

Review



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Multiple roles of ribosomal antimicrobial peptides in tackling global antimicrobial resistance

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In the last century, conventional antibiotics have played a significant role in global healthcare. Antibiotics support the body in controlling bacterial infection and simultaneously increase the tendency of drug resistance. Consequently, there is a severe concern regarding the regression of the antibiotic era. Despite the use of antibiotics, host defence systems are vital in fighting infectious diseases. In fact, the expression of ribosomal antimicrobial peptides (AMPs) has been crucial in the evolution of innate host defences and has been irreplaceable to date. Therefore, this valuable source is considered to have great potential in tackling the antimicrobial resistance (AMR) crisis. Furthermore, the possibility of bacterial resistance to AMPs has been intensively investigated. Here, we summarize all aspects related to the multiple applications of ribosomal AMPs and their derivatives in combating AMR.

1. Introduction

Antimicrobial resistance (AMR) occurs when microbial pathogens, including bacteria, viruses, fungi and parasites, change over time and no longer respond to drugs. AMR results in increased difficulties or even impossibility in treating infection and elevates the risk of spreading diseases, severe illness and death. Consequently, the cost of AMR to national economies and healthcare systems is serious, as it prolongs hospital stays, requires more expensive and intensive care, and

Table 1. Comparison of polypeptide antibiotics and antimicrobial peptides.

	antimicrobial peptides		polypeptide antibiotics
synthesizer	ribosomes		multifunctional enzymes
origins	bacteria, fungi, plants and animals		bacteria, fungi
structural properties	2nd structure	yes	rare
	branched structure	rare	common
	non-canonical amino acids	rare	common
	cyclization	not often (mostly via disulphide bonds)	often (results in oxazolines and thiazolines)
	other modifications	rare (mostly C-terminal amidation)	very common, including: <i>N</i> -methylation <i>N</i> -formylation glycosylation acylation halogenation hydroxylation oxidation and reduction
examples	LL37, magainins, indolicidins, Polybia-MP1, etc.		colistin, daptomycin, vancomycin, telavancin, etc.

significantly affects the productivity of patients and their caregivers [1,2]. Therefore, new antimicrobial agents are urgently required [3]. However, according to the World Health Organization (WHO), the clinical pipeline of newly developed antimicrobials is dry and insufficient to deal with the AMR challenge [4,5]. In 2020, among 26 antibiotics currently in clinical development, which are active against at least one of the priority pathogens in the WHO list, only seven candidates were classified as innovative [4]. However, even if all of these successfully pass clinical trials and enter the market, the new antibiotics will still suffer the same fate as the previous ones and soon become ineffective. In fact, the global threat of AMR needs to be tackled from many aspects at multiple levels and across different areas [6–8]. Recently, ribosomal antimicrobial peptides (AMPs) have emerged as a promising category that can effectively support many approaches for managing AMR [9–11].

Ribosomally synthesized AMPs are secreted as the first line of defence system by prokaryotes, plants and animals [11,12]. Unlike polypeptide antibiotics that have already been known and accepted for a long time [13], these AMPs have some significant differences in structural properties and biological profiles (table 1 and figures 1 and 2). These peptides mostly contain 7–50 amino acids [14–16] and usually share common structural properties, including a high content of cationic and hydrophobic residues, whereas their N-terminal is free and C-terminus is amidated. Such cationic amphipathic peptides show broad-spectrum activity, covering bacteria, fungi and viruses, as well as cancerous cells [16–19]. Interestingly, AMPs are well known for their fast and immediate killing effects [20–23]. Generally, the number of bacterial cells rapidly decreases after 15 min to 1 hour with at least 50% inhibition activity [24–27]. Furthermore, AMPs are well known for their immunomodulatory effects; on the one hand, they can recognize and activate the immune system to kill microbial pathogens, whereas on the other hand, they can suppress intense inflammation.

Over thousands of years of evolution, ribosomal AMPs still play crucial roles in the host defence systems of living organisms. However, several drawbacks, including poor absorption, distribution, metabolism and excretion (ADME), as well as high production costs compared with other commercially available antibiotics, have hindered their clinical development. Recently, owing to the urgent need for alternative antimicrobial therapies and the modern advances in peptide synthesis, chemical modification and bioengineering techniques, AMPs have attracted considerable attention for research and development. Therefore, these multifunctional molecules can be some of the most

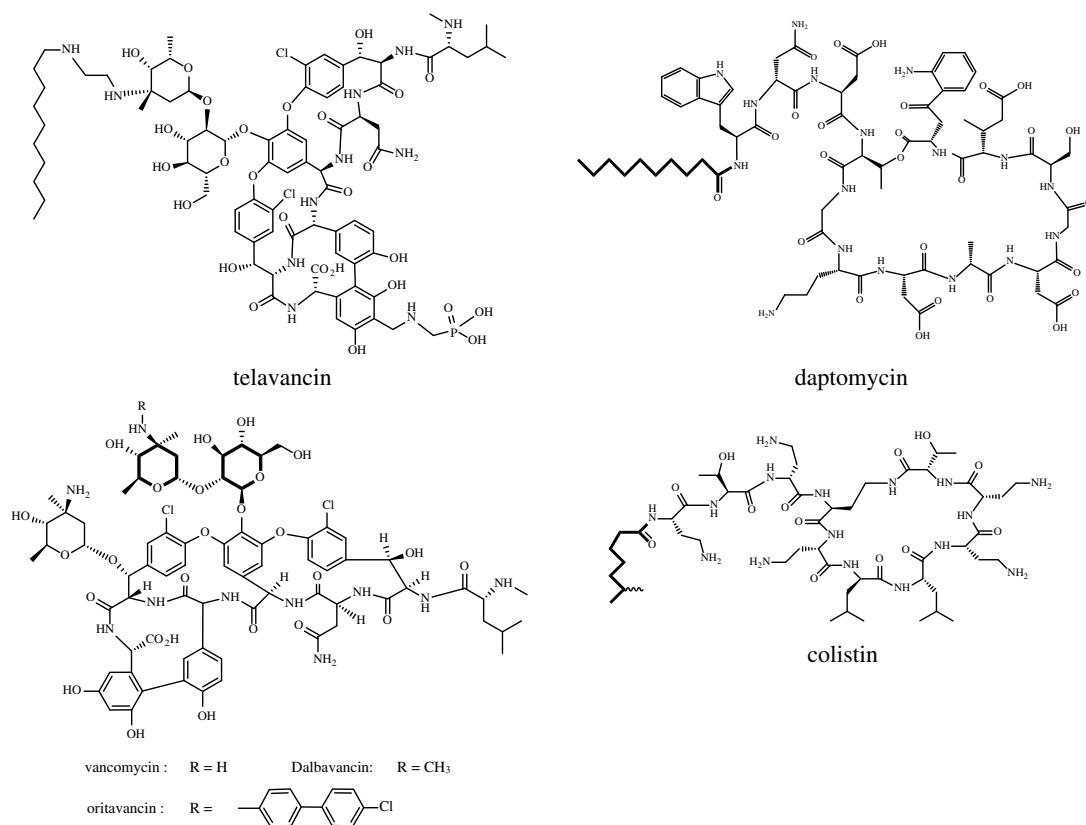


Figure 1. Examples of macrocyclic peptide-based antibiotic molecules.

important weapons in the battle against AMR. Starting from the diversity in the origins, structures and biological properties that lead to various therapeutic applications of AMPs, this review provides an overview and discusses their multiple roles in the management of AMR, as well as the opportunities and challenges in developing this emerging category.

2. Diversity of antimicrobial peptides

As an essential component of host defence systems, thousands of AMPs have been discovered and updated into many databases and bioinformatics resources (table 2 for more details). Overall, the sources and structural conformations of AMPs as well as their multiple biological actions are rather diverse (figure 3).

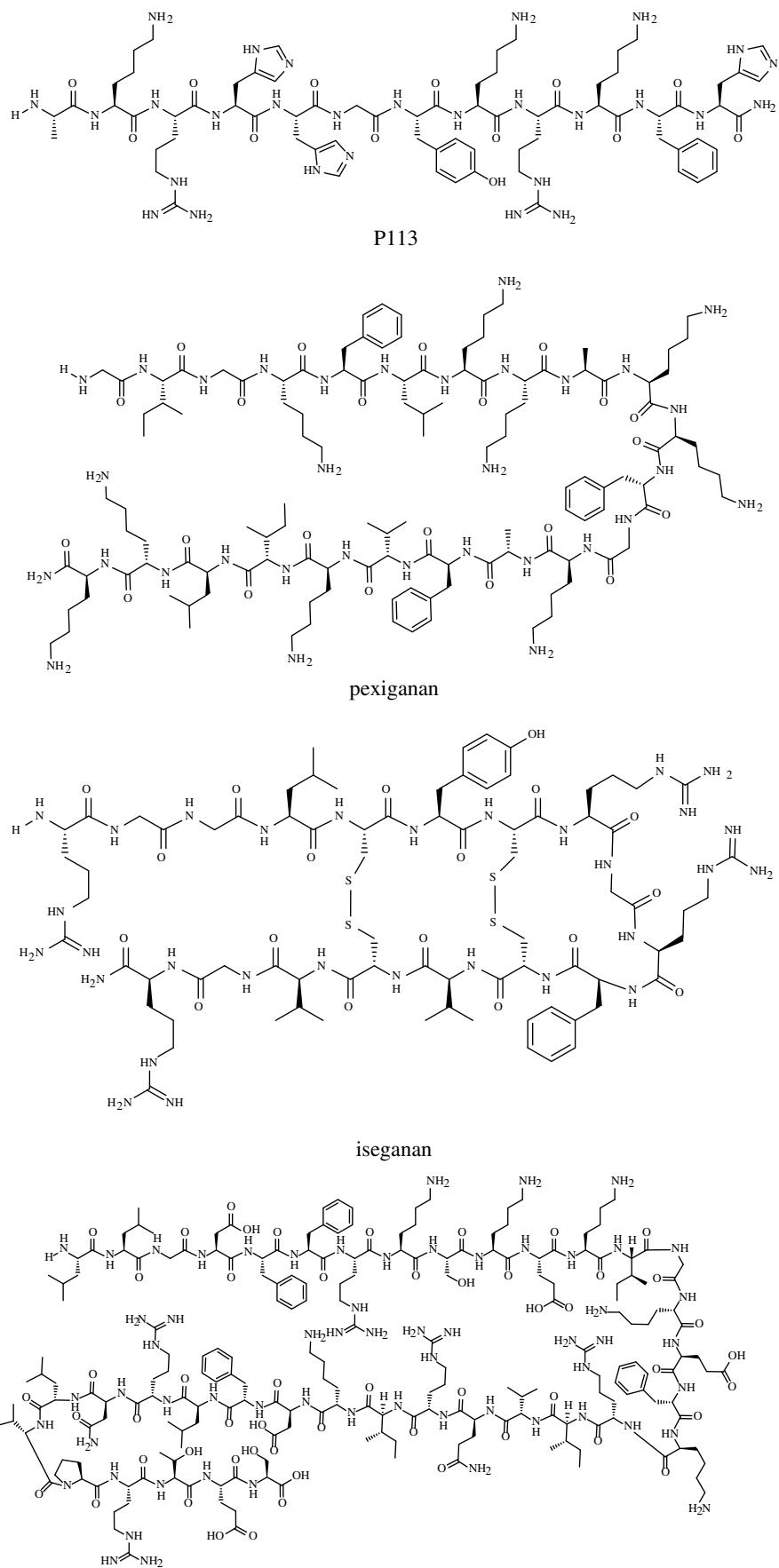
2.1. Diverse sources and secondary structures of antimicrobial peptides

2.1.1. Mammals, birds and fish

Defensins and cathelicidins are two large groups of AMPs isolated from mammals [32], birds [33] and fish [34,35]. Briefly, defensins are cysteine-rich peptides with a β -sheet structure and are divided into three subgroups: α -defensins, β -defensins and θ -defensins. In contrast to defensins, which are characterized and stabilized by three disulphide bridges, cathelicidins have a wide range of structures, number of residues and sequential differences. However, most of the known cathelicidins are linear with an α -helical conformation, including LL-37 [36], PMAP-36 [37,38] and CRAMP [39].

2.1.2. Amphibians

Currently, amphibians account for the largest proportion of AMPs found in nature, most of which are derived from skin secretions [22,40]. For example, 23 novel AMP sequences were discovered from a wild amphibian, *Hypsiboas pulchellus*, in Argentina [41]. Many other AMPs can be counted, such as magainins from the African frog *Xenopus laevis* [42], brevinins and esculentin found in *Rana* species



LL-37

Figure 2. Structure of selected antimicrobial peptides in clinical trials.

Table 2. Some databases and bioinformatics resources of antimicrobial peptides.

databases	major contents	last updated	references
data repository of antimicrobial peptides (DRAMP)	sequences, structures, classification, physicochemical properties, activities, patent and clinical information	4 November 2021 (latest v. DRAMP 3.0)	http://dramp.cpu-bioinfor.org/ [28]
antimicrobial peptide database (APD)	sequences, structures, classification, activities, timeline, prediction, design and statistics	20 July 2021 (latest v. APD3)	https://aps.unmc.edu/AP/ [14]
database of antimicrobial activity and structure of peptides (DBAASP)	sequences, structures, classification, activities, prediction, 3D structures and statistics	latest v. DBAASP v. 3.0	https://dbaasp.org/ [29]
yet another database of antimicrobial peptides (YADAMP)	sequences, structures, classification, physicochemical properties, activities and statistics	15 October 2018	http://yadamp.unisa.it/about.aspx [30]
LAMP (a database linking antimicrobial peptides)	sequences, structures, classification, activities and statistics	10 December 2016	http://biotechlab.fudan.edu.cn/database/lamp/index.php [31]

[43,44] and dermaseptin from the frog genus *Phyllomedusa* [45,46]. Despite large differences in size and sequence, these AMPs in most cases, still adopt an α -helix conformation in membrane-mimicking solutions to form a cationic amphiphilic helical structure [41].

2.1.3. Insects

Several AMP families have been reported to possess various secondary structures. For example, cecropins [47], lasioglossins [48], melittin [49] and Polybia-MP1 [50–52] form α -helical regions. Insect defensins form β -sheet conformations or proline/glycine-rich peptides [53], for example, drosocin [54], lebocins and attacin [55] show extended structures.

2.1.4. Plants

Many reports have indicated that plants produce different types of bioactive compounds to defend against the invasion of fungi, bacteria and insects [56]. Thus, AMPs have also been found in many plant components, such as fruits, flowers, leaves and stems [57,58], most of which contain cysteine residues and form disulphide bonds [57–59]. Recently, short disulphide-free AMPs were also found in green coconut water and proven to be multifunctional peptides without any sign of cytotoxicity to human cells [60–63].

