REVIEW ARTICLE



Hypertension and related diseases in the era of COVID-19: a report from the Japanese Society of Hypertension Task Force on COVID-19

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Abstract

Coronavirus disease-2019 (COVID-19), caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has affected more than seven million people worldwide, contributing to 0.4 million deaths as of June 2020. The fact that the virus uses angiotensin-converting enzyme (ACE)-2 as the cell entry receptor and that hypertension as well as cardiovascular disorders frequently coexist with COVID-19 have generated considerable discussion on the management of patients with hypertension. In addition, the COVID-19 pandemic necessitates the development of and adaptation to a "New Normal" lifestyle, which will have a profound impact not only on communicable diseases but also on noncommunicable diseases, including hypertension. Summarizing what is known and what requires further investigation in this field may help to address the challenges we face. In the present review, we critically evaluate the existing evidence for the epidemiological association between COVID-19 and hypertension. We also summarize the current knowledge regarding the pathophysiology of SARS-CoV-2 infection with an emphasis on ACE2, the cardiovascular system, and the kidney. Finally, we review evidence on the use of antihypertensive medication, namely, ACE inhibitors and angiotensin receptor blockers, in patients with COVID-19.

Keywords severe acute respiratory syndrome coronavirus $2 \cdot$ hypertension \cdot cardiovascular disease \cdot angiotensin converting enzyme 2

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Coronavirus disease-2019 (COVID-19) and hypertension: premonition of convergence of communicable diseases and noncommunicable diseases

In recent years, it has been well recognized that noncommunicable diseases (NCDs), including hypertension, are the main health issue, since ~70% of the causes of death in the world (57 million deaths/year) are attributed to NCDs. However, the sudden emergence of COVID-19, a communicable disease (CD), has changed our concept toward health and diseases. Indeed, we are harshly reminded that CDs can seriously threaten all aspects of life, including health, economics, education, and human relations. Furthermore, we must change our concept of a normal life to a "New Normal" one [1]. It is predicted that the pandemic of COVID-19 will create another pandemic of NCDs [2], and this prediction is plausible considering the similar situations manifested in the evacuation performed in the Great East Japan Earthquake on March 11, 2011. Even two years after the disaster, the blood pressure (BP) of the evacuees remained significantly elevated by an average of ~4~5 mmHg [3]. In this context, it is deeply concerning that a new disease entity is emerging, that is, the convergence of CDs and NCDs without borders. Thus far, we have been attempting to understand NCDs as lifestyle-related diseases, focusing upon each person's specific lifestyle in personalized medicine. However, from now on, we should also dissect out the pathophysiology of convergence of CDs and NCDs from each person's lifestyle in society where he/she is living, and should take care of life-environment as life-environment-related diseases. We also should pay more attention to virus-infected spaces of life as pressing matters, as well as global warming, which has been gradually recognized as an urgent issue.

To cope with this new disease entity, it is necessary to know the FACTS with regard to COVID-19 and NCDs. This is especially true for hypertension because severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), a novel virus that causes COVID-19, utilizes angiotensin-converting enzyme (ACE)-2, one of the components of the renin-angiotensin system (RAS), for its entry into the body and because hypertensionassociated diseases have been revealed to be factors contributing to the severity of COVID-19. We should also realize what is known and not yet known, and what requires further clarification. Here, we review recent literature regarding hypertension and COVID-19 available as of June 7th, 2020.

Epidemiological findings on the association between hypertension and COVID-19

Prevalence of hypertension among COVID-19 patients

Although emerging reports have demonstrated a high prevalence of hypertension among patients with COVID-19 [4–6], evidence is still insufficient (Table 1). For example, 58 of 191 Chinese patients with COVID-19 (30.4%; median age, 56.0 years) [4] and 509 of 1043 Italian patients with available data (48.8%, 95% confidence intervals, 46-52%; mean age among all 1591 study patients, 63 years) [5] had hypertension, and investigators in these studies [4, 5] reported a high prevalence of hypertension among patients with COVID-19. However, a Chinese nationwide survey showed that 44.6% of the population aged 55-64 years had hypertension [7], and 45.2% of the Italian population aged 60-69 years had hypertension according to an Italian inclusive general practitioner database [8]. Furthermore, in a US case series including 5700 sequentially hospitalized COVID-19 patients (median age, 63 years), 3026 (56.6%) had hypertension [6], with the prevalence of hypertension among COVID-19 patients also being lower than the US general population (estimated to be 63-77% of the population aged 55–64 years [9]). Furthermore, it should be noted that demographic data among patients, including comorbidities, were collected by different methods (by electronic medical records [4, 6] or by phone [5]). Most importantly, none of the currently available epidemiological studies regarding COVID-19 and hypertension (Table 1) clearly state the diagnostic methodology used or the criteria for hypertension, e.g., BP values based on office or out-of-office measurement, as well as self-reported or testimony of a family in serious situations. These variations would make the comparison of reported incidence with that in the general population inaccurate. Taking the high proportion of individuals with hypertension who are not aware of their condition [10] into account, it would be a reasonable assumption that the prevalence of hypertension in these reports [4-6]may be underestimated. Nevertheless, no supporting report was found that shows a higher rate of hypertension among patients with COVID-19, and there have been no reliable reports demonstrating an increased risk of SARS-CoV-2 infection in the presence of hypertension.

Impact of hypertension on the severity and mortality of COVID-19

Comparisons of COVID-19 patients with mild and severe clinical symptoms can be used to evaluate whether

Table 1 Summar	y of the epidemiol	logical studies conce	erning the association between h	ypertension and COVID-	19		
					Hypertension as a r	isk factor	
First author, country [Reference]	Design	Definition of hypertension	Number and population	Outcome	Univariate analysis	Multivariable-adjusted analysis	Note
Zhou, China [4]	Retrospective cohort study	N.A.	191 adult inpatients (>18 years), median age 56.0 years	In-hospital death	Yes (48% vs. 23%; <i>p</i> = 0.0008)	No (not selected)	137 patients were discharged and 54 died in hospital. Hypertension was the most common comorbidity ($n = 58$, 30%). Older age, high Sequential Organ Failure Assessment score, and high D-dimer level were selected as independent explanatory factors for inhospital death among 171 patients with complete data.
Grasselli, Italy [5]	Retrospective case series	И.А.	1591 patients admitted to ICU, median age 63 years	COVID-19	N.A.	. A . M	Hypertension was the most common comorbidity ($n = 509$, 49% of the 1,043 individuals with comorbidity data). Patients with hypertension were older than those without hypertension (median age, 66 vs. 62 years, $p <$ 0.001). ICU mortality was 26%. The mortality of older patients (264 years) was higher than that of younger patients (x65 years) (36% vs. 15%, $p < 0.01$).
Richardson, US [6]	Retrospective case series	ICD-10 coding No diagnostic criteria shown	5700 inpatients, median age 63 years	COVID-19	N.A.	N.A.	Hypertension was the most common comorbidity ($n = 3026$, 56.6%). 553 patients (21%) died in hospital; 134 were 18–65 years and 419 were >65 years.
Shi, China [11]	Retrospective cohort study	N.A.	487 inpatients, mean age 46 years	Severity on admission	Yes (53.1% vs. 16.7%; p<0.001)	Yes (OR, 2.71; CI,1.32–5.59; <i>p</i> = 0.007)	
Li, China [12]	Ambispective cohort study	ICD-10 coding No diagnostic criteria shown	548 inpatients, median age 60 years	 Severity on admission In-hospital death 	1) Yes (38.7% vs. 22.2%; <i>p</i> < 0.001) 2) N.A.	1) Yes (OR, 2.01; CI, 1.27–3.17) 2) No(not selected)	As independent predictors for mortality, male, 265 years, high white blood cell count, high lactate dehydrogenase value, cardiac injury, hyperglycemia, and high dose corticosteroid were selected.
Simonnet, France [13]	Retrospective cohort study	И.А.	124 patients admitted to ICU, median age 60 years	Need for IMV	Yes (OR, 2.81; CI, 1.25-6.30; $p = 0.012$)	No (OR, 2.29; CI, 0.89–5.84; <i>p</i> = 0.08)	Definition of obesity and severe obesity: BMI > 30 and 35 kg/m ² , respectively. The prevalence of obesity and severe obesity was 47.6% ($n = 59$) and 28.2% ($n = 35$), respectively. As BMI increased, the proportion of the patients who received IMV increased (chi-square test for trend, $p < 0.01$). On multivariable logistic regression analysis, severe obesity was an independent risk factor (OR, 7.36; CI, 1.63-33.14; $p = 0.021$).

