

SHORT COMMUNICATION



Fluoroenesulphonamides: *N*-sulphonylurea isosteres showing nanomolar selective cancer-related transmembrane human carbonic anhydrase inhibition

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ABSTRACT

After hydrofluorination of ynesulphonamides in superacid or in the presence of hydrofluoric acid/base reagents, a series of α -fluoroenamides has been synthesised and tested for the inhibition of carbonic anhydrase (CA, EC 4.2.1.1) isoforms. This study reveals a new, highly selective family of cancer-related transmembrane human (h) CA IX/XII inhibitors. These original fluorinated ureido isosteres do not inhibit the widespread cytosolic isoforms hCA I and II and selectively inhibit the transmembrane cancer-related hCA IX and XII, offering interesting new leads for future studies.

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Introduction

The elevated metabolic rate of solid cancer tumors leads frequently to acidosis and hypoxia¹, which can be directly related to spatial disorganisation and flow-based disruption of an abnormal microvascularisation initiated by the growing tumor². Under hypoxia stress exposition, tumor cells respond by transcription hypoxia inducible factor-1 (HIF-1 α -activates), reprogramming their metabolism to overcome the reduced supply of oxygen^{3,4}. The engaged nonoxygen-dependent glycolytic pathway results in increased production and export of lactic and carbonic acids to the extracellular proximal milieu, therefore decreasing extracellular pH⁵, which induces a variation of intracellular/extracellular pH ratio (pH_i/pH_e ratio). This is regulated by different players including transmembrane carbonic anhydrases IX and XII (CA IX and CA XII) which are overexpressed in human cancer cells⁶. As a consequence, CA IX and CA XII are now recognised as especially relevant targets for cancer therapy. Sulphonamides and their bioisosteres (sulphamates, sulphamides, etc.) constitute the most investigated inhibitors of these enzymes⁷, with useful therapeutic applications⁸. They act on their deprotonated forms and bind the Zn²⁺ ion of the active site, disrupting the catalytic process⁹. However, this class of inhibitors suffers from side-effects that are directly related to the undesired inhibition of the cytosolic isoform I and II, abundant in many tissues and involved in numerous physiological functions¹⁰. As a consequence, numerous efforts were dedicated over the last years to the evaluation of non-zinc binding inhibitors. This resulted for example in the discovery that coumarins, thiocoumarins¹¹ and, more recently, sulphocoumarins¹², located at the entrance of the enzyme active site, were

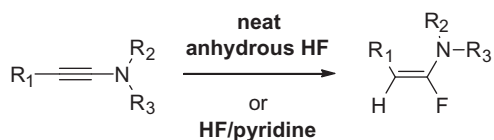
selective inhibitors of hCA IX isozyme. Our group recently contributed to this field by exploring the activity of tertiary benzenesulphonamides derivatives: substituted *N*-aryl-benzenesulphamides were found to act as selective nanomolar inhibitors of hCAs IX and XII^{13,14}. Despite good affinity/selectivity to hCA IX and excellent stability in plasma, a study with their ¹⁸F-labelled analogues however showed no significant uptake in HT-29 tumors compared to normal organs/tissues¹⁵.

Considering the recent discovery of the urea derivative SLC-0111 which successfully ended Phase I clinical programmes for the treatment of patients with advanced hypoxic tumors over-expressing the isoforms hCA IX/XII^{16,17} and by the impact of ureas on pharmacokinetic properties, the evaluation of the corresponding sulphonylurea analogues must find interest. However, recent studies on the exploitation of bisarenesulphonylureas as anti-cancer agents led to unsatisfactory results in advanced clinical trials^{18,19}, due to anemia and methemoglobinemia side effects that were correlated to the *in vivo* oxidative cleavage of the ureas and to the generation of the corresponding aniline-derived metabolites^{20,21}. Nevertheless, sulphonylurea analogues of SLC 0111, where the sulphonyl ureido is considered as a linker, showed recently promising hCA IX and XII inhibitory properties²². In this study, coumarinyl-substituted analogues showed even more promising profile, with nanomolar inhibition of cancer-related hCA IX and XII and low micromolar inhibition of off-targets hCA I and hCA II. Exploiting a strategy commonly used in medicinal chemistry, the use of isosteres of bioactive compounds^{23,24}, we recently developed a method to design new fluoroenesulphonamides as *N*-sulphonylureas isosteres²⁵ and demonstrated their similarities²⁶: these compounds, stable in solution, can be considered as good

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Scheme 1. Hydrofluorination of ynamides.

candidates to mimic unstable *N*-sulphonylureas. Therefore, following our seminal contribution on the use of tertiary benzene sulphonamides as selective cancer-related hCAs inhibitors, we considered that fluoroenesulphonamide group could represent an interesting novel selective chemotype and evaluated the activity of this new series against hCA I, hCA II, hCA IX and hCA XII.

