CLINICAL STUDY

OPEN ACCESS Check for updates

Taylor & Francis

Taylor & Francis Group

Helicobacter pylori infection is associated with elevated galactose-deficient IgA1 in IgA nephropathy

Xing-Zi Liu^{a,b}, Yue-Miao Zhang^{a,b}, Ni-Ya Jia^{a,b} and Hong Zhang^{a,b}

^aRenal Division, Peking University First Hospital, Peking University Institute of Nephrology, Beijing, People's Republic of China; ^bKey Laboratory of Renal Disease, Ministry of Health of China, Key Laboratory of Chronic Kidney Disease Prevention and Treatment (Peking University), Ministry of Education, Beijing, People's Republic of China

ABSTRACT

Background: Mucosal immunity plays an important role in the pathogenesis of IgA nephropathy (IgAN). This study aimed to investigate if infection of *Helicobacter pylori* (*H. pylori*), a common bacteria in the gastrointestinal tract, associated with IgAN.

Methods: This study included 261 patients with IgAN and 46 healthy controls. Clinical information and plasma samples were collected from patients and healthy controls. *H. pylori* infection was confirmed by western blot. Plasma IgA1 and galactose-deficient IgA1 (Gd-IgA1) levels were detected by specific enzyme-linked immunosorbent assay.

Results: Total *H. pylori* infection rates showed no statistical differences between IgAN patients and healthy controls, but the infection rates of type I *H. pylori* in IgAN patients were significantly higher than those in healthy controls (44.4 vs. 28.3%, p = 0.040). Compared with uninfected patients, the systolic blood pressure, 24-h proteinuria, and blood urea nitrogen levels were significantly higher in patients with *H. pylori* infection (126.0±15.5 vs. 119.6±14.5 mmHg, p = 0.010; 1.8±2.7 vs. 1.2±1.4 g/24h, p = 0.013; 7.9±5.4 vs. $6.7 \pm 3.9 \,\mu$ mol/L, p = 0.042), especially in patients with type I infection (126.5±15.4 vs. 119.6±14.5 mmHg, p = 0.002; 1.9 ± 2.9 vs. $1.2 \pm 1.4 \,\text{g}/24$ h, p = 0.033; 8.1 ± 5.6 vs. $6.7 \pm 3.9 \,\mu$ mol/L, p = 0.041). Similarly, patients with IgAN and type I *H. pylori* infection showed higher plasma Gd-IgA1 levels than uninfected patients (5.5±2.2 vs. $4.5 \pm 2.2 \,\mu$ g/mL, p = 0.037).

Conclusions: Virulent type I *H. pylori* infection is more common in patients with IgAN. Patients with IgAN and type I *H. pylori* infection showed lower renal function and higher underglycosylation of plasma IgA1.

ARTICLE HISTORY

Received 1 March 2020 Revised 14 May 2020 Accepted 14 May 2020

KEYWORDS

Immunoglobulin A nephropathy; *Helicobacter pylori*; cytotoxin-associated gene A/vacuolating cytotoxin A; galactosedeficient IgA1

Introduction

Immunoglobulin A nephropathy (IgAN), the most common cause of glomerulonephritis worldwide, is characterized by the predominant or codominant deposition of IgA in the glomerular mesangium [1]. Increasing evidences suggested that mucosal immunity played an important role in the pathogenesis of IgAN [2,3], and *Helicobacter pylori* (*H. pylori*) infection might be the most common factor [4,5]; however, the associations between *H. pylori* infection and clinical manifestations of IgAN and its possible mechanism have not been elucidated yet.

H. pylori, a gram-negative bacterium, colonizes the human gastric mucus layer. Recently, considerable studies changed the paradigm that *H. pylori* only participates in the pathogenesis of chronic gastritis and peptic ulcer disease, and demonstrated that several extra-intestinal diseases, such as renal-related diseases, were also caused by *H. pylori* [6–8]. The systemic antibody response to *H. pylori* was reported to be increased in patients with IgAN [9], suggesting that *H. pylori* might be involved in the pathogenesis and progression of IgAN. Cytotoxin-associated gene A (CagA) and vacuolating cytotoxin A (VacA) are the main virulence factors of *H. pylori* [10]. CagA has been observed in the tonsils of most *H. pylori* infected IgAN patients [11], and it was reported to promote glomerular mesangial cell proliferation and extracellular matrix secretion *via* suppressing the apoptotic signaling pathway [12]. VacA was found to play some roles in vacuolation, apoptosis, antigen presentation, and multiple cellular activities [13,14].