					Hypertension as a r	isk factor	
First author, country [Reference]	Design	Definition of hypertension	Number and population	Outcome	Univariate analysis	Multivariable-adjusted analysis	Note
Cai, China [14]	Retrospective cohort study	N.A.	383 inpatients, median age: nonsevere patients ($n = 292$, 76.2%) 44.5 years, severe patients ($n = 91, 23.8\%$) 61 years	Progression to severe COVID-19	Yes (23.08% vs. 12.67%; <i>p</i> = 0.02)	Done, but not shown	Definition of obesity: BMI > 28 kg/m ² . The prevalence of obesity was 10.7% ($n = 41$). Compared with the patients with BMI 18.5–23.9 kg/m ² , obese patients showed 3.40-fold odds (CI, 1.40–2.86; $p = 0.007$) of outcome by multiple logistic regression analysis after adjusting for several factors including hypertension.
Wu, China [15]	Retrospective cohort study	N.A.	201 inpatients with COVID-19 pneumonia, median age 51 years	1) Development of ARDS 2) Death among patients with ARDS ($n = 84$)	1) Yes (27.4% vs. 13.7%; p = 0.02) 2) Yes(36.4% vs. 17.5%; $p = 0.05)$	No (analysis unclear)	Hypertension raised the risk of ARDS development (HR, 1.82; CI, 1.13–2.95; $p = 0.01$), but not the risk of progression from ARDS to death (HR, 1.70; CI, 0.92–3.14; $p = 0.09$). However, these were based on bivariate Cox regression analysis, and the methodology, in particular whether other factors were adjusted, was unclear.
Zheng, China [19]	Retrospective cohort study	Consensus statement from the IDF	66 patients with metabolic associated fatty liver disease, mean age 47 years	Progression to Severe COVID-19	ЧЧ	Done, but not shown	Definition of obesity: BMI > 25 kg/m ² . The prevalence of obesity was 68% (n = 45). The hypertension prevalence between obese and non-obese patients did not differ significantly (35.6% vs. 14.3%, p = 0.089). In the logistic regression model, obesity was shown to be a risk factor after adjusting for age, sex, smoking, diabetes, hypertension, and dyslipidemia (OR, 6.32; CI, 1.16–34.54; $p = 0.033$).
<i>BMI</i> body mass International Cla	index, CDC Center assification of Diseas	s for Disease Control a es-version 10, <i>IDF</i> Inte	and Prevention, CI 95% confi ernational Diabetes Federation	idence interval, COVID-I) 1, IMV invasive mechanica	9 coronavirus dise I ventilation, ARD	ase 2019, <i>HR</i> hazard ra S acute respiratory distr	atio, <i>ICU</i> intensive care unit, <i>ICD-10</i> ess syndrome, <i>NA</i> . not available, <i>OR</i>

Table 1 (continued)

odds ratio

hypertension is a risk factor for aggravation of the disease. According to a retrospective study consisting of 487 COVID-19 patients in Zhejiang Province of China, the prevalence of hypertension was higher in the 49 severe cases than in the 438 mild cases (53.1% vs. 16.7%, p <0.0001) [11]. Further multivariable-adjusted analysis revealed that male sex, age ≥ 50 years old, and hypertension were independent factors for COVID-19 severity on admission (odds ratio, 2.71; 95% confidence intervals 1.32-5.59) [11]. In another study involving 548 inpatients in Wuhan, China, where the first COVID-19 outbreak occurred in December 2019 [12], the prevalence of hypertension was significantly higher in patients with severe COVID-19 than in nonsevere cases (38.7% vs. 22.2%, p <0.001). In a logistic model with adjustment for age, high lactate dehydrogenase (LDH), and D-dimer, hypertension was independently associated with the severity of COVID-19 on admission (odds ratio, 2.01; 95% confidence intervals, 1.27–3.17) [12].

However, it should be noted that hypertension is commonly accompanied by many comorbidities that are major determinant factors for the severity of COVID-19. The study in Wuhan [12] also reported that male sex, age ≥65 years old, high white blood cell count, LDH, cardiac injury, hyperglycemia, and high dose corticosteroid were independent predictive factors for death in a multivariable-adjusted Cox proportional hazard model. Nonetheless, hypertension was not included as a candidate explanatory factor in the multivariate analysis. A French single-center study [13] reported that hypertension was not significantly associated with progression of COVID-19, as defined as the requirement of invasive mechanical ventilation during hospitalization (OR, 2.29; 95% CI, 0.89–5.84; p = 0.08), even though the association was significant in a univariate model. In these and other studies, hypertension was not selected as an independent factor for COVID-19 severity based on multivariable-adjusted analysis, despite being identified as a risk factor by univariate [4, 12–14] or bivariate [15] survival analysis. Although some studies report that hypertension can be an independent risk factor for severe COVID-19 [11, 12], it would be plausible to interpret that the high prevalence of hypertension among patients with severe and fatal COVID-19 may be attributed to the vulnerability of older individuals to SARS-CoV-2 infection. At present, there is no clear epidemiological evidence supporting that hypertension itself is an independent risk factor for developing severe disease in patients with COVID-19. We, therefore, agree with the conclusion of the Centers for Disease Control and Prevention (CDC), which does not include hypertension in the list of risk factors for COVID-19 severity [16].

Key comorbidities of hypertension in relation to COVID-19

As mentioned above, age is the essential risk factor for increased severity and mortality in COVID-19. In Japan, the mortality rates among COVID-19 patients aged 70–79 and \geq 80 years were 6.8% and 14.8%, respectively, whereas COVID-19 mortality in all age groups was 2.6% as of 7 May 2020 [17]. In addition to age, available reports have suggested a variety of underlying medical conditions to be associated with COVID-19 disease severity and mortality [4–6, 11, 12, 15, 16]. Furthermore, the majority of these risk factors are observed in patients with hypertension [9, 18]. In clinical practice, one may not always have enough time to assess comorbidities in acute-phase patients with COVID-19. In such situations, hypertension diagnosed by various methods might be regarded as a marker of the severity of the disease.

Obesity is another crucial issue that needs to be discussed since it is often accompanied by hypertension and is recognized as a novel risk factor for COVID-19 [13]. It may also be a determinant of disease severity, independent of age and hypertension [14, 19]. Obesity might worsen the clinical course of COVID-19 by decreasing expiratory reserve volume, impeding diaphragm excursion, and restricting ventilation [20]. Furthermore, obesity, especially abdominal obesity, increases inflammatory cytokines and oxidative stress and causes hypertension, diabetes, and dyslipidemia [21]. These comorbidities accelerate atherosclerosis, which leads to cardiovascular complications and may increase the severity and mortality of COVID-19. Recently, patients with both obesity and hypertension have been increasing in Japan [22], and these patients are at particularly high risk for severe disease in COVID-19 and thus require careful observation and intensive treatment. Similar to hypertension, the definition of obesity is not consistent among studies/countries, which needs to be considered.

COVID-19 and ACE2

ACE2 as the entry receptor for SARS-CoV-2

The first step of SARS-CoV-2 infection in humans is contact of the virus with cell-surface ACE2 (Fig. 1). ACE2 interacts with external SARS-CoV-2 by binding to the receptor-binding domain (RBD) of the viral spike protein [23]. This process is followed by proteolytic cleavage of the spike protein, which allows fusion to cells, and transmembrane protease serine 2 (TMPRSS2) has been identified as a protease responsible for the reaction (Fig. 1) [24]. Although SARS-CoV, which was responsible for SARS outbreaks in



Fig. 1 Possible mechanism of SARS-CoV-2-induced vascular complications. ACE2 angiotensin-converting enzyme 2, ADAM17 a disintegrin and metalloprotease 17, AII angiotensin II, A1-7 angiotensin 1-7, ARDS acute respiratory distress syndrome, EC endothelial cell,

EpC epithelial cell, IFN interferon, SARS-CoV-2 severe acute respiratory syndrome coronavirus 2, TLRs Toll-like receptors, TMPRSS2 transmembrane serine protease 2, RAS renin-angiotensin system

2002–2004, uses the same ACE2 and TMPRSS2 for entry, our understanding of the differences between SARS-CoV and SARS-CoV-2 with regard to their cell entry mechanism has been rapidly developed by recent studies, including structural analysis of these viruses [25–29]. Interestingly, ACE2 binds with higher affinity to the RBD of SARS-CoV-2 than that of SARS-CoV. Paradoxically, however, the affinity of ACE2 for the entire SARS-CoV-2 spike protein is comparable to that of SARS-CoV, suggesting that the SARS-CoV-2 RBD is less exposed than is the SARS-CoV RBD [26].

