA) kits (Assay Designs, MI, USA) following the manufacturer's instructions. The results are presented as pg/mL.

2.4. Quantitative Determination of Serum TNF-Alpha Levels. Tumor necrosis factor-alpha rat ELISA kits produced by Biovendor Research and Diagnostic Products were utilised as described by the manufacturer. The results were expressed as pg/mL.

2.5. Quantitative Determination of Tissue Malondialdehyde Levels. Reaction with thiobarbituric acid (TBA) at $90-100^{\circ}\text{C}$ was used as the basis for the determination of tissue thiobarbituric acid-reactive substance (TBARS) levels [14]. In the TBA test, a pink pigment with a maximum absorption at 532 nm was produced from the reaction of MDA or MDA-like substances with TBA. The reaction took place at pH 2-3 and 90°C for 15 min. Protein was precipitated by mixing samples with two volumes of cold 10% (w/v) trichloroacetic acid. The precipitate was pelleted by centrifugation and an aliquot of the supernatant was combined with an equal volume of 0.67% (w/v) TBA and placed in a boiling water-bath for 10 min. The absorbance was read at 532 nm after cooling. Results were expressed as nmol per gram wet tissue, using the standard graphic prepared based on measurements with a standard solution.

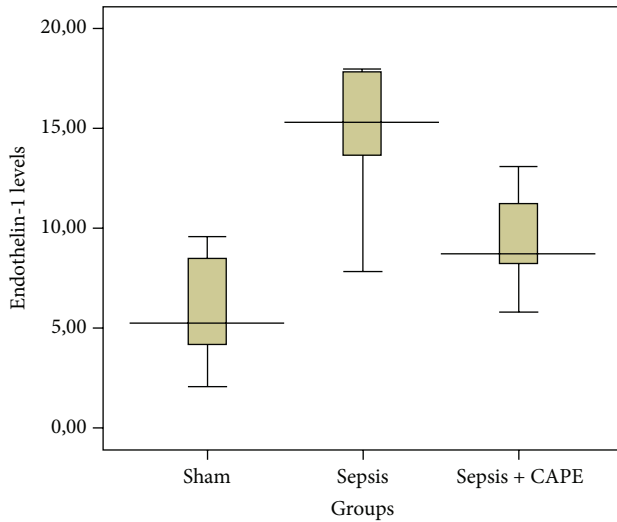
2.6. Quantitative Determination of Tissue Superoxide Dismutase Activities. SOD activity was determined according to the method defined by Sun et al. [15]. This method is based on the inhibition of nitroblue tetrazolium (NBT) reduction using a xanthine-xanthine oxidase system as a superoxide generator. Following the addition of 1.0 mL of an ethanol-chloroform mixture (5:3, v/v) to an equal volume of sample and centrifugation, activity was assessed in the ethanol phase of the supernatant. One unit of SOD was defined as the amount causing 50% inhibition in the NBT reduction rate. SOD activity is expressed as U mg^{-1} protein.

2.7. Statistical Analysis. For statistical evaluation, we used the statistical software package SPSS 15.0 and probability value of less than 0.05 was accepted as statistically significant. As the data were independent and showed normal distribution, statistical analysis was performed using analysis of variance (ANOVA) followed by Tukey's test when comparing groups. The results are given as the mean \pm standard deviation of the mean (SD).

3. Results

ET-1, TNF-alpha, and MDA levels and SOD activities for all groups are presented in Figures 1-4 as mean \pm SD.

