

Serum Bisphenol A Concentration in Postmenopausal Women with Osteoporosis

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= Abstract =

Objectives: Bishphenol A (BPA) is a representative endocrine disruptor and is also known as a xenoestrogen. The objective of the present study is to investigate how many patients are exposed to BPA and to analyze the relationships between serum BPA concentration, bone mineral density (BMD) and biochemical bone markers in postmenopausal women with osteoporosis.

Methods: Total 51 patients were enrolled for measuring BPA and clinical variables including BMD and bone markers. The relationship between BPA and clinical variables were analyzed by the Pearson's correlation test and the Kruskal-Wallis test. Serum BPA concentration was measured by enzyme linked immunosorbent assay (ELISA).

Results: BPA was detected in all samples. The mean BPA concentration was 1.44 ± 0.52 ng/mL. There was no statistically significant correlation between BPA and clinical variables.

Conclusion: There was no statistical significance between serum BPA concentration and clinical variables related to bone metabolism. To clarify the effect of BPA on bone metabolism, further large scaled and high risk group investigation may be needed.

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Key Words: Bisphenol A, Bone markers, Bone mineral density, Osteoporosis

INTRODUCTION

Bisphenol A (BPA) is well known as a representative endocrine disruptor interrupting body functions controlled by hormones.[1] Approximately 2.2 million tons of BPA are produced annually all around the world and widely applied in the manufacture of an epoxy resin, mainly used to produce a container or a package for foods and beverages, or

a polycarbonate plastic.[2] BPA is widely and continuously exposed to human through a diet and is also exposed by the other paths such as through skin and dust etc. In fact, according to a research performed in the US, measureable BPA was found in approximately 93-95% of the subjects through urine.[3,4]

The mechanism that BPA shows inappropriate effect in the body is known as estrogen like mechanism;[5] besides,

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a decrease of the function in a pancreatic beta cell,[6] an interruption of the function in a thyroid hormone,[7] an obesity acceleration effect,[8] and the induction of an oxidative stress and an inflammation[9] are included. According to recent studies, the relationship with endocrine related diseases, including diabetes, cardiovascular diseases, liver diseases, and semen abnormalities have brought up constantly,[10,11]

Osteoporosis is a skeletal disease occurred in postmenopausal women and elderly and increases the risk of a bone fracture due to a bone mass reduction and microstructure deterioration.[12] Of the known mechanisms up to date, an estrogen deficiency is the most important factor in the osteoporosis occurrence.[13]

Considering the relationship of an estrogen deficiency and an osteoporosis occurrence, a xenoestrogen is also thought to be related to the occurrence of osteoporosis; but until today some studies investigating the relationship of BPA, a xenoestrogen, and bone metabolism have only performed in in vitro or utilizing animals. Suzuki et al.[14] reported that tartrate-resistant acid phosphatase (TRAP) and alkaline phosphatase (ALP), the biochemical bone marker, decreased significantly when injecting BPA in in vitro experiment using goldfish scales. Also Seidlová-Wuttke et al.[15] examined that bone-forming factors increased and boneresorbing factors decreased in the rat group that a higher volume of BPA was injected but a bone mineral density (BMD) decreased. However there was no study investigating the relationship of a blood BPA concentration and osteoporosis in human until today. The objectives of the study were to examine how many patients were exposed to BPA by measuring serum BPA in the patients with osteoporosis and to analyze the relationships with a BMD and a biochemical bone marker.

SUBJECTS AND METHODS

1. Subjects

In order to examine the relationship between serum BPA concentration and a BMD and a bone marker, the study was carried out in 51 postmenopausal women who had a

treatment due to osteoporosis from October 2008 to July 2009 in Kyung Hee University Hospital at Gangdong; all patients were lived in Seoul or Hanam-si. Of the postmenopausal women who are more than 50 years old, the subjects were in the case where T-score of an average lumbar (L1-4), a femoral neck, or the total of BMD was less than -2.5 or T-score was less than -1.0 with at least more than a radiologic finding in a spine fracture. Menopause was defined in case where there was no natural menstruation more than a year at least or follicle stimulating hormone (FSH) exceeds 40 IU/L in case of the subjects with hysterectomy. There was no patient taking medications affecting bone or calcium metabolism or having a malignant tumor or other systemic diseases. The present study was performed after the review of the organization review committee and a written consent form was received from all the participants.

