

Greglist: a database listing potential G-quadruplex regulated genes

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ABSTRACT

The double helix is a conformation that genomic DNA usually assumes; under certain conditions, however, guanine-rich DNA sequences can form a four-stranded structure, G-quadruplex, which is found to play a role in regulating gene expression. Indeed, it has been demonstrated that the G-quadruplex formed in the *c-MYC* promoter suppresses its transcriptional activity. Recent studies suggest that G-quadruplex motifs (GQMs) are enriched in human gene promoters. To facilitate the research of G-quadruplex, we have constructed Greglist, a database listing potentially G-quadruplex regulated genes. Greglist harbors genes that contain promoter GQMs from genomes of various species, including humans, mice, rats and chickens. Many important genes are found to contain previously unreported promoter GQMs, such as *ATM*, *BAD*, *AKT1*, *LEPR*, *UCP1*, *APOE*, *DKK1*, *WT1*, *WEE1*, *WNT1* and *CLOCK*. Furthermore, we find that not only protein coding genes, 126 human microRNAs also contain promoter GQMs. Greglist therefore provides candidates for further studying G-quadruplex functions and is freely available at <http://tubic.tju.edu.cn/greglist>.

INTRODUCTION

The double helix structure is a conformation that genomic DNA usually assumes; however, DNA can form other non-classical structures as well (1). For instance, under certain conditions, guanine-rich DNA sequences can form a special structure called G-quadruplex. The discovery of G-quadruplex can be traced back to G-quartets, planar arrays of four guanines held together by hydrogen bonds, which were found by Davies and coworkers (2) about 5 decades ago. Later Sen and Gilbert (3) discovered G-quadruplex, a four-stranded structure that is stabilized by G-quartets. As an example, readers may visit

www.rcsb.org to view the 3-dimensional (3D) structure of a G-quadruplex (PDB code: 1XAV), which is formed in the promoter regions of the *c-MYC* gene (4). Sequences with high potential to form G-quadruplex have been found in many different genomic regions, suggesting diverse roles of G-quadruplexes (5–11). For instance, telomeric repeats in virtually all eukaryotes have the ability to form G-quadruplexes (10,11), offering a protection for the telomere 3' overhang (12,13), which is essential for cell survival.

Recent interests on G-quadruplexes have been focused on its role in transcriptional regulation. By using electron microscopy, Maizels and coworkers (14) observed that the G-quadruplex structure is formed cotranscriptionally *in vivo*. Indeed, Hurley and coworkers have demonstrated that the region upstream of the *c-MYC* promoter forms a G-quadruplex, removal of which results in an increase, whereas its stabilization results in a decrease in basal transcriptional activity of this promoter, suggesting promoter G-quadruplexes as transcriptional repressor elements (15).

Sequences containing G-quadruplex motifs (GQMs) in promoter regions have only been reported for about 10 genes, including *c-MYC* (15–17), *VEGF* (18), *BCL-2* (19), *c-KIT* (20,21) and some others (22,23). Recent bioinformatics studies, however, showed that GQMs are prevalent in the human genome (24,25). Furthermore, GQMs were found to be highly enriched in human gene promoters with more than 40% promoters containing at least 1 GQM (26).

To facilitate the study of the role of promoter G-quadruplexes, we constructed Greglist, a database listing potential G-quadruplex REGULATED Genes, i.e. genes that contain promoter GQMs. The database provides detailed information about the number, the position and the sequence of promoter GQMs from genes of various species. Many important genes are found to contain previously unreported promoter GQMs, such as *ATM*, *BAD*, *AKT1*, *LEPR*, *UCP1*, *APOE*, *DKK1*, *WT1*, *WEE1*, *WNT1* and *CLOCK*. Furthermore, we found that not only protein coding genes, 126 human microRNAs also contain promoter GQMs. Greglist contains

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Table 1. Descriptive statistics of genes in Greglist

