#### Check for updates

## OPEN ACCESS

EDITED BY Xiaofang Che, The First Affiliated Hospital of China Medical University, China

#### REVIEWED BY Alexey Tryakin, Russian Cancer Research Center NN Blokhin, Russia Vladimir Roudko, Mount Sinai Hospital, United States

\*CORRESPONDENCE Xiaoping Qian xiaopingqian@nju.edu.cn

<sup>†</sup>These authors have contributed equally to this work

#### SPECIALTY SECTION

This article was submitted to Cancer Immunity and Immunotherapy, a section of the journal Frontiers in Immunology

## RECEIVED 26 May 2022 ACCEPTED 20 July 2022 PUBLISHED 05 August 2022

#### CITATION

Zhang Q, Hu J, Zhang Y, Li L, Wang T and Qian X (2022) Case report: A colorectal cancer patient with microsatellite instability-high and *MSH2* germline mutation failed to respond to anti-PD-1 immunotherapy. *Front. Immunol.* 13:953421. doi: 10.3389/fimmu.2022.953421

#### COPYRIGHT

© 2022 Zhang, Hu, Zhang, Li, Wang and Qian. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and

that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

# Case report: A colorectal cancer patient with microsatellite instabilityhigh and *MSH2* germline mutation failed to respond to anti-PD-1 immunotherapy

# Qun Zhang<sup>1†</sup>, Jing Hu<sup>1†</sup>, Yaping Zhang<sup>2</sup>, Li Li<sup>1</sup>, Ting Wang<sup>3</sup> and Xiaoping Qian<sup>1\*</sup>

<sup>1</sup>Comprehensive Cancer Centre of Nanjing Drum Tower Hospital, Medical School of Nanjing University, Clinical Cancer Institute of Nanjing University, Nanjing, China, <sup>2</sup>The Comprehensive Cancer Center, Nanjing Drum Tower Hospital Clinical College of Xuzhou Medical University, Nanjing, China, <sup>3</sup>Department of Pathology, Nanjing University Medical School Affiliated Nanjing Drum Tower Hospital, Nanjing, China

Lynch syndrome (LS) is characterized by germline mutations in the DNA mismatch repair (MMR) genes. In colorectal cancer (CRC), germline mutations of DNA MMR genes commonly lead to microsatellite instability-high (MSI-H) subtype formation. Recent studies have demonstrated that CRC patients with MSI-H or mismatch repair-deficient (dMMR) status can benefit from anti-PD1 immunotherapy. However, almost 50% of CRC patients with MSI-H status do not respond to it. It is reported that heterogeneity of tumor and abnormal activation of cancer-related signaling pathways contribute to resistance to anti-PD1 therapy. To improve the clinical efficacy of such patients, the underlying mechanisms of resistance to anti-PD1 treatment must be explored. In this case, we describe an LS-associated CRC patient with MSI-H who suffered resistance to anti-PD1 therapy. Here, we attempted to elucidate the potential reasons, and thus appropriate strategies may be derived to overcome this clinical problem.

### KEYWORDS

microsatellite instability-high, colorectal cancer, anti-PD1 therapy, lynch syndrome, resistance

# Introduction

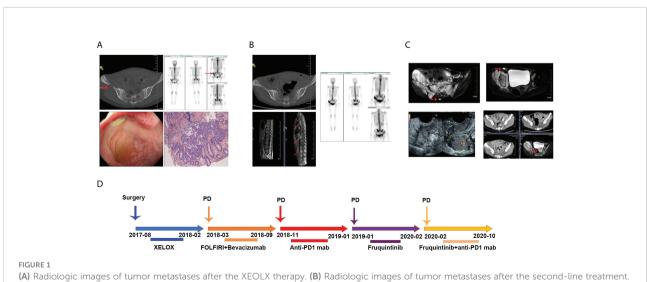
Lynch syndrome (LS) is caused by germline inactivation of one allele of genes involved in the mismatch repair (MMR) system, namely MLH1, MSH2, MSH6, and PMS2 (1). Inactivation of MLH1 or PMS2 alleles is the most frequent and is associated with approximately 80% of LS cases (2). LSassociated CRC usually presents as a microsatellite instabilityhigh (MSI-H) subtype. Several clinical trials have confirmed CRC patients with MSI-H/dMMR are the beneficiaries of anti-PD1 therapy, and the overall response rate varies from 40% to 60%. Based on these data, anti-PD1 monoclonal antibody (mAb) was approved for first-line treatment of advanced MSI-H CRC, which revolutionized the treatment mode of metastatic colorectal cancer (mCRC) (3). Clinical data have demonstrated the long-lasting and stable antitumor efficacy of anti-PD1 therapy in MSI-H mCRC (4).