Organs responsible for SARS-CoV-2 entry

In the human lung, type II alveolar epithelial cells coexpress ACE2 and TMPRSS2 and are considered to be primarily responsible for virus entry in both SARS and COVID-19 [30, 31]. It was recently reported that ACE2 and TMPRSS2 are highly coexpressed in nasal and corneal epithelial cells [31]. Enterocytes also coexpress ACE2 and TMPRSS2 [31], and SARS-CoV-2 as well as SARS-CoV rapidly infect human small intestinal organoids [31]. These findings suggest alternative pathways of viral entry via the upper airways, eyes and intestinal organs. A recent report by Bunyavanich et al. found that ACE2 mRNA expression in the nasal epithelium increased with age when data were

stratified into groups of younger children (<10 years), older children (10–17 years), young adults (18–24 years), and adults (\geq 25 years), which might help to explain the relatively low prevalence of COVID-19 in children [32].

The role of ACE2 in COVID-19

The argument about the role of ACE2 in the pathogenesis of COVID-19 has been complicated by the multifunctionality of ACE2 in addition to serving as the entry receptor for SARS-CoV-2. ACE2 was identified in 2000, and accumulating evidence has established diverse roles of ACE2, including as a negative regulator of the RAS, interacting protein with apelin peptides, and as a chaperone protein for the amino acid transporter B0AT1 (SLC6A19) [33]. Similar to SARS-CoV [34], infection with SARS-CoV-2 may downregulate cell-surface ACE2, leading to the reduced activity of ACE2 in infected organs. Moreover, the binding of ACE2 to SARS-CoV, and probably to SARS-CoV-2, increases the activity of disintegrin and metalloproteinase domain-containing protein 17 (ADAM17), also named tumor necrosis factor- α convertase (TACE) [35]. ADAM17 induces ectodomain shedding of ACE2 and produces circulating soluble ACE2, which can be detected in blood tests (Fig. 1) [36]. This mechanism could also reduce the cellsurface ACE2 [35]. The decrease in cell-surface ACE2 by

these pathways appears to reduce the chance of further invasion by the virus. However, it might also attenuate the inhibition of RAS by ACE2 in infected organs. It was reported that depletion of ACE2 worsened acute lung inflammation induced by acid aspiration, sepsis, or endotoxin in mice [37]. In that study, severe lung inflammation induced by acid aspiration was alleviated in mice genetically lacking ACE or angiotensin II type 1a receptor, and also by losartan, an angiotensin II type 1 receptor antagonist, suggesting that local activation of RAS contributes to lung inflammation in pneumonia [37]. Moreover, a research group reported that the SARS spike protein binds to pulmonary ACE2, exacerbating acid-induced pneumonia accompanied by an increase in angiotensin II concentration, and that the pathologic alterations were rescued by losartan in wild-type mice [34]. Given the evidence that angiotensin II is a key mediator of tissue inflammation [38, 39], these findings suggest that downregulation of ACE2 in response to the binding of SARS-CoV, and probably SARS-CoV2, may serve as a mechanism to counteract viral infection at the expense of an increase in angiotensin II.

Interestingly, Ziegler et al. recently reported that ACE2 is upregulated by stimulation by interferon, an antiviral cytokine, in SARS-CoV-2 target cells such as in the lung, nose, and small intestine [40]. Given the abovementioned roles of ACE2 in the respiratory system, upregulation of ACE2 appears to be involved in the series of tissueprotective responses induced by interferon. However, SARS-CoV-2 may exploit this innate immune mechanism to enhance infection [40], and the process would eventually lead to a decrease in cell-surface ACE2. Although this proposed mechanism requires further validation, including in vivo experiments, these data indicate that the pathophysiology of COVID-19 depends on the critical balance between the SARS-CoV-2 viral load and host defense mechanism.

The potential influence of RAS inhibitors on COVID-19

In the early phase of the COVID-19 pandemic, it was proposed that antihypertensive treatment using ACE inhibitors or angiotensin receptor blockers (ARBs) may contribute to adverse outcomes in patients with hypertension and COVID-19 [41]. This hypothesis was primarily based on previous experimental studies showing that RAS inhibitors could alter tissue activity or expression of ACE2. While these studies were mainly conducted using heart [42–51], arterial [52–55], and kidney [43, 45, 46, 56–59] tissues, some papers showed that changes in ACE2 expression also occurred in the lung [60–62] (Table 2). There are also several reports indicating that mineralocorticoid receptor antagonists (MRAs)

increase the expression or activity of ACE2 in the heart and kidney [48, 49, 63-65] (Table 2). Most of these studies have suggested that these antihypertensive drugs might increase tissue expression and/or activity of ACE2, though some have reported no alternation or a decrease of ACE2 in response to the drug [48-50, 59] (Table 2). Nevertheless, increased membrane expression of ACE2 induced by these drugs can theoretically increase the chance of virus entry into organs, but it is also conceivable that RAS inhibition contributes to organ protection against respiratory infection, as mentioned above [34, 37]. In addition, cellular entry of SARS-CoV-2 depends not only on ACE2 but also on proteolytic cleavage primarily by TMPRSS2. Therefore, while the increase in ACE2 by these drugs can be harmful at least in terms of viral invasion in organs that coexpress ACE2 and TMPRSS2, it remains unknown how increased tissue levels of ACE2 would alter the severity of COVID-19 in organs that lack proteases mediating fusion of the virus. Finally, there is little evidence that the therapeutic dose of RAS inhibitors influences tissue expression or activity of ACE2 in humans. Further investigation is required to clarify whether the influence of RAS inhibitors on COVID-19 is beneficial, neutral, or harmful.

Circulating ACE2 as a biomarker of SARS-CoV-2 infection?

Given the essential role of ACE2 in SARS-CoV-2 infection, it has been postulated that the circulating plasma concentration of ACE2 can serve as a biomarker to predict susceptibility to COVID-19 or disease severity. Circulating ACE2 levels are theoretically modulated by the activity of ADAM17, which cleaves cellular ACE2 in the cardiovascular system, as well as ACE2 abundance in each organ. Given the previous in vitro finding that ACE2 binding to SARS-CoV increases the truncated form of ACE2 by activating ADAM17 [35], it is theoretically conceivable that SARS-CoV and SARS-CoV-2 infection can alter circulating ACE2 levels. Nevertheless, it remains unknown whether circulating ACE2 levels have any relationship with ACE2 abundance in the respiratory system or intestinal tissues. Moreover, circulating ACE2 levels are increased in patients with cardiovascular diseases (CVDs), including heart failure [66, 67] and arterial fibrillation [68], chronic kidney disease (CKD) [69], atherosclerosis [70], and stroke [71]. In addition, circulating ACE2 levels are reported to be higher in male than in female patients with heart failure [72]. These data indicate that circulating levels of ACE2 can largely be affected by cardiovascular comorbidities or other characteristics. Further studies are needed to clarify whether circulating ACE2 is indeed associated with susceptibility to or disease severity of COVID-19.