2.1.5. Microorganisms

Bacteria and fungi produce a wide range of antimicrobial agents. However, non-ribosomal peptide antibiotics such as polymyxins, vancomycin or teixobactin are generally referred as polypeptide antibiotics, whereas the term 'AMPs' is more commonly used for ribosomal antibiotics. Hence, some well-known AMPs include nisin, microsin and pediocin from bacteria [64–66] and plectasin from fungi [67].

2.1.6. Synthetic sources

In addition to ribosomally synthesized molecules isolated from nature, several artificial AMPs have been created [27,68–70]. These peptides can be designed and synthesized based on the structure–activity relationships (SARs) of natural antimicrobial agents, aimed at improving one or more pharmacological properties [71,72]. Moreover, owing to the vast number of natural peptides with diverse lengths,

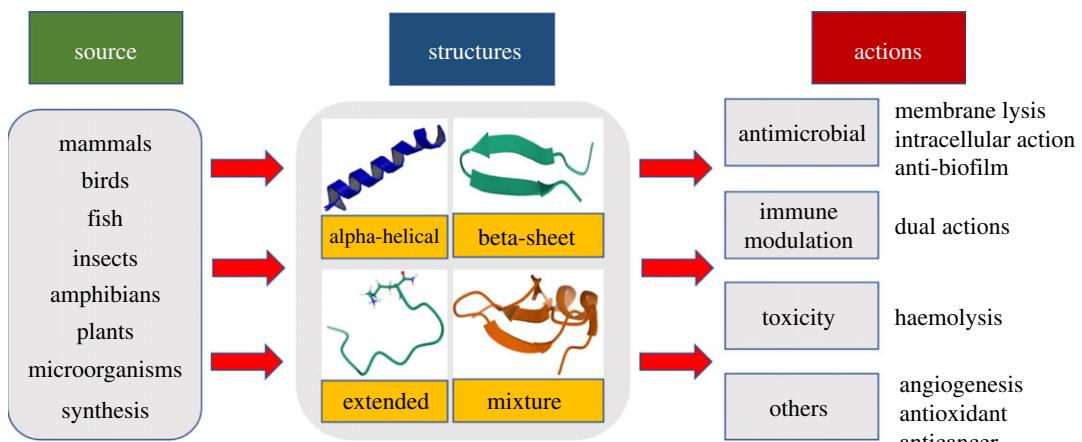


Figure 3. Diverse sources, structures and actions of antimicrobial peptides in nature.

structures and mechanisms of action, it is difficult to obtain reliable and complete SAR data. Therefore, the de novo design of AMPs based on general structural requirements was applied, and promising data were obtained [69,73,74].

2.2. Diverse biological effects of AMPs

Ribosomally synthesized AMPs have been demonstrated to be active against various types of pathogens, including bacteria, fungi, viruses, protozoa and cancer cells [17,75,76]. The most common mode of action of AMPs is membrane lysis via pore formation, leading to bacterial cell death [10,77,78]. The net positive charge, which is provided by cationic residues (arginine and lysine), the free amino group at the N-terminus, and the charge distribution are important for initial binding to the bacterial membrane [79]. Notably, amidation of the C-terminus benefits the increased net charge as well as structural stability and enhances membrane activity [48,80–82]. Next, the hydrophobic interaction between multiple hydrophobic residues and lipid bilayers can induce membrane hyperpolarization [83,84], membrane permeation and destruction through various models, including the barrel stave, carpet model, membrane thinning or thickening, electroporation, and toroidal and disordered toroidal pores [10,32,76]. This mechanism of action is selective for the negatively charged outer bacterial membrane over zwitterionic mammalian membranes [85,86]. Furthermore, in contrast to the lack of cholesterol in the bacterial membrane, the presence of cholesterol in eukaryotic cell membranes was demonstrated to reduce the interaction with AMPs and to suppress the disruption of lipid bilayers [87–91]. Furthermore, after entering bacterial cells without membrane disruption, other mechanisms of action have also been explored, as AMPs have been reported to inhibit some intracellular functions [17,92–94]. They can interact with negatively charged nucleic acids to interfere with their synthesis, replication and translation [95–99]. AMPs also target the biosynthesis [100–103], folding [104–106] and enzymatic activity [107–109] of proteins as well as metabolic processes [110–112] to achieve bacterial cell killing. Interestingly, AMPs have recently been reported to sequester and restrict the access of essential metals in invading pathogens [113].

Regarding immune system modulation, many AMPs are known to recruit and activate other immune components to clear infection. However, they can also act as suppressors when the inflammation becomes too strong by neutralizing bacterial products such as the endotoxin lipopolysaccharide (LPS) and lipoteichoic acid (LTA), and can control the Toll-like receptor (TLR) response [114,115]. Thus, AMPs are considered safer than conventional antibiotics. Consequently, several AMPs are currently in clinical trials for the management of septic shock [116,117].

Moreover, AMPs can also play active roles in wound healing [17,118–120] via antimicrobial activity as well as modulation of cytokine production, cell migration, proliferation, collagen synthesis and in some cases, angiogenesis [120–123]. Notably, wound healing is not unique to mammals and can also be observed in other species, such as fruit flies [124,125] and amphibians [126,127]. Furthermore, AMPs with antioxidant potential have also been found in fishes [128–130], frogs [131,132] and molluscs [133], thus demonstrating their protective effect against reactive oxygen species (ROS) in anti-ageing strategies. In the near future, AMPs are promising candidates for fighting resistant bacteria as well as promoting wound healing and skin regeneration [118,134,135].

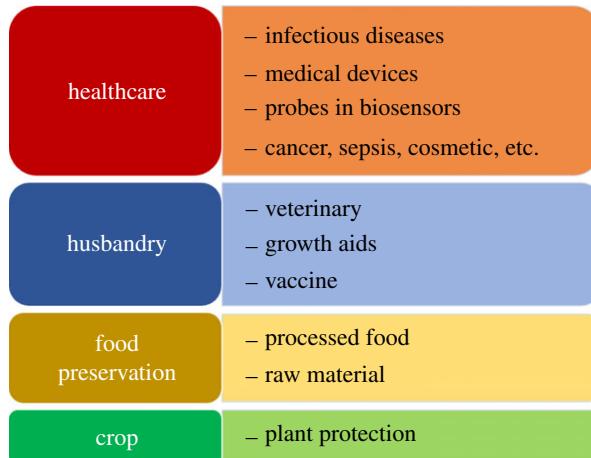


Figure 4. The diverse applications of antimicrobial peptides.

2.3. Diverse therapeutic applications of antimicrobial peptides

The current applications of AMPs are divided into four main groups: human healthcare, husbandry, food preservation and plant protection (figure 4). The majority of research effort to date has been focused on developing new therapies for various aspects of human health, such as infectious diseases [9,10,136], medical devices [17,137], cosmetology [118,135,138,139], cancer [51,140–144] and septic shock [114,145–147]. The broad-spectrum, along with the fast and selective actions of antimicrobial peptides can also benefit the development of biosensors that can rapidly detect pathogenic threats or monitor bacterial contaminations [148–150].

Many natural AMPs have been exploited for livestock, including veterinary medicines such as nisin. The FDA has approved nisin for dairy animals suffering from mastitis [151], as an adjuvant for new vaccine formulations [9,152,153] and as a growth aid [154,155]. Moreover, after nisin was the first bacteriocin approved for use as a food additive with code E234 [156–158], AMPs are now considered a new generation of food preservatives in both processed foods and their raw materials [159–161]. Interestingly, several studies have examined the potential of AMPs in plant protection, and some of these are introduced in table 3 [59,69,162–164].

Conventional antibiotics, as well as other antifungal, antiviral and antiparasitic drugs, are becoming increasingly ineffective as AMR has spread globally among humans, animals and in the environment [165]. This means that it is more challenging to manage infectious diseases and that the number of relevant deaths will increase, especially in patients with high-risk medical conditions, and undergoing treatments such as cancer chemotherapy, organ transplantation and other surgical indications [166,167]. Ribosomal AMPs with diverse mechanisms of action have become one of the most promising alternative antimicrobial agents to address the crisis of antibiotic resistance. Accordingly, they have been demonstrated to be active against a wide range of multi-drug-resistant pathogens, including those in the list of WHO priority pathogens for the research and development of new antibiotics (table 4 for some recent AMPs demonstrated to be active against antibiotic-resistant bacteria).

2.4. Resistance strategies against antimicrobial peptides

Broad-spectrum antibiotics are usually more susceptible to drug resistance. Therefore, it is possible that AMPs with notably broad-spectrum activity can suffer from microbial resistance compared with conventional antibiotics. Such resistance mechanisms have been reported for both types of bacterial species [189–191] and can be classified into four major groups:

- Membrane modification: bacteria can decrease the attraction and insertion of AMPs into their membrane by reducing the overall negative charges (such as by *D*-alanylation of teichoic acid [192], addition of 4-aminoarabinose to lipid A [193]), and enhancing rigidity (for example, by biofilm formation [194] or lipid A acylation [195,196]).
- Efflux pumps: export of antimicrobial agents out of the cell is an important strategy to remove several conventional antibiotics as well as antimicrobial peptides, such as LL-37, defensins and CRAMP [197,198].

Table 3. Sequence of some potential antimicrobial peptides for plant protection.

peptide	sequence	references
SP1-D	RKKRLKLLKRLV-NH ₂	[69]
SP7-D	LLIKFLKRFIKH-NH ₂	
SP10-D	LRFLKKILKHLF-NH ₂	
SP13-D	KRRRIARILRLAARALVKKR-NH ₂	
BP100	KKLFKKILKYL-NH ₂	[162]
BP134	KKLFKKILKYL-OH	[163]
BP203	KKLFKKILKYL-KKLFKKILKYL-OH	
BP209	G-KKLFFKKILKYL-AGPA-GIGKFLHSAK-OH	
BP210	S-KKLFFKKILKYL-AGPA-GIGKFLHSAK-OH	

Table 4. Recent antimicrobial peptides that active against some drug-resistant species.

priority ^a	pathogens ^a	antibiotic resistance ^a	antimicrobial peptides
critical	<i>Acinetobacter baumannii</i>	carbapenem-resistant	Cec4 [168], Ω276 [169], ZY4 [170], Hp1404 [171], TP4 derivatives dC4 and dN4 [172], AMPR-11 [114].
	<i>Pseudomonas aeruginosa</i>	carbapenem-resistant	ZY4 [170], P5 [173], AMPR-11 [114], ΔM2 [174], Ci-MAM-A24 [175].
	<i>Enterobacteriaceae</i>	carbapenem-resistant, ESBL-producing	AMPR-11 [114], DRGN-6 [176], ΔM2 [174], A-athanatin [177], Arenicin-3 [178], AA139 [178], Ci-MAM-A24 [175].
	<i>Enterococcus faecium</i>	vancomycin-resistant	Ci-MAM-A24 [175], Bip-P-113 [179], SLAY-P1 [180], Nisin [181], Lacticin 3147 [181].
high	<i>Staphylococcus aureus</i>	methicillin-resistant, vancomycin-intermediate and resistant	Ci-MAM-A24 [175], Nisin [181], Lacticin 3147 [181], WR12 [182], D-IK8 [182]. Mellittin [183]
	<i>Helicobacter pylori</i>	clarithromycin-resistant	Cbf-K ₁₆ [184], CRAMP [185], LL-37 [185], sLL-37 [185], TP4 [186]

^aAccording to the list of WHO priority pathogens [187,188].

- Proteolytic degradation: bacteria use proteases such as metalloproteinase [199–201], cysteine protease [202,203], or the omptin family of aspartate proteases [204,205] to break AMPs and avoid their killing action. For example, aureolysin is a zinc metalloprotease belonging to the thermolysin family that cleaves peptide bonds between Leu₃₁-Val₃₂, Arg₂₃-Ile₂₄, and Arg₁₉-Ile₂₀ in LL-37 [201,206]. Hence, it is suggested that *S. aureus* strains with significant secretion of this proteinase are less susceptible than those that do not express aureolysin activity [201].
- Sequestration: bacteria block AMPs from outside the cell, thus preventing them from reaching the bacterial cell membrane [207–209].

2.5. Low propensity to induce resistance and cross-resistance to antimicrobial peptides

Recently, a polypeptide antibiotic, colistin and an antimicrobial peptide, ZY4, were evaluated for their propensity to develop resistance in several *P. aeruginosa* and *A. baumannii* strains [170]. Accordingly, bacteria were exposed to ZY4 or colistin in the presence of sub-inhibitory concentrations. After the

first 20 passages, the minimum inhibitory concentrations (MICs) of colistin steadily increased, whereas no appreciable change was observed in the antimicrobial activity of ZY4. This difference became more significant after 60 passages, by which the MICs of colistin increased by 16–25 times compared with those of ZY4, which increased by 4.0–4.5 times [170]. Further investigation suggested that there was no observed cross-resistance between ZY4 and two similar antimicrobial peptides, ZY13 [210] and LZ1 [211] with other antibiotics, including colistin, tobramycin and levofloxacin [170].

In a different experimental approach, the integrated evolutionary analysis of two antimicrobial peptides, Tachypleasin II and Cecropin P1, confirmed the relatively low frequency of resistance through point mutations and gene amplification [212]. Furthermore, an investigation of the AMPs and antibiotic resistance genes in the human gut microbiota revealed that these two kinds of genes are different in adapting to new bacterial hosts. Consequently, the transfer tendency between the members of antimicrobial peptide resistance genes is less frequent than the others [213].

Although bacteria can develop resistance to antimicrobial peptides, it is suggested that the diversity in their mechanisms of action can provide an effective therapy to control bacterial growth [214]. For example, bacteria can increase electrostatic repulsion to defend themselves against some high cationic charge antimicrobial peptides; however, this strategy may not be practical for peptides with low net charge or multiple anionic residues (e.g. Polybia-MP1 [72]). Moreover, there is a minority group of AMPs with an anionic net charge [215–217] that can avoid this membrane modification. In the case of biofilms or capsule formation, there are always other AMPs with anti-biofilm properties [218] or the ability to destroy the protective capsule [219]. It is thus possible that the activity of an individual antimicrobial peptide is significantly reduced owing to a specific resistance mechanism in microbial pathogens; however, using a combination of AMPs or of AMPs with current antibiotics can promote synergistic action and overcome resistance [17,22]. It is also worth mentioning that innate host defence systems usually contain multiple types of AMPs with some differences in structure and function. Furthermore, the rapid action of AMPs [17] and the energy costs for developing multiple defence capacities are major obstacles in the proliferation and growth of microbial pathogens, thus limiting their resistance to antimicrobial peptide therapies.