Despite the remarkable efficacy of anti-PD1 therapy in advanced MSI-H/dMMR CRC patients, nearly half of MSI-H/ dMMR CRC patients do not respond to it. Studies have indicated that the underlying resistance mechanisms and the most straightforward reason are the absence of tumor antigens leads to a lack of recognition by T cells (5). Recently, abnormal activation of multiple tumor-associated signaling has been identified to contribute to the resistance of anti-PD1 mAb. The mitogen-activated protein kinase (MAPK) pathway, the PTEN expression, the PI3K signaling, and the WNT/b-catenin signaling pathway are the main immuneevasive oncogenic signaling pathways. Moreover, loss of interferon-gamma signaling pathways and lack of tumor antigen expression were also involved in tumor immune escape (6-11). With the development of gene sequencing platforms, sequencing analysis of MSI-H/dMMR subtypes of CRC revealed that some specific gene mutations also lead to resistance to PD1 mab therapy. Here, we present a CRC patient with MSI-H and germline MSH2 mutation who failed to respond to anti-PD1 treatment. Immunohistochemical (IHC), PCR, and nextgeneration (NGS) sequencing assays were used to explore the underlying mechanism of this resistance.

# **Case description**

## Case presentation and treatment

A 27-year-old young female patient presented with abdominal pain and with no excrement for 1day and was sent to the hospital for physical examination in July 2017. Colonoscopy examination revealed a space-occupying lesion 30cm from the anal margin. Pathological examination suggested poorly differentiated adenocarcinoma. Computed Tomography (CT) examination showed no metastasis. Then, this patient underwent a radical resection of left colon cancer. The postoperative pathological stage was pT3N2aM0. Postoperative baseline assessment showed no recurrence and metastasis in this patient. We chose the XELOX regimen as postoperative adjuvant chemotherapy. However, this patient had progressive disease (PD) after six cycles of XELOX chemotherapy. ECT and CT examination suggested bone destruction of the right iliac crest, which was considered metastasis. Colonoscopy revealed a neoplasm at the top of the anastomosis, and pathology revealed a moderately differentiated adenocarcinoma (Figure 1A). Since genetic test results suggested a KRAS gene mutation in this patient, bevacizumab combined with the FOLFIRI regimen was used as a second-line treatment for her. In addition, we performed local radiotherapy on her



(C) Radiologic images of tumor metastases after the xEOLX therapy. (D) Radiologic images of tumor metastases after the second-time treat (C) Radiologic images of tumor metastases after anti-PD1 immunotherapy. (D) The entire treatment process of this patient.

right iliac crest metastatic lesion. However, after two cycles of combined targeted therapy with chemotherapy treatment, she developed disease progression again, presenting with enlarged tissue mass shadow around the right iliac crest and multiple metastases in the thoracolumbosacral vertebral body and appendages (Figure 1B). We reviewed her tumor tissue immunohistochemical (IHC) results and found a loss of MSH2 protein (Figure 3B). PCR and NGS tests further confirmed an MSI-H status in this patient. NGS tests also indicated a high tumor mutation burden (TMB) in the patient's peripheral blood and tumor tissue, suggesting that the patient is likely to benefit from PD1 mab treatment. According to it, we chose anti-PD1 therapy (Tislelizumab 200mg every three weeks) as the third-line treatment. Unfortunately, this patient experienced a rapid progression again with widespread metastases including bone, ovary, and retroperitoneal lymph nodes after three cycles of anti-PD1 therapy (Figure 1C). When that, we chose furoquitinib and furoquitinib combined with sindilizumab as the follow-up treatment for her. During this period, this patient's lesions were in a state of slow progression. Due to her poor physical condition, she discontinued therapy in October 2020 and received the best supportive care. She died in April 2021. To intuitively express this patient's treatment process and efficacy, we listed the entire treatment process in Figure 1D.