CONTACT Hong Zhang Shongzh@bjmu.edu.cn 🗈 Renal Division, Peking University First Hospital, Peking University Institute of Nephrology, No.8 Xi Shi Ku Street, Xi Cheng District Beijing 100034, China

B Supplemental data for this article can be accessed here.

© 2020 Peking University First Hospital.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

However, the association of the virulent *H. pylori* strains and IgAN clinical manifestations and possible mechanism remained to be investigated.

In this study, we investigated the infection rates of different *H. pylori* types, particularly virulent strains, and the correlation between *H. pylori* infection and clinical manifestations in patients with IgAN. Moreover, the levels of plasma IgA1 and galactose-deficient IgA1 (Gd-IgA1) in patients with different *H. pylori* type infection were detected to explore the possible mechanism.

Materials and methods

Study population

A total of 261 patients with IgAN (mean age: 37.7 ± 12.3 years, male ratio: 51.7%) were enrolled in this study from Peking University First Hospital. Forty-six ageand sex-matched healthy individuals (mean age: 38.0 ± 12.2 years, male ratio: 47.8%) were recruited as healthy controls. The patients with IgAN were diagnosed by renal biopsy and confirmed by the deposition of IgA in the glomerular mesangium using immunofluorescence and ultrastructural examination. Patients with secondary causes of IgAN, such as IgA vasculitis, systemic lupus erythematosus, or liver cirrhosis were excluded.

Plasma samples were collected from patients (at renal biopsy) and healthy controls and stored at -80 °C for further use. Baseline demographic and clinical data were collected at the time of renal biopsy, including age, sex, systolic blood pressure (SBP), diastolic blood pressure (DBP), gross/microscopic hematuria, serum creatinine (Scr), serum IgA/IgG/IgM, serum complement 3 (C3), 24-h proteinuria, blood urea nitrogen (BUN), treatment regimes, and histological characteristics. Estimated glomerular filtration rate (eGFR) was calculated using the Chronic Kidney Disease Epidemiology Collaboration equation [15]. Twenty-four hour creatinine clearance (CrCl) was done by the Cockcroft–Gault formula [16]. The histological characteristics were scored according to the Oxford classification [17].

Detection of H. pylori infection

Serum *H. pylori*-IgG antibodies (CagA, VacA, UreA, and UreB) were detected in all participants using the Typing Detection Kit (Blot Biotech, Shenzhen, China), according to the manufacturer's protocol. Seropositivity for UreA and/or UreB IgG confirmed *H. pylori* infection. Type I *H. pylori* infection was diagnosed if seropositivity for CagA and/or VacA IgG, otherwise, the type II *H. pylori* infection was diagnosed. The diagnosis of *H. pylori* infection is summarized in Supplementary Table 1.

Detection of plasma IgA1 and galactosedeficient IgA1

The levels of plasma IgA1 in patients with IgAN were detected by enzyme-linked immunosorbent assay (ELISA), according to a previously described protocol [18]. Briefly, 96-well plates were coated with $F(ab')_2$ fragments of goat IgG anti-human IgA overnight at 4°C, followed by blocking with 1% bovine serum albumin for 1 h at 37 °C. Next, plasma samples (1:80,000 dilution) and standard samples were added to the 96well plates and incubated for 1 h at 37 °C, followed by treatment with horseradish peroxidase-conjugated monoclonal anti-human IgA1 antibodies and tetramethylbenzidine liquid substrate. The absorbance was then measured at 450/570 nm with a microplate reader (Bio-Rad, Japan). Plasma Gd-IgA1 levels were detected using the KM55 ELISA kit (IBL, Japan) [19,20]. Briefly, the ELISA plates were incubated with plasma samples (1:400 dilution in EIA buffer) and standard samples for 1 h at 37 °C, washed four times with wash buffer, incubated with prepared-labeled antibodies, and then treated with 50 µL TMA solution for 30 min in the dark. The absorbance was measured at 450/630 nm by an ELISA reader (Bio-Rad, Japan). The reference range of IgA1 and Gd-IgA1 levels was calculated according to their respective standard curves generated from parallel working standards.