2.6. AMPs and current therapies for antimicrobial resistance

AMR is a complicated problem that requires various strategic solutions; fortunately, AMPs are closely related to most of resistance mechanisms (see the summary in figure 5).

2.6.1. Antibiotics management

One of the main drivers of the multi-drug resistance crisis is the misuse and overuse of antimicrobials. Therefore, one of the important objectives is to promote the proper use of current drugs, which AMPs can support through several approaches such as replacing or at least reducing the traditional doses of antimicrobials, for example, by increasing antimicrobial peptide use for topical applications and in combination therapy [17,138,220]. Notably, according to the Food and Drug Administration (FDA), approximately 80% of all antibiotics were sold for use on livestock farms in 2014—the same year that the WHO published its first ever report on global AMR [221]. Accordingly, medically important antibiotics are limited or even banned in animal food in the USA and other countries [222]. Therefore, AMPs can be considered as one of the most promising antibiotic alternatives for both human and animal use.

2.6.2. New antimicrobial agents

High rates of resistance against frequently used antimicrobials have been observed everywhere, indicating that the world is running out of effective drugs for infectious disease. In drug development, ribosomal AMPs and their derivatives can serve as alternative antimicrobial agents, anti-biofilm agents or even both [21,223]. In fact, many AMPs, such as Histatin [224], Plectasin [67], Omiganan [10], IMX942 [225], Iseganan [9], LL-37 [9] and P113 [9], are currently in pre-clinical or clinical studies for various anti-infectious applications.

Additionally, efforts are underway in drug repurposing (or drug repositioning) to address the absence of new antimicrobial agents and limit the risk of failure and high costs required for development [226,227]. Thus, AMP development and drug repurposing are complementary in the fight against antibiotic-resistant bacteria. Moreover, AMPs can inspire the investigation of possible

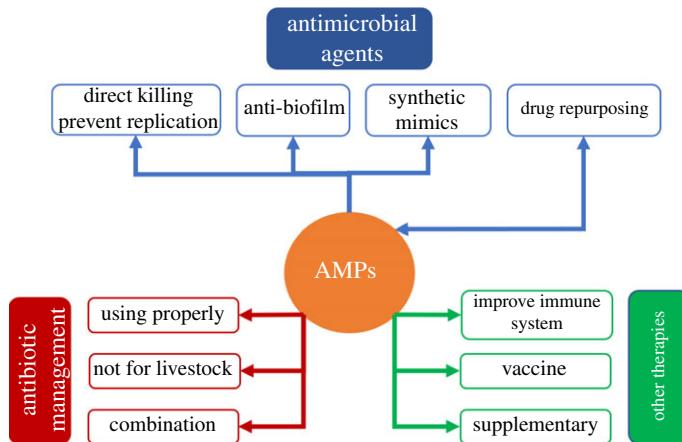


Figure 5. Diagram illustrating the correlation of antimicrobial peptides and the current therapies in antibiotic resistance management.

drug repurposing for antimicrobial discovery. For example, glatiramer acetate (GA), also referred to as COP-1, is a popular and safe treatment for multiple sclerosis. The structure of GA is similar to the cationic amphiphilic property of standard antimicrobial peptides, thus leading to the investigation of its antibacterial activity. GA was thus demonstrated to be active against both Gram-negative and Gram-positive bacterial species, including *Pseudomonas aeruginosa*, *Escherichia coli*, *Acinetobacter baumannii* and *Staphylococcus aureus* [228–231]. In particular, it displayed higher potency towards Gram-negative *Pseudomonas aeruginosa* compared with the human antimicrobial peptide LL-37 [228].

Notably, synthetic mimics of cationic AMPs (CAMPs) have been demonstrated to generate promising compounds for further development of new anti-infectious diseases [232–235]. Based on the common pharmacophore of short antimicrobial peptides, this strategy provides an attractive option that can avoid protease degradation and salt sensitivity, whereas large-scale production can be simple and cheap [236,237]. Alpha-mangostin, a xanthone extracted from *Garcinia mangostana*, showed potent and rapid antibacterial activity against Gram-positive species [238]. However, this compound has major disadvantages for clinical applications, including unsatisfactory cytotoxicity and poor aqueous solubility. Therefore, a series of amphipathic xanthones were designed as membrane-targeting antimicrobial peptidomimetics [237] (figure 6). Through systematic modifications of the cationic and hydrophobic moieties, some optimized compounds have displayed excellent and fast antibacterial activity against Gram-positive bacteria, including vancomycin-resistant enterococci and methicillin-resistant *S. aureus*, with higher selectivity and low propensity to develop resistance [239–241]. In another approach, inspired by the pharmacophore model CAMPs and marine antimicrobials, eusynstyelamides, a novel series of amphipathic barbiturates were designed with two cationic groups and two lipophilic side chains (figure 6). The obtained data suggested potent lead peptidomimetic compounds with broad-spectrum *in vitro* activity against 30 multi-drug resistant clinical isolates and exhibited promising *in vivo* efficacy in a mouse model infected with *Klebsiella pneumoniae* and *E. coli* [242].

2.6.3. Other therapies

In addition to their potential as new antibacterial agents, the benefits of AMPs have also been exploited to improve human and animal health, mainly by improving the immune system and intestinal morphology [243–247]. Nonetheless, most of these applications are currently in livestock and veterinary medicine [155].

3. Advantages, limitations and solutions

As described above, natural AMPs have various advantages in replacing traditional antibiotics. Broad-spectrum activity, diversity of mechanisms, fast action, lower risk of resistance and low propensity to develop toxicity are some of the most notable advantages [248]. Interestingly, in contrast to the high toxicity of available polypeptide antibiotics, many recent *in vivo* studies have reported that AMPs are safe for animal models [70,223,249]. Notably, by testing the lysis of human renal proximal tubular

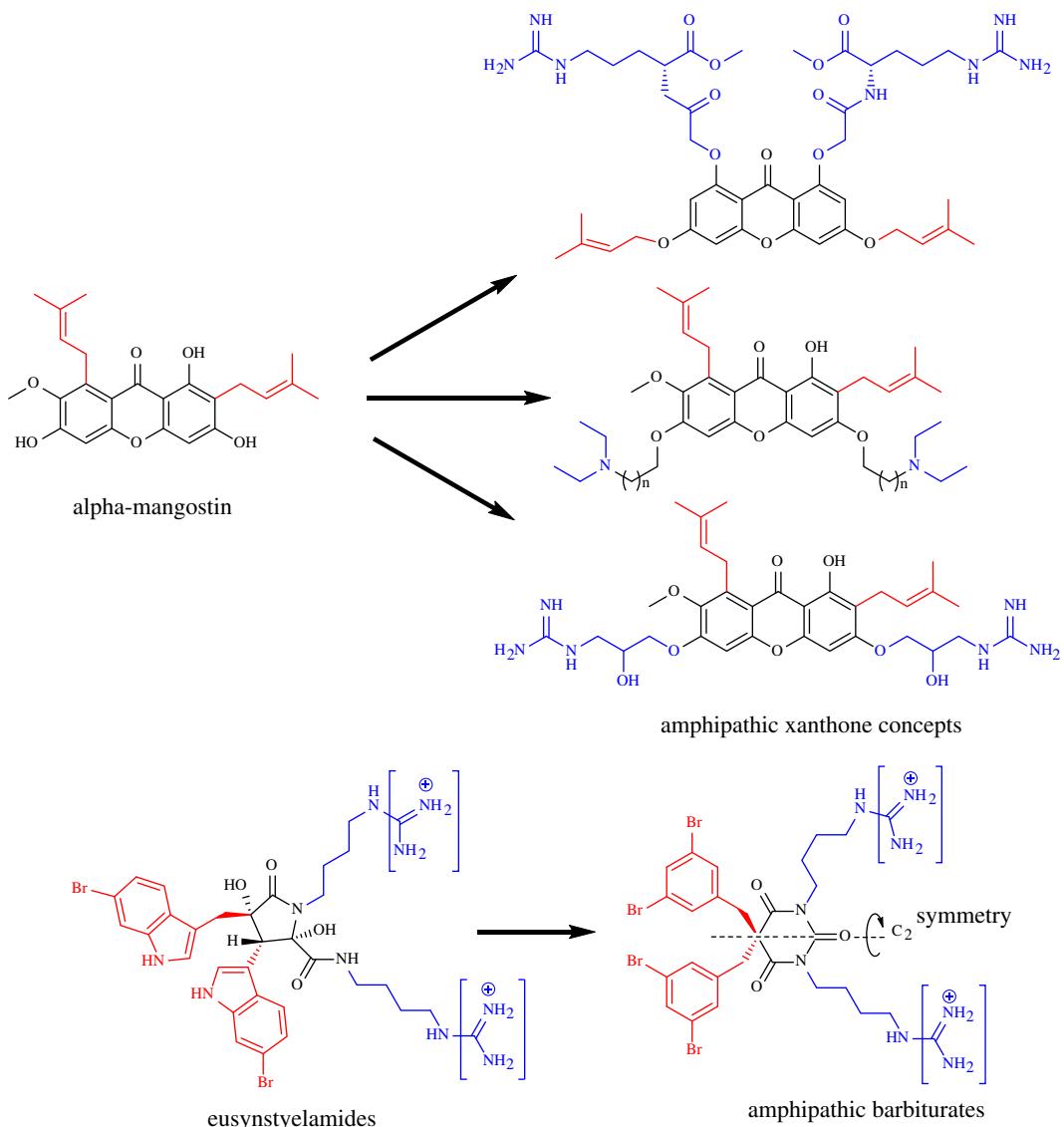


Figure 6. Synthetic mimics of cationic antimicrobial peptides.

epithelial cells (HRPTEC) *in vitro* and applying some sequence modifications, the possibility of reduced renal function can be avoided both *in vitro* and *in vivo* [70]. Furthermore, despite cytotoxicity to human red blood cells *in vitro*, there were no significant anaemia symptoms reported in animal models. In fact, even melittin, a well-known antimicrobial peptide with high haemolytic properties ($\text{EC}_{50} < 1 \mu\text{M}$), was confirmed to be safe for use in mice [250].

It should be noted that AMPs also have several limitations, including moderate antimicrobial activity, large size and poor *in vivo* bioavailability [17,248]. Thus, it is essential to find practical solutions to overcome these. To date, numerous strategies have been proposed including combination therapies [220,251–253], chemical modification approaches [254–258], optimization of peptide synthesis and structure [259,260] and formulation strategies [261–263]. A summary of some factors in the development of small molecules and AMPs as new antimicrobial drugs is presented in table 5.

4. Production of commercial antimicrobial peptides

There are currently two major technologies for producing commercial peptides, including chemical and microbial production, each with different strengths and weaknesses. The chemical method requires less time to develop and is easier for purification. However, its disadvantages include high production costs, difficulty in synthesizing long peptide sequences, and the use of environmentally unfriendly solvents [264,265]. Recombinant production can overcome the weakness of its chemical counterpart; however, it is more complex, labour-consuming, has difficult purification, and is greatly restricted by natural

Table 5. Summary of the advantages, limitations and solutions of small molecules and AMPs in the development of novel antimicrobial agents.

	small antibiotic molecules	natural AMPs
advantages	lower cost	broad spectrum
	stable	various mechanisms for each
	good permeability	fast action
	good oral bioavailability	lower propensity to develop toxicity or resistance
limitations	narrow spectrum	high cost
	mostly one mechanism for each	unstable
	higher propensity to resistance	low permeability
	high risk of drug–drug interaction	sensitive to environmental changes (pH, salts, fluids, ...)
solutions	management of undesirable outcomes	optimize the synthesis process
	combination therapy	
	biological and chemical strategies	
	choose proper routes of administration	

amino acids and vectors used [266,267]. Moreover, the high expression of AMPs could induce a killing effect on yeast and bacteria, thus resulting in a low yield and endotoxin release [268]. Therefore, it is suggested that chemical technology is more suitable for human use with high purity requirements, especially for producing AMPs with non-canonical amino acids and other chemical modifications. In addition, recombinant technology is widely applied for veterinary, animal growth aid and plant protection owing to the balance between cost and efficacy [243,267,269].

5. Conclusion

The emergence and spread of AMR will be accelerated without effective tools for the adequate treatment of infectious diseases and antimicrobial stewardship. This review provides an overview of the potential and recent advances in the research and development of AMPs to resolve the current global crisis of antimicrobial drug resistance. The diverse origins and mechanisms of action of natural AMPs can be favourable for developing alternative antimicrobial agents [9,10,270] and can provide widespread support for many other aspects in the management of AMR. In addition to latest findings that suggested a low propensity to develop resistance and toxicity, AMPs can be some of the most potent weapons in the war against resistant microbial pathogens.

Data accessibility. This article does not contain any additional data.