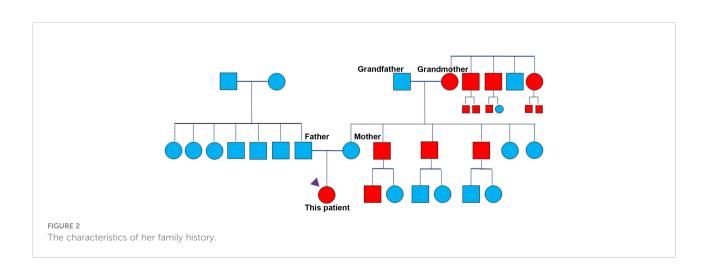
## Family history

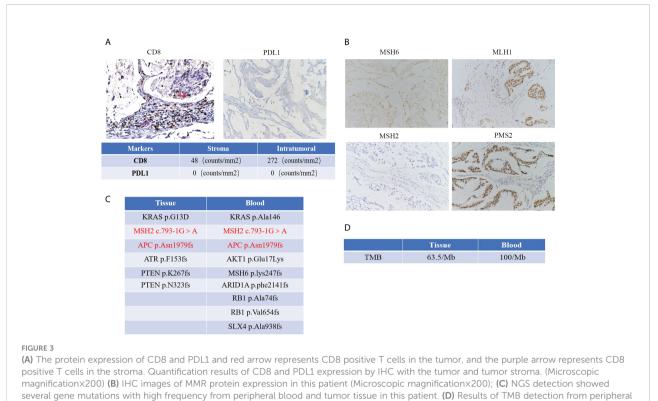
Interestingly, when we reviewed her family history, it was worth noting that her grandmother had CRC, three uncles had CRC, and her grandmother's siblings and their children all had CRC. Her family history fulfilled Amsterdam criteria. We presented her family history in Figure 2. NGS tests on blood and tumor tissue showed this patient had an *MSH2* germline mutation. Based on the above clinical and laboratory findings, she was diagnosed as a Lynch syndrome (LS) associated MSI-H CRC patient.

## IHC test and gene analysis

We used IHC assay to detect the protein expression of MLH1, MSH2, MSH6, PMS2, CD8, and PD-L1 in the formalin-fixed, paraffin-embedded (FFPE) tumor tissues. For quantitative analysis, we used ipp software to analyze the density of CD8 and PDL1. These data showed a small number of CD8+T cells were infiltrated in the tumor, and a large number of CD8+T cells were infiltrated in the stroma (Figure 3A). And we found a negative protein expression of PDL1 in this patient (Figure 3A).

The tissue DNA was extracted with a QIAamp DNA FFPE tissue kit (Qiagen, Valencia, CA, USA). Circulating tumor DNA (ctDNA) was extracted from plasma using the QIAamp Circulating Nucleic Acid Kit (Qiagen, Valencia, CA, USA). The tissue DNA and ctDNA were measured by Qubit 2.0 Fluorometer with a Qubit double-stranded DNA assay kit (Life Technologies, Carlsbad, CA, USA). Capture-based targeted sequencing was performed on tumor tissue and plasma samples using a panel (OncoScreen, Burning Rock Biotech, Guangzhou, China) consisting of 520 cancer-related genes. Sequencing data analysis was performed by OncoScreen Plus<sup>TM</sup>. The result from the peripheral blood test revealed nine gene mutations with high frequency, including KRAS p.Ala146, MSH2 c.793-1G>A, MSH6 p.lys247fs, AKT1 p.Glu17Lys, APC p.Asn1979fs, ARID1A p.phe2141fs, RB1 p.Ala74fs, RB1 p.Val654fs and SLX4 p.Ala938fs. The result from FFPE showed six gene mutations with high frequency, including KRAS p.G13D, MSH2 c.793-1G>A, APC p.Asn1979fs, ATR p.F153fs, PTEN p.K267fs and PTEN p.N323fs. These mutations were showed in Figure 3C. In addition, the results of tumor mutation burden from peripheral blood and tumor tissue were 100/Mb and 63.5/Mb, respectively (Figure 3D).





blood and tumor tissue in this patient.