Species (Latin name)	Species	Genome version	Number of genes having promoter GQMs	Total gene number	Percentage of genes having promoter GQMs	Average GQMs a gene has	GQM density in promoter regions (GQMs/Kb)	Average GQM length (mean \pm SD)
<i>Homo sapiens</i>	Human	NCBI36	10 277	31 524	32.60%	1.93	0.63	29.19 \pm 13.57
<i>Mus musculus</i>	Mouse	NCBIM36	8962	28 390	31.57%	1.61	0.51	28.18 \pm 12.57
<i>Rattus norvegicus</i>	Rat	RGSC3.4	7013	27 302	25.69%	1.43	0.37	26.39 \pm 8.82
<i>Gallus gallus</i>	Chicken	WASHUC2	5949	17 438	34.12%	1.75	0.60	28.70 \pm 14.44

candidates for further studying G-quadruplex functions and is another device added to the existing online G-quadruplex toolbox.

DATABASE CONSTRUCTION AND DESCRIPTION

Greglist of the current version contains genes that have promoter GQMs in the genomes of human, mouse, rat and chicken. Table 1 provides a descriptive statistics of the content of the database. We generally defined sequences 1 kb upstream of transcription start site (TSS) as promoter regions. These sequences were downloaded from Ensembl using the software BioMart. The dataset used was Ensembl 45 and human, mouse, rat and chicken genome sequences were based on the versions of NCBI36, NCBIM36, RGSC3.4 and WASHUC2, respectively. The software Quadparser (26) was used to find the promoter GQM, which is $G_3+N_{1-7}G_3+N_{1-7}G_3+N_{1-7}G_3+$, where N denotes any nucleotide. In addition, the G-quadruplex structure can be formed on either of the two DNA strands; therefore the motif of $C_3+N_{1-7}C_3+N_{1-7}C_3+N_{1-7}C_3+$ was also used, which suggests the capability of the G-quadruplex formation on the complementary strand.

So far, only about 10 genes have been reported to contain promoter GQMs. In Greglist, however, a lot more genes that contain promoter GQMs are listed. For instance, these genes include *ATM*, *BAD*, *AKT1*, *LEPR*, *UCP1*, *APOE*, *DKK1*, *WT1*, *WEE1*, *WNT1*, *CLOCK*, *ATF1* and *BMP2*, which have critical functions in various cellular processes, such as apoptosis and transcriptional regulation. Table 2 lists a sample of 30 genes that contain promoter GQMs with the position of GQMs and gene functions.

In addition, we found that not only protein coding genes, many microRNAs, such as hsa-mir-639 and hsa-mir-381, also contain promoter GQMs. Totally 126 human microRNAs were found to have promoter GQMs. To get a full list of these microRNAs, refer to the Supplementary Table 1. MicroRNAs have emerged as important regulators of gene expression. The finding that promoter regions of microRNA genes contain GQMs necessitates further studies to address the role of G-quadruplexes in microRNA regulation.

Of note, the presence of a GQM only suggests the potential of a sequence to form G-quadruplex. In addition, the G-quadruplex structure is a dynamic structure that is formed upon denaturation of the DNA duplex. Therefore caution must be taken to interpret the data in

Greglist. In other words, gene records in Greglist provide a starting point for further analysis of the potential G-quadruplex structure in these genes. Furthermore, Huppert *et al.* (26) reported that more than 40% of human genes contain promoter GQMs, however, in Greglist, ~32% human genes do. This is likely because in Ref. (26), only less than 20 000 known genes were used, whereas in the current study, more than 30 000 human genes, including those classified as novel and those encode RNAs were included. Therefore, Greglist is made to be inclusive, not exclusive.

Gene names, Ensembl IDs, RefSeq IDs, numbers of GQMs, distance of the GQM to TSS, functional description of gene ontology, sequences containing the GQM and coding sequences of the gene, were extracted from Ensembl database and Quadparser output files. All the data were then organized by using an open-source management system, MySQL, which allows rapid data retrieval. All gene records have been linked directly to corresponding entries in Ensembl. Users can browse each entry or download all records. Because of the large volume of data, a good searching function is important for this database. In Greglist, users can perform searches by inputting gene accession numbers or names at the homepage, and then click 'Go'. To perform more detailed searches, users can click 'Search', and then in the new page, more detailed searching options are provided. For instance, users can search by gene ontology terms to get a list of genes that have desired functions. To further facilitate searching the gene of interest, we installed Blast program locally. So users can input the coding sequence of their gene of interest and perform Blast searches to find homologous ones.