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References

- Hofer U. 2019 The cost of antimicrobial resistance. *Nat. Rev. Microbiol.* **17**, 3. (doi:10.1038/s41579-018-0125-x)
- Roope LS *et al.* 2019 The challenge of antimicrobial resistance: what economics can contribute. *Science* **364**, eaau4679. doi:10.1126/science.aau4679
- Theuretzbacher U, Bush K, Harbarth S, Paul M, Rex JH, Tacconelli E, Thwaites GE. 2020 Critical analysis of antibacterial agents in clinical development. *Nat. Rev. Microbiol.* **18**, 286–298. (doi:10.1038/s41579-020-0340-0)
- Venkatesan P. 2021 WHO 2020 report on the antibacterial production and development

- pipeline. *Lancet Microbe.* **2**, e239. (doi:10.1016/S2666-5247(21)00124-5)
5. World Health Organization. 2019 *Antibacterial agents in clinical development: an analysis of the antibacterial clinical development pipeline*. Geneva, Switzerland: World Health Organization.
 6. World Health Organization. 2012 *The evolving threat of antimicrobial resistance: options for action*. Geneva, Switzerland: World Health Organization.
 7. World Health Organization. 2018 *Antimicrobial resistance and primary health care*. Geneva, Switzerland: World Health Organization.
 8. Tacconelli E *et al.* 2018 Surveillance for control of antimicrobial resistance. *Lancet Infect. Dis.* **18**, e99–e106. (doi:10.1016/S1473-3099(17)30485-1)
 9. Mookherjee N, Anderson MA, Haagsman HP, Davidson DJ. 2020 Antimicrobial host defence peptides: functions and clinical potential. *Nat. Rev. Drug Discovery* **19**, 311–332. (doi:10.1038/s41573-019-0058-8)
 10. Magana M *et al.* 2020 The value of antimicrobial peptides in the age of resistance. *Lancet Infect. Dis.* **20**, e216–e230. (doi:10.1016/S1473-3099(20)30327-3)
 11. Mahlapuu M, Björn C, Ekblom J. 2020 Antimicrobial peptides as therapeutic agents: opportunities and challenges. *Crit. Rev. Biotechnol.* **40**, 978–992. (doi:10.1080/07388551.2020.1796576)
 12. Jaspreet Kaur B, Pushpendra Kumar S. 2020 Mini review on antimicrobial peptides, sources, mechanism and recent applications. *Protein & Peptide Lett.* **27**, 4–16. (doi:10.2174/09298665266619022165812)
 13. Wanger A, Chavez V, Huang RSP, Wahed A, Actor JK, Dasgupta A. 2017 Chapter 7 - Antibiotics, antimicrobial resistance, antibiotic susceptibility testing, and therapeutic drug monitoring for selected drugs. In *Microbiology and molecular diagnosis in pathology* (eds A Wanger, V Chavez, RSP Huang, A Wahed, JK Actor, A Dasgupta), pp. 119–153. Amsterdam, The Netherlands: Elsevier.
 14. Wang G, Li X, Wang Z. 2016 APD3: the antimicrobial peptide database as a tool for research and education. *Nucleic Acids Res.* **44**, D1087–D1093. (doi:10.1093/nar/gkv1278)
 15. Waghu FH, Barai RS, Gurung P, Idicula-Thomas S. 2016 CAMPR3: a database on sequences, structures and signatures of antimicrobial peptides. *Nucleic Acids Res.* **44**, D1094–D1097. (doi:10.1093/nar/gkv1051)
 16. Kang X *et al.* 2019 DRAMP 2.0, an updated data repository of antimicrobial peptides. *Sci. Data* **6**, 148. (doi:10.1038/s41597-019-0154-y)
 17. Luong HX, Thanh TT, Tran TH. 2020 Antimicrobial peptides – advances in development of therapeutic applications. *Life Sci.* **260**, 118407. (doi:10.1016/j.lfs.2020.118407)
 18. Haney EF, Mansour SC, Hancock REW. 2017 Antimicrobial peptides: an introduction. In *Antimicrobial peptides: methods and protocols* (ed. PR Hansen), pp. 3–22. New York, NY: Springer.
 19. Strom MB, Haug BE, Skar ML, Stensen W, Stiberg T, Svendsen JS. 2003 The pharmacophore of short cationic antibacterial peptides. *J. Med. Chem.* **46**, 1567–1570. (doi:10.1021/jm0340039)
 20. Li J, Koh J-J, Liu S, Lakshminarayanan R, Verma CS, Beuerman RW. 2017 Membrane active antimicrobial peptides: translating mechanistic insights to design. *Front. Neurosci.* **11**, 73. (doi:10.3389/fnins.2017.00073)
 21. Raheem N, Straus SK. 2019 Mechanisms of action for antimicrobial peptides with antibacterial and antibiofilm functions. *Front. Microbiol.* **10**, 2866. (doi:10.3389/fmicb.2019.02866)
 22. Lazzaro BP, Zasloff M, Rolff J. 2020 Antimicrobial peptides: application informed by evolution. *Science* **368**, eaau5480. (doi:10.1126/science.aau5480)
 23. Bellavita R *et al.* 2021 First-in-class cyclic temporin I analogue: design, synthesis, and antimicrobial assessment. *J. Med. Chem.* **64**, 11 675–11 694. (doi:10.1021/acs.jmedchem.1c01033)
 24. Nam J, Yun H, Rajasekaran G, Kumar SD, Kim JI, Min HJ, Shin SY, Lee CW. 2018 Structural and functional assessment of mbBJAMP1, an antimicrobial peptide from *Branchiostoma japonicum*, revealed a novel α -Hairpin-like scaffold with membrane permeable and DNA binding activity. *J. Med. Chem.* **61**, 11 101–11 113. (doi:10.1021/acs.jmedchem.8b01135)
 25. Irazabal LN *et al.* 2019 Fast and potent bactericidal membrane lytic activity of PaDBS1R1, a novel cationic antimicrobial peptide. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1861**, 178–190. (doi:10.1016/j.bbamem.2018.08.001)
 26. Hu F *et al.* 2016 Antimicrobial activity and safety evaluation of peptides isolated from the hemoglobin of chickens. *BMC Microbiol.* **16**, 287. (doi:10.1186/s12866-016-0904-3)
 27. Akbari R, Hakemi Vala M, Hashemi A, Aghazadeh H, Sabatier J-M, Pooshang Bagheri K. 2018 Action mechanism of melittin-derived antimicrobial peptides, MDP1 and MDP2, de novo designed against multidrug resistant bacteria. *Amino Acids* **50**, 1231–1243. (doi:10.1007/s00726-018-2596-5)
 28. Shi G, Kang X, Dong F, Liu Y, Zhu N, Hu Y, Xu H, Lao X, Zheng H. 2021 DRAMP 3.0: an enhanced comprehensive data repository of antimicrobial peptides. *Nucleic Acids Res.* **50**, D488–D496. (doi:10.1093/nar/gka651)
 29. Pirtskhalava M *et al.* 2016 DBAASP v.2: an enhanced database of structure and antimicrobial/cytotoxic activity of natural and synthetic peptides. *Nucleic Acids Res.* **44**, D1104–D1112. (doi:10.1093/nar/gkv1174)
 30. Piotto SP, Sessa L, Conclilio S, Iannelli P. 2012 YADAMP: yet another database of antimicrobial peptides. *Int. J. Antimicrob. Agents* **39**, 346–351. (doi:10.1016/j.ijantimicag.2011.12.003)
 31. Zhao X, Wu H, Lu H, Li G, Huang Q. 2013 LAMP: a database linking antimicrobial peptides. *PLoS ONE* **8**, e66557. (doi:10.1371/journal.pone.0066557)
 32. Ageitos JM, Sánchez-Pérez A, Calo-Mata P, Villa TG. 2017 Antimicrobial peptides (AMPs): ancient compounds that represent novel weapons in the fight against bacteria. *Biochem. Pharmacol.* **133**, 117–138. (doi:10.1016/j.bcp.2016.09.018)
 33. Zhang G, Sunkara LT. 2014 Avian antimicrobial host defense peptides: from biology to therapeutic applications. *Pharmaceuticals* **7**, 220–247. (doi:10.3390/ph7030220)
 34. Masso-Silva JA, Diamond G. 2014 Antimicrobial peptides from fish. *Pharmaceuticals (Basel)* **7**, 265–310. (doi:10.3390/ph7030265)
 35. Shabir U, Ali S, Magray AR, Ganai BA, Firdous P, Hassan T, Nazir R. 2018 Fish antimicrobial peptides (AMP's) as essential and promising molecular therapeutic agents: a review. *Microb. Pathog.* **114**, 50–56. (doi:10.1016/j.micpath.2017.11.039)
 36. Xhindoli D, Pacor S, Benincasa M, Scocchi M, Gennaro R, Tossi A. 2016 The human cathelicidin LL-37—a pore-forming antibacterial peptide and host-cell modulator. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1858**, 546–566. (doi:10.1016/j.bbamem.2015.11.003)
 37. Scheenstra MR, van den Belt M, Tjeerdsma-van Bokhoven JLM, Schneider VAF, Ordonez SR, van Dijk A, Veldhuijen EJA, Haagsman HP. 2019 Cathelicidins PMAP-36, LL-37 and CATH-2 are similar peptides with different modes of action. *Sci. Rep.* **9**, 4780. (doi:10.1038/s41598-019-41246-6)
 38. Scocchi M, Zelezetsky I, Benincasa M, Gennaro R, Mazzoli A, Tossi A. 2005 Structural aspects and biological properties of the cathelicidin PMAP-36. *FEBS J.* **272**, 4398–4406. (doi:10.1111/j.1742-4658.2005.04852.x)
 39. Coorens M, Scheenstra MR, Veldhuijen EJA, Haagsman HP. 2017 Interspecies cathelicidin comparison reveals divergence in antimicrobial activity, TLR modulation, chemokine induction and regulation of phagocytosis. *Sci. Rep.* **7**, 40874. (doi:10.1038/srep40874)
 40. Jiri P, Eugenie N, Blanka K, Qinghua W, Kamil K. 2019 Antimicrobial peptides: amphibian host defense peptides. *Curr. Med. Chem.* **26**, 5924–5946. (doi:10.2174/092986732566180713125314)
 41. Siano A, Humpola MV, de Oliveira E, Albericio F, Simonetta AC, Lajmanovich R, Tonarelli GG. 2014 Antimicrobial peptides from skin secretions of *Hypsiboas pulchellus* (Anura: Hylidae). *J. Nat. Prod.* **77**, 831–841. (doi:10.1021/np4009317)
 42. Zasloff M. 1987 Magainins, a class of antimicrobial peptides from *Xenopus* skin: isolation, characterization of two active forms, and partial cDNA sequence of a precursor. *Proc. Natl Acad. Sci. USA* **84**, 5449–5453.
 43. Zohrab F, Askarian S, Jalili A, Kazemi Oskuee R. 2019 Biological properties, current applications and potential therapeutic applications of brevinin peptide superfamily. *Int. J. Pept. Res. Ther.* **25**, 39–48. (doi:10.1007/s10989-018-9723-8)
 44. Rollins-Smith LA, Carey C, Longcore J, Doersam JK, Boutte A, Bruzgal JE, Conlon JM. 2002 Activity of antimicrobial skin peptides from ranid frogs against *Batrachochytrium dendrobatidis*, the chytrid fungus associated with global amphibian declines. *Dev. Comp. Immunol.* **26**, 471–479. (doi:10.1016/S0145-305X(01)00088-X)