3.1. Serum Endothelin Levels. ET-1 levels were determined as it is one of the major peptides in the pathogenesis of sepsis



Group 1 (sham)	5.89 ± 3.10
Group 2 (sepsis)	14.64 ± 3.85
Group 3 (sepsis + CAPE)	10.31 ± 3.00
<i>P</i> values: groups 1-2	<i>P</i> = 0.001
groups 1-3	<i>P</i> > 0.05
groups 2-3	<i>P</i> = 0.028

ET-1: endothelin-1 CAPE: caffeic acid phenethyl ester

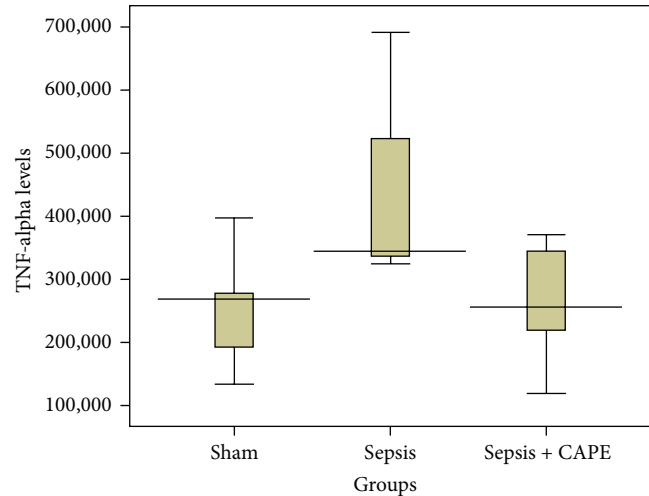
FIGURE 1: Serum ET-1 level (pg/mL) was described as mean value ± SD for all groups.

(Figure 1). ET-1 levels in sepsis group treated with CAPE (group 3) were lower than in sepsis group (group 2) ($P = 0.028$). Similarly, ET-1 levels in sham group (G1) were lower than in sepsis group (group 2) ($P = 0.001$).

3.2. Serum TNF-Alpha Levels. TNF-alpha levels were determined since they have an important role in the pathogenesis of sepsis (Figure 2). TNF-alpha levels in group 2 were significantly higher compared to group 1 ($P = 0.035$). When sepsis was induced in rat pretreated with CAPE (group 3), TNF-alpha levels were found to be similar to group 1 ($P > 0.05$) and significantly lower compared to group 2 ($P = 0.039$).

3.3. Tissue Malondialdehyde Levels. Tissue MDA levels, used as a marker of oxidative stress, were determined (Figure 3). Tissue MDA levels were significantly increased in the sepsis group (group 2) compared to group 1 ($P = 0.001$). When rats were pretreated with CAPE before sepsis induction, MDA levels were significantly lower compared to those in group 2 ($P = 0.002$).

3.4. Tissue Superoxide Dismutase Levels. SOD activities were used as a marker of oxidative stress (Figure 4). Tissue SOD activities after sepsis induction (group 2) were significantly decreased relative to those in group 1 ($P = 0.001$). When rats were pretreated with CAPE prior to sepsis induction, SOD activities were significantly higher compared to group



Group 1 (sham)	256.94 ± 88.88
Group 2 (sepsis)	440.31 ± 159.34
Group 3 (sepsis + CAPE)	283.98 ± 120.92
<i>P</i> values: groups 1-2	<i>P</i> = 0.035
groups 1-3	<i>P</i> > 0.05
groups 2-3	<i>P</i> = 0.039

TNF-alpha: tumor necrosis factor-alpha CAPE: caffeic acid phenethyl ester

FIGURE 2: Serum TNF-alpha level (pg/mL) was described as mean value ± SD for all groups.

2 ($P = 0.002$). Tissue SOD activities in group 1 were higher than group 3 ($P = 0.001$).

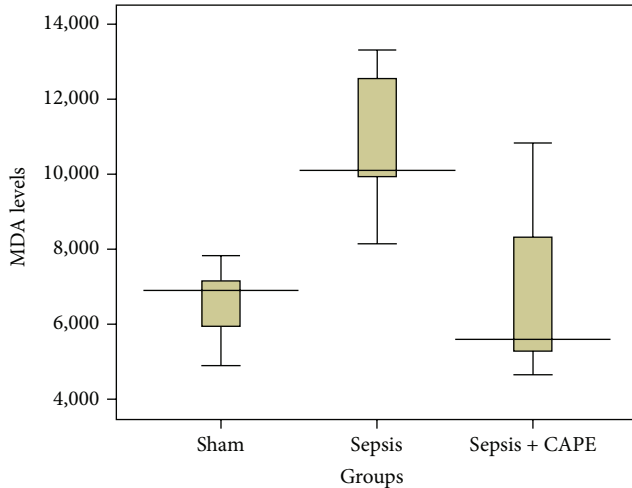
4. Discussion

Overall, results revealed that pretreatment with CAPE significantly reduced serum ET-1, TNF-alpha, and MDA levels while increasing SOD activities, when sepsis was induced in a rat sepsis model.