Statistical analysis

Continuous variables with normal distribution were expressed as mean ± standard deviation (SD) and nonnormal variables were presented as median and interquartile range (IQR). Categorical variables were reported as absolute frequencies and percentages. For comparison of normally distributed continuous variables, the independent samples t-test was used. For analysis of non-normally distributed data, Mann–Whitney U test or Kruskal-Wallis test was used. The chi-squared test was performed for comparison of categorical variables. Pearson's correlation and linear regression analyses were used to determine the association between two continuous variables. All results were analyzed by SPSS version 22.0 (SPSS Inc., Chicago, USA) and expressed as hazard ratios with 95% confidence intervals. A two-tailed P value less than 0.05 was considered statistically significant.

Results

The baseline characteristics of patients with IgAN

Two hundred and sixty-one patients with IgAN were enrolled in this study. The baseline characteristics,



Figure 1. The *Helicobacter pylori* infection rates in IgA nephropathy patients and healthy controls. IgAN: IgA nephropathy; *H. pylori*: *Helicobacter pylori*.

including age, sex, SBP, DBP, gross/microscopic hematuria, Scr, serum IgA/IgG/IgM, serum C3, 24-h proteinuria, eGFR, CrCl, BUN, treatment regimes, and histological characteristics, were demonstrated in Supplementary Table 2.

H. pylori infection rates

H. pylori infection rates in patients with IgAN tended to be higher than those in healthy controls (157/261, 60.2% vs. 22/46, 47.8%; p = 0.118). Subgroup analyses showed that there were significant higher type I *H. pylori* infection rates in patients with IgAN, as compared with healthy controls (116/261, 44.4% vs. 13/46, 28.3%; p = 0.040). While, type II *H. pylori* infection rates were comparable between IgAN patients and healthy controls (41/261, 15.7% vs. 9/46, 19.5%, p = 0.514) (Figure 1).

Association between H. pylori infection and clinical manifestations

Compared with uninfected IgAN patients, patients with *H. pylori* infection showed significantly higher SBP (126.0 ± 15.5 vs. 119.6 ± 14.5 mmHg; p = 0.010), 24-h proteinuria (1.8 ± 2.7 vs. 1.2 ± 1.4 g/24h; p = 0.013), and BUN (7.9 ± 5.4 vs. 6.7 ± 3.9 µmol/L, p = 0.042) levels. Further subgroup analyses showed that the above indicators were significantly higher in IgAN patients with

pylori infection (126.5 ± 15.4) type 1 Н. vs. 119.6 ± 14.5 mmHg, p = 0.002; 1.9 ± 2.9 vs. 1.2 ± 1.4 g/ 24h, p = 0.033; 8.1 ± 5.6 vs. 6.7 ± 3.9 μ mol/L, p = 0.041), but not in IgAN patients with type II H. pylori infection $(124.5 \pm 16.0 \text{ vs.} 119.6 \pm 14.5 \text{ mmHg}, p = 0.081; 1.8 \pm 2.0$ vs. $1.2 \pm 1.4 \text{ g}/24\text{h}$, p = 0.088; $7.7 \pm 4.7 \text{ vs. } 6.7 \pm 3.9 \,\mu\text{mol}/$ L, p = 0.211). And no significant differences were observed in age, sex, SBP, DBP, gross/microscopic hematuria, Scr, serum IgA/IgG/IgM, serum C3, 24-h proteinuria, eGFR, CrCl, BUN, treatment regimes, and histological characteristics between patients with type I infection and patients with type II infection (Table 1).