Materials and methods

Chemistry

Two methods were equally used to generate the fluoroenamides from their corresponding ynamides, as shown in [Scheme 1](#).

General procedure A

To a solution of HF (6 ml) maintained at -50°C or -78°C , was added very slowly ynamide derivative (1 mmol). The mixture was magnetically stirred at the same temperature during 5 min. The reaction mixture was then neutralised with water-ice- Na_2CO_3 , extracted with ethyl acetate ($\times 3$). The combined organic phases were dried (MgSO_4) and concentrated *in vacuo*. Products were isolated by column chromatography on silica gel.

General procedure B

To a mixture of hydrofluoric acid and pyridine (4 ml, 70/30 w/w) maintained at the required temperature was added the starting ynamide (1 mmol). The mixture was magnetically stirred at the same temperature during the required time. The reaction mixture was then neutralised with water-ice-sodium carbonate solution, extracted with dichloromethane ($3\times$). The combined organic phases were dried over anhydrous magnesium sulphate, filtered and concentrated *in vacuo*. Products were isolated by column chromatography on silica gel. The NMR spectra of the products and their detailed characterisation can be found in literature^{25,26}.

CA inhibition assay

An SX.18Mv-R Applied Photophysics (Oxford, UK) stopped-flow instrument has been used to assay the catalytic activity of various CA isozymes for CO_2 hydration reaction²⁷. Phenol red (at a concentration of 0.2 mM) was used as indicator, working at the absorbance maximum of 557 nm, with 10 mM HEPES (pH 7.5) as buffer, and 0.1 M Na_2SO_4 (for maintaining constant ionic strength, which is not inhibitory against these CAs), following the CA-catalysed CO_2 hydration reaction for a period of 10 s at 25°C . The CO_2 concentrations ranged from 1.7 to 17 mM for the determination of the kinetic parameters and activation constants. For each inhibitor, at least six traces of the initial 5–10% of the reaction have been used for determining the initial rate. The uncatalysed rates were determined in the same manner and subtracted from the total observed rates. Stock solutions of inhibitors (10 mM) were prepared in distilled deionized water and the solution diluted to 1 nM using the assay buffer. Inhibitor and enzyme solutions were pre-incubated together for 15 min (standard assay at room

temperature) prior to assay, in order to allow for the formation of the enzyme-inhibitor complex. The inhibition constant (K_i), was obtained by considering the classical Michaelis-Menten equation and the Cheng-Prusoff algorithm by using non-linear least squares fitting as reported earlier^{28–30}.

Results and discussion

A series of α -fluoroenesulphonamides and imides were therefore synthesised from the corresponding ynesulphonamides and imides according to our previously reported procedures^{25–26}. α -Fluoroenesulphonamide analogue **1** was therefore tested as a carbonic anhydrase inhibitor and found to be inactive toward hCAI and II, a poor micromolar inhibitor of HCAIX but, most interestingly, a nanomolar inhibitor of HCAXII. This result was especially encouraging as it reinforces our initial hypothesis and revealed a very selective inhibitor profile for the α -fluoroenesulphonamide pharmacophore (to be compared to acetazolamide reference compound AAZ, [Table 1](#): entry 1).