45. Nicolas P, El Amri C. 2009 The dermaseptin superfamily: a gene-based combinatorial library of antimicrobial peptides. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1788**, 1537–1550. (doi:10.1016/j.bbamem.2008.09.006)
46. Savage JM, Heyer WR. 1967 Variation and distribution in the tree-frog genus *Phyllomedusa* in Costa Rica, Central America. *Beitrage zur Neotropischen Fauna*. **5**, 111–131. (doi:10.1080/01650526709360400)
47. Baek M-H *et al.* 2016 Lipopolysaccharide-bound structure of the antimicrobial peptide cecropin P1 determined by nuclear magnetic resonance spectroscopy. *J. Pept. Sci.* **22**, 214–221. (doi:10.1002/psc.2865)
48. Čeřovský V *et al.* 2009 Lasioglossins: three novel antimicrobial peptides from the venom of the eusocial bee *Lasioglossum laticeps* (Hymenoptera: Halictidae). *Chembiochem* **10**, 2089–2099. (doi:10.1002/cbic.200900133)
49. Leveritt III JM, Pino-Angeles A, Lazaridis T. 2015 The structure of a melittin-stabilized pore. *Biophys. J.* **108**, 2424–2426. (doi:10.1016/j.bpj.2015.04.006)
50. Alvares DS, Fanani ML, Ruggiero Neto J, Wilke N. 2016 The interfacial properties of the peptide Polybia-MP1 and its interaction with DPPC are modulated by lateral electrostatic attractions. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1858**, 393–402. (doi:10.1016/j.bbamem.2015.12.010)
51. Batista Martins D, Fadel V, Oliveira FD, Gaspar D, Alvares DS, Castanho MARB, dos Santos Cabrera MP. 2021 Protonectin peptides target lipids, act at the interface and selectively kill metastatic breast cancer cells while preserving morphological integrity. *J. Colloid Interface Sci.* **601**, 517–530. (doi:10.1016/j.jcis.2021.05.115)
52. Martins IBS, Viegas TG, dos Santos Alvares D, de Souza BM, Palma MS, Ruggiero Neto J, de Araujo AS. 2021 The effect of acidic pH on the adsorption and lytic activity of the peptides Polybia-MP1 and its histidine-containing analog in anionic lipid membrane: a biophysical study by molecular dynamics and spectroscopy. *Amino Acids* **53**, 753–767. (doi:10.1007/s00726-021-02982-0)
53. Li W, Tailhades J, O'Brien-Simpson NM, Separovic F, Otvos L, Hossain MA, Wade JD. 2014 Proline-rich antimicrobial peptides: potential therapeutics against antibiotic-resistant bacteria. *Amino Acids* **46**, 2287–2294. (doi:10.1007/s00726-014-1820-1)
54. McManus AM, Otvos L, Hoffmann R, Craik DJ. 1999 Conformational studies by NMR of the antimicrobial peptide, drosocin, and its non-glycosylated derivative: effects of glycosylation on solution conformation. *Biochemistry* **38**, 705–714. (doi:10.1021/bi981956d)
55. Yi H-Y, Chowdhury M, Huang Y-D, Yu X-Q. 2014 Insect antimicrobial peptides and their applications. *Appl. Microbiol. Biotechnol.* **98**, 5807–5822. (doi:10.1007/s00253-014-5792-6)
56. Tang S-S, Prodhan ZH, Biswas SK, Le C-F, Sekaran SD. 2018 Antimicrobial peptides from different plant sources: isolation, characterisation, and purification. *Phytochemistry* **154**, 94–105. (doi:10.1016/j.phytochem.2018.07.002)
57. Campos ML, de Souza CM, de Oliveira KBS, Dias SC, Franco OL. 2018 The role of antimicrobial peptides in plant immunity. *J. Exp. Bot.* **69**, 4997–5011. (doi:10.1093/jxb/ery294)
58. Meneguetti BT, Machado LD, Oshiro KGN, Nogueira ML, Carvalho CME, Franco OL. 2017 Antimicrobial peptides from fruits and their potential use as biotechnological tools—a review and outlook. *Front Microbiol.* **7**, 2136. (doi:10.3389/fmicb.2016.02136)
59. Phazang P, Negi NP, Raina M, Kumar D. 2020 Plant antimicrobial peptides: next-generation bioactive molecules for plant protection. In *Phyto-microbiome in stress regulation* (eds M Kumar, V Kumar, R Prasad), pp. 281–293. Singapore: Springer Singapore.
60. Silva ON *et al.* 2012 Cn-AMP1: a new promiscuous peptide with potential for microbial infections treatment. *Pept. Sci.* **98**, 322–331. (doi:10.1002/bip.22071)
61. Mandal SM, Dey S, Mandal M, Sarkar S, Maria-Neto S, Franco OL. 2009 Identification and structural insights of three novel antimicrobial peptides isolated from green coconut water. *Peptides* **30**, 633–637. (doi:10.1016/j.peptides.2008.12.001)
62. Santana MJ, de Oliveira AL, Queiroz Júnior LHK, Mandal SM, Matos CO, de O. Dias R, Franco OL, Liao LM. 2015 Structural insights into Cn-AMP1, a short disulfide-free multifunctional peptide from green coconut water. *FEBS Lett.* **589**, 639–644. (doi:10.1016/j.febslet.2015.01.029)
63. Anaya K, Podszun M, Franco OL, de Almeida Gadelha CA, Frank J. 2020 The coconut water antimicrobial peptide CnAMP1 is taken up into intestinal cells but does not alter P-glycoprotein expression and activity. *Plant Foods Hum. Nutr.* **75**, 396–403. (doi:10.1007/s11130-020-00826-y)
64. Yang S-C, Lin C-H, Sung CT, Fang J-Y. 2014 Antibacterial activities of bacteriocins: application in foods and pharmaceuticals. *Front. Microbiol.* **5**, 241. (doi:10.3389/fmicb.2014.00241)
65. Martínez B, Rodríguez A, Suárez E. 2016 Antimicrobial peptides produced by bacteria: the bacteriocins. In *New weapons to control bacterial growth* (eds TG Villa, M Vinas), pp. 15–38. Cham, Switzerland: Springer International Publishing.
66. Hassan M, Kjos M, Nes IF, Diep DB, Lotfipour F. 2012 Natural antimicrobial peptides from bacteria: characteristics and potential applications to fight against antibiotic resistance. *J. Appl. Microbiol.* **113**, 723–736. (doi:10.1111/j.1365-2672.2012.05338.x)
67. Mygind PH *et al.* 2005 Plectasin is a peptide antibiotic with therapeutic potential from a saprophytic fungus. *Nature* **437**, 975–980. (doi:10.1038/nature04051)
68. Luong HX, Kim D-H, Lee B-J, Kim Y-W. 2018 Effects of lysine-to-arginine substitution on antimicrobial activity of cationic stapled heptapeptides. *Arch. Pharm. Res.* **41**, 1092–1097. (doi:10.1007/s12272-018-1084-5)
69. Zeitler B, Herrera Diaz A, Dangel A, Thellmann M, Meyer H, Sattler M, Lindermayr C. 2013 De-novo design of antimicrobial peptides for plant protection. *PLoS ONE* **8**, e71687. (doi:10.1371/journal.pone.0071687)
70. Mountada R, Herce HD, Yin DJ, Moroco JA, Wales TE, Engen JR, Walensky LD. 2019 Design of stapled antimicrobial peptides that are stable, nontoxic and kill antibiotic-resistant bacteria in mice. *Nat. Biotechnol.* **37**, 1186–1197. (doi:10.1038/s41587-019-0222-z)
71. Hirano M, Saito C, Yokoo H, Goto C, Kawano R, Misawa T, Demizu Y. 2021 Development of antimicrobial stapled peptides based on magainin 2 sequence. *Molecules* **26**, 444. (doi:10.3390/molecules2620444)
72. Xuan HL, Duc TD, Thuy AM, Chau PM, Tung TT. 2021 Chemical approaches in the development of natural nontoxic peptide Polybia-MP1 as a potential dual antimicrobial and antitumor agent. *Amino Acids* **53**, 843–852. (doi:10.1007/s00726-021-02995-9)
73. Chen Z, Yu X, Zhang A, Wang F, Xing Y. 2020 De novo hydrocarbon-stapling design of single-turn α -helical antimicrobial peptides. *Int. J. Pept. Res. Ther.* **26**, 1711–1719. (doi:10.1007/s10989-019-09964-7)
74. Dinh TT, Kim D-H, Lee B-J, Kim Y-W. 2014 De novo design and their antimicrobial activity of stapled amphiphatic helices of heptapeptides. *Bull. Korean Chem. Soc.* **35**, 3632–3636.
75. Zhang L-J, Gallo RL. 2016 Antimicrobial peptides. *Curr. Biol.* **26**, R14–R19. (doi:10.1016/j.cub.2015.11.017)
76. Nguyen LT, Haney EF, Vogel HJ. 2011 The expanding scope of antimicrobial peptide structures and their modes of action. *Trends Biotechnol.* **29**, 464–472. (doi:10.1016/j.tibtech.2011.05.001)
77. da Cunha NB *et al.* 2017 The next generation of antimicrobial peptides (AMPs) as molecular therapeutic tools for the treatment of diseases with social and economic impacts. *Drug Discov. Today* **22**, 234–248. (doi:10.1016/j.drudis.2016.10.017)
78. Melo MN, Ferre R, Castanho MARB. 2009 Antimicrobial peptides: linking partition, activity and high membrane-bound concentrations. *Nat. Rev. Microbiol.* **7**, 245–250. (doi:10.1038/nrmicro2095)
79. Leite NB, da Costa LC, dos Santos Alvares D, dos Santos Cabrera MP, de Souza BM, Palma MS, Ruggiero Neto J. 2011 The effect of acidic residues and amphipathicity on the lytic activities of mastoparan peptides studied by fluorescence and CD spectroscopy. *Amino Acids* **40**, 91–100. (doi:10.1007/s00726-010-0511-9)
80. Zhu S, Li W, O'Brien-Simpson N, Separovic F, Sani M-A. 2021 C-terminus amidation influences biological activity and membrane interaction of maculatin 1.1. *Amino Acids* **53**, 769–777. (doi:10.1007/s00726-021-02983-z)
81. Mura M, Wang J, Zhou Y, Pinna M, Zvelindovsky AV, Dennison SR, Phoenix DA. 2016 The effect of amidation on the behaviour of antimicrobial peptides. *Eur. Biophys. J.* **45**, 195–207.
82. Li W *et al.* 2017 C-Terminal modification and multimerization increase the efficacy of a proline-rich antimicrobial peptide. *Chem. A Eur. J.* **23**, 390–396. (doi:10.1002/chem.201604172)
83. Yasir M, Dutta D, Willcox MDP. 2019 Comparative mode of action of the antimicrobial peptide melimine and its derivative Mel4

- against *Pseudomonas aeruginosa*. *Sci. Rep.* **9**, 7063. (doi:10.1038/s41598-019-42440-2)
84. Li W *et al.* 2015 Multimerization of a proline-rich antimicrobial peptide, Chex-Arg20, alters its mechanism of interaction with the *Escherichia coli* membrane. *Chem. Biol.* **22**, 1250–1258. (doi:10.1016/j.chembiol.2015.08.011)
 85. Lis M, Tew GN. 2012 9.16 – Polymer–membrane Interactions. In *Polymer science: a comprehensive reference* (eds K Matyjaszewski, M Möller), pp. 289–315. Amsterdam, The Netherlands: Elsevier.
 86. Southam HM, Butler JA, Chapman JA, Poole RK. 2017 Chapter one – The microbiology of ruthenium complexes. In *Advances in microbial physiology* (ed. RK Poole), pp. 1–96. New York, NY: Academic Press.
 87. Brender JR, McHenry AJ, Ramamoorthy A. 2012 Does cholesterol play a role in the bacterial selectivity of antimicrobial peptides? *Front. Immunol.* **3**, 195. (doi:10.3389/fimmu.2012.00195)
 88. Ramamoorthy A, Lee D-K, Narasimhaswamy T, Nanga RPR. 2010 Cholesterol reduces pardaxin's dynamics – a barrel-stave mechanism of membrane disruption investigated by solid-state NMR. *Biochim. Biophys. Acta* **1798**, 223–227. (doi:10.1016/j.bbapm.2009.08.012)
 89. McHenry AJ, Sciacca MFM, Brender JR, Ramamoorthy A. 2012 Does cholesterol suppress the antimicrobial peptide induced disruption of lipid raft containing membranes? *Biochim. Biophys. Acta* **1818**, 3019–3024. (doi:10.1016/j.bbapm.2012.07.021)
 90. Sharma VK, Mamontov E, Anunciado DB, O'Neill H, Urban VS. 2015 Effect of antimicrobial peptide on the dynamics of phosphocholine membrane: role of cholesterol and physical state of bilayer. *Soft Matter* **11**, 6755–6767. (doi:10.1039/CSSM01562F)
 91. Alvares DS, Monti MR, Ruggiero Neto J, Wilke N. 2021 The antimicrobial peptide Polybia-MP1 differentiates membranes with the hopanoid, diploterol from those with cholesterol. *BBA Adv.* **1**, 100002. (doi:10.1016/j.bbadv.2021.100002)
 92. Muñoz-Camargo C, Salazar VA, Barrero-Guevara L, Camargo S, Mosquera A, Groot H, Boix E. 2018 Unveiling the multifaceted mechanisms of antibacterial activity of buforin II and frenatin 2.3 S peptides from skin micro-organs of the orinoco lime treefrog (*Sphaenorhynchus lacteus*). *Int. J. Mol. Sci.* **19**, 2170.
 93. Benfield AH, Henriques ST. 2020 Mode-of-action of antimicrobial peptides: membrane disruption vs. intracellular mechanisms. *Front. Med. Technol.* **2**, 1–10. (doi:10.3389/fmedt.2020.610997)
 94. Paredes-Gamero EJ, Martins MNC, Cappabianco FAM, Ide JS, Miranda A. 2012 Characterization of dual effects induced by antimicrobial peptides: regulated cell death or membrane disruption. *Biochimica et Biophysica Acta (BBA) - General Subjects* **1820**, 1062–1072. (doi:10.1016/j.bbagen.2012.02.015)
 95. Park CB, Kim HS, Kim SC. 1998 Mechanism of action of the antimicrobial peptide buforin II: buforin II kills microorganisms by penetrating the cell membrane and inhibiting cellular functions. *Biochem. Biophys. Res. Commun.* **244**, 253–257.
 96. Hsu C-H, Chen C, Jou M-L, Lee AY-L, Lin Y-C, Yu Y-P, Huang W-T, Wu S-H. 2005 Structural and DNA-binding studies on the bovine antimicrobial peptide, indolicidin: evidence for multiple conformations involved in binding to membranes and DNA. *Nucleic Acids Res.* **33**, 4053–4064.
 97. Limoli DH, Rockel AB, Host KM, Jha A, Kopp BT, Hollis T, Wozniak DJ. 2014 Cationic antimicrobial peptides promote microbial mutagenesis and pathoadaptation in chronic infections. *PLoS Pathog.* **10**, e1004083.
 98. Snoussi M, Talledo JP, Del Rosario N-A, Mohammadi S, Ha B-Y, Košmrlj A, Taheri-Araghi S. 2018 Heterogeneous absorption of antimicrobial peptide LL37 in *Escherichia coli* cells enhances population survivability. *Elife* **7**, e38174.
 99. Friedrich CL, Rozek A, Patrzykat A, Hancock REW. 2001 Structure and mechanism of action of an indolicidin peptide derivative with improved activity against Gram-positive bacteria. *J. Biol. Chem.* **276**, 24 015–24 022. (doi:10.1074/jbc.M009691200)
 100. Mardirossian M, Grzela R, Giglione C, Meinell T, Gennaro R, Mergaert P, Scocchi M. 2014 The host antimicrobial peptide bac71-35 binds to bacterial ribosomal proteins and inhibits protein synthesis. *Chem. Biol.* **21**, 1639–1647. (doi:10.1016/j.chembiol.2014.10.009)
 101. Cole AM, Weis P, Diamond G. 1997 Isolation and characterization of pleurocidin, an antimicrobial peptide in the skin secretions of winter flounder. *J. Biol. Chem.* **272**, 12 008–12 013. (doi:10.1074/jbc.272.18.12008)
 102. Le C-F, Fang C-M, Sekaran SD. 2017 Intracellular targeting mechanisms by antimicrobial peptides. *Antimicrob. Agents Chemother.* **61**, e02340-16. (doi:10.1128/AAC.02340-16)
 103. Krizsan A, Volke D, Weinert S, Sträter N, Knappe D, Hoffmann R. 2014 Insect-derived proline-rich antimicrobial peptides kill bacteria by inhibiting bacterial protein translation at the 70 S ribosome. *Angew. Chem. Int. Ed.* **53**, 12 236–12 239. (doi:10.1002/anie.201407145)
 104. Kragol G, Lovas S, Varadi G, Condie BA, Hoffmann R, Otvos L. 2001 The antibacterial peptide pyrrhocoricin inhibits the atpase actions of DnaK and prevents chaperone-assisted protein folding. *Biochemistry* **40**, 3016–3026. (doi:10.1021/bi002656a)
 105. Scocchi M, Lüthy C, Decarli P, Mignogna G, Christen P, Gennaro R. 2009 The proline-rich antibacterial peptide Bac7 binds to and inhibits in vitro the molecular chaperone DnaK. *Int. J. Pept. Res. Ther.* **15**, 147–155.
 106. Knappe D, Zahn M, Sauer U, Schiffer G, Sträter N, Hoffmann R. 2011 Rational design of oncocin derivatives with superior protease stabilities and antibacterial activities based on the high-resolution structure of the oncocin-DnaK complex. *Chembiochem.* **12**, 874–876. (doi:10.1002/cbic.201000792)
 107. Nishikata M, Kanehira T, Oh H, Tani H, Tazaki M, Kuboki Y. 1991 Salivary histatin as an inhibitor of a protease produced by the oral bacterium *Bacteroides gingivalis*. *Biochem. Biophys. Res. Commun.* **174**, 625–630. (doi:10.1016/0006-291X(91)9463-M)
 108. Couto MA, Harwig SS, Lehrer RI. 1993 Selective inhibition of microbial serine proteases by eNAP-2, an antimicrobial peptide from equine neutrophils. *Infect. Immun.* **61**, 2991–2994. (doi:10.1128/la.61.7.2991-2994.1993)
 109. Huan Y, Kong Q, Mou H, Yi H. 2020 Antimicrobial peptides: classification, design, application and research progress in multiple fields. *Front. Microbiol.* **11**, 582779. (doi:10.3389/fmicb.2020.582779)
 110. Ho Y-H, Sung T-C, Chen C-S. 2012 Lactoferricin B inhibits the phosphorylation of the two-component system response regulators BasR and CreB. *Mol. Cell. Proteomics* **11**, M111.014720. (doi:10.1074/mcp.M111.014720)
 111. Tu Y-H, Ho Y-H, Chuang Y-C, Chen P-C, Chen C-S. 2011 Identification of Lactoferricin B intracellular targets using an *Escherichia coli* proteome chip. *PLoS ONE* **6**, e28197. (doi:10.1371/journal.pone.0028197)
 112. Ho Y-H, Shah P, Chen Y-W, Chen C-S. 2016 Systematic analysis of intracellular-targeting antimicrobial peptides, Bactenecin 7, hybrid of Pleurocidin and Dermaseptin, proline-arginine-rich peptide, and Lactoferricin B, by using *Escherichia coli* proteome microarrays. *Mol. Cell. Proteomics* **15**, 1837–1847. (doi:10.1074/mcp.M115.054999)
 113. Damo S, Kehl-Fie TE. 2016 Metal sequestration: an important contribution of antimicrobial peptides to nutritional immunity. In *Antimicrobial peptides: role in human health and disease* (eds J Harder, J-M Schröder), pp. 89–100. Cham, Switzerland: Springer International Publishing.
 114. Lee H-R, You D-G, Kim HK, Sohn JW, Kim MJ, Park JK, Lee GY, Yoo YD. 2020 Romo1-derived antimicrobial peptide is a new antimicrobial agent against multidrug-resistant bacteria in a murine model of sepsis. *mBio* **11**, e03258-19. (doi:10.1128/mBio.03258-19)
 115. Brandenburg K, Heinbockel L, Correa W, Lohner K. 2016 Peptides with dual mode of action: killing bacteria and preventing endotoxin-induced sepsis. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1858**, 971–979. (doi:10.1016/j.bbapm.2016.01.011)
 116. Fan L, Sun J, Zhou M, Zhou J, Tao X, Zheng H, Xu H. 2016 DRAMP: a comprehensive data repository of antimicrobial peptides. *Sci. Rep.* **6**, 24482. (doi:10.1038/srep24482)
 117. Koo HB, Seo J. 2019 Antimicrobial peptides under clinical investigation. *Pept. Sci.* **111**, e24122. (doi:10.1002/pep2.24122)
 118. Alencar-Silva T, Braga MC, Santana GOS, Saldanha-Araujo F, Pogue R, Dias SC, Franco OL, Carvalho JL. 2018 Breaking the frontiers of cosmetology with antimicrobial peptides. *Biotechnol. Adv.* **36**, 2019–2031. (doi:10.1016/j.bioteadv.2018.08.005)
 119. Wang Y, Ouyang J, Luo X, Zhang M, Jiang Y, Zhang F, Zhou J, Wang Y. 2021 Identification and characterization of novel bi-functional cathelicidins from the black-spotted frog (*Pelophylax nigromaculata*) with both anti-infective and antioxidant activities. *Dev. Comp.*