Plasma IgA1 and galactose-deficient IgA1 levels

Plasma IgA1 and Gd-IgA1 levels were both significantly higher in patients with IgAN than those in healthy controls $(3.3 \pm 1.6 \text{ vs. } 2.1 \pm 1.0 \text{ g/L}; p < 0.001; 5.0 \pm 2.2 \text{ vs.}$ $3.9 \pm 2.4 \mu\text{g/mL}, p = 0.033$), which were the common disease characteristics (Figure 2(A)). Plasma Gd-IgA1 levels were significantly higher in patients with type I infection than in uninfected patients $(5.5 \pm 2.2 \text{ vs.}$ $4.5 \pm 2.2 \mu\text{g/mL}, p = 0.037$). But there was no significance between patients with type I and type II *H.pylori* infection $(5.5 \pm 2.2 \text{ vs. } 4.9 \pm 1.9 \mu\text{g/mL}, p = 0.344)$. Conversely, compared with uninfected patients, plasma IgA1 levels were comparable with patients with type I infection $(3.2 \pm 1.7 \text{ vs. } 3.4 \pm 1.7 \text{ g/L}, p = 0.693)$ and patients with type II *H. pylori* infection $(3.2 \pm 1.7 \text{ vs.})$

	Uninfected ($n = 104$)	<i>H. pylori</i> infection $(n = 157)$	Type I <i>H. pylori</i> infection $(n = 116)$	Type II <i>H. pylori</i> infection $(n = 41)$	p ^a value	p ^b value	p ^c value	p ^d value
Age, years	37.1±11.9	38.1 ± 12.5	39.0 ± 13.4	35.6 ± 9.3	0.500	0.258	0.473	0.075
Sex, male, <i>n</i> (%)	50 (48.1)	85 (54.1)	61 (52.6)	24 (58.5)	0.337	0.504	0.257	0.511
SBP, mmHg	119.6 ± 14.5	126.0 ± 15.5	126.5 ± 15.4	124.5 ± 16.0	0.010	0.002	0.081	0.101
DBP, mmHg	75.1 ± 11.3	77.3 ± 10.5	77.2 ± 9.8	77.5 ± 12.4	0.109	0.138	0.257	0.593
Hypertension, <i>n</i> (%)	42 (40.4)	73 (46.5)	59 (50.9)	14 (34.1)	0.330	0.119	0.487	0.065
Gross hematuria, n (%)	26 (25.0)	37 (23.6)	28 (24.1)	9 (22.0)	0.791	0.882	0.699	0.777
Microscopic hematuria, n (%)	87 (83.7)	137 (87.3)	100 (86.2)	37 (90.2)	0.246	0.399	0.221	0.505
Serum lgG, g/L	10.8 ± 3.2	10.9 ± 3.1	11.0 ± 3.3	10.1 ± 2.8	0.916	0.909	0.248	0.397
Serum IgA, g/L	3.3 ± 1.2	3.4 ± 1.1	3.3 ± 1.1	3.4 ± 1.2	0.510	0.674	0.930	0.989
Serum IgM, g/L	1.3 ± 0.7	1.4 ± 3.3	1.5 ± 3.8	1.1 ± 0.6	0.671	0.464	0.231	0.550
Serum C3, g/L	0.9 ± 0.2	0.9 ± 0.2	0.9 ± 0.2	0.9 ± 0.2	0.672	0.736	0.800	0.879
Proteinuria, g/24 h	1.2 ± 1.4	1.8 ± 2.7	1.9 ± 2.9	1.8 ± 2.0	0.013	0.033	0.088	0.592
Scr, µmol/L	126.4 ± 116.0	149.8 ± 144.4	148.5 ± 128.9	153.5 ± 183.0	0.171	0.187	0.292	0.851
eGFR, mL/min/1.73m ²	77.8 ± 36.7	72.3 ± 37.0	71.8 ± 37.8	73.7 ± 34.9	0.240	0.236	0.546	0.803
CrCl, mL/min	82.8 ± 43.4	77.1 ± 40.4	76.9 ± 42.4	77.4 ± 34.9	0.284	0.322	0.477	0.952
BUN, µmol/L	6.7 ± 3.9	7.9 ± 5.4	8.1 ± 5.6	7.7 ± 4.7	0.042	0.041	0.211	0.687
Treatment regimes, <i>n</i> (%)								
ACE inhibitors or ARBs	100 (96.2)	153 (97.5)	114 (98.3)	39 (95.1)	0.551	0.335	0.779	0.271
Immunosuppressive agents	34 (32.7)	48 (30.6)	36 (31.3)	12 (29.3)	0.718	0.792	0.690	0.833
Prednisone	39 (37.5)	52 (33.1)	42 (36.6)	10 (24.4)	0.467	0.843	0.133	0.167
Oxford Score, n (%)								
M0/1	30/74 (28.8/71.2)	53/104 (33.8/66.2)	41/75 (35.3/64.7)	12/29 (29.3/70.7)	0.404	0.303	0.960	0.479
E0/1	66/38 (63.5/36.5)	104/53 (66.2/33.8)	75/41 (64.7/35.3)	29/12 (70.7/29.3)	0.644	0.854	0.407	0.479
S0/1	31/73 (29.8/70.2)	49/108 (31.2/68.8)	40/76 (34.5/65.5)	9/32 (22.0/78.0)	0.810	0.459	0.340	0.132
T0/1/2	54/41/9 (49.0/	72/72/13 (45.9/	50/55/11 (43.1/	22/17/2 (53.7/41.5/4.8)	0.581	0.418	0.741	0.418
	39.4/11.6)	45.9/8.2)	47.4/9.5)					
C0/1/2	86/13/5 (82.7/12.5/4.8)	130/20/7 (82.8/ 127/15/	96/16/7 (80.2/13.8/6.0)	37/4/0 (90.2/9.8/0.0)	066.0	0.876	0.308	0.214
ACE: angiotensin-converting enzym	ne; ARB: angiotensin II recepto	or blocker; BUN: blood urea	nitrogen; C: crescents; CrCl: 24	-h creatinine clearance; DBP: c	liastolic blood pr	ressure; E: Endoo	capillary prolifera	ation; eGFR:
estimated glomerular filtration rate	;; SBP: systolic blood pressure;	Scr: serum creatinine; M: m	esangial hypercellularity; S: seg	mental sclerosis; T: interstitial	fibrosis and tubu	llar atrophy.	- - -	
<i>p</i> value: <i>H. pylori</i> infection group	versus uninfected group; p ² v	alue: type I <i>H. pylori</i> infectio	n group versus unintected gro	up; p^ value: type II H. pylori I	Intection group	/ersus uninfecte	d group; <i>p</i> " valu	ie: type I H.
pylori intection group versus type	ll <i>H. pylori</i> intection group.							