Many online resources for G-quadruplexes are available. These include G4P calculator (14), QGRS Mapper (27), Quadfinder (28), which are online programs or web servers for predicting G-quadruplexes. GRSDB (29) is a database of quadruplex forming G-rich sequences in alternatively processed mammalian pre-mRNA sequences. Greglist is another device added to the existing online G-quadruplex toolbox.

We plan to include more species in future versions of Greglist. In addition, with the availability of more experimental data, we plan to integrate experimental evidence in corresponding entries. Furthermore, although the GQM used in Quadparser is quite commonly used, there are other motifs that have potential to form G-quadruplexes, and we also plan to include these motifs in future versions of the database. We welcome

Table 2. A list of 30 human genes that have not been previously reported to contain promoter G-quadruplex motifs

No.	Abbreviation	Gene name	Ensembl ID	Function or associated disease	Reference	Number of GQM	Distance to TSS
1	WNT1	Wingless-type MMTV integration site family, member 1	ENSG00000125084	The Wnt signaling pathway, CNS development	(30)	1	193
2	WNT5A	Wingless-type MMTV integration site family, member 5A	ENSG00000114251	The Wnt signaling pathway, vertebrate development	(31)	2	567, 936
3	LEPR	LEPTIN receptor	ENSG00000116678	Energy metabolism	(32)	3	310, 372, 495
4	UCP1	Uncoupling protein 1	ENSG00000109424	Energy metabolism	(33)	2	89, 224
5	APOE	Apolipoprotein E	ENSG00000130203	Alzheimer's disease	(34)	4	46, 65, 407, 739
6	ATM	Ataxia telangiectasia mutated	ENSG00000149311	Ataxia telangiectasia	(35)	1	59
7	PAX8	Paired box gene 8	ENSG00000125618	Permanent congenital hypothyroidism	(36)	1	133
8	SOX1	SRY (sex determining region Y)-box 1	ENSG00000203883	Lens development	(37)	3	80, 726, 826
9	SOX10	SRY (sex determining region Y)-box 10	ENSG00000100146	Waardenburg-Hirschsprung disease	(38)	2	130, 313
10	HDAC1	Histone deacetylase 1	ENSG00000116478	Histone modification	(39)	1	34
11	TGFβ1	Transforming growth factor, beta 1	ENSG00000105329	TGFβ signaling	(40)	1	151
12	SMAD2	MAD homolog 2	ENSG00000175387	TGFβ signaling	(41)	2	235, 450
13	DKK1	Dickkopf homolog 1	ENSG00000107984	TGFβ signaling	(42)	1	136
14	CLOCK	Clock homolog	ENSG00000134852	Circadian rhythms	(43)	3	147, 341, 692
15	WEE1	WEE1 homolog	ENSG00000166483	Cell cycle control	(44)	1	542
16	BAD	BCL2-antagonist of cell death	ENSG00000002330	Apoptosis	(45)	3	116, 628, 756
17	AKT1	V-akt murine thymoma viral oncogene homolog 1	ENSG00000142208	Apoptosis	(46)	1	61
18	GATA4	GATA-binding protein 4	ENSG00000136574	Heart development	(47)	1	314
19	MYOD1	Myogenic differentiation 1	ENSG00000129152	Muscle development	(48)	2	128, 216
20	WT1	Wilms tumor 1	ENSG00000184937	Kidney development	(49)	2	168, 900
21	GDF1	Growth differentiation factor 1	ENSG00000135414	Left-right patterning	(50)	4	78, 166, 327, 766
22	BMP2	Bone morphogenetic protein 2	ENSG00000125845	Bone development	(51)	1	163
23	MEF2D	MADS box transcription enhancer factor 2D	ENSG00000116604	Heart development	(52)	4	18, 85, 169, 232
24	STAT6	Signal transducer and activator of transcription 6	ENSG00000166888	Immunity	(53)	1	505
25	SOCS1	Suppressor of cytokine signaling 1	ENSG00000185338	Immunity	(54)	5	112, 211, 534, 578, 758
26	MMP2	Matrix metalloproteinase 2	ENSG00000167346	Function of extracellular matrix	(55)	1	576
27	MAPK2	Mitogen-activated protein kinase 2	ENSG00000162889	MAP kinase pathway	(56)	2	100, 137
28	ATF1	Activating transcription factor 1	ENSG00000123268	Transcriptional regulation	(57)	1	36
29	TAF2	TAF2 RNA polymerase II	ENSG00000064313	Transcriptional regulation	(58)	1	296
30	RING1	Ring finger protein 1	ENSG00000204227	Transcriptional regulation	(59)	4	501, 559, 677, 938