To further explore substituent effect on the inhibitory activity/selectivity of this new class of hCA inhibitors, a brief structure activity relationships study was initiated. Exceptionally, all the tested fluoroenesulphonamides were found to be ineffective as offtarget hCA I and hCA II inhibitors and are selective inhibitor of the tumor associated isoforms IX and XII. Replacement of the phenyl ring on the alkene in **1** by a phenanthrene or a thiophene (compounds **2** and **3**) revealed a strong influence of this substituent on the efficiency and selectivity of the inhibitors. Introduction of a phenanthrene (compound **2**) was indeed detrimental to the activity while the presence of a thiophene dramatically modified the inhibitory profile. 1-Thiophenyl-substituted fluoroenesulphonamide **3** was found to be a micromolar inhibitor for hCA IX and not active for hCA XII and the presence of the heteroaromatic ring, in place of the tolyl group, shifts the inhibitor from a highly selective hCA XII inhibitor to a selective hCAIX inhibitor. These results suggest a non-zinc binding mode of action for these new chemotypes and evidence a variation of binding mode for these inhibitors⁸. To further verify this hypothesis, we next modified the position of the heteroatom in the thiophenyl substituent to impact eventual intra and inter molecular hydrogen bonding, analogously to what has been observed for aromatic ureas in solution³¹. In this case, compound **4** exhibited hCA IX nanomolar inhibition and low micromolar hCA XII inhibition. By increasing the distance between the fluoroenamide ureidoisoster moiety and the hydrophobic phenyl group, while maintaining a linear rigidity thanks to electronic conjugation between π electrons, a dual nanomolar selective inhibitor of hCAIX and hCAXII, compound **5**, could be discovered. Previous report of *in vivo* experiments nicely demonstrated that when silencing hCA IX alone leads to a 40% reduction in xenograft tumor volume, the concomitant inhibition of both transmembrane isoforms IX and XII leads to 85% reduction of tumor growth⁶. As a consequence, compound **5** can be considered as an interesting lead compound for further studies in this direction. To further explore the potential of fluoroenamide as a new chemotype for hCA selective inhibitors quest, fluoroenimides **6** and **7** were synthesised and tested. As for previously tested *N*-sulphonyl analogue **1**, compound **6** was shown to be nanomolar selective inhibitor of hCAXII. This result suggests that the α -fluoroolefin core, the ureido isoster, is the essential pharmacophore for these compounds, thus confirming our initial hypothesis. Again, shifting from phenyl to alkyl chains dramatically modify the selectivity of these compounds with compound **7** being now a hCA IX selective inhibitor at the micromolar level.

Table 1. CA inhibition with acetazolamide (AAZ) as standard and compounds 1–7, against isoforms hCA I, II, IX and XII, by a stopped flow CO₂ hydrase adssay [27].

Compound		K _i * (μM)*				Selectivity ratios			
		hCA I ^a	hCA II ^a	hCA IX ^b	hCA XII ^b	I/IX	I/XII	II/IX	II/XII
	AAZ**	0.25 ^c	0.012 ^c	0.025 ^c	0.006 ^c	10.0	41.6	0.48	2
	1	^e	^e	>50	0.120	NC ^f	>1000	NC ^f	>1000
	2	^e	^e	>50	>50	NC ^f	NC ^f	NC ^f	NC ^f
	3	^e	^e	1.0	>50	>1000	NC ^f	>1000	NC ^f
	4	^e	^e	0.038	4.078	>1000	>1000	>1000	>1000
	5	^e	^e	0.021	0.366	>1000	>1000	>1000	>1000
	6	^e	^e	>10	0.116	NC ^f	>1000	NC ^f	>1000
	7	^e	^e	2.834	>50	>1000	NC ^f	>1000	NC ^f

*Errors in the range of ±5% of the reported data from three different assays.

**Acetazolamide (AAZ) was used as a standard inhibitor for all CAs investigated here.

^aRecombinant isoforms, from Ref. [9a].

^bCatalytic domain.

^cData collected from Ref. [9a].

^dNot determined.

^eNot active, K_i > 100 μM.

^fNot calculated.

On the other hand, considering the substantial interest that bacterial/fungal/protozoan CA inhibition raised ultimately^{32–34}, it would be of great interest to test some of these new CA inhibitors for their interaction with such enzymes belonging to other classes than the α-CAs investigated here.

Conclusions

This study reveals a new, highly selective family of cancer-related transmembrane CA inhibitors. The tested α-fluoroenamides ureidoisosters did not inhibit widespread cytosolic isoforms hCA I and II, and selectively inhibited the transmembrane cancer-related ones, hCA IX and XII. The simple modification of the C-substituent of the α-fluoroenesulphonamide and α-fluoroenimide revealed the possibility to either generate selective hCA IX, selective hCA XII or dual hCA IX and hCA XII isoform confirming the strong potential of these new pharmacophores for further studies.

Disclosure statement

No potential conflict of interest was reported by the authors.