Table 1. Association of *H. pylori* infection and clinical manifestation in patients with IgA nephropathy.



Figure 2. Levels of plasma IgA1 and galactose-deficient IgA1 in participants. (A) The plasma IgA1 and Gd-IgA1 levels in patients with IgAN and healthy controls. (B) The Plasma IgA1 and Gd-IgA1 levels in IgAN patients with or without *H. pylori* infection. (C) The Plasma IgA1 and Gd-IgA1 levels in healthy controls with or without *H. pylori* infection.

3.1 ± 1.4 g/L, p = 0.723). Further correlation analyses showed a significantly positive correlation between plasma lgA1 and Gd-lgA1 levels (correlation coefficient = 0.61, $p = 1.38 \times 10^{-6}$) in patients with lgAN (Figure 2(B)). While, in controls, there were no significant differences both in plasma lgA1 and Gd-lgA1 levels whether between uninfected controls and controls with type I infection $(1.9\pm0.7 \text{ vs. } 2.1\pm0.7 \text{ g/L}, p=0.535; 4.0\pm2.7 \text{ vs. } 3.4\pm1.9 \,\mu\text{g/mL}, p=0.608)$ or between controls and controls with type II infection $(1.9\pm0.7 \text{ vs. } 1.8\pm0.8 \text{ g/L}, p=0.869; 4.0\pm2.7 \text{ vs. } 4.6\pm2.4 \,\mu\text{g/mL}, p=0.682)$ or between controls with I infection and controls with II

infection (2.1 ± 0.7 vs. 1.8 ± 0.8 g/L, p = 0.552; 3.4 ± 1.9 vs. 4.6 ± 2.4 µg/mL, p = 0.363) (Figure 2(C)).