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Los(Lis)Drinking water10 mg/kg/d12 dmRen2 tg ratsLosDrinking water $25 mg/kg/d$ $3m$ Male new ZeallaLosOral gavage10 or $30 mg/kg/d$ $-$ SD rats with acuLosOral gavage10 or $30 mg/kg/d$ $-$ SD rats with acuLosJohnSubcutaneous $2.5 mg/kg/d$ $4 w$ Male new ZeallaLosOral gavage10 or $30 mg/kg/d$ $4 w$ Male stroke-prorOhnSubcutaneous $0.5 mg/kg/d$ $4 w$ Male stroke-prorOhnSubcutaneous $0.5 mg/kg/d$ $4 w$ Male stroke-prorOhnDrinking water $10 mg/kg/d$ $4 w$ Male stroke-prorOhnDrinking water $10 mg/kg/d$ $21 d$ Lewis rats withTelOralGastric gavage $10 mg/kg/d$ $21 d$ Lewis rats withTelDrinking water $2 mg/kg/d$ $10 w$ Male DS ratsEpr(spi)Intraperitoneal $5 0 mg/kg/d$ $10 w$ Male DS ratsEpr(spi)Intraperitoneal $5 mg/kg/d$ $12 d$ Lewis rats withIfDrinking water $10 mg/kg/d$ $2 w$ Lewis ratsLis/(Los)Drinking water $10 mg/kg/d$ $2 w$ Lewis ratsLis/(Los)Drinkin	os/(Lis)	Drinking water	10 mg/kg/d	2 w	Lewis rats	Kidney	I	Ι	←	56
LosDrinking water $25 mg/kg/d$ $3m$ Male new ZealaLosIntravenous $25 mg/kg/d$ -SD rats with acuLosOral gavage10 or $30 mg/kg/d$ -SD rats exposedLosOral gavage10 or $30 mg/kg/d$ -SD rats exposedLos / OlmSubcutaneousLos: $10 mg/kg/d$ 28 dLewis rats withOlmSubcutaneous $0.5 mg/kg/d$ 4 wMale stroke-prorOlmDrinking water $0.5 mg/kg/d$ 4 wMale stroke-prorTelOral $10 ng/kg/d$ 2 uFemale Stroke-prorTelOral $10 ng/kg/d$ 2 wMale hRN/hANTelOral $10 ng/kg/d$ 2 wMale bR/mANTelOral $10 ng/kg/d$ 2 wMale bR rats withTelOral $10 ng/kg/d$ $14 d$ SHRTelOral $10 ng/kg/d$ $14 o$ $28 d$ TelDrinking water $10 ng/kg/d$ $14 d$ SHRCan/(epl)Gastric gavage $10 ng/kg/d$ $14 d$ $28 m$ TelDrinking water $10 ng/kg/d$ $14 d$ $28 m$ $28 m$ Can/(epl)Gastric gavage $10 ng/kg/d$ $14 d$ $28 m$ $28 m$ Castric gavage $10 ng/kg/d$ $12 d$ Lewis rats withTelDrinking water $10 ng/kg/d$ $2 w$ Lewis ratsCastric gavage $10 ng/kg/d$ $2 w$ Lewis ratsLis/(Los)Drinking water $10 ng/kg/d$ $2 w$ Lewis rats </td <td>os/(Lis)</td> <td>Drinking water</td> <td>10 mg/kg/d</td> <td>12 d</td> <td>mRen2 tg rats</td> <td>Heart/Kidney</td> <td>←</td> <td>Ι</td> <td>←</td> <td>43</td>	os/(Lis)	Drinking water	10 mg/kg/d	12 d	mRen2 tg rats	Heart/Kidney	←	Ι	←	43
Los Intravenous 2.5 mg/kg/h - SD rats with acu Los Oral gavage 10 or 30 mg/kg/d - SD rats exposed Los Oral gavage 10 or 30 mg/kg/d - SD rats exposed Los Olm Subcutaneous Los: 10 mg/kg/d 28 d Lewis rats with Olm Subcutaneous 0.5 mg/kg/d 4 w Male stroke-pror Olm Drinking water 10 mg/kg/d 4 w Male stroke-pror Olm Drinking water 10 mg/kg/d 4 w Male stroke-pror Tel Oral Inors/kg/d 4 w Male stroke-pror Tel Oral Inorg/kg/d 4 w Male stroke-pror Tel Oral Inorg/kg/d 2 w Female C57BLk Tel Oral Drinking water 10 mg/kg/d 14 or 28 d With Strats with Tel Oral Drinking water 10 mg/kg/d 17 d C57BL/6 mice ACE inhibitors Intraperitoneal 50 mg/kg/d 12 d Lewis rats wit	SO	Drinking water	25 mg/kg/d	$3\mathrm{m}$	Male new Zealand white rabbits	Aorta	Ι	←	Ι	52
LosOral gavage10 or $30 \mathrm{mg/kg/d}$ -SD rats exposedLos / OlmSubcutaneousLos: $10 \mathrm{mg/kg/d}$ 28 dLewis rats withOlmSubcutaneous $0.5 \mathrm{mg/kg/d}$ 4 wMale stroke-prorOlmDrinking water $0.5 \mathrm{mg/kg/d}$ 4 wMale stroke-prorOlmDrinking water $10 \mathrm{mg/kg/d}$ 4 wMale stroke-prorTelOral $10 \mathrm{mg/kg/d}$ 2 uFemale C57BLKTelDrinking water $2 \mathrm{mg/kg/d}$ 2 wMale SHRTelDrinking water $2 \mathrm{mg/kg/d}$ 14 or 28 dWistar rats withTelDrinking water $2 \mathrm{mg/kg/d}$ 14 or 28 dWistar rats withTelDrinking water $10 \mathrm{mg/kg/d}$ 14 or 28 dWistar rats withTelDrinking water $10 \mathrm{mg/kg/d}$ 14 or 28 dWistar rats withTelDrinking water $10 \mathrm{mg/kg/d}$ 12 dLewis ratsLis/(Los)Drinking water $10 \mathrm{mg/kg/d}$ 2 wMale DS ratsLis/(Los)Drinking water $10 \mathrm{mg/kg/d}$ 2 wMale SHRLis/(Los)Drinking water $10 \mathrm{mg/kg/d}$ 2 wLewis ratsLis/(Los)Drinking water $10 \mathrm{mg/kg/d}$ 2 wMale C57BL6Lis/(Los)Drinking water $10 \mathrm{mg/kg/d}$ 2 wLewis ratsLis/(Los)Drinking water $10 \mathrm{mg/kg/d}$ 2 wMale C57BL6RamGavage $10 \mathrm{mg/kg/d}$ 2 wMale C57BL6 <td< td=""><td>SO</td><td>Intravenous</td><td>2.5 mg/kg/h</td><td>I</td><td>SD rats with acute lung injury induced by LPS</td><td>Lung</td><td>I</td><td>Ι</td><td>←</td><td>09</td></td<>	SO	Intravenous	2.5 mg/kg/h	I	SD rats with acute lung injury induced by LPS	Lung	I	Ι	←	09
Los / Olm Subcutaneous Los: 10 mg/kg/d 28 d Lewis rats with Olm: 0.1 mg/kg/d Olm Subcutaneous 0.5 mg/kg/d 4 w Male stroke-proronon Olm: 0.1 mg/kg/d Olm Drinking water 10 mg/kg/d 4 w Male stroke-prorononon Olm: 0.1 mg/kg/d Azi/Olm Dral gavage 1 or 5 mg/kg/d 4 w Male stroke-proronononononononononononononononononon	SO	Oral gavage	10 or 30 mg/kg/d	I	SD rats exposed to cigarettes for 6 months	Lung	I	←	I	61
OlmSubcutaneous0.5 mg/kg/d4 wMale stroke-provOlmDrinking water10 mg/kg/d4 w.Male stroke-provAzi/OlmOral1 or 5 mg/kg/d4 w.Male stroke-provTelOral1 or 5 mg/kg/d4 w.Male stroke-provTelOralDrinking water2 mg/kg/d2 wFemale C57BLKTelDrinking water2 mg/kg/d10 wMale SHRTelGastric gavage10 mg/kg/d8 wMale SHRTelGastric gavage10 mg/kg/d14 or 28 dWistar rats withTelDrinking water5 mg/kg/d14 or 28 dWistar rats withLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wMale SD ratsLis/(Los)Drinking water10 mg/kg/d2 wMale SD ratsLis/(Los)Drinking water10 mg/kg/d2 wSD rats with MIRamOral1 mg/kg/d2 wSD rats with MIRamOral1 mg/kg/d2 wSD rats with MIFis/(Los)Drinking water1 mg/kg/d2 wSD rats with MIFis/(Los)Drinking water1 mg/kg/d3 wSD rats with MIFis/(Los)Drinking water1 mg/kg/d5 wMale C57bl6 micLis/(Los)Drinking water1 mg/kg/d2 wSD rats with MIFis/(Los)Drinking water1 mg/kg/d5 wMale C57bl6 micFisDrinking wat	os / Olm	Subcutaneous	Los: 10 mg/kg/d Olm: 0.1 mg/kg/d	28 d	Lewis rats with MI induced by coronary artery ligation	Heart	←	I	I	4
OlmDrinking water10 mg/kg/d14 dSHRAzi/OlmOral gavage1 or 5 mg/kg/d4 w.Male hRN/hANGTelOralI or 5 mg/kg/d2 wHewis rats withTelDrinking water2 mg/kg/d2 wFemale C57BLKTelDrinking water2 mg/kg/d10 wMale SHRTelGastric gavage5 or 10 mg/kg/d10 wMale SHRTelGastric gavage10 mg/kg/d14 or 28 dWistar rats withThDrinking water50 mg/kg/d17 dC57BL/6 miceEpr/(spi)Intraperitoneal5 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wMale SD ratsLis/(Los)Drinking water10 mg/kg/d2 wMale C57bl6 miceACE inhlibrosDrinking water10 mg/kg/d2 wMale C57bl6 miceLis/(Los)Drinking water10 mg/kg/d2 wSD rats with MIRamOral1 mg/kg/d5 wMale C57bl6 miceLis/(Los)Drinking water1 mg/kg/d5 wMale C57bl6 miceFisCavage<	JIm	Subcutaneous	0.5 mg/kg/d	4 w	Male stroke-prone SHR	Heart/Kidney	\leftarrow	I	I	45
Azi/OlmOral gavage1 or 5 mg/kg/d4 w.Male hRN/hANTelOral10 mg/kg/d21 dLewis rats withTelDrinking water $2 mg/kg/d$ 2 wFemale C57BLKTelGastric gavage $5 or 10 mg/kg/d$ $2 w$ Male SHRTelGastric gavage $5 or 10 mg/kg/d$ $14 or 28 d$ Wistar rats withTroDrinking water $5 mg/kg/d$ $14 or 28 d$ Wistar rats withIrbDrinking water $50 mg/kg/d$ $17 d$ $C57BL/6$ miceACE inhibitors $10 mg/kg/d$ $12 d$ Lewis ratsLis/(Los)Drinking water $10 mg/kg/d$ $2 w$ Lewis ratsLis/(Los)Drinking water $10 mg/kg/d$ $2 w$ Lewis ratsLis/(Los)Drinking water $10 mg/kg/d$ $2 w$ Male SD ratsLis/(Los)Drinking water $10 mg/kg/d$ $2 w$ Male C57BL/6 miceACE inhibitors $1 mg/kg/d$ $2 w$ Lewis ratsLis/(Los)Drinking water $10 mg/kg/d$ $2 w$ Male C57bl6 miceACE inhibitors $1 mg/kg/d$ $2 w$ Male C57bl6 miceLis/(Los)Drinking water $1 mg/kg/d$ $2 w$ SD rats with MIRamGavage $1 mg/kg/d$ $8 w$ SD rats with MIRamGavage $1 mg/kg/d$ $8 w$ SD rats with MIEnaGavage $1 mg/kg/d$ $8 w$ SD rats with MIEnaGavage $1 mg/kg/d$ $8 w$ SD rats with MIEnaGavage	Olm	Drinking water	10 mg/kg/d	14 d	SHR	Thoracic aorta Carotid artery	← ↑		1 1	53