# Discussion

This is a case of failure from anti-PD1 therapy in LSassociated MSI-H CRC. This is also a CRC case to describe characteristics such as TMB, tumor immune infiltration factors, and some gene alterations in LS-associated MSI-H CRC patients. In this case, we tried to elucidate the reasons for resistance to ant-PD1 immunotherapy.

Two forms of testing are commonly used in screening MMR or MSI status. IHC was used for detecting MMR proteins and PCR testing for MSI. Due to the development of gene sequencing platforms, NGS has been applied more and more in gene detection. In this case, the IHC assay revealed a loss of MSH2 protein expression. Besides, the PCR test also showed all five single nucleotide sites (BAT-25, MONO-27, CAT-25, BAT-26, and NR-24) were changed. Thus, there is no doubt that this patient is an MSI-H/dMMR CRC patient. As a germline mutation, LS-associated tumors are commonly microsatellite unstable. In this case, the NGS test showed this patient had an MSH2 germline mutation and high TMB. Combined with her family history, she was diagnosed with an LS-associated MSI-H CRC.

Recent studies have demonstrated that solid tumors with MSI-H/dMMR subset commonly obtained a favorable response from anti-PD1 mAb, including CRC (12–14). The excellent efficacy of anti-PD1 therapy in treating MSI-H/dMMR CRCs

is associated with the high expression levels of CD8 positive T cells within the tumor tissues (15, 16). The MSI-H CRC exhibits an active immune microenvironment probably due to recognizing many tumor neoantigens (17, 18). In this patient, the IHC assay showed a small number of CD8 positive T cells infiltrated within the tumor tissue, and many CD8 positive T cells were expressed in the tumor stroma, which may have contributed to the failure of immunotherapy.

The keynote-158 clinical trial has confirmed that TMB is a robust biomarker for predicting the efficacy of PD1 mab, with higher TMB indicating better efficacy (19). For TMB detection, peripheral blood and tumor tissues are generally selected. In this case, we found that the TMB in the peripheral blood was higher than in the tumor tissue. We speculated that a large amount of ctDNA in the lesion was released into the peripheral blood after the rapid tumor progression. In contrast, the tumor tissue only represented the TMB in this site.

Previous reports have suggested that MSI-H or immuneinfiltrated tumors have evolved mutations that may confer resistance to recognition by the immune system in untreated samples (5). It is observed that *APC* biallelic mutations associate with increased WNT signaling and decreased TILs in MSS and MSI-H tumors (20). Cen et al. reported that mutant *APC* promotes tumor immune evasion *via* PD-L1 in CRC (21). In our case, we found that this patient had an *APC* mutation, which may lead to resistance to anti-PD1 therapy. However, the protein

expression of PDL1 was negative in the tumor tissue of this patient, suggesting that in addition to APC gene mutation, there may be other gene dysfunction leading to immune tolerance. It is reported that PTEN gene mutation also correlated with response to anti-PD1 therapy. Chida K et al. showed that PTEN gene mutations in MSI-H/dMMR gastrointestinal tumors often did not respond to PD1 mab therapy (22). In addition, KRAS gene mutations are associated with poor anti-PD1 efficacy (23). In this patient, PTEN mutations were detected in the tumor tissue, and KRAS mutations were detected in the tumor tissue and peripheral blood. Therefore, the above gene mutations may be related to the failure of PD1 mab treatment. This suggests that the tumor immune microenvironment is very complex, and the factors determining immunotherapy's efficacy still need further exploration. Therefore, the patient's tumor microenvironment characteristics and detailed genomic status must be comprehensively evaluated before anti-PD1 mAb treatment, even for MSI-H tumors. In addition to MSI status, other gene mutations that may affect the therapeutic effect should also be considered comprehensively in screening the population with anti-PD1 treatment advantage among CRC patients.