Discussion

In this study, patients with IgAN showed higher infection rates of *H. pylori*, especially type I *H. pylori*, than healthy controls. And IgAN patients infected with *H. pylori*, especially type I *H. pylori*, showed significantly higher SBP, 24-h proteinuria, and BUN levels than uninfected patients. Similarly, higher plasma Gd-IgA1 levels were observed in IgAN patients with type I *H. pylori* infection.

Mucosal infection was reported to be involved in the development of IgAN through triggering innate and adaptive immune responses and the alternative complement pathway [21,22]. Recently, H. pylori infection and its linkage with extra-gastric diseases has been widely studied [23,24]. However, whether the infection rates of H. pylori are higher in patients with IgAN remains controversial [5,9]. In this study, we recruited 261 patients with IgAN and 46 healthy controls, and results showed that there were slight differences in the infection rates of *H. pylori* between these two groups, but not reach statistical significance; however, the infection rates of type I H. pylori, the main virulent strain of H. pylori, were significantly higher in patients with IgAN. H. pylori was reported to increase proteinuria and renal vasoconstriction in several renal disease as an infection factor through altering the permeability of the glomerular basement membrane, damaging endothelial dysfunction, and promoting production of inflammatory cytokines [25,26], suggesting that H. pylori was involved in progression of renal disease. Previous study [27] and our clinical practice (data not included) showed that H. pylori eradication reduced proteinuria in patients with IgAN. Decrease trend of eGFR levels were observed in IgAN patients with H. pylori infection [4], but the association between different types of H. pylori infection and clinical manifestations of IgAN remained to be elucidated. In this study, a trend of lower eGFR was showed in patients with type I infection, but it does not reach statistical significance. And significantly higher 24-h proteinuria, SBP, and BUN were showed in IgAN patients with type I infection, but not in patients with type II infection, implying that IgAN patients with virulent *H. pylori* infection may have lower renal function.

IgA1 with high lectin binding was produced in response to mucosal *H. pylori* infection [28], and exaggerated systemic antibody response, mainly serum anti-*H. pylori* IgA1, to mucosal infection was demonstrated in IgAN patients with *H. pylori* infection [9]. CagA, a crucial virulence factor of type I H. pylori, was reported to promote the production and undergalactosylation of IgA1 in the B cell line, DAKIKI [9,29]. It was reported that aberrant glycosylation of IgA1 was involved in the pathogenesis of IgAN [30]. However, the effects of different H. pylori strains on plasma Gd-IgA1 levels of IgAN patients remain unclear. In present study, patients with IgAN displayed significantly higher plasma IgA1 and Gd-IgA1 levels than controls, which are the common disease characteristics. And the plasma Gd-IgA1 levels were significantly higher in patients with type I H. pylori infection than those in uninfected patients, while the levels in controls and controls with H. pylori infection were comparable, indicating that the higher plasma Gd-IgA1 observed in IgAN patients with virulent H. pylori infection was associated with the disease, and not a general change in healthy controls. This was consistent with previous studies that the levels of serum mucosal-type IgA1 against H. pylori were significantly higher in IgAN patients than healthy controls [9] and the degree of H. pylori antigen and CagA deposition were obviously severe in IgAN patients than patients with non-IgAN primary glomerulonephritis [4]. The immune response to H. pylori infection may be more stronger in IgAN patients than controls.

The main limitations in our study were as followed. We evaluated the serum anti-H. pylori IgG to identify individuals exposed to H. pylori infection rather than testing the current infection status using ¹³C-UBT. However, the seropositivity of anti-H. pylori IgG could not be converted to seronegativity without H. pylori eradication therapy. To a certain extent, the serum anti-H. pylori IgG represents current infection and has been used for several epidemiological investigations [31, 32]. Although Gd-IgA1 was suggested to play a central role in pathogenesis of IgAN. Some studies reported that Gd-IgA1 was unrelated with proteinuria in IgAN patients [33,34]. While others observed a trend of lower eGFR, higher proteinuria, and increased use of immunosuppressives in IgAN patients with higher Gd-IgA1 levels [18]. In this observation study, we found that the IgAN patients with type I H. pylori infection showed lower renal function and higher plasma Gd-lgA1. Further cohort and H. pylori eradication studies are needed to verify whether H. pylori infection participate in the pathogenesis and progression of IgAN through the elevated plasma Gd-IgA1.