2. Methods

Body mass index (BMI), blood 25-hydroxyvitamin D (25[OH]D; DiaSorin Corporation, Stillwater, MN, USA), C-telopeptide of collagen cross-links (CTX; Roche Diagnostics, Mannheim, Germany), and osteocalcin (OC; CIS Bio International, Gif-sur-Yvette, France) of the subjects were investigated at the time of measuring the BMD. The BMD was measured in the lumber number 1 through 4, a total femur, and a femoral neck using dual energy X-ray absorptiometry (DXA; Lunar Prodigy Advance, GE Lunar, Madison, WI, USA) and was expressed as g/cm².

In the study, a pre-processing was carried out in order to measure BPA concentration of the serum separated from blood in the subjects. The samples were passed through a SPE column (Isolute M-M cartridge) in order to remove high molecular proteins and lipid components in the serum and concentrate the samples. Then the samples were washed with 200 μ L of 10% methanol. An extraction was carried out using 200 μ L of an extraction solution (methanol: acetonitrile, 3:1). Once evaporated, the samples were reconstituted in 40 μ L of analysis buffer (0.01 M phosphate buffer, pH 7.4).

After such procedures, the blood BPA concentration was

Table 1. Baseline characteristics of the patients

Patients (n = 51)	
64.5 ± 7.0	
None	
1.44 ± 0.52 (Range: $0.26-2.51$)	
-2.87 ± 0.45	
23.0 ± 2.7	
17.8 ± 6.6	
0.55 ± 0.24	
15.1 ± 3.8	

Data are expressed as means \pm SD

BMD, bone mineral density; BMI, body mass index; 25(OH)D, 25-hydroxyvitamin D; CTX, C-telopeptide of collagen cross-links

measured utilizing Ecologiena BPA enzyme linked immunosorbent assay (ELISA) Kit (Japan Environchemicals, Tokyo, Japan). The measurement method was developed by utilizing the characteristic that a monoclonal antibody bound to BPA only and didn't bind to any other chemical substances with a similar structure. A quantitative analysis range was 0.05-10 μ g/L (ppb) and sensitivity was good enough to measure BPA present in the serum. The results were shown a high reproducibility and coefficient variation (CV) was less than 10% in general.

3. Statistical analysis

Results were expressed in an average and a standard deviation and significance of the relationship between clinical factors including blood BPA concentration of the experimental group, a BMD, and a biochemical bone marker etc were verified utilizing the Pearson's correlation test. Additionally, partial correlation analysis was used to control the confounding variables such as age and BMI that could have an influence on the blood BPA concentration. Also the subjects were classified into three groups based upon several clinical factors and then Kruskal-Wallis test was used in order to compare average BPA concentration of each group. It was considered statistically significant when *P* is less than 0.05 in all cases. Statistical analysis was

Table 2. Correlation coefficients between bisphenol A concentration and clinical variables

Bisphenol A	Correlation coefficients	P-value
Age (year)	0.144	0.312
BMI (kg/m ²)	0.008	0.955
BMD (T-score)	0.120	0.402
25(OH)D (ng/mL)	0.095	0.505
CTX (U/L)	0.029	0.840
Osteocalcin (µg/L)	-0.024	0.867

Correlation is significant at the 0.05 level

BMI, body mass index; BMD, bone mineral density; 25(OH)D, 25-hydroxyvitamin D; CTX, C-telopeptide of collage cross-links

performed utilizing SPSS version 18.0 (SPSS Inc., Chicago, IL, USA).

RESULTS

The average age of 51 subjects participated in the study was 64.5 (50-82 years old) and all of them were non-smokers. The subjects showed the pattern of overweight with BMI 23 and the BMD T-score was -2.87. Average 25(OH)D concentration was 17.8 ng/mL (Table 1).

All subjects showed BPA of more than measureable concentration. The average concentration of the blood BPA was 1.44 ng/mL and didn't show statistically significant relationship with the BMD. Also 25(OH)D, CTX, and OC measured as clinical variables related to bone metabolism had no statistically significant relationship with the blood BPA concentration (Table 2). The results were examined in the same manner when correcting based upon age and BMI.