users' comments, corrections and new information, which will be used for updating.

Greglist is freely available at the website: <http://tubic.tju.edu.cn/greglist>, and should be cited with the present publication as reference.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

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REFERENCES

- Perez-Martin, J. and de Lorenzo, V. (1997) Clues and consequences of DNA bending in transcription. *Annu. Rev. Microbiol.*, **51**, 593–628.
- Gellert, M., Lipsett, M.N. and Davies, D.R. (1962) Helix formation by guanylic acid. *Proc. Natl Acad. Sci. USA*, **48**, 2013–2018.
- Sen, D. and Gilbert, W. (1988) Formation of parallel four-stranded complexes by guanine-rich motifs in DNA and its implications for meiosis. *Nature*, **334**, 364–366.
- Ambrus, A., Chen, D., Dai, J., Jones, R.A. and Yang, D. (2005) Solution structure of the biologically relevant G-quadruplex element in the human c-MYC promoter. Implications for G-quadruplex stabilization. *Biochemistry*, **44**, 2048–2058.
- Simonsson, T. (2001) G-quadruplex DNA structures—variations on a theme. *Biol. Chem.*, **382**, 621–628.
- Gilbert, D.E. and Feigon, J. (1999) Multistranded DNA structures. *Curr. Opin. Struct. Biol.*, **9**, 305–314.
- Maizels, N. (2006) Dynamic roles for G4 DNA in the biology of eukaryotic cells. *Nat. Struct. Mol. Biol.*, **13**, 1055–1059.
- Han, H. and Hurley, L.H. (2000) G-quadruplex DNA: a potential target for anti-cancer drug design. *Trends Pharmacol. Sci.*, **21**, 136–142.
- Burge, S., Parkinson, G.N., Hazel, P., Todd, A.K. and Neidle, S. (2006) Quadruplex DNA: sequence, topology and structure. *Nucleic Acids Res.*, **34**, 5402–5415.
- Blackburn, E.H. (1991) Structure and function of telomeres. *Nature*, **350**, 569–573.
- Williamson, J.R. (1994) G-quartet structures in telomeric DNA. *Annu. Rev. Biophys. Biomol. Struct.*, **23**, 703–730.
- Henderson, E., Hardin, C.C., Walk, S.K., Tinoco, I. Jr and Blackburn, E.H. (1987) Telomeric DNA oligonucleotides form novel intramolecular structures containing guanine-guanine base pairs. *Cell*, **51**, 899–908.
- Sundquist, W.I. and Klug, A. (1989) Telomeric DNA dimerizes by formation of guanine tetrads between hairpin loops. *Nature*, **342**, 825–829.
- Duquette, M.L., Handa, P., Vincent, J.A., Taylor, A.F. and Maizels, N. (2004) Intracellular transcription of G-rich DNAs induces formation of G-loops, novel structures containing G4 DNA. *Genes Dev.*, **18**, 1618–1629.
- Siddiqui-Jain, A., Grand, C.L., Bearss, D.J. and Hurley, L.H. (2002) Direct evidence for a G-quadruplex in a promoter region and its targeting with a small molecule to repress c-MYC transcription. *Proc. Natl Acad. Sci. USA*, **99**, 11593–11598.
- Simonsson, T., Pecinka, P. and Kubista, M. (1998) DNA tetraplex formation in the control region of c-myc. *Nucleic Acids Res.*, **26**, 1167–1172.