Conclusions

In conclusion, our study revealed that virulent type I H. pylori infection was more common in patients with

IgAN. And *H. pylori* infection, especially type I *H. pylori* infection, was associated with higher 24-h proteinuria, SBP, and BUN, as well as higher plasma Gd-IgA1 levels in patients with IgAN.

Acknowledgements

The authors thank all the patients and healthy controls who participated in this study.

Ethical approval

The study was approved by the Medical Ethics Committee of Peking University, and informed written consent was obtained from all patients.

Disclosure statement

All the authors report no conflicts of interest in this work.

Funding

This work was supported by National Natural Science Foundation of China [81800636], Beijing Natural Science Foundation [7184253], and Peking University Medicine Fund of Fostering Young Scholars' Scientific & Technological Innovation [BMU2017PY007].

References

- [1] Wyatt RJ, Julian BA. IgA nephropathy. N Engl J Med. 2013;368(25):2402–2414.
- [2] Perse M, Veceric-Haler Z. The role of IgA in the pathogenesis of IgA nephropathy. Int J Mol Sci. 2019;20(24): 6199.
- [3] Zhang YM, Zhang H. Insights into the role of mucosal immunity in IgA nephropathy. Clin J Am Soc Nephrol. 2018;13(10):1584–1586.
- [4] Zhu TT, Wang L, Wang HL, et al. *Helicobacter pylori* participates in the pathogenesis of IgA nephropathy. Ren Fail. 2016;38(9):1398–1404.
- [5] Kusano K, Inokuchi A, Fujimoto K, et al. Coccoid Helicobacter pylori exists in the palatine tonsils of patients with IgA nephropathy. J Gastroenterol. 2010; 45(4):406–412.
- [6] Niknam R, Barfei M, Mahmoudi L. *Helicobacter pylori*, endoscopic, and histologic features among kidney transplant candidates in southern Iran. IDR. 2019; 12(1):3687–3693.
- [7] Pan W, Zhang H, Wang L, et al. Association between *Helicobacter pylori* infection and kidney damage in patients with peptic ulcer. Ren Fail. 2019;41(1): 1028–1034.
- [8] Shin SP, Bang CS, Lee JJ, et al. *Helicobacter pylori* infection in patients with chronic kidney disease: a systematic review and meta-analysis. Gut Liver. 2019; 13(6):628–641.