Since few subjects were participated in the study, the clinical variables including a BMD and the concentration of a biochemical bone marker etc were classified into three groups (average - under 1 standard deviation [SD], average \pm 1SD, average + 1SD excess group) based upon averages and then analyzed if there was a difference in blood BPA concentration. No significant difference was observed in the blood BPA concentration of all indexes among the groups (Table 3).



Table 3. Kruskal-Wallis test of the bisphenol A according to clinical variables

	< Mean - 1SD	Mean ± 1SD	> Mean - 1SD	<i>P</i> -value
•	Mean cond	•		
Age (year)	< 58	58-71	> 71	0.020
	1.37	1.44	1.50	0.938
BMI (kg/m ²)	< 20.3	20.3-25.7	> 25.7	0.567
	1.26	1.53	1.35	0.567
BMD (T-score)	< -3.20	-3.302.50	> -2.50	0.060
	1.37	1.45	1.50	0.869
25(OH)D (ng/mL)	< 11.2	11.2-24.4	> 24.4	0.500
	1.58	1.40	1.48	0.509
CTX (U/L)	< 0.31	0.31-0.80	> 0.80	0.420
	1.15	1.50	1.40	0.429
Osteocalcin (µg/L)	< 11.3	11.3-18.9	> 19.9	0.704
	1.45	1.46	1.29	0.724

Statistically significant at the 0.05 level

BMI, body mass index; BMD, bone mineral density; 25(OH)D, 25-hydroxyvitamin D; CTX, C-telopeptide of collagen cross-links

DISCUSSION

Environmental estrogens are parts of xenoestrogens and bind to estrogen receptors thereby exhibiting their functions. If organs with estrogen receptors are continuously exposed to such xenoestrogens, they may either act like inherent estrogens that are synthesized in the body or represent opposite effects as xenoestrogens inhibit the bindings between inherent estrogens and receptors. In the former cases, xenoestrogens act agonistically while they affect antagonistically in the later cases. Of the xenoestrogens, representative environmental estrogens include BPA and benzopyrene which are classified as endocrine disruptors

A environmental estrogen, BPA which is a compound of interests in the study, shows various exposure routes. It is mainly detected from plastic products and widely used for baby bottles, bowls, food containers, and cans as well as the production of epoxyresin which is in the amalgam for the dental clinic.[16] Lately, the exposure of BPA via medical

devices has been an issue; it is known that there are highly frequent exposures for the patients with hemodialysis as circulating blood contact the polycarbonate devices of hemodialysis and extract BPA when performing the dialysis. [17] Furthermore, according to Le et al.[18], if boiling water was added, there was a 55 fold increase in the BPA concentrations compared to that of room temperature. It is also known that repetitive usages of containers are correlated with increase of BPA concentrations.[19] These absorbed BPA in the body are biological active unconjugated form but quickly conjugated in the liver so that excreted via bile or urine; the half life of this compound is 5.3 hr which is relatively short.[20-22]

The fast excretion of BPA due to its short half life has been adopted for the scientific evidences of BPA safety, but β-glucuronidases in multiples organs, especially lung, liver, and kidneys of rats and placenta of human deconjugate BPA thereby excreting active BPA.[23] Furthermore, according to recent studies, BPA represents 3.3 of a coefficient of correlation which is lipophilic and exhibits approximately 3 times higher affinity compared to kidneys and muscles, meaning that there are higher chances of accumulation and it may continuously affect the body metabolism.[29]

The concentrations of BPA in blood of postmenopausal women with osteoporosis were 1.44 ng/mL, which are significant. Compared to the values (0.437 \pm 0.036 ng/mL, 83.9% detection rate) of population from the domestic study with uterine myoma patients, [27] these were relatively high levels. Such differences might because of higher exposure and accumulation of BPA in the body owing to relatively high ages and BMI of subjects as well as differences in residential areas. In addition, methods for BPA measurements, subjects, and results from domestic and foreign studies, are listed in Table 4. Depending upon the research subjects and various conditions, concentrations of BPA in the blood were different; yet it represented the significant association with female hormones related diseases including endometrial hyperplasia, habitual abortion, endometriosis, and polycystic ovary syndrome except for the domestic study with uterine myoma patients