In summary, this case may have significant clinical implications for MSI-H CRC patients' resistance to anti-PD1 mab therapy, especially those with LS-related MSI-H CRC.

## Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

# **Ethics statement**

Ethical review and approval was not required for the study involving human participants in accordance with the local legislation and institutional requirements. Written informed

# References

1. Latham A, Srinivasan P, Kemel Y, Shia J, Bandlamudi C, Mandelker D, et al. Microsatellite instability is associated with the presence of lynch syndrome pancancer. *J Clin Oncol* (2019) 37(4):286–95. doi: 10.1200/JCO.18.00283

2. Lynch HT, Kimberling W, Albano W, Lynch JF, Schuelke G, Sandberg AA, et al. Hereditary nonpolyposis colorectal cancer (Lynch syndromes I and II). *Clin Description Resource Cancer* (1985) 56:934–8. doi: 10.1002/1097-0142(19850815) 56:4<934::AID-CNCR2820560439>3.0.CO;2-I

3. oudko V, Cimen Bozkus C, Greenbaum B, Lucas A, Samstein R, Bhardwaj N. Lynch syndrome and MSI-h cancers: From mechanisms to "Off-The-Shelf" cancer vaccines. *Front Immunol* (2021) 12:757804. doi: 10.3389/fimmu.2021.757804

4. Ganesh K, Stadler ZK, Cercek A, Mendelsohn RB, Shia J, Segal NH, et al. Immunotherapy in colorectal cancer: rationale, challenges and potential. *Nat Rev Gastroenterol Hepatol* (2019) 16(6):361–75. doi: 10.1038/s41575-019-0126-x consent was obtained from the patient for the publication of any potentially identifiable images or data included in this article.

## Author contributions

XQ conceived the idea of the article. QZ composed the manuscript and figure-making. JH and LL supported the clinical data. TW and YZ provided the IHC test. All authors contributed to the article and approved the submitted version.

# Funding

This study was supported by Jiangsu Provincial Natural Science Foundation (SBK2021021543), the Nanjing Science and Technology Development Project (ZKX21028), and the High-level Innovation and Entrepreneurship Talent Introduction Plan of Jiangsu Province (JSSCBS20211532).

# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

# Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

5. Sharma P, Hu-Lieskovan S, Wargo JA, Ribas A. Primary, adaptive, and acquired resistance to cancer immunotherapy. *Cell* (2017) 168(4):707-23. doi: 10.1016/j.cell.2017.01.017

6. Peng W, Chen JQ, Liu C, Malu S, Creasy C, Tetzlaff MT, et al. Loss of PTEN promotes resistance to T cell-mediated immunotherapy. *Cancer Discovery* (2016) 6 (2):202–16. doi: 10.1158/2159-8290.CD-15-0283

7. De Henau O, Rausch M, Winkler D, Campesato LF, Liu C, Cymerman DH, et al. Overcoming resistance to checkpoint blockade therapy by targeting PI3K $\gamma$  in myeloid cells. *Nature* (2016) 539(7629):443–7. doi: 10.1038/nature20554

8. Liu C, Peng W, Xu C, Lou Y, Zhang M, Wargo JA, et al. BRAF inhibition increases tumor infiltration by T cells and enhances the antitumor activity of adoptive immunotherapy in mice. *Clin Cancer Res* (2013) 19(2):393–403. doi: 10.1158/1078-0432.CCR-12-1626

9. Luke JJ, Bao R, Sweis RF, Spranger S, Gajewski TF. WNT/β-catenin pathway activation correlates with immune exclusion across human cancers. *Clin Cancer Res* (2019) 25(10):3074–83. doi: 10.1158/1078-0432.CCR-18-1942

10. Gubin MM, Zhang X, Schuster H, Caron E, Ward JP, Noguchi T, et al. Checkpoint blockade cancer immunotherapy targets tumour-specific mutant antigens. *Nature* (2014) 515(7528):577–81. doi: 10.1038/nature13988

11. Gao J, Shi LZ, Zhao H, Chen J, Xiong L, He Q, et al. Loss of IFN-γ pathway genes in tumor cells as a mechanism of resistance to anti-CTLA-4 therapy. *Cell* (2016) 167(2):397–404.e9. doi: 10.1016/j.cell.2016.08.069

12. Motta R, Cabezas-Camarero S, Torres-Mattos C, Riquelme A, Calle A, Figueroa A, et al. Immunotherapy in microsatellite instability metastatic colorectal cancer: Current status and future perspectives. J Clin Transl Res (2021) 7(4):511–22.