- [9] Barratt J, Bailey EM, Buck KS, et al. Exaggerated systemic antibody response to mucosal *Helicobacter pylori* infection in IgA nephropathy. Am J Kidney Dis. 1999;33(6):1049–1057.
- [10] Chmiela M, Kupcinskas J. Review: pathogenesis of *Helicobacter pylori* infection. Helicobacter. 2019; 24(Suppl 1):e12638.
- [11] Kusano K, Tokunaga O, Ando T, et al. *Helicobacter pylori* in the palatine tonsils of patients with IgA nephropathy compared with those of patients with recurrent pharyngotonsillitis. Hum Pathol. 2007;38(12): 1788–1797.
- [12] Wang L, Tan RZ, Chen Y, et al. CagA promotes proliferation and secretion of extracellular matrix by inhibiting signaling pathway of apoptosis in rat glomerular mesangial cells. Ren Fail. 2016;38(3):458–464.
- [13] Galmiche A, Rassow J, Doye A, et al. The N-terminal 34 kDa fragment of *Helicobacter pylori* vacuolating cytotoxin targets mitochondria and induces cytochrome c release. Embo J. 2000;19(23):6361–6370.
- [14] Molinari M, Salio M, Galli C, et al. Selective inhibition of li-dependent antigen presentation by *Helicobacter pylori* toxin VacA. J Exp Med. 1998;187(1):135–140.
- [15] Levey AS, Stevens LA, Schmid CH, et al.; for the CKD-EPI (Chronic Kidney Disease Epidemiology Collaboration). A new equation to estimate glomerular filtration rate. Ann Intern Med. 2009;150(9):604–612.
- [16] Doolan PD, Alpen EL, Theil GB. A clinical appraisal of the plasma concentration and endogenous clearance of creatinine. Am J Med. 1962;32:65–79.
- [17] Cattran DC, Coppo R, Cook HT, et al.; Working Group of the International Ig ANN. The Oxford classification of IgA nephropathy: rationale, clinicopathological correlations, and classification. Kidney Int. 2009;76(5): 534–545.
- [18] Zhao N, Hou P, Lv J, et al. The level of galactose-deficient IgA1 in the sera of patients with IgA nephropathy is associated with disease progression. Kidney Int. 2012;82(7):790–796.
- [19] Zhang K, Li Q, Zhang Y, et al. Clinical significance of galactose-deficient IgA1 by KM55 in patients with IgA nephropathy. Kidney Blood Press Res. 2019;44(5): 1196–1206.
- [20] Suzuki H, Yasutake J, Makita Y, et al. IgA nephropathy and IgA vasculitis with nephritis have a shared feature involving galactose-deficient IgA1-oriented pathogenesis. Kidney Int. 2018;93(3):700–705.
- [21] Kiryluk K, Novak J, Gharavi AG. Pathogenesis of immunoglobulin A nephropathy: recent insight from genetic studies. Annu Rev Med. 2013;64(1):339–356.
- [22] Novak J, Moldoveanu Z, Julian BA, et al. Aberrant glycosylation of IgA1 and anti-glycan antibodies in IgA nephropathy: role of mucosal immune system. Adv Otorhinolaryngol. 2011;72(1):60–63.
- [23] Bravo D, Hoare A, Soto C, et al. *Helicobacter pylori* in human health and disease: mechanisms for local gastric and systemic effects. World J Gastroenterol. 2018; 24(28):3071–3089.
- [24] Robinson K. *Helicobacter pylori*-mediated protection against extra-gastric immune and inflammatory disorders: the evidence and controversies. Diseases. 2015; 3(2):34–55.

- [25] Kanbay M, Kasapoglu B, Akcay A. An occult risk factor for proteinuria: *Helicobacter pylori* infection. Med Hypotheses. 2007;69(3):709–710.
- [26] Migneco A, Ojetti V, Specchia L, et al. Eradication of *Helicobacter pylori* infection improves blood pressure values in patients affected by hypertension. Helicobacter. 2003;8(6):585–589.
- [27] Caliskan B, Yazici H, Caliskan Y, et al. The effects of *Helicobacter pylori* eradication on proteinuria in patients with primary glomerulonephritis. Int J Nephrol. 2014;2014(1):180690.
- [28] Smith AC, Molyneux K, Feehally J, et al. O-glycosylation of serum IgA1 antibodies against mucosal and systemic antigens in IgA nephropathy. J Am Soc Nephrol. 2006;17(12):3520–3528.
- [29] Yang M, Li FG, Xie XS, et al. CagA, a major virulence factor of *Helicobacter pylori*, promotes the production and underglycosylation of IgA1 in DAKIKI cells. Biochem Biophys Res Commun. 2014;444(2):276–281.

- [30] Novak J, Barratt J, Julian BA, et al. Aberrant glycosylation of the IgA1 molecule in IgA nephropathy. Semin Nephrol. 2018;38(5):461–476.
- [31] Somily AM, Morshed MG. An update of laboratory diagnosis of *Helicobacter pylori* in the Kingdom of Saudi Arabia. J Infect Dev Ctries. 2015;9(8):806–814.
- [32] Chey WD, Wong BC. Practice Parameters Committee of the American College of Gastroenterology. American College of Gastroenterology guideline on the management of *Helicobacter pylori* infection. Am J Gastroenterol. 2007;102(8):1808–1825.
- [33] Bagchi S, Lingaiah R, Mani K, et al. Significance of serum galactose deficient IgA1 as a potential biomarker for IgA nephropathy: a case control study. PLoS One. 2019;14(3):e0214256.
- [34] Hastings MC, Afshan S, Sanders JT, et al. Serum galactose-deficient IgA1 level is not associated with proteinuria in children with IgA nephropathy. Int J Nephrol. 2012;2012:315467.