Sepsis syndromes range from the systemic inflammatory response syndrome to severe sepsis and septic shock. These syndromes are the major causes of death in critical care units worldwide [16]. Although none performed it in a systematic way, some studies have correlated inflammatory markers, oxidative stress, and clinical markers of organ failure [17].

ET-1 levels increase with a septic process in correlation with the circulatory dysfunction and sepsis severity [2, 18]. Its levels are thought to correlate with circulatory dysfunction during sepsis and with sepsis severity [19]. Several ET-1 receptor antagonists have already been reported to have possible beneficial effects on cardiovascular performance and survival in experimental sepsis models [20, 21]. Our results reveal that pretreatment with CAPE may also have an effect to decrease ET-1 levels during sepsis.

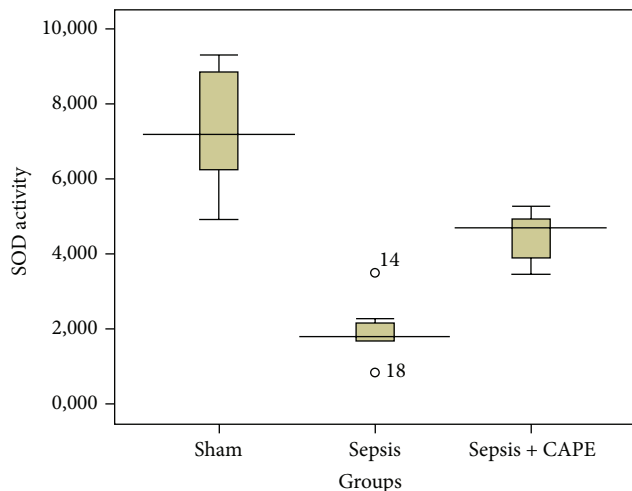
Increased oxidative stress, one of many factors involved in development of multiorgan dysfunction syndrome during sepsis, is the result of imbalance between antioxidant and oxidant status. ROS levels rise due to increased production as a part of the proinflammatory response during sepsis [17,



Group 1 (sham)	6.55 ± 0.99
Group 2 (sepsis)	10.93 ± 1.89
Group 3 (sepsis + CAPE)	6.90 ± 2.31
<i>P</i> values: groups 1-2	<i>P</i> = 0.001
groups 1-3	<i>P</i> > 0.05
groups 2-3	<i>P</i> = 0.002

MDA: malondialdehyde CAPE: caffeic acid phenethyl ester

FIGURE 3: Tissue MDA level (nmol g⁻¹ protein) was described as mean value ± SD for all groups.



Group 1 (sham)	7.29 ± 1.66
Group 2 (sepsis)	2.56 ± 1.21
Group 3 (sepsis + CAPE)	4.02 ± 1.21
<i>P</i> values: groups 1-2	<i>P</i> = 0.001
groups 1-3	<i>P</i> = 0.001
groups 2-3	<i>P</i> = 0.002

SOD: superoxide dismutase CAPE: caffeic acid phenethyl ester

FIGURE 4: Tissue SOD activity (U mg⁻¹ protein) was described as mean value ± SD for all groups.

22]. Moreover, during sepsis, major endogenous antioxidant systems including glutathione, vitamins A, C, and E, and several other antioxidant enzymes such as SOD are commonly depleted [22, 23]. SOD activity, an enzyme that scavenges the superoxide radicals and catalyzes them to hydrogen peroxide and oxygen, is commonly used to provide information on the antioxidant status [24]. MDA is also used to determine oxidant status, as it is a product of lipid peroxidation [25].