References

- (a) Neri D, Supuran CT. Interfering with pH regulation in tumours as a therapeutic strategy. *Nat Rev Drug Discov* 2011;10:767–77. (b) Supuran CT. Carbonic anhydrase inhibition and the management of hypoxic tumors. *Metabolites* 2017;7:E48. (c) Supuran CT. Carbonic anhydrases and metabolism. *Metabolites* 2018;8:E25.
- (a) McDonald PC, Winum JY, Supuran CT, Dedhar S. Recent developments in targeting carbonic anhydrase IX for cancer therapeutics. *Oncotarget* 2012;3:84–7. (b) Iessi E, Logozzi M, Mizzone D, et al. Rethinking the combination of proton exchanger inhibitors in cancer therapy. *Metabolites* 2018;8:E2.
- Supuran CT, Alterio V, Di Fiore A, et al. Inhibition of carbonic anhydrase IX targets primary tumors, metastases and cancer

- stem cells: three for the price of one. *Med Res Rev* 2018. doi:10.1002/med.21497
4. (a) Spugnini EP, Sonveaux P, Stock C, et al. Proton channels and exchangers in cancer. *Biochim Biophys Acta* 2015;1848:2715–26. (b) Kusuzaki K, Matsubara T, Murata H, et al. Natural extracellular nanovesicles and photodynamic molecules: is there a future for drug delivery? *J Enzyme Inhib Med Chem* 2017;32:908–16. (c) Ward C, Langdon SP, Mullen P, et al. New strategies for targeting the hypoxic tumour microenvironment in breast cancer. *Cancer Treat Rev* 2013;39:171–9.
 5. (a) Parks SK, Chiche J, Pouyssegur J. pH control mechanisms of tumor survival and growth. *J Cell Physiol* 2011;226:299–308. (b) Chiche J, Ilc K, Laferrière J, et al. Hypoxia-inducible carbonic anhydrase IX and XII promote tumor. *Cancer Res* 2009;69:358–68.
 6. (a) Pacchiano F, Aggarwal M, Avvaru BS, et al. Selective hydrophobic pocket binding observed within the carbonic anhydrase II active site accommodate different 4-substituted-ureido-benzenesulfonamides and correlate to inhibitor potency. *Chem Commun* 2010;4:8371–3. (b) Garaj V, Puccetti L, Fasolis G, et al. Carbonic anhydrase inhibitors: synthesis and inhibition of cytosolic/tumor-associated carbonic anhydrase isozymes I, II and IX with sulfonamides incorporating 1,2,4-triazine moieties. *Bioorg Med Chem Lett* 2004;14:5427–33.
 7. (a) Alterio V, Di Fiore A, D'Ambrosio K, et al. X-ray crystallography of CA inhibitors and its importance in drug design. In: Supuran CT, Winum JY. eds. *Drug design of zinc-enzyme inhibitors: functional, structural, and disease applications*. Wiley: Hoboken (NJ); 2009:73–138. (b) Temperini C, Cecchi A, Scozzafava A, Supuran CT. Carbonic anhydrase inhibitors. Sulfonamide diuretics revisited—old leads for new applications? *Org Biomol Chem* 2008;6:2499–2506; (c) Di Fiore A, Maresca A, Alterio V, et al. Carbonic anhydrase inhibitors: X-ray crystallographic studies for the binding of N-substituted benzenesulfonamides to human isoform II. *Chem Commun* 2011;47:11636–11638.
 8. (a) Alterio V, Di Fiore A, D'Ambrosio K, et al. Multiple binding modes of inhibitors to carbonic anhydrases: how to design specific drugs targeting 15 different isoforms? *Chem Rev* 2012;112:4421–68. (b) Supuran CT. Structure and function of carbonic anhydrases. *Biochem J* 2016;473:2023–32. (c) Scozzafava A, Supuran CT, Carta F. Antiobesity carbonic anhydrase inhibitors: a literature and patent review. *Expert Opin Ther Pat* 2013;23:725–35.
 9. (a) Supuran CT. Carbonic anhydrases: novel therapeutic applications for inhibitors and activators. *Nat Rev Drug Discov* 2008;7:168–81. (b) Supuran CT. Structure-based drug discovery of carbonic anhydrase inhibitors. *J Enzyme Inhib Med Chem* 2012;27:759–72. (c) Supuran CT. How many carbonic anhydrase inhibition mechanisms exist? *J Enzyme Inhib Med Chem* 2016;31:345–60. (c) Casey JR, Morgan PE, Vullo D, et al. Carbonic anhydrase inhibitors. Design of selective, membrane-impermeant inhibitors targeting the human tumor-associated isozyme IX. *J Med Chem* 2004;47:2337–47.