TelOral10 mg/kg/d21 dLewis rats withTelDrinking water2 mg/kg/d2 wFemale C57BLKTelGastric gavage5 or 10 mg/kg/d10 wMale SHRCan/(epl)Gastric gavage10 mg/kg/d14 or 28 dWistar rats withIrbDrinking water50 mg/kg/d17 dC57BL/6 miceACE inhibitors10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wMale C57BL/6 miceACE inhibitors10 mg/kg/d2 w12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wMale C57bl6 miceACE inhibitors1 mg/kg/d8 wSD rats with MIRamGavage1 mg/kg/d8 wSD rats with MIRamGavage1 mg/kg/d5 wMale C57bl6 miceFar<	Azi/Olm	Oral gavage	1 or 5 mg/kg/d	4 w.	Male hRN/hANG-Tg mice	Heart/Kidney	Azi† Olm→			46
TelDrinking water2 mg/kg/d2 wFemale C57BLKTelGastric gavage5 or 10 mg/kg/d10 wMale SHRCan/(epl)Gastric gavage10 mg/kg/d14 or 28 dWistar rats withEpr/(spi)Intraperitoneal5 mg/kg/d17 dC57BL/6 miceEpr/(spi)Intraperitoneal5 mg/kg/d17 dC57BL/6 miceLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wRamsCadaOral1 mg/kg/d2 wMale C57bl6 miRamGavage1 mg/kg/d5 wMale C57bl6 miFerDrinking water2 mg/kg/d8 wSD rats with MIRamGavage10 mg/kg/d5 wMale C57bl6 miFerDrinking water2 mg/kg/d10 dFemale SD ratsRamGavage10 mg/kg/d8 wSD rats with MIFerDrinking water2 mg/kg/d10 dFemale SD ratsRamGavage10 mg/kg/d5 wSD rats with atFerDrinking water2 mg/kg/d10 mg/kg/d10 d <td>[e]</td> <td>Oral</td> <td>10 mg/kg/d</td> <td>21 d</td> <td>Lewis rats with EAM</td> <td>Heart</td> <td>I</td> <td>←</td> <td>I</td> <td>47</td>	[e]	Oral	10 mg/kg/d	21 d	Lewis rats with EAM	Heart	I	←	I	47
TelGastric gavage5 or 10 mg/kg/d10 wMale SHRCan/(epl)Gastric gavage10 mg/kg/d8 wMale DS ratsEpr/(spi)Intraperitoneal5 mg/kg/d14 or 28 dWistar rats withIrbDrinking water50 mg/kg/d17 dC57BL/6 miceLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wRansCanOral1 mg/kg/d28 dSD rats with MIRamGavage1 mg/kg/d5 wMale C57bl6 miPerDrinking water1 mg/kg/d8 wSD rats with MIRamGavage10 mg/kg/d5 wMale C57bl6 miCapIntraperitoneal50 mg/kg/d6 wSD rats with MIEnaGavage10 mg/kg/d7 wSD rats with MIEnaGavage10 mg/kg/d7 wSD rats with MIEnaGavage10 mg/kg/d7 w8 wSD rats with MIEnaGavage10 mg/kg/d7 w8 wSD rats with MIEnaGavage10 mg/kg/d8 wSD rats with MIEnaGavage10 mg/kg/d-SD rats with with actEnaIntraperitoneal50 mg/kg-SD rats with act<	[el	Drinking water	2 mg/kg/d	2 w	Female C57BLKS/J mice	Kidney (renal vasculature)	- (←	I	57
Can/(epl)Gastric gavage10 mg/kg/d8 wMale DS ratsEpr/(spi)Intraperitoneal5 mg/kg/d14 or 28 dWistar rats withIrbDrinking water50 mg/kg/d17 dC57BL/6 miceACE inhibitors50 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wMale C57bl6 miRamOral1 mg/kg/d28 dSD rats with MIRamGavage1 mg/kg/d8 wSD rats with MIEnaGavage10 mg/kg/d8 wSD rats with MICapDrinking water2 mg/kg/d8 wSD rats with MICapIntraperitoneal50 mg/kg/d7 wSD rats with MI	Cel	Gastric gavage	5 or 10 mg/kg/d	10 w	Male SHR	Aorta	I	~	I	54
Epr/(spi)Intraperitoneal5 mg/kg/d14 or 28 dWistar rats withIrbDrinking water50 mg/kg/d17 dC57BL/6 miceACE inhibitors10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 dSD rats with MIRamOral1 mg/kg/d28 dSD rats with MIRamGavage1 mg/kg/d28 dSD rats with MIRamGavage1 mg/kg/d8 wMale C57bl6 miFerDrinking water2 mg/kg/d8 wSD rats with MICapIntraperitoneal50 mg/kg/d9 wSD rats with MICapIntraperitoneal50 mg/kg/d-SD rats with actMineralocorticoid receptor antagonists-SD rats with act	Can/(epl)	Gastric gavage	10 mg/kg/d	8 w	Male DS rats	Heart	\leftarrow	←	I	48
IrbDrinking water50 mg/kg/d17 dC57BL/6 miceACE inhibitorsLis/(Los)Drinking water10 mg/kg/d12 dLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d2 wLewis ratsLis/(Los)Drinking water10 mg/kg/d12 dmRen2 tg ratsRamOral1 mg/kg/d28 dSD rats with MIRamGavage1 mg/kg/d10 dFemale SD ratsPerDrinking water2 mg/kg/d5 wMale C57bl6 miEnaGavage10 mg/kg/d8 wSD rats with MIEnaGavage10 mg/kg/d5 wMale C57bl6 miCapIntraperitoneal50 mg/kg/d8 wSD rats with MICapIntraperitoneal50 mg/kg-SD rats with act	Jpr/(spi)	Intraperitoneal	5 mg/kg/d	14 or 28 d	Wistar rats with experimental CHF induced by ACF	Heart	Ι	←	←	49
ACE inhibitors Is/Los) Drinking water 10 mg/kg/d 12 d Lewis rats Lis/(Los) Drinking water 10 mg/kg/d 2 w Lewis rats Lis/(Los) Drinking water 10 mg/kg/d 2 w Lewis rats Lis/(Los) Drinking water 10 mg/kg/d 2 w Lewis rats Ram Oral 1 mg/kg/d 28 d SD rats with MI Ram Gavage 1 mg/kg/d 28 d SD rats with MI Ram Gavage 1 mg/kg/d 28 d SD rats with MI Ram Gavage 1 mg/kg/d 8 d SD rats with MI Ram Gavage 1 mg/kg/d 8 w SD rats with MI Enal Gavage 10 mg/kg/d 8 w SD rats with MI Enal Gavage 10 mg/kg/d 8 w SD rats with MI Cap Intraperitoneal 50 mg/kg/d 8 w SD rats with act Mineralocorticoid receptor antagonists - SD rats with act	rb	Drinking water	50 mg/kg/d	17 d	C57BL/6 mice	Aorta	←	←	I	55
Lis/(Los) Drinking water 10 mg/kg/d 12 d Lewis rats Lis/(Los) Drinking water 10 mg/kg/d 2 w Lewis rats Lis/(Los) Drinking water 10 mg/kg/d 12 d mRen2 tg rats Ram Oral 1 mg/kg/d 28 d SD rats with MI Ram Gavage 1 mg/kg/d 10 d Female SD rats Per Drinking water 2 mg/kg/d 5 w Male C57bl6 mi Ena Gavage 10 mg/kg/d 8 w SD rats with MI Cap Intraperitoneal 50 mg/kg - SD rats with act Mineralocorticoid receptor antagonists	ACE inhibi	tors								
Lis/(Los) Drinking water 10 mg/kg/d 2 w Lewis rats Lis/(Los) Drinking water 10 mg/kg/d 12 d mRen2 tg rats Ram Oral 1 mg/kg/d 28 d SD rats with MI Ram Gavage 1 mg/kg/d 10 d Female SD rats Per Drinking water 2 mg/kg/d 5 w Male C57bl6 mi Ena Gavage 10 mg/kg/d 8 w SD rats with MI Cap Intraperitoneal 50 mg/kg – SD rats with act Mineralocorticoid receptor antagonists	is/(Los)	Drinking water	10 mg/kg/d	12 d	Lewis rats	Heart	←	I	Ŷ	42
Lis/(Los)Drinking water10 mg/kg/d12 dmRen2 tg ratsRamOral1 mg/kg/d28 dSD rats with MIRamGavage1 mg/kg/d10 dFemale SD ratsPerDrinking water2 mg/kg/d5 wMale C57bl6 miEnaGavage10 mg/kg/d8 wSD rats with MIEnaGavage10 mg/kg/d5 wMale c57bl6 miCapIntraperitoneal50 mg/kg/d8 wSD rats with MICapIntraperitoneal50 mg/kg-SD rats with actMineralocorticoid receptor antagonists-SD rats with act	is/(Los)	Drinking water	10 mg/kg/d	2 w	Lewis rats	Kidney	I	Ι	←	56
RamOral1 mg/kg/d28 dSD rats with MIRamGavage1 mg/kg/d10 dFemale SD ratsPerDrinking water2 mg/kg/d5 wMale C57bl6 miEnaGavage10 mg/kg/d8 wSD rats with MICapIntraperitoneal50 mg/kg-SD rats with actMineralocorticoid receptor antagonists	is/(Los)	Drinking water	10 mg/kg/d	12 d	mRen2 tg rats	Heart/Kidney	←	Ι	←	43
RamGavage1 mg/kg/d10 dFemale SD ratsPerDrinking water2 mg/kg/d5 wMale C57bl6 miEnaGavage10 mg/kg/d8 wSD rats with MICapIntraperitoneal50 mg/kg-SD rats with actMineralocorticoid receptor antagonists	tam	Oral	1 mg/kg/d	28 d	SD rats with MI induced by ligation of the left coronary artery	Heart	ſ	Ι	Ŷ	50
PerDrinking water2 mg/kg/d5 wMale C57bl6 miEnaGavage10 mg/kg/d8 wSD rats with MICapIntraperitoneal50 mg/kg-SD rats with actMineralocorticoid receptor antagonists	Ram	Gavage	1 mg/kg/d	10 d	Female SD rats with acute kidney injury induced by STNx.	Kidney Plasma			$\leftarrow \rightarrow$	58
Ena Gavage 10 mg/kg/d 8 w SD rats with MI Cap Intraperitoneal 50 mg/kg – SD rats with acu Mineralocorticoid receptor antagonists	ber	Drinking water	2 mg/kg/d	5 w	Male C57bl6 mice with STZ-induced diabetes	Kidney Plasma	\rightarrow	\rightarrow	$\rightarrow \rightarrow$	59
Cap Intraperitoneal 50 mg/kg – SD rats with acu Mineralocorticoid receptor antagonists	ina	Gavage	10 mg/kg/d	8 w	SD rats with MI induced by ligation of the left coronary artery	Heart Plasma	←		$\leftarrow \leftarrow$	51
	Cap Mineraloco	Intraperitoneal rticoid receptor antago	50 mg/kg nists	I	SD rats with acute lung injury induced by LPS	Lung	I	←	I	62
Spi Water 40 mg/kg/d 4 or 11 d SD rats with BL	òpi	Water	40 mg/kg/d	4 or 11 d	SD rats with BDL as an obstructive jaundice model	Kidney	Ι	←	Ι	63