Likewise, the osteoporosis is one of the representative

Table 4. Bisphenol A levels in female serum

Author	Year	Subject	Method	Levels (ng/mL)
Hiroi et al.[24]	2004	Healthy female & Simple endometrial hyperplasia	ELISA	$\begin{array}{c} 2.5 \pm 0.452 \\ 2.9 \pm 0.932 \end{array}$
Sugiura-Ogasawara et al.[25]	2005	Women with recurrent miscarriage	ELISA	2.59 ± 0.780
Cobellis et al.[26]	2009	Endometriotic female	RP-HPLC/MS	2.91 ± 1.74
Byun et al.[27]	2010	Normal female Leiomyoma female	ELISA	0.557 ± 0.086 0.401 ± 0.038
Kandaraki et al.[28]	2011	Normal female PCOS female	ELISA	0.72 ± 0.37 1.05 ± 0.56

ELISA, enzyme linked immunosorbent assay; RP-HPLC-MS, reversed-phase-high-pressure liquid chromatography/mass spectrometry; PCOS, polycystic ovary syndrome

female hormones related diseases. So far, however there are only limited numbers of *in vivo* studies have been done with regard to the association between bone metabolism and BPA, a xenoestrogen. This study is the first study investigated the association between the blood BPA concentration and bone metabolism. Considering the effects of BPA, as a xenoestrogen, it was expected that there would either positive or negative correlation between the BPA level and biochemical markers that represent dynamic changes, but any significant correlation was not observed.

There were several limitations to draw significant results. First, there were a relatively small number of patient subjects. Second, although aging and deficits of female hormones are known risk factors for osteoporosis, they are only a part of multiple risk factors, meaning that this environmental estrogen, BPA would be just one of the many factors for such diseases; thus a consideration should be taken account regarding its limited effects on such diseases. Third, other than estrogen related one, the mode of action of BPA that affects the bone metabolism includes various mechanisms including the lipid metabolism as well as oxidative stress. Fourth, there is an issue in regards to the measurement of BPA concentration. That is, it would be necessary to understand better 1) how to measure the BPA 2) whether blood samples would reflect the accumulation of BPA in the body better than urine samples, or vice versa 3) if a single measurement of blood BPA would accurately show the accumulation level of BPA in the body. It is

considered that there is a limitation to conclude about the causal relationship between the blood BPA concentration and the BMD as well as bone factors. Lastly, although we performed the partial correlation analysis for potential confounding variables (e.g., age, and BMI) regarding the blood BPA concentration, it might not be corrected enough and analyzed. Given the limitations, further additional large scale studies with various subject groups are warranted in order to clarify such relationships.

CONCLUSION

It has been emerged that the association between an endocrine disruptor such as BPA, a xenoestrogen, and various diseases. To the best of our knowledge, it was the first time to investigate the relationship between bone metabolism and BPA in human, but there were no significant correlations between the blood BPA concentration and a BMD as well as a biochemical bone marker. Further investigations are warranted for future.

REFERENCES

 Talsness CE, Andrade AJ, Kuriyama SN, et al. Components of plastic: experimental studies in animals and relevance for human health. Philos Trans R Soc Lond B Biol Sci 2009;364: 2079-96.