 10. (a) Supuran CT. Carbonic anhydrase inhibitors. *Bioorg Med Chem Lett* 2010;20:3467–74. (b) Supuran CT. Advances in structure-based drug discovery of carbonic anhydrase inhibitors. *Expert Opin Drug Discov* 2017;12:61–88.
 11. (a) Maresca A, Temperini C, Vu H, et al. Non-zinc mediated inhibition of carbonic anhydrases: coumarins are a new class of suicide inhibitors. *J Am Chem Soc* 2009;131:3057–62. (b) Maresca A, Temperini C, Pochet L, et al. Deciphering the mechanism of carbonic anhydrase inhibition with coumarins and thiocoumarins. *J Med Chem* 2010;53:335–44. (c) Carta F, Maresca A, Scozzafava A, Supuran CT. Novel coumarins and 2-thioxo-coumarins as inhibitors of the tumor-associated carbonic anhydrases IX and XII. *Bioorg Med Chem* 2012;20:2266–73. (d) Supuran CT. Carbonic anhydrase activators. *Future Med Chem* 2018;10:561–73.
 12. (a) Tars K, Vullo D, Kazaks K, et al. Sulfo-coumarins (1,2-benzoxathiine-2,2-dioxides): a class of potent and isoform-selective inhibitors of tumor-associated carbonic anhydrases. *J Med Chem* 2013;56:293–300. (b) Supuran CT. Carbon- versus sulphur-based zinc binding groups for carbonic anhydrase inhibitors? *J Enzyme Inhib Med Chem* 2018;33:485–95.
 13. Métayer B, Mingot A, Vullo D, et al. New superacid synthesized (fluorinated) tertiary benzenesulfonamides acting as selective hCA IX inhibitors: toward a new mode of carbonic anhydrase inhibition by sulfonamides. *Chem Commun* 2013;49:6015–7.
 14. Métayer B, Mingot A, Vullo D, et al. Superacid synthesized tertiary benzenesulfonamides and benzofused sultams act as selective hCA IX inhibitors: toward understanding a new mode of inhibition by tertiary sulfonamides. *Org Biomol Chem* 2013;11:7540–9.
 15. Lau J, Pan J, Zhang Z, et al. Synthesis and evaluation of 18F-labeled tertiary benzenesulfonamides for imaging carbonic anhydrase IX expression in tumours with positron emission tomography. *Bioorg Med Chem Lett* 2014;24:3064–8.
 16. (a) Lomelino CL, Mahon BP, McKenna R, et al. Kinetic and X-ray crystallographic investigations on carbonic anhydrase isoforms I, II, IX and XII of a thioureido analog of SLC-0111. *Bioorg Med Chem* 2016;24:976–81. (b) Mastrolorenzo A, Rusconi S, Scozzafava A, et al. Inhibitors of HIV-1 protease: current state of the art 10 years after their introduction. From antiretroviral drugs to antifungal, antibacterial and antitumor agents based on aspartic protease inhibitors. *Curr Med Chem* 2007;14:2734–48.
 17. Pacchiano F, Carta F, McDonald PC, et al. Ureido-substituted benzenesulfonamides potentially inhibit carbonic anhydrase IX and show antimetastatic activity in a model of breast cancer metastasis. *J Med Chem* 2011;54:1896–902.
 18. Abdel-Aziz AAM, El-Azab AS, El-Subbagh HI, et al. Design, synthesis, single-crystal and preliminary antitumor activity of novel arenesulfonylimidazolidin-2-ones. *Bioorg Med Chem Lett* 2012;22:2008–14.
 19. Howbert JJ, Grossman CS, Crowell TA, et al. Novel agents effective against solid tumors: the diarylsulfonylureas. Synthesis, activities, and analysis of quantitative structure-activity relationships. *J Med Chem* 1990;33:2393–407.
 20. Guan X, Hoffman BN, McFarland DC, et al. Glutathione and mercapturic acid conjugates of sulofenur and their activity against a human colon cancer cell line. *Drug Metab Dispos* 2002;30:331–5.
 21. (a) Fourouzes B, Takimoto CH, Goetz A, et al. A phase I and pharmacokinetic study of ILX-295501, an oral diarylsulfonylurea, on a weekly for 3 weeks every 4-week schedule in patients with advanced solid malignancies. *Clin Cancer Res* 2003;9:5540–9. (b) Boyd NH, Walker K, Fried J, et al. Addition of carbonic anhydrase 9 inhibitor SLC-0111 to temozolomide treatment delays glioblastoma growth *in vivo*. *JCI Insight* 2017;2:92928.
 22. Bozdag M, Ferraroni M, Carta F, et al. Structural insights on carbonic anhydrase inhibitory action, isoform selectivity, and