13. Gelsomino F, Barbolini M, Spallanzani A, Pugliese G, Cascinu S. The evolving role of microsatellite instability in colorectal cancer: A review. *Cancer Treat Rev* (2016) 51:19–26. doi: 10.1016/j.ctrv.2016.10.005

14. Schrock AB, Ouyang C, Sandhu J, Sokol E, Jin D, Ross JS, et al. Tumor mutational burden is predictive of response to immune checkpoint inhibitors in MSI-high metastatic colorectal cancer. *Ann Oncol* (2019) 30(7):1096–103. doi: 10.1093/annonc/mdz134

15. Jenkins MA, Hayashi S, O'Shea AM, Burgart LJ, Smyrk TC, Shimizu D, et al. Pathology features in Bethesda guidelines predict colorectal cancer microsatellite instability: a population-based study. *Gastroenterology* (2007) 133(1):48–56. doi: 10.1053/j.gastro.2007.04.044

16. Phillips SM, Banerjea A, Feakins R, Li SR, Bustin SA, Dorudi S. Tumourinfiltrating lymphocytes in colorectal cancer with microsatellite instability are activated and cytotoxic. *Br J Surg* (2004) 91(4):469–75. doi: 10.1002/bjs.4472 17. Raeker MO, Carethers JM. Immunological features with DNA microsatellite alterations in patients with colorectal cancer. *J Cancer Immunol (Wilmington)* (2020) 2(3):116–27. doi: 10.33696/cancerimmunol.2.024

18. Kim JH, Seo MK, Lee JA, Yoo SY, Oh HJ, Kang H, et al. Genomic and transcriptomic characterization of heterogeneous immune subgroups of microsatellite instability-high colorectal cancers. *J Immunother Cancer* (2021) 9 (12):e003414. doi: 10.1136/jitc-2021-003414

19. Marabelle A, Fakih M, Lopez J, Shah M, Shapira-Frommer R, Nakagawa K, et al. Association of tumour mutational burden with outcomes in patients with advanced solid tumours treated with pembrolizumab: prospective biomarker analysis of the multicohort, open-label, phase 2 KEYNOTE-158 study. *Lancet Oncol* (2020) 21(10):1353–65. doi: 10.1016/S1470-2045(20)30445-9

20. Hankey W, McIlhatton MA, Ebede K, Kennedy B, Hancioglu B, Zhang J, et al. Mutational mechanisms that activate wnt signaling and predict outcomes in colorectal cancer patients. *Cancer Res* (2018) 78(3):617–30. doi: 10.1158/0008-5472.CAN-17-1357

21. Cen B, Wei J, Wang D, Xiong Y, Shay JW, DuBois RN. Mutant APC promotes tumor immune evasion *via* PD-L1 in colorectal cancer. *Oncogene* (2021) 40(41):5984–92. doi: 10.1038/s41388-021-01972-6

22. Chida K, Kawazoe A, Kawazu M, Suzuki T, Nakamura Y, Nakatsura T, et al. A low tumor mutational burden and PTEN mutations are predictors of a negative response to PD-1 blockade in MSI-H/dMMR gastrointestinal tumors. *Clin Cancer Res* (2021) 27(13):3714–24. doi: 10.1158/1078-0432.CCR-21-0401

23. Liao W, Overman MJ, Boutin AT, Shang X, Zhao D, Dey P, et al. KRAS-IRF2 axis drives immune suppression and immune therapy resistance in colorectal cancer. *Cancer Cell* (2019) 35(4):559–72. doi: 10.1016/j.ccell.2019.02.008