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/	× .								
Drugs ^a	Route of administra	tion Dose ^b	Duration ^c	Experimental animal ^d	Tissues ^e	ACE	5		Ref.
						mRN	VA Prote	in Activit	× ا
Spi	Drinking water	80 mg/kg/d	12 w	Wistar rats with MI induced by left coronary artery ligation	Heart	Ι	~	I	64
Spi/(Epr)	Intraperitoneal	15 mg/kg/d	14 or 28 d	Wistar rats with experimental CHF induced by ACF	Heart	Ι	\leftarrow	ſ	49
Epl/(Can)	Oral gavage	100 mg/kg/d	18 w	Male DS rats	Heart	ſ	ſ	Ι	48
Epl	Drinking water	200 mg/kg/d	2 w	Patients with CHF	МФ(h)	~	Ι	←	65
	•)		Balb/C mice	$M\Phi(m)$		I	←	
					Heart(m)	~	Ι	~	
					Kidney(m)	¢	Ι	1	
^a Los losar	tan, Lis lisinopril, Olm	olmesartan, Azi azilsa	ırtan, <i>Tel</i> telmi	:artan, Can candesartan, Epl eplerenone, Epr eprosartan, Spi sp	ironolactone, Ram rar	nipril, Per perii	ndopril, E	<i>na</i> enalapı	il, <i>Cap</i>
captopril									
^b d day, h	hour								
^с d day, <i>w</i>	week, m month								