- potency of sulfonamides and coumarins incorporating aryl-sulfonylureido groups. *J Med Chem* 2014;57:9152–67.
23. Lima JM, Barreiro EJ. Bioisosterism: a useful strategy for molecular modification and drug design. *Curr Med Chem* 2005;12:23–49.
 24. Angeli A, Tanini D, Peat TS, et al. Discovery of new selenour-*eido* analogues of 4-(4-fluorophenylureido)benzenesulfonamide as carbonic anhydrase inhibitors. *ACS Med Chem Lett* 2017;8:963–8.
 25. Compain G, Jouvin K, Martin-Mingot A, et al. Stereoselective hydrofluorination of ynamides: a straightforward synthesis of novel α -fluoroenamides. *Chem Commun* 2012;48:5196–8.
 26. Métayer B, Compain G, Jouvin K, et al. Chemo- and stereoselective synthesis of fluorinated enamides from ynamides in HF/pyridine: second-generation approach to potent ureas bioisosteres. *J Org Chem* 2015;80:3397–410.
 27. Khalifah RG. The carbon dioxide hydration activity of carbonic anhydrase. I. Stop-flow kinetic studies on the native human isoenzymes B and C. *J Biol Chem* 1971;246:2561–73.
 28. (a) Menchise V, De Simone G, Alterio V, et al. Carbonic anhydrase inhibitors: stacking with Phe131 determines active site binding region of inhibitors as exemplified by the X-ray crystal structure of a membrane-impermeant antitumor sulfonamide complexed with isozyme II. *J Med Chem* 2005;48:5721–7. (b) Supuran CT, Mincione F, Scozzafava A, et al. Carbonic anhydrase inhibitors—part 52. Metal complexes of heterocyclic sulfonamides: a new class of strong topical intraocular pressure-lowering agents in rabbits. *Eur J Med Chem* 1998;33:247–54. (c) Garaj V, Puccetti L, Fasolis G, et al. Carbonic anhydrase inhibitors: novel sulfonamides incorporating 1,3,5-triazine moieties as inhibitors of the cytosolic and tumour-associated carbonic anhydrase isozymes I, II and IX. *Bioorg Med Chem Lett* 2005;15:3102–8. (d) Şentürk M, Gülçin İ, Beydemir Ş, et al. *In vitro* inhibition of human carbonic anhydrase I and II isozymes with natural phenolic compounds. *Chem Biol Drug Des* 2011;77:494–9. (e) Fabrizi F, Mincione F, Somma T, et al. A new approach to antiglaucoma drugs: carbonic anhydrase inhibitors with or without NO donating moieties. Mechanism of action and preliminary pharmacology. *J Enzyme Inhib Med Chem* 2012;27:138–47. (f) Dogne JM, Hanson J, Supuran C, Pratico D. Coxibs and cardiovascular side-effects: from light to shadow. *Curr Pharm Des* 2006;12:971–5.
 29. (a) Krall N, Pretto F, Decurtins W, et al. A small-molecule drug conjugate for the treatment of carbonic anhydrase IX expressing tumors. *Angew Chem Int Ed Engl* 2014;53:4231–5. (b) Rehman SU, Chohan ZH, Gulnaz F, Supuran CT. *In-vitro* antibacterial, antifungal and cytotoxic activities of some coumarins and their metal complexes. *J Enzyme Inhib Med Chem* 2005;20:333–40. (c) Clare BW, Supuran CT. Carbonic anhydrase activators. 3: Structure-activity correlations for a series of isozyme II activators. *J Pharm Sci* 1994;83:768–73. (d) Dubois L, Peeters S, Lieuwes NG, et al. Specific inhibition of carbonic anhydrase IX activity enhances the *in vivo* therapeutic effect of tumor irradiation. *Radiother Oncol* 2011;99:424–31. (e) Chohan ZH, Munawar A, Supuran CT. Transition metal ion complexes of Schiff-bases. Synthesis, characterization and antibacterial properties. *Met Based Drugs* 2001;8:137–43. (f) Zimmerman SA, Ferry JG, Supuran CT. Inhibition of the archaeal β -class (Cab) and γ -class (Cam) carbonic anhydrases. *Curr Top Med Chem* 2007;7:901–8. (g) De Simone G, Supuran CT. (In)organic anions as carbonic anhydrase inhibitors. *J Inorg Biochem* 2012;111:117–29.