- Grand, C.L., Han, H., Munoz, R.M., Weitman, S., Von Hoff, D.D., Hurley, L.H. and Bearss, D.J. (2002) The cationic porphyrin TMPyP4 down-regulates c-MYC and human telomerase reverse transcriptase expression and inhibits tumor growth in vivo. *Mol. Cancer Ther.*, **1**, 565–573.
- Sun, D., Guo, K., Rusche, J.J. and Hurley, L.H. (2005) Facilitation of a structural transition in the polypurine/polypyrimidine tract within the proximal promoter region of the human VEGF gene by the presence of potassium and G-quadruplex-interactive agents. *Nucleic Acids Res.*, **33**, 6070–6080.
- Dai, J., Dexheimer, T.S., Chen, D., Carver, M., Ambrus, A., Jones, R.A. and Yang, D. (2006) An intramolecular G-quadruplex structure with mixed parallel/antiparallel G-strands formed in the human BCL-2 promoter region in solution. *J. Am. Chem. Soc.*, **128**, 1096–1098.
- Rankin, S., Reszka, A.P., Huppert, J., Zloh, M., Parkinson, G.N., Todd, A.K., Ladame, S., Balasubramanian, S. and Neidle, S. (2005) Putative DNA quadruplex formation within the human c-kit oncogene. *J. Am. Chem. Soc.*, **127**, 10584–10589.
- Fernando, H., Reszka, A.P., Huppert, J., Ladame, S., Rankin, S., Venkitaraman, A.R., Neidle, S. and Balasubramanian, S. (2006) A conserved quadruplex motif located in a transcription activation site of the human c-kit oncogene. *Biochemistry*, **45**, 7854–7860.
- Howell, R.M., Woodford, K.J., Weitzmann, M.N. and Usdin, K. (1996) The chicken beta-globin gene promoter forms a novel "cinched" tetrahelical structure. *J. Biol. Chem.*, **271**, 5208–5214.
- De Armond, R., Wood, S., Sun, D., Hurley, L.H. and Ebbinghaus, S.W. (2005) Evidence for the presence of a guanine quadruplex forming region within a polypurine tract of the hypoxia inducible factor 1 alpha promoter. *Biochemistry*, **44**, 16341–16350.
- Huppert, J.L. and Balasubramanian, S. (2005) Prevalence of quadruplexes in the human genome. *Nucleic Acids Res.*, **33**, 2908–2916.
- Todd, A.K., Johnston, M. and Neidle, S. (2005) Highly prevalent putative quadruplex sequence motifs in human DNA. *Nucleic Acids Res.*, **33**, 2901–2907.
- Huppert, J.L. and Balasubramanian, S. (2007) G-quadruplexes in promoters throughout the human genome. *Nucleic Acids Res.*, **35**, 406–413.
- Kikin, O., D'Antonio, L. and Bagga, P.S. (2006) QGRS Mapper: a web-based server for predicting G-quadruplexes in nucleotide sequences. *Nucleic Acids Res.*, **34**, W676–W682.
- Scaria, V., Hariharan, M., Arora, A. and Maiti, S. (2006) Quadfinder: server for identification and analysis of quadruplex-forming motifs in nucleotide sequences. *Nucleic Acids Res.*, **34**, W683–W685.
- Kostadinov, R., Malhotra, N., Viotti, M., Shine, R., D'Antonio, L. and Bagga, P. (2006) GRSDb: a database of quadruplex forming G-rich sequences in alternatively processed mammalian pre-mRNA sequences. *Nucleic Acids Res.*, **34**, D119–D124.