TNF-alpha is one of the proinflammatory cytokines which has been accepted as important mediators of sepsis [26–28]. Therefore, we wanted to investigate that therapeutic strategies may reduce the severity of sepsis by decreasing proinflammatory cytokines like TNF-alpha. In previous investigations, Pascual et al. described inhibition of the tumor necrosis factors pathway by propolis and Jung WK et al. reported decreasing serum levels of tumor necrosis factor-alpha and interleukin-1 beta by treatment of CAPE in lipopolysaccharide-induced septic shock model of mice [29, 30]. Our study results also have shown that TNF-alpha levels are lower upon pretreatment with CAPE. This finding also supports that CAPE may have beneficial effects during sepsis.

Death from sepsis has decreased recently depending on various adjunctive therapies. However, it remains high when compared with other critical illnesses [31]. In addition to fluid resuscitation, antibiotic therapy, and source control to remove the sepsis-inducing insult, inotropic-vasopressor therapy, glycemic control, prophylaxis for deep vein thrombosis, and stress ulcer prophylaxis are involved in sepsis treatment [16]. However, widespread research for a new drug that is effective in reducing mortality in sepsis has been sustaining.

A growing number of reports demonstrate that a proinflammatory and oxidative condition is related to the pathogenesis and the progression of endotoxin-induced septic shock and that antioxidants may have therapeutic potential in lipopolysaccharide- (LPS-) induced sepsis [32].

Because of this, in current study we investigate the roles of antioxidant therapy using CAPE in the treatment of sepsis. CAPE is a major component of honey bee propolis which is commonly used in Chinese traditional medicine. CAPE was also investigated in various organs such as intestine, lung, and spinal cord in a number of animal models and it has been reported that it has anti-inflammatory, radical scavenging and, therefore, antioxidant activities [10, 11, 33–37]. It also has immunomodulatory actions such as inhibition of ROS production and it is also demonstrated to exhibit cytostatic, antiviral, antibacterial, and antifungal properties [10, 11, 38–45]. Korish and Arafa showed that CAPE decreased the inflammatory cytokines and increased the anti-inflammatory cytokines levels in septic shock model and suggested that CAPE could help the prophylaxis and treatment of septic shock [46]. Similarly, Teke et al. reported that CAPE significantly decreased oxidative stress in another intraperitoneal sepsis model [47]. Moreover, there have been no reported side effects of CAPE on normal cells [45].

5. Conclusion

Our study demonstrated that effects of CAPE seemed to depend not only on the diminution of oxidative damage but also on its anti-inflammatory activity in rats *in vivo*. Therefore, it is reasonable to propose CAPE as a molecule with therapeutic potential for the treatment of systemic inflammation by interfering at the earliest steps of activation of the oxidative and proinflammatory cascade.

These data demonstrate the protective effect of CAPE treatment in experimental sepsis by reducing the inflammatory process and oxidative stress.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