- 2. Burridge E. Bisphenol A: product profile. Eur Chem News 2003;17:14-20.
- Calafat AM, Kuklenyik Z, Reidy JA, et al. Urinary concentrations of bisphenol A and 4-nonylphenol in a human reference population. Environ Health Perspect 2005;113:391-5.
- Calafat AM, Ye X, Wong LY, et al. Exposure of the U.S. population to bisphenol A and 4-tertiary-octylphenol: 2003-2004. Environ Health Perspect 2008;116:39-44.
- Wozniak AL, Bulayeva NN, Watson CS. Xenoestrogens at picomolar to nanomolar concentrations trigger membrane estrogen receptor-alpha-mediated Ca2+ fluxes and prolactin release in GH3/B6 pituitary tumor cells. Environ Health Perspect 2005;113:431-9.
- Ropero AB, Alonso-Magdalena P, Garcia-Garcia E, et al. Bisphenol-A disruption of the endocrine pancreas and blood glucose homeostasis. Int J Androl 2008;31:194-200.
- Moriyama K, Tagami T, Akamizu T, et al. Thyroid hormone action is disrupted by bisphenol A as an antagonist. J Clin Endocrinol Metab 2002;87:5185-90.
- Newbold RR, Padilla-Banks E, Jefferson WN, et al. Effects of endocrine disruptors on obesity. Int J Androl 2008;31:201-8.
- Yang YJ, Hong YC, Oh SY, et al. Bisphenol A exposure is associated with oxidative stress and inflammation in postmenopausal women. Environ Res 2009;109:797-801.
- Lang IA, Galloway TS, Scarlett A, et al. Association of urinary bisphenol A concentration with medical disorders and laboratory abnormalities in adults. JAMA 2008;300:1303-10.
- 11. Li DK, Zhou Z, Miao M, et al. Urine bisphenol-A (BPA) level in relation to semen quality. Fertil Steril 2011;95:625-30 e1-4.
- Lirani-Galvão AP, Lazaretti-Castro M. Physical approach for prevention and treatment of osteoporosis. Arq Bras Endocrinol Metabol 2010;54:171-8.
- ESHRE Capri Workshop Group. Bone fractures after menopause. Hum Reprod Update 2010;16:761-73.
- Suzuki N, Hattori A. Bisphenol A suppresses osteoclastic and osteoblastic activities in the cultured scales of goldfish. Life Sci 2003;73:2237-47.
- Seidlová-Wuttke D, Jarry H, Wuttke W. Pure estrogenic effect of benzophenone-2 (BP2) but not of bisphenol A (BPA) and dibutylphtalate (DBP) in uterus, vagina and bone. Toxicology 2004;205:103-12.
- 16. Haishima Y, Hayashi Y, Yagami T, et al. Elution of bisphenol-A

- from hemodialyzers consisting of polycarbonate and polysulfone resins. J Biomed Mater Res 2001;58:209-15.
- Murakami K, Ohashi A, Hori H, et al. Accumulation of bisphenol A in hemodialysis patients. Blood Purif 2007;25: 290-4.
- Le HH, Carlson EM, Chua JP, et al. Bisphenol A is released from polycarbonate drinking bottles and mimics the neurotoxic actions of estrogen in developing cerebellar neurons. Toxicol Lett 2008;176:149-56.
- Nam SH, Seo YM, Kim MG. Bisphenol A migration from polycarbonate baby bottle with repeated use. Chemosphere 2010;79:949-52.
- Yokota H, Iwano H, Endo M, et al. Glucuronidation of the environmental oestrogen bisphenol A by an isoform of UDPglucuronosyltransferase, UGT2B1, in the rat liver. Biochem J 1999;340 (Pt 2):405-9.
- Völkel W, Colnot T, Csanády GA, et al. Metabolism and kinetics of bisphenol a in humans at low doses following oral administration. Chem Res Toxicol 2002;15:1281-7.
- 22. Inoue H, Yuki G, Yokota H, et al. Bisphenol A glucuronidation and absorption in rat intestine. Drug Metab Dispos 2003;31: 140-4.
- Ginsberg G, Rice DC. Does rapid metabolism ensure negligible risk from bisphenol A? Environ Health Perspect 2009;117: 1639-43.
- 24. Hiroi H, Tsutsumi O, Takeuchi T, et al. Differences in serum bisphenol a concentrations in premenopausal normal women and women with endometrial hyperplasia. Endocr J 2004;51: 595-600.
- Sugiura-Ogasawara M, Ozaki Y, Sonta S, et al. Exposure to bisphenol A is associated with recurrent miscarriage. Hum Reprod 2005;20:2325-9.
- Cobellis L, Colacurci N, Trabucco E, et al. Measurement of bisphenol A and bisphenol B levels in human blood sera from healthy and endometriotic women. Biomed Chromatogr 2009; 23:1186-90.
- Byun JC, Han MS. Serum bisphenol-A concentrations from uterine leiomyoma patients. Korean J Obstet Gynecol 2010; 53:915-20.
- 28. Kandaraki E, Chatzigeorgiou A, Livadas S, et al. Endocrine disruptors and polycystic ovary syndrome (PCOS): elevated serum levels of bisphenol A in women with PCOS. J Clin

Endocrinol Metab 2011;96:E480-4.

29. Csanády GA, Oberste-Frielinghaus HR, Semder B, et al. Distribution and unspecific protein binding of the xeno-

estrogens bisphenol A and daidzein. Arch Toxicol 2002;76: 299-305.