SD Sprague–Dawley. LPS lipopolysaccharide, MI myocardial infarction, SHR spontaneously hypertensive rats. EAM experimental autoimmune myocarditis, hRN/hANG-Tg human renin/ human

angiotensinogen transgenic, DS Dahl salt-sensitive, CHF congestive heart failure, ACF aortocaval fistula, STNx subtotal nephrectomy, STZ streptozotocin, BDL bile duct ligation

m mouse

 $M\Phi$ macrophage, h human,

SPRINGER NATURE

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ACE2 in COVID-19 from a therapeutic point of view

From a therapeutic point of view, supplementation with soluble exogenous ACE2 can theoretically be favorable for protection against COVID-19, as it can inhibit interaction of the virus with endogenous ACE2. In fact, it was recently reported that human recombinant soluble ACE2 can inhibit infection of SARS-CoV-2 in human blood vessel organoids and human kidney organoids (Fig. 1) [73].

Cardiovascular and cerebrovascular complications in COVID-19

COVID-19 and thromboembolic complications

The risk of venous and arterial thromboembolic complications has been reported to be higher in patients with COVID-19. Klok et al. demonstrated the cumulative incidence of venous thromboembolism (VTE) in 27% and ischemic stroke in 3.7% of patients with COVID-19 pneumonia [74]. Lodigiani et al. also reported that among 388 COVID-19 inpatients, the ratio of thromboembolic events, including VTE, ischemic stroke, and ischemic heart disease, was higher in intensive care unit (ICU) patients (27.6%) than in patients in the general ward (6.6%) [75]. Regarding stroke, patients with severe infection exhibited neurologic manifestations such as acute cerebrovascular diseases (5.7% in severe vs 0.8% in nonsevere, respectively) [76]. In SARS, a case of VTE in multiple organs was described [77], but there are very few reports on SARS-induced thrombotic complications. Large-artery ischemic strokes occurred in 0.7% of Taiwanese [78] and 2% of Singaporean [79] SARS patients. For cases in Singapore, the authors considered that stroke occurred as a side effect of intensive treatment, such as intravenous immunoglobulin; thus, the incidence of VTE and ischemic stroke in COVID-19 patients appears to be remarkably higher than that in SARS patients. According to several reports of "stroke cases" with COVID-19 [80-84], almost all showed elevated plasma D-dimer levels. Additionally, higher D-dimer levels on admission effectively predicted in-hospital mortality in patients with COVID-19 [4, 85, 86]. Among thromboembolic complications, VTE was a common complication in hospitalized patients (observed in 20%) with COVID-19 and was associated with death (adjusted hazard ratio [HR]: 2.4) [87]. In particular, lower extremity deep-vein thrombosis (DVT) was detected in 85.4% of critically ill COVID-19 patients [88]. These findings indicate that abnormalities in the coagulation cascade result in VTE and stroke after SARS-CoV-2 infection. Therefore, systemic anticoagulation therapy may improve outcomes in COVID-19 patients [89].

Possible mechanisms of SARS-CoV-2-induced endothelial injury

SARS-CoV-2-induced ischemic organ damage appears to be associated with a hyperinflammatory state, cytokine storm, vascular endothelial damage or fibrinogen consumption coagulopathy (Fig. 1) [84]. Yang et al. clearly reviewed the underlying mechanisms of the dysfunctional coagulatory response in the pathogenesis of influenza A virus [90]. Toll-like receptors (TLRs) have a central role in innate immunity. Viral pathogen-associated molecular patterns such as viral proteins, double-stranded RNA, and single-stranded RNA initially activate the innate immune system [91]. Using in silico studies on the interaction of the SARS-CoV-2 spike glycoprotein with human TLRs, Choudhury et al. demonstrated that TLR4 is most likely to be involved in recognizing molecular patterns from SARS-CoV-2 to induce inflammatory responses [92]. Other TLRs, such as TLR5, TLR7, and TLR8, have been also reported to also be involved in SARS-CoV-2 infection [93, 94]. TLRs activate a common signaling pathway via MyD88, leading to the production of proinflammatory cytokines [95]. Further activation of the innate immune response to eradicate the virus induces overproduction of pro-inflammatory cytokines, resulting in a "cytokine storm" [96] of overactivated neutrophils, monocytes, and lymphocytes. Coagulation is a highly organized process that involves endothelial cells (ECs), platelets and coagulation factors in the sequential action of primary and secondary hemostasis and fibrinolysis [97]. Inflammatory cytokines and leukocyte activation lead to EC activation and endothelial dysfunction via multiple mechanisms, including direct damage, loss of tight junctions, and hyperpermeability induced by inflammatory factors [90]. Activated or injured ECs initiate coagulation by activating platelets and expression of coagulation components (Fig. 1). Visseren et al. found that respiratory virus-infected ECs exhibit procoagulant activity associated with the induction of tissue factor (TF) expression [98]. The coagulation cascade is initiated after exposure of TF to the blood. Recently, Varga et al. reported the pathology of EC dysfunction in COVID-19 based on postmortem analysis of three cases [99]. As ECs express ACE2 [30], viral inclusion structures in ECs and diffuse endothelial inflammation, such as endotheliitis, has been observed. These results suggest that SARS-CoV-2 virus infection directly and indirectly injures ECs and may activate the coagulation pathway. Very recently, Ackermann et al. performed morphologic and molecular analyses in the peripheral lung of seven cases of COVID-19 at autopsy [100], and severe EC injury associated with the presence of intracellular virus, disrupted cell membranes, widespread thrombosis with microangiopathy, and new vessel growth were observed. Ma et al. reported that human umbilical vein endothelial cells (HUVECs) express both ACE2 and TMPRSS2 mRNAs [101]. Human dermal microvascular endothelial cells are also known to express TMPRSS2 [102]. However, whether SARS-CoV-2 induces EC injury directly or indirectly remains unclear. An increase in vascular permeability induced by EC injury plays a pathogenic role in the development of pulmonary fibrosis [103], and acute respiratory distress syndrome (ARDS) is characterized by acute respiratory failure, bilateral pulmonary infiltrates, and noncardiogenic pulmonary edema resulting from vascular hyperpermeability [104]. Thus, EC injury can lead to respiratory dysfunction and may be a key factor that determines the clinical outcome of COVID-19 patients.

COVID-19 and Kawasaki disease

Verdoni et al. surprisingly reported a 30-fold increase in the incidence of Kawasaki-like disease (KLD) during the SARS-CoV-2 epidemic [105]. In that study, 10 patients were diagnosed with KLD between February 18, 2020 and April 20, 2020, compared with 19 patients in the five years before the beginning of the COVID-19 pandemic [105]. Two of 10 patients were positive for SARS-CoV-2 by reverse-transcriptase quantitative PCR assay, and eight of 10 patients diagnosed with KLD were positive for SARS-CoV-2 IgG and/or IgM antibodies. Kawasaki disease (KD) is an acute systemic vasculitis with coronary artery abnormalities that predominantly affects young children [106]. Necrotizing arteritis, subacute chronic vasculitis, and luminal myofibroblastic proliferation are three linked processes associated with KD [107]. Although a correlation between viral infections and KD was reported [108, 109], it is not clear whether KD-associated infections are causal or incidental. KD might also be associated with a dysregulated innate immune response;[110] however, a link between SARS-CoV-2 infection and KLD has not been demonstrated.

Myocardial injury associated with COVID-19

Clinical manifestations of cardiovascular disorders associated with COVID-19 are diverse, with heart failure, arrhythmia, cardiogenic shock, acute myocardial infarction (AMI), and myocarditis having been reported [4, 85, 111– 114]. In an initial report from Wuhan, China, Huang et al. documented that acute cardiac injury, as defined by the elevation of cardiac biomarkers (e.g., troponin I) or new abnormalities in electrocardiography and echocardiography, was present in 12.2% of 41 inpatients with COVID-19 [115]. In another study, acute cardiac injury (7.2%; using the same definition as above) and arrhythmia (16.7%) were common complications among 138 hospitalized patients with COVID-19 [85]. Cardiac complications in COVID-19 are associated with a poor clinical outcome [116, 117]. In one study, 19.7% of 416 hospitalized patients with COVID-19 had abnormally high levels of troponin I, and those with elevated troponin I levels had higher mortality (51.2%) than did those with lower levels (4.5%) [117]. Another study showed that elevation in troponin T levels is associated with the occurrence of malignant arrhythmias and fatal outcomes [116].

As described above, SARS-CoV-2 infection promotes vascular endothelial injury and thromboembolism, which can result in ischemic heart disease and stroke [84, 112]. However, cases of myocardial injury with no evidence of obstructive coronary disease have also been reported [114, 118, 119], suggesting that COVID-19 interacts with the cardiovascular system through multiple mechanisms.

Possible mechanisms of cardiovascular injury in COVID-19

Although the molecular causes of myocardial injury in COVID-19 have remained elusive, several mechanisms can be considered. Severe viral infection triggers systemic inflammatory response syndrome, which increases the risk of plaque rupture and thrombus formation, resulting in the occurrence of atherosclerotic diseases. For example, there is a significant association between the incidence of AMI and influenza [120]. In addition, several papers have reported the occurrence of AMI in COVID-19 patients [112, 121], though more studies are necessary to determine the actual prevalence. Cytokine storms can both cause plaque instability and promote cardiovascular inflammation and myocardial depression [122, 123], which may be involved in myocarditis and stress-induced cardiomyopathy in COVID-19 patients [114, 124].