- Immunol.* **116**, 103928. (doi:10.1016/j.dci.2020.103928)
120. Thapa RK, Diep DB, Tønnesen HH. 2020 Topical antimicrobial peptide formulations for wound healing: current developments and future prospects. *Acta Biomater.* **103**, 52–67. (doi:10.1016/j.actbio.2019.12.025)
 121. Mangoni ML, McDermott AM, Zasloff M. 2016 Antimicrobial peptides and wound healing: biological and therapeutic considerations. *Exp. Dermatol.* **25**, 167–173. (doi:10.1111/exd.12929)
 122. Pfalzgraaf A, Brandenburg K, Weindl G. 2018 Antimicrobial peptides and their therapeutic potential for bacterial skin infections and wounds. *Front. Pharmacol.* **9**, 281. (doi:10.3389/fphar.2018.00281)
 123. Mi B, Liu J, Liu Y, Hu L, Liu Y, Panayi AC, Zhou W, Liu G. 2018 The designer antimicrobial peptide A-hBD-2 facilitates skin wound healing by stimulating keratinocyte migration and proliferation. *Cell. Physiol. Biochem.* **51**, 647–663. (doi:10.1159/000495320)
 124. Anderson AE, Gallo MJ. 2014 Will the wound-healing field earn its wings? *Exp. Dermatol.* **23**, 809–810. (doi:10.1111/exd.12498)
 125. Onfelt Tingvall T, Roos E, Engström Y. 2001 The *imd* gene is required for local *Cecropin* expression in *Drosophila* barrier epithelia. *EMBO Rep.* **2**, 239–243. (doi:10.1093/embo-reports/kve048)
 126. Mu L, Tang J, Liu H, Shen C, Rong M, Zhang Z, Lai R. 2014 A potential wound-healing-promoting peptide from salamander skin. *FASEB J.* **28**, 3919–3929. (doi:10.1096/fj.13-248476)
 127. Satoh A, Graham GMC, Bryant SV, Gardiner DM. 2008 Neurotrophic regulation of epidermal dedifferentiation during wound healing and limb regeneration in the axolotl (*Ambystoma mexicanum*). *Dev. Biol.* **319**, 321–335. (doi:10.1016/j.ydbio.2008.04.030)
 128. Guillén G, López Caballero M, Alemán A, Lacey ALD, Giménez B, Montero García P. 2010 Antioxidant and antimicrobial peptide fractions from squid and tuna skin gelatin.
 129. Harnedy PA, Fitzgerald RJ. 2012 Bioactive peptides from marine processing waste and shellfish: a review. *J. Funct. Foods* **4**, 6–24. (doi:10.1016/j.jff.2011.09.001)
 130. Najafian L, Babji AS. 2012 A review of fish-derived antioxidant and antimicrobial peptides: their production, assessment, and applications. *Peptides* **33**, 178–185. (doi:10.1016/j.peptides.2011.11.013)
 131. Guo C, Hu Y, Li J, Liu Y, Li S, Yan K, Wang X, Liu J, Wang H. 2014 Identification of multiple peptides with antioxidant and antimicrobial activities from skin and its secretions of *Hyalarana taipehensis*, *Amolops lisanensis*, and *Amolops granulosus*. *Biochimie* **105**, 192–201. (doi:10.1016/j.biochi.2014.07.013)
 132. Wang X *et al.* 2017 Identification and functional analyses of novel antioxidant peptides and antimicrobial peptides from skin secretions of four East Asian frog species. *Acta Biochim. Biophys. Sin.* **49**, 550–559. (doi:10.1093/abbs/gmx032)
 133. Borquaye LS, Darko G, Ocansey E, Ankomah E. 2015 Antimicrobial and antioxidant properties of the crude peptide extracts of *Galatea paradoxa* and *Patella rustica*. *SpringerPlus* **4**, 500. (doi:10.1186/s40064-015-1266-2)
 134. Rahnamaeian M, Vilcinskas A. 2015 Short antimicrobial peptides as cosmetic ingredients to deter dermatological pathogens. *Appl. Microbiol. Biotechnol.* **99**, 8847–8855. (doi:10.1007/s00253-015-6926-1)
 135. Woody B, Pambianchi E, Ferrara F, Therrien J-P, Pecorelli A, Messano N, Lila MA, Valacchi G. 2021 Cutaneous antimicrobial peptides: new 'actors' in pollution related inflammatory conditions. *Redox Biol.* **41**, 101952. (doi:10.1016/j.redox.2021.101952)
 136. Costa F, Teixeira C, Gomes P, Martins MCL. 2019 Clinical application of AMPs. In *Antimicrobial peptides: basics for clinical application* (ed. K Matsuzaki), pp. 281–298. Singapore: Springer Singapore.
 137. Kazemzadeh-Narbat M, Cheng H, Chabok R, Alvarez MM, de la Fuente-Nunez C, Phillips KS, Khademhosseini A. 2020 Strategies for antimicrobial peptide coatings on medical devices: a review and regulatory science perspective. *Crit. Rev. Biotechnol.* **41**, 94–120. (doi:10.1080/07388551.2020.1828810)
 138. Woodburn WK, Jaynes J, Clemens EL. 2020 designed antimicrobial peptides for topical treatment of antibiotic resistant acne vulgaris. *Antibiotics* **9**, 23. (doi:10.3390/antibiotics9010023)
 139. Nyonsoba F, Kiatsurayanan C, Chieilosilapatham P, Ogawa H. 2017 Friends or foes? Host defense (antimicrobial) peptides and proteins in human skin diseases. *Exp. Dermatol.* **26**, 989–998. (doi:10.1111/exd.13314)
 140. da Silva AMB, Silva-Gonçalves LC, Oliveira FA, Arcisio-Miranda M. 2018 Pro-necrotic activity of cationic mastoparan peptides in human glioblastoma multiforme cells via membranolytic action. *Mol. Neurobiol.* **55**, 5490–5504. (doi:10.1007/s12035-017-0782-1)
 141. Roudi R, Syn NL, Roudbari M. 2017 Antimicrobial peptides as biologic and immunotherapeutic agents against cancer: a comprehensive overview. *Front. Immunol.* **8**, 1320. (doi:10.3389/fimmu.2017.01320)
 142. Xie M, Liu D, Yang Y. 2020 Anti-cancer peptides: classification, mechanism of action, reconstruction and modification. *Open Biol.* **10**, 200004. (doi:10.1098/rsob.200004)
 143. Tornesello AL, Borrelli A, Buonaguro L, Buonaguro FM, Tornesello ML. 2020 Antimicrobial peptides as anticancer agents: functional properties and biological activities. *Molecules (Basel, Switzerland)* **25**, 2850. (doi:10.3390/molecules25122850)
 144. Riolo M, de Breij A, Drijfhout JW, Nibbering PH, Zaaij SAJ. 2017 Antimicrobial peptides in biomedical device manufacturing. *Front. Chem.* **5**, 63. (doi:10.3389/fchem.2017.00063)
 145. Schuerholz T, Brandenburg K, Marx G. 2012 Antimicrobial peptides and their potential application in inflammation and sepsis. In *Annual update in intensive care and emergency medicine 2012* (ed. J-L Vincent), pp. 85–97. Berlin, Germany: Springer.
 146. Papareddy P, Kasetty G, Kalle M, Bhongir RKV, Mörgelin M, Schmidtchen A, Malmsten M. 2016 NLF20: an antimicrobial peptide with therapeutic potential against invasive *Pseudomonas aeruginosa* infection. *J. Antimicrob. Chemother.* **71**, 170–180. (doi:10.1093/jac/dkv322)
 147. Martin L *et al.* 2016 The synthetic antimicrobial peptide 19–25 attenuates septic cardiomyopathy and prevents down-regulation of SERCA2 in polymicrobial sepsis. *Sci. Rep.* **6**, 37277. (doi:10.1038/srep37277)
 148. Pardoux É, Roux A, Mathey R, Botury D, Roupoiz Y. 2019 Antimicrobial peptide arrays for wide spectrum sensing of pathogenic bacteria. *Talanta* **203**, 322–327. (doi:10.1016/j.talanta.2019.05.062)
 149. Yuan K *et al.* 2018 Antimicrobial peptide based magnetic recognition elements and Au@Ag-GO SERS tags with stable internal standards: a three in one biosensor for isolation, discrimination and killing of multiple bacteria in whole blood. *Chem. Sci.* **9**, 8781–8795. (doi:10.1039/C8SC04637A)
 150. Kulagina NV, Lassman ME, Ligler FS, Taitt CR. 2005 Antimicrobial peptides for detection of bacteria in biosensor assays. *Anal. Chem.* **77**, 6504–6508. (doi:10.1021/ac050639r)
 151. Ahmad V, Khan MS, Jamal QMS, Alzohairy MA, Al Karaawi MA, Siddiqui MU. 2017 Antimicrobial potential of bacteriocins: in therapy, agriculture and food preservation. *Int. J. Antimicrob. Agents* **49**, 1–11. (doi:10.1016/j.ijantimicag.2016.08.016)
 152. Piyush B, Santi MM. 2019 Antimicrobial peptides and vaccine development to control multi-drug resistant bacteria. *Protein & Peptide Lett.* **26**, 324–331. (doi:10.2174/0929866526666190228162751)
 153. Fritz JH, Brunner S, Birnstiel ML, Buschle M, Gabain AV, Mattner F, Zauner W. 2004 The artificial antimicrobial peptide KLKLLLLLKLK induces predominantly a TH2-type immune response to co-injected antigens. *Vaccine* **22**, 3274–3284. (doi:10.1016/j.vaccine.2004.03.007)
 154. Xiao H, Shao F, Wu M, Ren W, Xiong X, Tan B, Yin Y. 2015 The application of antimicrobial peptides as growth and health promoters for swine. *J. Anim. Sci. Biotechnol.* **6**, 19. (doi:10.1186/s40104-015-0018-z)
 155. Wang S, Zeng X, Yang Q, Qiao S. 2016 Antimicrobial peptides as potential alternatives to antibiotics in food animal industry. *Int. J. Mol. Sci.* **17**, 603.
 156. Delves-Broughton J, Blackburn P, Evans RJ, Hugenholtz J. 1996 Applications of the bacteriocin, nisin. *Antonie Van Leeuwenhoek* **69**, 193–202. (doi:10.1007/BF00399424)
 157. de Arauz LJ, Jozala AF, Mazzola PG, Vessoni Penna TC. 2009 Nisin biotechnological production and application: a review. *Trends Food Sci. Technol.* **20**, 146–154. (doi:10.1016/j.tifs.2009.01.056)
 158. Shin JM, Gwak JW, Kamarajan P, Fenno JC, Rickard AH, Kapila YL. 2016 Biomedical applications of nisin. *J. Appl. Microbiol.* **120**, 1449–1465. (doi:10.1111/jam.13033)
 159. Ben said L, Fliss I, Offret C, Beaulieu L. 2019 Antimicrobial peptides: the new generation of food additives. In *Encyclopedia of food chemistry* (eds L Melton, F Shahidi, P Varelis), pp. 576–582. Oxford, UK: Academic Press.