References

- [1] A. J. Dare, A. R. J. Phillips, A. J. R. Hickey et al., "A systematic review of experimental treatments for mitochondrial dysfunction in sepsis and multiple organ dysfunction syndrome," *Free Radical Biology and Medicine*, vol. 47, no. 11, pp. 1517–1525, 2009.
- [2] M. Piechota, M. Banach, R. Izmanski et al., "Plasma endothelin-1 levels in septic patients," *Journal of Intensive Care Medicine*, vol. 22, no. 4, pp. 232–239, 2007.
- [3] R. Shah, "Endothelins in health and disease," *European Journal of Internal Medicine*, vol. 18, no. 4, pp. 272–282, 2007.
- [4] V. M. Victor, M. Rocha, J. V. Esplugues, and M. de la Fuente, "Role of free radicals in sepsis: antioxidant therapy," *Current Pharmaceutical Design*, vol. 11, no. 24, pp. 3141–3158, 2005.
- [5] T. Iba and T. Miyasho, "Danaparoid sodium attenuates the increase in inflammatory cytokines and preserves organ function in endotoxemic rats," *Critical Care*, vol. 12, no. 4, article R86, 2008.
- [6] Y. Ren, Y. Xie, G. Jiang et al., "Apoptotic cells protect mice against lipopolysaccharide-induced shock," *The Journal of Immunology*, vol. 180, no. 7, pp. 4978–4985, 2008.
- [7] R. F. Guo and P. A. Ward, "Role of oxidants in lung injury during sepsis," *Antioxidants and Redox Signaling*, vol. 9, no. 11, pp. 1991–2002, 2007.
- [8] P. H. Zapelini, G. T. Rezin, M. R. Cardoso et al., "Antioxidant treatment reverses mitochondrial dysfunction in a sepsis animal model," *Mitochondrion*, vol. 8, no. 3, pp. 211–218, 2008.
- [9] M. E. Andrades, C. Ritter, and F. Dal-Pizzol, "The role of free radicals in sepsis development," *Frontiers in Bioscience*, vol. 1, pp. 277–287, 2009.
- [10] A. Ilhan, U. Koltuksuz, S. Ozen, E. Uz, H. Ciralik, and O. Akyol, "The effects of caffeic acid phenethyl ester (CAPE) on spinal cord ischemia/reperfusion injury in rabbits," *European Journal of Cardio-Thoracic Surgery*, vol. 16, no. 4, pp. 458–463, 1999.
- [11] M. F. Tolba, S. S. Azab, A. E. Khalifa, S. Z. Abdel-Rahman, and A. B. Abdel-Naim, "Caffeic acid phenethyl ester, a promising component of propolis with a plethora of biological activities: a review on its anti-inflammatory, neuroprotective, hepatoprotective, and cardioprotective effects," *IUBMB Life*, vol. 65, no. 8, pp. 699–709, 2013.
- [12] R. Ghiselli, A. Giacometti, O. Cirioni et al., "Cecropin B enhances betalactams activities in experimental rat models of gram-negative septic shock," *Annals of Surgery*, vol. 239, no. 2, pp. 251–256, 2004.
- [13] D. Grunberger, R. Banerjee, K. Eisinger et al., "Preferential cytotoxicity on tumor cells by caffeic acid phenethyl ester isolated from propolis," *Experientia*, vol. 44, no. 3, pp. 230–232, 1988.
- [14] H. Ohkawa, N. Ohishi, and K. Yagi, "Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction," *Analytical Biochemistry*, vol. 95, no. 2, pp. 351–358, 1979.