 30. (a) Supuran CT, Nicolae A, Popescu A. Carbonic anhydrase inhibitors. Part 35. Synthesis of Schiff bases derived from sulfanilamide and aromatic aldehydes: the first inhibitors with equally high affinity towards cytosolic and membrane-bound isozymes. *Eur J Med Chem* 1996;31:431–8. (b) Pacchiano F, Aggarwal M, Avvaru BS, et al. Selective hydrophobic pocket binding observed within the carbonic anhydrase II active site accommodate different 4-substituted-ureido-benzenesulfonamides and correlate to inhibitor potency. *Chem Commun*. 2010;46:8371–3. (c) Ozensoy Guler O, Capasso C, Supuran CT. A magnificent enzyme superfamily: carbonic anhydrases, their purification and characterization. *J Enzyme Inhib Med Chem* 2016;31:689–94. (d) De Simone G, Langella E, Esposito D, et al. Insights into the binding mode of sulphamates and sulphamides to hCA II: crystallographic studies and binding free energy calculations. *J Enzyme Inhib Med Chem* 2017;32:1002–11. (e) Di Fiore A, De Simone G, Alterio V, et al. The anticonvulsant sulfamide JNJ-26990990 and its S, S-dioxide analog strongly inhibit carbonic anhydrases: solution and X-ray crystallographic studies. *Org Biomol Chem* 2016;14:4853–8.
 31. Giannicchi I, Jouvelet B, Isare B, et al. Orthohalogen substituents dramatically enhance hydrogen bonding of aromatic ureas in solution. *Chem Commun* 2014;50:611.
 32. (a) Capasso C, Supuran CT. An overview of the alpha-, beta- and gamma-carbonic anhydrases from bacteria: can bacterial carbonic anhydrases shed new light on evolution of bacteria? *J Enzyme Inhib Med Chem* 2015;30:325–32. (b) Supuran CT, Scozzafava A, Mastrolorenzo A. Bacterial proteases: current therapeutic use and future prospects for the development of new antibiotics. *Exp Opin Ther Pat* 2001;11:221–59. (c) Del Prete S, De Luca V, Vullo D, et al. A new procedure for the cloning, expression and purification of the β -carbonic anhydrase from the pathogenic yeast *Malassezia globosa*, an anti-dandruff drug target. *J Enzyme Inhib Med Chem* 2016;31:1156–61.
 33. (a) Supuran CT, Capasso C. New light on bacterial carbonic anhydrases phylogeny based on the analysis of signal peptide sequences. *J Enzyme Inhib Med Chem* 2016;31:1254–60. (b) Supuran CT, Capasso C. Carbonic anhydrase from *Porphyromonas gingivalis* as a drug target. *Pathogens* 2017;6:E30. (c) Del Prete S, De Luca V, De Simone G, et al. Cloning, expression and purification of the complete domain of the η -carbonic anhydrase from *Plasmodium falciparum*. *J Enzyme Inhib Med Chem* 2016;31:54–9.
 34. (a) Maresca A, Carta F, Vullo D, Supuran CT. Dithiocarbamates strongly inhibit the β -class carbonic anhydrases from *Mycobacterium tuberculosis*. *J Enzyme Inhib Med Chem* 2013;28:407–11. (b) Chohan ZH, Arif M, Shafiq Z, et al. *In vitro* antibacterial, antifungal & cytotoxic activity of some isonicotinoylhydrazide Schiff's bases and their cobalt (II), copper (II), nickel (II) and zinc (II) complexes. *J Enzyme Inhib Med Chem* 2006;21:95–103. (c) De Menezes Dda R, Calvet CM, Rodrigues GC, et al. Hydroxamic acid derivatives: a promising scaffold for rational compound optimization in Chagas disease. *J Enzyme Inhib Med Chem* 2016;31:964–73.