- Megason, S.G. and McMahon, A.P. (2002) A mitogen gradient of dorsal midline Wnts organizes growth in the CNS. *Development*, **129**, 2087–2098.
- Yamaguchi, T.P., Bradley, A., McMahon, A.P. and Jones, S. (1999) A Wnt5a pathway underlies outgrowth of multiple structures in the vertebrate embryo. *Development*, **126**, 1211–1223.
- Tartaglia, L.A., Dembski, M., Weng, X., Deng, N., Culpepper, J., Devos, R., Richards, G.J., Campfield, L.A., Clark, F.T. et al. (1995) Identification and expression cloning of a leptin receptor, OB-R. *Cell*, **83**, 1263–1271.
- Ricquier, D. and Bouillaud, F. (2000) The uncoupling protein homologues: UCP1, UCP2, UCP3, StUCP and AtUCP. *Biochem J.*, **345**(Pt 2), 161–179.
- Corder, E.H., Saunders, A.M., Strittmatter, W.J., Schmechel, D.E., Gaskell, P.C., Small, G.W., Roses, A.D., Haines, J.L. and Pericak-Vance, M.A. (1993) Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science*, **261**, 921–923.
- Savitsky, K., Bar-Shira, A., Gilad, S., Rotman, G., Ziv, Y., Vanagaite, L., Tagle, D.A., Smith, S., Uziel, T. et al. (1995) A single ataxia telangiectasia gene with a product similar to PI-3 kinase. *Science*, **268**, 1749–1753.
- Macchia, P.E., Lapi, P., Krude, H., Pirro, M.T., Missero, C., Chiovato, L., Souabni, A., Baserga, M., Tassi, V. et al. (1998) PAX8 mutations associated with congenital hypothyroidism caused by thyroid dysgenesis. *Nat. Genet.*, **19**, 83–86.
- Nishiguchi, S., Wood, H., Kondoh, H., Lovell-Badge, R. and Episkopou, V. (1998) Sox1 directly regulates the gamma-crystallin genes and is essential for lens development in mice. *Genes Dev.*, **12**, 776–781.
- Pingault, V., Bondurand, N., Kuhlbrodt, K., Goerich, D.E., Prehu, M.O., Puliti, A., Herbarth, B., Hermans-Borgmeyer, I., Legius, E. et al. (1998) SOX10 mutations in patients with Waardenburg-Hirschsprung disease. *Nat. Genet.*, **18**, 171–173.
- Brehm, A., Miska, E.A., McCance, D.J., Reid, J.L., Bannister, A.J. and Kouzarides, T. (1998) Retinoblastoma protein recruits histone deacetylase to repress transcription. *Nature*, **391**, 597–601.
- Grainger, D.J., Heathcote, K., Chiano, M., Snieder, H., Kemp, P.R., Metcalfe, J.C., Carter, N.D. and Spector, T.D. (1999) Genetic control of the circulating concentration of transforming growth factor type beta1. *Hum. Mol. Genet.*, **8**, 93–97.
- Labbe, E., Silvestri, C., Hoodless, P.A., Wrana, J.L. and Attisano, L. (1998) Smad2 and Smad3 positively and negatively regulate TGF beta-dependent transcription through the forkhead DNA-binding protein FAST2. *Mol. Cell*, **2**, 109–120.
- Mao, B., Wu, W., Li, Y., Hoppe, D., Stanek, P., Glinka, A. and Niehrs, C. (2001) LDL-receptor-related protein 6 is a receptor for Dickkopf proteins. *Nature*, **411**, 321–325.
- Reppert, S.M. and Weaver, D.R. (2002) Coordination of circadian timing in mammals. *Nature*, **418**, 935–941.