- [15] Y. Sun, L. W. Oberley, and Y. Li, "A simple method for clinical assay of superoxide dismutase," *Clinical Chemistry*, vol. 34, no. 3, pp. 497–500, 1988.
- [16] J. X. Wilson, "Mechanism of action of vitamin C in sepsis: ascorbate modulates redox signaling in endothelium," *BioFactors*, vol. 35, no. 1, pp. 5–13, 2009.
- [17] M. Andrades, C. Ritter, M. R. de Oliveira, E. L. Streck, J. C. Fonseca Moreira, and F. Dal-Pizzol, "Antioxidant treatment reverses organ failure in rat model of sepsis: role of antioxidant enzymes imbalance, neutrophil infiltration, and oxidative stress," *Journal of Surgical Research*, vol. 167, no. 2, pp. e307–e313, 2011.
- [18] J. Figueras-Aloy, L. Gómez-Lopez, M. J. Rodríguez-Miguélez et al., "Plasma endothelin-1 and clinical manifestations of neonatal sepsis," *Journal of Perinatal Medicine*, vol. 32, no. 6, pp. 522–526, 2004.
- [19] M. Forni, S. Mazzola, L. A. Ribeiro et al., "Expression of endothelin-1 system in a pig model of endotoxic shock," *Regulatory Peptides*, vol. 131, no. 1–3, pp. 89–96, 2005.
- [20] A. B. Iskit and M. O. Guç, "A new therapeutic approach for the treatment of sepsis," *Medical Hypotheses*, vol. 62, no. 3, pp. 342–345, 2004.
- [21] D. Konrad, M. Haney, G. Johansson, M. Wanecek, E. Weitzberg, and A. Oldner, "Cardiac effects of endothelin receptor antagonism in endotoxemic pigs," *American Journal of Physiology—Heart and Circulatory Physiology*, vol. 293, no. 2, pp. H988–H996, 2007.
- [22] C. Ribeiro Nogueira, A. Ramalho, E. Lameu, C. A. da Silva Franca, C. David, and E. Accioli, "Serum concentrations of vitamin A and oxidative stress in critically ill patients with sepsis," *Nutricion Hospitalaria*, vol. 24, no. 3, pp. 312–317, 2009.
- [23] S. Rinaldi, F. Landucci, and A. R. de Gaudio, "Antioxidant therapy in critically septic patients," *Current Drug Targets*, vol. 10, no. 9, pp. 872–880, 2009.
- [24] H. Lu, J. Zhen, T. Wu et al., "Superoxide dismutase mimetic drug tempol aggravates anti-GBM antibody-induced glomerulonephritis in mice," *The American Journal of Physiology—Renal Physiology*, vol. 299, no. 2, pp. F445–F452, 2010.
- [25] I. Hünová, R. Novotný, H. Uhlířová et al., "The impact of ambient ozone on mountain spruce forests in the Czech Republic as indicated by malondialdehyde," *Environmental Pollution*, vol. 158, no. 7, pp. 2393–2401, 2010.
- [26] W. Li, M. Ashok, J. Li, H. Yang, A. E. Sama, and H. Wang, "A major ingredient of green tea rescues mice from lethal sepsis partly by inhibiting HMGB1," *PLoS ONE*, vol. 2, no. 11, Article ID e1153, 2007.
- [27] R. Saueremann, C. Marsik, I. Steiner et al., "Immunomodulatory effects of fosfomycin in experimental human endotoxemia," *Antimicrobial Agents and Chemotherapy*, vol. 51, no. 5, pp. 1879–1881, 2007.
- [28] H. Zhou, G. Ding, W. Liu et al., "Lipopolysaccharide could be internalized into human peripheral blood mononuclear cells