160. Woraprayote W, Malila Y, Sorapukdee S, Svetiwathana A, Benjakul S, Visessanguan W. 2016 Bacteriocins from lactic acid bacteria and their applications in meat and meat products. *Meat Sci.* **120**, 118–132. (doi:10.1016/j.meatsci.2016.04.004)
161. Abdulhussain Kareem R, Razavi SH. 2020 Plantaricin bacteriocins: as safe alternative antimicrobial peptides in food preservation—a review. *J. Food Saf.* **40**, e12735. (doi:10.1111/jfs.12735)
162. Badosa E, Ferre R, Planas M, Feliu L, Besalú E, Cabrefiga J, Bardaji E, Montesinos E. 2007 A library of linear undecapeptides with bactericidal activity against phytopathogenic bacteria. *Peptides* **28**, 2276–2285. (doi:10.1016/j.peptides.2007.09.010)
163. Badosa E, Moiset G, Montesinos L, Talleda M, Bardaji E, Feliu L, Planas M, Montesinos E. 2013 Derivatives of the antimicrobial peptide BP100 for expression in plant systems. *PLoS ONE* **8**, e85515. (doi:10.1371/journal.pone.0085515)
164. Powell WA, Catranis CM, Maynard CA. 2000 Design of self-processing antimicrobial peptides for plant protection. *Lett. Appl. Microbiol.* **31**, 163–168. (doi:10.1046/j.1365-2672.2000.00782.x)
165. Rousham EK, Unicomb L, Islam MA. 2018 Human, animal and environmental contributors to antibiotic resistance in low-resource settings: integrating behavioural, epidemiological and one health approaches. *Proc. Biol. Sci.* **285**, 20180332. (doi:10.1098/rspb.2018.0332)
166. World Health Organization. 2014 *Antimicrobial resistance: global report on surveillance*. Geneva, Switzerland: World Health Organization.
167. World Health Organization. 2015 *Global action plan on antimicrobial resistance*. Geneva, Switzerland: World Health Organization.
168. Liu W, Wu Z, Mao C, Guo G, Zeng Z, Fei Y, Wan S, Peng J, Wu J. 2020 Antimicrobial peptide Cec4 eradicates the bacteria of clinical carbapenem-resistant *Acinetobacter baumannii* biofilm. *Front. Microbiol.* **11**, 1532. (doi:10.3389/fmicb.2020.01532)
169. Nagarajan D *et al.* 2019 L276: A designed antimicrobial peptide to combat carbapenem- and tigecycline-resistant *Acinetobacter baumannii*. *Sci. Adv.* **5**, eaax1946. (doi:10.1126/sciadv.aax1946)
170. Mwangi J, Yin Y, Wang G, Yang M, Li Y, Zhang Z, Lai R. 2019 The antimicrobial peptide ZY4 combats multidrug-resistant *Pseudomonas aeruginosa* and *Acinetobacter baumannii* infection. *Proc. Natl Acad. Sci. USA* **116**, 26516. (doi:10.1073/pnas.1909585117)
171. Hong MJ, Kim MK, Park Y. 2021 Comparative antimicrobial activity of Hp404 peptide and its analogs against *Acinetobacter baumannii*. *Int. J. Mol. Sci.* **22**, 5540.
172. Jung C-J, Liao Y-D, Hsu C-C, Huang T-Y, Chuang Y-C, Chen J-W, Kuo Y-M, Chia J-S. 2021 Identification of potential therapeutic antimicrobial peptides against *Acinetobacter baumannii* in a mouse model of pneumonia. *Sci. Rep.* **11**, 7318. (doi:10.1038/s41598-021-86844-5)
173. Martinez M, Gonçalves S, Felício MR, Maturana P, Santos NC, Semorile L, Hollmann A, Maffia PC. 2019 Synergistic and antibiofilm activity of the antimicrobial peptide P5 against carbapenem-resistant *Pseudomonas aeruginosa*. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1861**, 1329–1337. (doi:10.1016/j.bbamem.2019.05.008)
174. Rivera-Sánchez SP *et al.* 2020 Antibacterial activity of a cationic antimicrobial peptide against multidrug-resistant Gram-negative clinical isolates and their potential molecular targets. *Molecules* **25**, 5035.
175. Fedders H, Podschun R, Leippe M. 2010 The antimicrobial peptide Ci-MAM-A24 is highly active against multidrug-resistant and anaerobic bacteria pathogenic for humans. *Int. J. Antimicrob. Agents* **36**, 264–266. (doi:10.1016/j.ijantimicag.2010.04.008)
176. Hitt SJ, Bishop BM, van Hoek ML. 2020 Komodo-dragon cathelicidin-inspired peptides are antibacterial against carbapenem-resistant *Klebsiella pneumoniae*. *J. Med. Microbiol.* **69**, 1262–1272. (doi:10.1099/jmm.0.001260)
177. Hou Z, Lu J, Fang C, Zhou Y, Bai H, Zhang X, Xue X, Chen Y, Luo X. 2011 Underlying mechanism of in vivo and in vitro activity of C-terminal-amidated thanatin against clinical isolates of extended-spectrum beta-lactamase-producing *Escherichia coli*. *J. Infect. Dis.* **203**, 273–282. (doi:10.1093/infdis/jiq029)
178. Elliott AG *et al.* 2020 An amphiphatic peptide with antibiotic activity against multidrug-resistant Gram-negative bacteria. *Nat. Commun.* **11**, 3184. (doi:10.1038/s41467-020-16950-x)
179. Wu C-L, Hsueh J-Y, Yip B-S, Chih Y-H, Peng K-L, Cheng J-W. 2020 Antimicrobial peptides display strong synergy with vancomycin against vancomycin-resistant *E. faecium*, *S. aureus*, and Wild-Type *E. coli*. *Int. J. Mol. Sci.* **21**, 4578. (doi:10.3390/ijms21134578)
180. Liu Y, Jia Y, Yang K, Li R, Xiao X, Wang Z. 2020 Antagonizing vancomycin resistance in enterococcus by surface localized antimicrobial display-derived peptides. *ACS Infect. Dis.* **6**, 761–767. (doi:10.1021/acsinfecdis.9b00164)
181. Piper C, Draper LA, Cotter PD, Ross RP, Hill C. 2009 A comparison of the activities of lacticin 3147 and nisin against drug-resistant *Staphylococcus aureus* and *Enterococcus* species. *J. Antimicrob. Chemother.* **64**, 546–551. (doi:10.1093/jac/dkp221)
182. Mohamed MF, Abdelkalek A, Seleem MN. 2016 Evaluation of short synthetic antimicrobial peptides for treatment of drug-resistant and intracellular *Staphylococcus aureus*. *Sci. Rep.* **6**, 29707. (doi:10.1038/srep29707)
183. Ravensdale J, Wong Z, O'Brien F, Gregg K. 2016 Efficacy of antibacterial peptides against peptide-resistant MRSA is restored by permeabilization of bacteria membranes. *Front. Microbiol.* **7**, 1745. (doi:10.3389/fmicb.2016.01745)
184. Jiang M, Ma L, Huang Y, Wu H, Dou J, Zhou C. 2020 Antimicrobial activities of peptide Cbf-K16 against drug-resistant *Helicobacter pylori* infection in vitro and in vivo. *Microb. Pathog.* **138**, 103847. (doi:10.1016/j.micpath.2019.103847)
185. Zhang L *et al.* 2016 Critical role of antimicrobial peptide Cathelicidin for controlling *Helicobacter pylori* survival and infection. *J. Immunol.* **196**, 1799. (doi:10.4049/jimmunol.1500021)
186. Narayana JL, Huang H-N, Wu C-J, Chen J-Y. 2015 Efficacy of the antimicrobial peptide TP4 against *Helicobacter pylori* infection: *in vitro* membrane perturbation via micellization and *in vivo* suppression of host immune responses in a mouse model. *Oncotarget* **6**, 12 936–12 954. (doi:10.18632/oncotarget.4101)
187. Tacconelli E *et al.* 2018 Discovery, research, and development of new antibiotics: the WHO priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect. Dis.* **18**, 318–327. (doi:10.1016/S1473-3099(17)30753-3)
188. Asokan GV, Vanitha A. 2018 WHO global priority pathogens list on antibiotic resistance: an urgent need for action to integrate One Health data. *Perspect. Public Health* **138**, 87–88.
189. Joo H-S, Fu C-I, Otto M. 2016 Bacterial strategies of resistance to antimicrobial peptides. *Phil. Trans. R Soc. B* **371**, 20150292. (doi:10.1098/rstb.2015.0292)
190. Andersson DI, Hughes D, Kubicek-Sutherland JZ. 2016 Mechanisms and consequences of bacterial resistance to antimicrobial peptides. *Drug Resist. Updat.* **26**, 43–57. (doi:10.1016/j.drup.2016.04.002)
191. Assoni L, Milani B, Carvalho MR, Nepomuceno LN, Waz NT, Guerra MES, Converso TR, Darrieux M. 2020 Resistance mechanisms to antimicrobial peptides in Gram-positive bacteria. *Front. Microbiol.* **11**, 2362. (doi:10.3389/fmicb.2020.593215)
192. Kristian SA, Datta V, Weidenmaier C, Kansal R, Fedtke I, Peschel A, Gallo RL, Nizet V. 2005 d-Alanylation of teichoic acids promotes Group A *Streptococcus* antimicrobial peptide resistance, neutrophil survival, and epithelial cell invasion. *J. Bacteriol.* **187**, 6719. (doi:10.1128/JB.187.19.6719-6725.2005)
193. Gunn JS, Lim KB, Krueger J, Kim K, Guo L, Hackett M, Miller SI. 1998 PmrA–PmrB-regulated genes necessary for 4-aminoarabinose lipid A modification and polymyxin resistance. *Mol. Microbiol.* **27**, 1171–1182. (doi:10.1046/j.1365-2958.1998.00757.x)
194. Gooderham WJ, Bains M, McPhee JB, Wiegand I, Hancock REW. 2008 Induction by cationic antimicrobial peptides and involvement in intrinsic polymyxin and antimicrobial peptide resistance, biofilm formation, and swarming motility of PsrA in *Pseudomonas aeruginosa*. *J. Bacteriol.* **190**, 5624–5634. (doi:10.1128/JB.00594-08)
195. Boll JM, Tucker AT, Klein DR, Beltran AM, Brodbelt JS, Davies BW, Trent MS. 2015 Reinforcing lipid A acylation on the cell surface of *Acinetobacter baumannii* promotes cationic antimicrobial peptide resistance and desiccation survival. *mBio* **6**, e00478-15. (doi:10.1128/mBio.00478-15)
196. Anaya-López JL, López-Meza JE, Ochoa-Zarzosa A. 2013 Bacterial resistance to cationic antimicrobial peptides. *Crit. Rev. Microbiol.* **39**, 180–195. (doi:10.3109/1040841X.2012.699025)
197. LaRock CN, Nizet V. 2015 Cationic antimicrobial peptide resistance mechanisms of streptococcal pathogens. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1848**, 3047–3054. (doi:10.1016/j.bbamem.2015.02.010)
198. Zähner D, Zhou X, Chancey ST, Pohl J, Shafer WM, Stephens DS. 2010 Human antimicrobial

- peptide LL-37 induces MefE/Mel-mediated macrolide resistance in *Streptococcus pneumoniae*. *Antimicrob. Agents Chemother.* **54**, 3516–3519. (doi:10.1128/AAC.01756-09)
199. Kooi C, Sokol PA. 2009 *Burkholderia cenocepacia* zinc metalloproteases influence resistance to antimicrobial peptides. *Microbiology* **155**, 2818–2825. (doi:10.1099/mic.0.028969-0)
200. Gruenheid S, Le Moual H. 2012 Resistance to antimicrobial peptides in Gram-negative bacteria. *FEMS Microbiol. Lett.* **330**, 81–89. (doi:10.1111/j.1574-6968.2012.02528.x)
201. Sieprawska-Lupa M *et al.* 2004 Degradation of human antimicrobial peptide LL-37 by *Staphylococcus aureus*-derived proteinases. *Antimicrob. Agents Chemother.* **48**, 4673–4679. (doi:10.1128/AAC.48.12.4673-4679.2004)
202. Starr CG, Wimley WC. 2017 Antimicrobial peptides are degraded by the cytosolic proteases of human erythrocytes. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1859**, 2319–2326. (doi:10.1016/j.bbamem.2017.09.008)
203. Shin YP, Park HJ, Shin SH, Lee YS, Park S, Jo S, Lee YH, Lee IH. 2010 Antimicrobial activity of a halocidin-derived peptide resistant to attacks by proteases. *Antimicrob. Agents Chemother.* **54**, 2855–2866. (doi:10.1128/AAC.01790-09)
204. Hritonenko V, Stathopoulos C. 2007 OmpT proteins: an expanding family of outer membrane proteases in Gram-negative *Enterobacteriaceae* (Review). *Mol. Membr. Biol.* **24**, 395–406. (doi:10.1080/09687680701443822)
205. Brannon JR, Burk DL, Leclerc J-M, Thomassin J-L, Portt A, Berghuis AM, Gruenheid S, Le Moual H. 2015 Inhibition of outer membrane proteases of the OmpT family by aprotinin. *Infect. Immun.* **83**, 2300–2311. (doi:10.1128/IAI.00136-15)
206. Strömstedt AA, Pasupuleti M, Schmidtchen A, Malmsten M. 2009 Evaluation of strategies for improving proteolytic resistance of antimicrobial peptides by using variants of EFK17, an internal segment of LL-37. *Antimicrob. Agents Chemother.* **53**, 593–602. (doi:10.1128/AAC.00477-08)
207. Sabnis A, Ledger EVK, Pader V, Edwards AM. 2018 Antibiotic interceptors: creating safe spaces for bacteria. *PLoS Pathog.* **14**, e1006924. (doi:10.1371/journal.ppat.1006924)
208. Kuo HH, Chan C, Burrows LL, Deber CM. 2007 Hydrophobic interactions in complexes of antimicrobial peptides with bacterial polysaccharides. *Chem. Biol. Drug Des.* **69**, 405–412.
209. Chan C, Burrows LL, Deber CM. 2005 Alginate as an auxiliary bacterial membrane: binding of membrane-active peptides by polysaccharides. *J. Peptide Res.* **65**, 343–351. (doi:10.1111/j.1399-3011.2005.00217.x)
210. Jin L *et al.* 2016 A designed tryptophan- and lysine/arginine-rich antimicrobial peptide with therapeutic potential for clinical antibiotic-resistant *Candida albicans* Vaginitis. *J. Med. Chem.* **59**, 1791–1799. (doi:10.1021/acs.jmedchem.5b01264)
211. Zhang Z, Mu L, Tang J, Duan Z, Wang F, Wei L, Rong M, Lai R. 2013 A small peptide with therapeutic potential for inflammatory acne vulgaris. *PLoS ONE* **8**, e72923.
212. Spohn R *et al.* 2019 Integrated evolutionary analysis reveals antimicrobial peptides with limited resistance. *Nat. Commun.* **10**, 4538. (doi:10.1038/s41467-019-12364-6)
213. Kintses B *et al.* 2019 Phylogenetic barriers to horizontal transfer of antimicrobial peptide resistance genes in the human gut microbiota. *Nat. Microbiol.* **4**, 447–458. (doi:10.1038/s41564-018-0313-5)
214. Jahnson RD, Haney EF, Franzky H, Hancock REW. 2013 Characterization of a proteolytically stable multifunctional host defense peptidomimetic. *Chem. Biol.* **20**, 1286–1295. (doi:10.1016/j.chembiol.2013.09.007)
215. Frederick H, Sarah RD, David AP. 2009 Anionic antimicrobial peptides from eukaryotic organisms. *Curr. Protein Pept. Sci.* **10**, 585–606. (doi:10.2174/138920309789630589)
216. Frederick H, Sarah RD, David AP. 2011 Anionic antimicrobial peptides from eukaryotic organisms and their mechanisms of action. *Curr. Chem. Biol.* **5**, 142–153. (doi:10.2174/2212796811105020142)
217. Almarwani B, Phambu N, Hamada YZ, Sunda-Meya A. 2020 Interactions of an anionic antimicrobial peptide with zinc(II): application to bacterial mimetic membranes. *Langmuir* **36**, 14 554–14 562. (doi:10.1021/acs.langmuir.0c02306)
218. Lin Q, Deslouches B, Montelaro RC, Di YP. 2018 Prevention of ESKAPE pathogen biofilm formation by antimicrobial peptides WLBU2 and LL37. *Int. J. Antimicrob. Agents* **52**, 667–672. (doi:10.1016/j.ijantimicag.2018.04.019)
219. Fleeman RM, Macias LA, Brodbelt JS, Davies BW. 2020 Defining principles that influence antimicrobial peptide activity against encapsulated *Klebsiella pneumoniae*. *Proc. Natl. Acad. Sci. USA* **117**, 27620. (doi:10.1073/pnas.2007036117)
220. Zharkova MS, Orlov DS, Golubeva OY, Chakchir OB, Eliseev IE, Grinchuk TM, Shamova OV. 2019 Application of antimicrobial peptides of the innate immune system in combination with conventional antibiotics—a novel way to combat antibiotic resistance? *Front. Cell. Infect. Microbiol.* **9**, 128. (doi:10.3389/fcimb.2019.00128)
221. Li Z, Hu Y, Yang Y, Lu Z, Wang Y. 2018 Antimicrobial resistance in livestock: antimicrobial peptides provide a new solution for a growing challenge. *Animal Front.* **8**, 21–29. (doi:10.1093/af/fyy005)
222. Woolhouse M, Ward M, van Bunnik B, Farrar J. 2015 Antimicrobial resistance in humans, livestock and the wider environment. *Phil. Trans. R. Soc. B* **370**, 20140083. (doi:10.1098/rstb.2014.0083)
223. Volejníková A, Melicherčík P, Neštař O, Váňková E, Bednářová L, Rybáček J, Čeřovský V. 2019 Antimicrobial peptides prevent bacterial biofilm formation on the surface of polymethylmethacrylate bone cement. *J. Med. Microbiol.* **68**, 961–972. (doi:10.1099/jmm.0.001000)
224. Paquette DW, Simpson DM, Friden P, Braman V, Williams RC. 2002 Safety and clinical effects of topical histatin gels in humans with experimental gingivitis. *J. Clin. Periodontol.* **29**, 1051–1058. (doi:10.1038/j.1600-051X.2002.291201.x)
225. Easton DM, Nijnik A, Mayer ML, Hancock REW. 2009 Potential of immunomodulatory host defense peptides as novel anti-infectives. *Trends Biotechnol.* **27**, 582–590. (doi:10.1016/j.tibtech.2009.07.004)
226. Pushpakom S *et al.* 2019 Drug repurposing: progress, challenges and recommendations. *Nat. Rev. Drug Discovery* **18**, 41–58. (doi:10.1038/nrd.2018.168)
227. Farha MA, Brown ED. 2019 Drug repurposing for antimicrobial discovery. *Nat. Microbiol.* **4**, 565–577. (doi:10.1038/s41564-019-0357-1)
228. Christiansen SH *et al.* 2017 The immunomodulatory drug glatiramer acetate is also an effective antimicrobial agent that kills Gram-negative bacteria. *Sci. Rep.* **7**, 15653. (doi:10.1038/s41598-017-15969-3)
229. Skovdal SM, Christiansen SH, Johansen KS, Viborg O, Bruun NH, Jensen-Fangel S, Holm IE, Vorup-Jensen T, Petersen E. 2019 Inhaled nebulized glatiramer acetate against Gram-negative bacteria is not associated with adverse pulmonary reactions in healthy, young adult female pigs. *PLoS ONE* **14**, e0223647. (doi:10.1371/journal.pone.0223647)
230. Christiansen SH *et al.* 2017 The random co-polymer glatiramer acetate rapidly kills primary human leukocytes through sialic-acid-dependent cell membrane damage. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1859**, 425–437. (doi:10.1016/j.bbamem.2017.01.001)
231. Abdelaziz HB, Lemaire S, Carryn S, Van Bambeke F, Mingeot-Leclercq M-P, Tulkens PM. 2004 Inhibition of TNF- α production in THP-1 macrophages by glatiramer acetate does not alter their susceptibility to infection by *Listeria monocytogenes* and does not impair the efficacy of ampicillin or moxifloxacin against intracellular bacteria. *J. Antimicrob. Chemother.* **54**, 288–289. (doi:10.1093/jac/dkh288)
232. Thaker HD, Som A, Ayaz F, Lui D, Pan W, Scott RW, Anguita J, Tew GN. 2012 Synthetic mimics of antimicrobial peptides with immunomodulatory responses. *J. Am. Chem. Soc.* **134**, 11088–11 091. (doi:10.1021/ja303304j)
233. Scott RW, DeGrado WF, Tew GN. 2008 De novo designed synthetic mimics of antimicrobial peptides. *Curr. Opin. Biotechnol.* **19**, 620–627. (doi:10.1016/j.copbio.2008.10.013)
234. Gabriel GJ, Madkour AE, Dabkowski JM, Nelson CF, Nüsslein K, Tew GN. 2008 Synthetic mimic of antimicrobial peptide with nonmembrane-disrupting antibacterial properties. *Biomacromolecules* **9**, 2980–2983. (doi:10.1021/bm800855t)
235. Ghosh C, Haldar J. 2015 Membrane-active small molecules: designs inspired by antimicrobial peptides. *ChemMedChem.* **10**, 1606–1624. (doi:10.1002/cmdc.201500299)
236. Rotem S, Mor A. 2009 Antimicrobial peptide mimics for improved therapeutic properties. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1788**, 1582–1592. (doi:10.1016/j.bbamem.2008.10.020)