44. Russell,P. and Nurse,P. (1987) Negative regulation of mitosis by *wee1+*, a gene encoding a protein kinase homolog. *Cell*, **49**, 559–567.
45. Datta,S.R., Dudek,H., Tao,X., Masters,S., Fu,H., Gotoh,Y. and Greenberg,M.E. (1997) Akt phosphorylation of BAD couples survival signals to the cell-intrinsic death machinery. *Cell*, **91**, 231–241.
46. Nicholson,K.M. and Anderson,N.G. (2002) The protein kinase B/Akt signalling pathway in human malignancy. *Cell. Signal.*, **14**, 381–395.
47. Molkentin,J.D., Lin,Q., Duncan,S.A. and Olson,E.N. (1997) Requirement of the transcription factor GATA4 for heart tube formation and ventral morphogenesis. *Genes Dev.*, **11**, 1061–1072.
48. Tapscott,S.J., Davis,R.L., Thayer,M.J., Cheng,P.F., Weintraub,H. and Lassar,A.B. (1988) MyoD1: a nuclear phosphoprotein requiring a Myc homology region to convert fibroblasts to myoblasts. *Science*, **242**, 405–411.
49. Kreidberg,J.A., Sariola,H., Loring,J.M., Maeda,M., Pelletier,J., Housman,D. and Jaenisch,R. (1993) WT-1 is required for early kidney development. *Cell*, **74**, 679–691.
50. Rankin,C.T., Bunton,T., Lawler,A.M. and Lee,S.J. (2000) Regulation of left-right patterning in mice by growth/differentiation factor-1. *Nat. Genet.*, **24**, 262–265.
51. Hogan,B.L. (1996) Bone morphogenetic proteins: multifunctional regulators of vertebrate development. *Genes Dev.*, **10**, 1580–1594.
52. Edmondson,D.G., Lyons,G.E., Martin,J.F. and Olson,E.N. (1994) *Mef2* gene expression marks the cardiac and skeletal muscle lineages during mouse embryogenesis. *Development*, **120**, 1251–1263.
53. Takeda,K., Tanaka,T., Shi,W., Matsumoto,M., Minami,M., Kashiwamura,S., Nakanishi,K., Yoshida,N., Kishimoto,T. and Akira,S. (1996) Essential role of Stat6 in IL-4 signalling. *Nature*, **380**, 627–630.
54. Alexander,W.S., Starr,R., Fenner,J.E., Scott,C.L., Handman,E., Sprigg,N.S., Corbin,J.E., Cornish,A.L., Darwiche,R. *et al.* (1999) SOCS1 is a critical inhibitor of interferon gamma signaling and prevents the potentially fatal neonatal actions of this cytokine. *Cell*, **98**, 597–608.
55. Giannelli,G., Falk-Marzillier,J., Schiraldi,O., Stetler-Stevenson,W.G. and Quaranta,V. (1997) Induction of cell migration by matrix metalloprotease-2 cleavage of laminin-5. *Science*, **277**, 225–228.
56. Jordan,J.D., Carey,K.D., Stork,P.J. and Iyengar,R. (1999) Modulation of rap activity by direct interaction of Galpha(o) with Rap1 GTPase-activating protein. *J. Biol. Chem.*, **274**, 21507–21510.
57. Hummler,E., Cole,T.J., Blendy,J.A., Ganss,R., Aguzzi,A., Schmid,W., Beermann,F. and Schutz,G. (1994) Targeted mutation of the CREB gene: compensation within the CREB/ATF family of transcription factors. *Proc. Natl Acad. Sci. USA*, **91**, 5647–5651.
58. Tzukerman,M.T., Esty,A., Santiso-Mere,D., Danielian,P., Parker,M.G., Stein,R.B., Pike,J.W. and McDonnell,D.P. (1994) Human estrogen receptor transactivational capacity is determined by both cellular and promoter context and mediated by two functionally distinct intramolecular regions. *Mol. Endocrinol.*, **8**, 21–30.
59. Satijn,D.P., Gunster,M.J., van der Vlag,J., Hamer,K.M., Schul,W., Alkema,M.J., Saurin,A.J., Freemont,P.S., van Driel,R. *et al.* (1997) RING1 is associated with the polycomb group protein complex and acts as a transcriptional repressor. *Mol. Cell. Biol.*, **17**, 4105–4113.