- and elicit TNF- α release, but not via the pathway of toll-like receptor 4 on the cell surface," *Cellular & Molecular Immunology*, vol. 1, no. 5, pp. 373–377, 2004.
- [29] C. Pascual, R. Gonzalez, and R. G. Torricella, "Scavenging action of propolis extract against oxygen radicals," *Journal of Ethnopharmacology*, vol. 41, no. 1-2, pp. 9–13, 1994.
- [30] W.-K. Jung, I. Choi, D.-Y. Lee et al., "Caffeic acid phenethyl ester protects mice from lethal endotoxin shock and inhibits lipopolysaccharide-induced cyclooxygenase-2 and inducible nitric oxide synthase expression in RAW 264.7 macrophages via the p38/ERK and NF- κ B pathways," *International Journal of Biochemistry and Cell Biology*, vol. 40, no. 11, pp. 2572–2582, 2008.
- [31] C. E. Sandrock and T. E. Albertson, "Controversies in the treatment of sepsis," *Seminars in Respiratory and Critical Care Medicine*, vol. 31, no. 1, pp. 66–78, 2010.
- [32] D. de Filippis, T. Iuvone, G. Esposito et al., "Melatonin reverses lipopolysaccharide-induced gastro-intestinal motility disturbances through the inhibition of oxidative stress," *Journal of Pineal Research*, vol. 44, no. 1, pp. 45–51, 2008.
- [33] B. T. Bilen, H. Kiliç, N. Alaybeyoglu et al., "Effect of caffeic acid phenethyl ester on survival of axial pattern flaps in rats with ischaemia-reperfusion injuries," *Scandinavian Journal of Plastic and Reconstructive Surgery and Hand Surgery*, vol. 40, no. 2, pp. 73–78, 2006.
- [34] H. Fidan, O. Sahin, Y. Yavuz et al., "Caffeic acid phenethyl ester reduces mortality and sepsis-induced lung injury in rats," *Critical Care Medicine*, vol. 35, no. 12, pp. 2822–2829, 2007.
- [35] C. Ara, A. Dirican, S. Erdoğan et al., "The effect of caffeic acid phenethyl ester on bacterial translocation and intestinal damage after intestinal obstruction," *Turkish Journal of Medical Sciences*, vol. 40, no. 6, pp. 897–903, 2010.
- [36] O. G. Yildiz, S. Soyuer, R. Saraymen, and C. Eroglu, "Protective effects of caffeic acid phenethyl ester on radiation induced lung injury in rats," *Clinical and Investigative Medicine*, vol. 31, no. 5, pp. E242–E247, 2008.
- [37] J. J. Haddad and C. S. Fahlman, "Nuclear factor- κ B-independent regulation of lipopolysaccharide-mediated interleukin-6 biosynthesis," *Biochemical and Biophysical Research Communications*, vol. 291, no. 4, pp. 1045–1051, 2002.
- [38] F. Armutcu, A. Gürel, M. Hoşnuter, O. Pabuçcu, and C. Altinyazar, "Caffeic acid phenethyl ester improves oxidative erythrocyte damage in a rat model of thermal injury," *Journal of Burn Care & Rehabilitation*, vol. 25, no. 2, pp. 171–178, 2004.
- [39] A. Gürel, F. Armutcu, M. Hosnuter, M. Unalacak, E. Kargi, and C. Altinyazar, "Caffeic acid phenethyl ester improves oxidative organ damage in rat model of thermal trauma," *Physiological Research*, vol. 53, no. 6, pp. 675–682, 2004.
- [40] O. Koksel, M. B. Kaplan, A. Ozdulger et al., "Oleic acid-induced lung injury in rats and effects of caffeic acid phenethyl ester," *Experimental Lung Research*, vol. 31, no. 5, pp. 483–496, 2005.
- [41] M. K. Ozer, H. Parlakpınar, N. Vardi, Y. Cigremis, M. Ucar, and A. Acet, "Myocardial ischemia/reperfusion-induced oxidative renal damage in rats: protection by caffeic acid phenethyl ester (CAPE)," *Shock*, vol. 24, no. 1, pp. 97–100, 2005.
- [42] A. M. Mohamadin, L. N. A. Hammad, M. F. El-Bab, and H. S. Abdel Gawad, "Attenuation of oxidative stress in plasma and tissues of rats with experimentally induced hyperthyroidism by caffeic acid phenylethyl ester," *Basic and Clinical Pharmacology and Toxicology*, vol. 100, no. 2, pp. 84–90, 2007.
- [43] H. Ozyurt, B. Ozyurt, K. Koca, and S. Ozgocmen, "Caffeic acid phenethyl ester (CAPE) protects rat skeletal muscle against ischemia-reperfusion-induced oxidative stress," *Vascular Pharmacology*, vol. 47, no. 2-3, pp. 108–112, 2007.
- [44] S. C. Barber, A. Higginbottom, R. J. Mead, S. Barber, and P. J. Shaw, "An in vitro screening cascade to identify neuroprotective antioxidants in ALS," *Free Radical Biology and Medicine*, vol. 46, no. 8, pp. 1127–1138, 2009.
- [45] B. Ozyurt, H. Ozyurt, N. Akpolat, H. Erdogan, and M. Sarsilmaz, "Oxidative stress in prefrontal cortex of rat exposed to MK-801 and protective effects of CAPE," *Progress in Neuro-Psychopharmacology and Biological Psychiatry*, vol. 31, no. 4, pp. 832–838, 2007.
- [46] A. A. Korish and M. M. Arafa, "Propolis derivatives inhibit the systemic inflammatory response and protect hepatic and neuronal cells in acute septic shock," *Brazilian Journal of Infectious Diseases*, vol. 15, no. 4, pp. 332–338, 2011.
- [47] Z. Teke, E. B. Bostanci, C. Yenisey et al., "Effects of caffeic acid phenethyl ester on anastomotic healing in secondary peritonitis," *Journal of Investigative Surgery*, vol. 25, no. 5, pp. 301–310, 2012.