237. Lin S, Wade JD, Liu S. 2021 De novo design of flavonoid-based mimetics of cationic antimicrobial peptides: discovery, development, and applications. *Acc. Chem. Res.* **54**, 104–119. (doi:10.1021/acs.accounts.0c00550)
238. Koh J-J *et al.* 2013 Rapid bactericidal action of alpha-mangostin against MRSA as an outcome of membrane targeting. *Biochimica et Biophysica Acta (BBA) – Biomembranes* **1828**, 834–844. (doi:10.1016/j.bbamem.2012.09.004)
239. Lin S *et al.* 2017 Symmetrically substituted xanthone amphiphiles combat Gram-positive bacterial resistance with enhanced membrane selectivity. *J. Med. Chem.* **60**, 1362–1378. (doi:10.1021/acs.jmedchem.6b01403)
240. Koh J-J *et al.* 2015 Amino acid modified xanthone derivatives: novel, highly promising membrane-active antimicrobials for multidrug-resistant Gram-positive bacterial infections. *J. Med. Chem.* **58**, 739–752. (doi:10.1021/jm501285x)
241. Zou H *et al.* 2013 Design and synthesis of amphiphilic xanthone-based, membrane-targeting antimicrobials with improved membrane selectivity. *J. Med. Chem.* **56**, 2359–2373. (doi:10.1021/jm301683j)
242. Paulsen MH *et al.* 2021 Amphiphatic barbiturates as mimics of antimicrobial peptides and the marine natural products eusynstyelamides with activity against multi-resistant clinical isolates. *J. Med. Chem.* **64**, 11 395–11 417. (doi:10.1021/acs.jmedchem.1c00734)
243. Tai H-M, Huang H-N, Tsai T-Y, You M-F, Wu H-Y, Rajanbabu V, Chang H-Y, Pan C-Y, Chen J-Y. 2020 Dietary supplementation of recombinant antimicrobial peptide *Epinephelus lanceolatus* piscidin improves growth performance and immune response in *Gallus gallus domesticus*. *PLoS ONE* **15**, e0230021. (doi:10.1371/journal.pone.0230021)
244. Choi SC, Ingale SL, Kim JS, Park YK, Kwon IK, Chae BJ. 2013 An antimicrobial peptide-A3: effects on growth performance, nutrient retention, intestinal and faecal microflora and intestinal morphology of broilers. *Br. Poult. Sci.* **54**, 738–746. (doi:10.1080/00071668.2013.838746)
245. Xiong X, Yang HS, Li L, Wang YF, Huang RL, Li FN, Wang SP, Qiu W. 2014 Effects of antimicrobial peptides in nursery diets on growth performance of pigs reared on five different farms. *Livestock Sci.* **167**, 206–210. (doi:10.1016/j.livsci.2014.04.024)
246. Peng Z, Wang A, Xie L, Song W, Wang J, Yin Z, Zhou D, Li F. 2016 Use of recombinant porcine β -defensin 2 as a medicated feed additive for weaned piglets. *Sci. Rep.* **6**, 26790. (doi:10.1038/srep26790)
247. Cuperus T, van Dijk A, Matthijs MGR, Veldhuizen EJA, Haagsman HP. 2016 Protective effect of in ovo treatment with the chicken cathelicidin analog D-CATH-2 against avian pathogenic *E. coli*. *Sci. Rep.* **6**, 26622. (doi:10.1038/srep26622)
248. Mwangi J, Hao X, Lai R, Zhang Z-Y. 2019 Antimicrobial peptides: new hope in the war against multidrug resistance. *Zool. Res.* **40**, 488–505. (doi:10.24272/j.issn.2095-8137.2019.062)
249. Liu Y *et al.* 2020 A novel amphibian antimicrobial peptide, phyllosopeptin-PV1, exhibits effective anti-staphylococcal activity without inducing either hepatic or renal toxicity in mice. *Front. Microbiol.* **11**, 2713. (doi:10.3389/fmicb.2020.565158)
250. Duffy C *et al.* 2020 Honeybee venom and melittin suppress growth factor receptor activation in HER2-enriched and triple-negative breast cancer. *npj Precis. Oncol.* **4**, 24. (doi:10.1038/s41698-020-00129-0)
251. Sheard DE, O'Brien-Simpson NM, Wade JD, Separovic F. 2019 Combating bacterial resistance by combination of antibiotics with antimicrobial peptides. *Pure Appl. Chem.* **91**, 199–209. (doi:10.1515/pac-2018-0707)
252. Pizzolato-Cezar LR, Okuda-Shinagawa NM, Machini MT. 2019 Combinatory therapy antimicrobial peptide-antibiotic to minimize the ongoing rise of resistance. *Front. Microbiol.* **10**, 1703. (doi:10.3389/fmicb.2019.01703)
253. Naghmouchi K, Le Lay C, Baah J, Drider D. 2012 Antibiotic and antimicrobial peptide combinations: synergistic inhibition of *Pseudomonas fluorescens* and antibiotic-resistant variants. *Res. Microbiol.* **163**, 101–108. (doi:10.1016/j.resmic.2011.11.002)
254. Luong HX, Kim D-H, Lee B-J, Kim Y-W. 2017 Antimicrobial activity and stability of stapled helices of polybia-MP1. *Arch. Pharm. Res.* **40**, 1414–1419. (doi:10.1007/s12272-017-0963-5)
255. Li FF, Brimble MA. 2019 Using chemical synthesis to optimise antimicrobial peptides in the fight against antimicrobial resistance. *Pure Appl. Chem.* **91**, 181–198. (doi:10.1515/pac-2018-0704)
256. Shigeki H, Seiichi T. 2010 Activity improvement of antimicrobial peptides by a chemical modification approach: toward the creation of novel types of antimicrobial agents. *Mini-Rev. Organic Chem.* **7**, 282–289. (doi:10.2174/157019310792246373)
257. Di YP, Lin Q, Chen C, Montelaro RC, Doi Y, Deslouches B. 2020 Enhanced therapeutic index of an antimicrobial peptide in mice by increasing safety and activity against multidrug-resistant bacteria. *Sci. Adv.* **6**, eaay6817. (doi:10.1126/sciadv.aay6817)
258. Li W, Separovic F, O'Brien-Simpson NM, Wade JD. 2021 Chemically modified and conjugated antimicrobial peptides against superbugs. *Chem. Soc. Rev.* **50**, 4932–4973. (doi:10.1039/DOCS01026J)
259. Mijalis AJ, Thomas DA, Simon MD, Adamo A, Beaumont R, Jensen KF, Pentelute BL. 2017 A fully automated flow-based approach for accelerated peptide synthesis. *Nat. Chem. Biol.* **13**, 464–466. (doi:10.1038/nchembio.2318)
260. Hartrampf N *et al.* 2020 Synthesis of proteins by automated flow chemistry. *Science* **368**, 980. (doi:10.1126/science.abb2491)
261. Wang C, Hong T, Cui P, Wang J, Xia J. 2021 Antimicrobial peptides towards clinical application: delivery and formulation. *Adv. Drug Deliv. Rev.* **175**, 113818. (doi:10.1016/j.addr.2021.05.028)
262. Ndayishimiye J, Popat A, Blaskovich M, Falconer JR. 2020 Formulation technologies and advances for oral delivery of novel nitroimidazoles and antimicrobial peptides. *J. Control Release* **324**, 728–749. (doi:10.1016/j.jconrel.2020.05.002)
263. Tan P, Fu H, Ma X. 2021 Design, optimization, and nanotechnology of antimicrobial peptides: from exploration to applications. *Nano Today* **39**, 101229. (doi:10.1016/j.nantod.2021.101229)
264. Isidro-Llobet A, Kenworthy MN, Mukherjee S, Kopach ME, Wegner K, Gallou F, Smith AG, Roschangar F. 2019 Sustainability challenges in peptide synthesis and purification: from R&D to production. *J. Org. Chem.* **84**, 4615–4628. (doi:10.1021/acs.joc.8b03001)
265. Petrou C, Sarigiannis Y. 2018 1 - Peptide synthesis: methods, trends, and challenges. In *Peptide applications in biomedicine, biotechnology and bioengineering* (ed. S Koutsopoulos), pp. 1–21. Woodhead Publishing.
266. Pina AS, Lowe CR, Roque ACA. 2014 Challenges and opportunities in the purification of recombinant tagged proteins. *Biotechnol. Adv.* **32**, 366–381. (doi:10.1016/j.biotechadv.2013.12.001)
267. Sampaio de Oliveira KB *et al.* 2020 Strategies for recombinant production of antimicrobial peptides with pharmacological potential. *Expert Rev. Clin. Pharmacol.* **13**, 367–390. (doi:10.1080/17512433.2020.1764347)
268. Tahaeian A, Habibi Najafi MB, Rahnama P, Azghandi M. 2020 Production of a recombinant peptide (Lasioglossin LL III) and assessment of antibacterial and antioxidant activity. *Int. J. Pept. Res. Ther.* **26**, 1021–1029. (doi:10.1007/s10989-019-09904-5)
269. Silveira RF, Roque-Borda CA, Vicente EF. 2021 Antimicrobial peptides as a feed additive alternative to animal production, food safety and public health implications: an overview. *Anim. Nutr.* **7**, 896–904. (doi:10.1016/j.aninu.2021.01.004)
270. Annunziato G, Costantino G. 2020 Antimicrobial peptides (AMPs): a patent review (2015–2020). *Expert Opin. Ther. Patents* **30**, 931–947. (doi:10.1080/13543776.2020.1851679)