Furthermore, recent data suggest that a substantial portion of patients with COVID-19 have risk factors for atherosclerotic disease, which include hypertension, obesity, smoking, and diabetes mellitus [6, 85, 111, 115]. Because severe respiratory viral infection induces hypoxemia and abnormal hemodynamic changes, enhanced BP variability may also trigger cardiovascular events in COVID-19 patients with atherosclerotic risk factors. These mechanisms likely act in parallel to cause cardiac damage, resulting in severe clinical manifestations such as AMI, ventricular arrhythmias, and congestive heart failure.

It is currently unknown whether SARS-CoV-2 infects cardiomyocytes. Autopsy studies have reported the detection of SARS-CoV-2 RNA in the heart [125]. However, the cell types in which viral RNA was detected were not defined. Endomyocardial biopsy in a case of COVID-19 with cardiogenic shock revealed viral particles in interstitial macrophages but not in cardiomyocytes [126]. Although ACE2 is expressed in human cardiomyocytes [127], there

appears to be little expression of TMPRSS2 in these cells [31]. Therefore, the direct pathogenic role of SARS-CoV-2 in cardiomyocytes needs further investigation.

Acute kidney injury (AKI) in COVID-19: incidence and clinical significance

In addition to cardiac complications, accumulating data indicate that renal abnormalities frequently accompany COVID-19, though there does seem to be a geographic difference in the occurrence of AKI. Several single-center studies from Wuhan report that AKI developed in 3-7% of patients with COVID-19 [85, 115, 128], and the reported incidence of AKI in a multicenter study involving 1099 inhospital patients in 30 provinces across mainland China was 0.5% [129]. Studies in the US found AKI incidence to be higher than that in the aforementioned reports. For instance, the incidence of AKI was 19.1% among critically ill patients in a small single-center study in Seattle [111], and in a multicenter study conducted in the largest academic health system in New York [6, 130], AKI occurred in 36.6% of 5449 patients admitted with COVID-19, of whom 14.3% required renal replacement therapy (RRT). This geographic difference in the occurrence of AKI might be explained by several factors, including disease severity, ethnicity, and comorbid conditions (such as diabetes and coronary artery disease), all of which are reported to be independent risk factors for AKI in COVID-19 [130].

AKI in COVID-19 is highly associated with respiratory failure. In one study, as many as 89.7% of mechanically ventilated patients developed AKI, whereas AKI occurred in 21.7% of nonventilated patients [130]. The demand for RRT may significantly increase during the COVID-19 pandemic, and clinicians should be aware of the possibility of facing challenges in delivering RRT to COVID-19 patients [131]. Moreover, in line with findings for SARS [132], the mortality rate seems much higher among COVID-19 patients with AKI than among those without AKI. In a report by Zhou et al. [4], AKI was observed only in 0.7% of survivors, whereas 50.0% of nonsurvivors had AKI. Consistently, a dose-dependent relationship between mortality and the severity of AKI was observed in a different cohort, with stage 3 AKI being associated with a fourfold increase in mortality risk [133]. A poor prognosis for COVID-19 patients with AKI was also confirmed in a US cohort [130]. In addition, renal abnormalities associated with COVID-19 may not necessarily be limited to AKI. Pei et al. reported that proteinuria and hematuria were seen in 65.7% and 41.7% of 333 patients with COVID-19 pneumonia, respectively [134]. Glomerulopathy associated with COVID-19 has also been reported [135, 136].

Mechanisms of kidney injury associated with COVID-19

Multiple mechanisms explaining the occurrence of kidney injury in COVID-19 have been proposed, which include but are not limited to the following: hemodynamic instability and renal ischemia, ARDS and the cytokine storm, rhabdomyolysis, hypercoagulability and thrombosis, cardiac failure and kidney congestion, and direct renal infection of SARS-CoV-2. In one study, AKI was more common in patients with cardiac injury than in those without cardiac injury, suggesting a cardio-renal interaction [117].

In autopsy data for 26 cases, diffuse proximal tubule injury was prominent, along with erythrocyte aggregates in peritubular capillaries, ischemic changes with fibrin thrombi in glomeruli, and pigmented casts indicative of rhabdomyolysis [137]. Of note, this study identified virus particles with crown-like morphology in electron microscopy within renal tubules and podocytes, which was accompanied by degenerative changes such as vacuolization and necrotic epithelia. Another study also identified abundant viral forms in the area of vacuolated tubules in a case of COVID-19 at autopsy [138]. These reports were followed by the demonstration of SARS-CoV-2 RNA in the kidney [125, 139]. Such histopathological analyses of cases postmortem indicate that SARS-CoV-2 may have tropism for the kidney, especially in severe cases, though the pathogenic role of direct infection remains undetermined. ACE2 is abundant in renal proximal tubules [31, 140], and its levels are altered in disease states such as diabetes mellitus [140–142]. However, TMPRSS2 is highly expressed in the more distal portion of the renal tubules [31]. At present, it is unclear how SARS-CoV-2 enters renal cells and whether comorbid conditions affect the cellular tropism of the virus in the kidney. It also remains to be determined whether renal infection with SARS-CoV-2 indeed contributes to kidney injury.

COVID-19 in advanced CKD

Several lines of evidence suggest that CKD patients, especially those at advanced stages, are vulnerable to SARS-CoV-2 infection. In a multicenter study of 5700 patients hospitalized with COVID-19, end-stage kidney disease (ESKD) was present in 3.5% of all cases [6]. Conversely, the prevalence of laboratory-confirmed COVID-19 was 2.1% in 7154 patients undergoing hemodialysis in Wuhan, which was apparently higher than the morbidity of the general population in that area (~0.5% as of March 2020) [143]. In a cross-sectional study of 3802 cases with SARS-CoV-2 test results, CKD was associated with a positive SARS-CoV-2 result after adjustment for potential confounding variables [144]. Moreover, a higher baseline serum creatinine level is an independent risk factor for in-hospital death in COVID-19 [133], suggesting that ESKD patients are at high risk of developing severe disease. In recent reports, the overall mortality rate for ESKD patients with COVID-19 was 29% (out of 94 cases) in Italy [145] and 31% (of 59 cases) in New York [146]. The basis for such vulnerability is likely multifactorial, and both medical (older age, immune cell dysfunction, cardiovas-cular and pulmonary comorbidities) and environmental factors need to be considered [147, 148]. Currently, several guiding principles have been proposed to mitigate the risk of COVID-19 in ESKD patients [149–151].

Antihypertensive agents during the COVID-19 pandemic

Use of ACE inhibitors and ARBs in patients with COVID-19

Because ACE inhibitors and ARBs may increase the amount of ACE2, whether these drugs should be discontinued during the COVID-19 pandemic has been a topic of discussion [152, 153]. The role of ACE2 in the pathophysiology of COVID-19 as well as experimental evidence for ACE2 and RAS inhibitors are reviewed in detail in the previous sections. In the following sections, we discuss the clinical evidence for COVID-19 and antihypertensive agents.

Although several reports in an early phase of the COVID-19 pandemic have suggested the relationship between COVID-19 and ACE inhibitors or ARBs [154, 155], it is possible that the association between COVID-19 and hypertensive medication results from reverse causality because older patients, who are at the highest risk for COVID-19, tend to have multiple comorbidities, including hypertension and CVD. Moreover, adjustments for age and other possible confounding factors were not performed in most of the early studies [156–158].

Several recent reports analyze the effect of ACE inhibitors and ARBs on clinical outcomes in patients with COVID-19 and hypertension [159, 160]. A retrospective, single-site, cohort study from Wuhan compared clinical outcomes among 126 COVID-19 patients with pre-existing hypertension (43 of whom were taking either ACE inhibitors or ARBs; 83 of whom were not taking these agents) and 125 age- and sexmatched COVID-19 control patients without hypertension [159]. In that study, it was found that ACE inhibitors or ARBs did not increase the risk of morbidity or mortality in patients with SARS-CoV-2 infection. Moreover, the study showed a nonsignificant trend toward marginally lower critical illness and death rates in patients taking ACE inhibitors or ARBs compared to those taking other antihypertensive agents. In a study from Spain, a case-population study (1139 cases and 11,390 population controls) showed that users of ACE inhibitors or ARBs had an adjusted odds ratio for COVID-19 requiring admission to hospital of 0.94 (95% CI 0.77–1.15) compared to users of other antihypertensive drugs, with no increased risk for ACE inhibitors or ARBs [160]. This study concluded that ACE inhibitors or ARBs do not increase the risk of COVID-19 requiring admission to the hospital, including fatal cases and those admitted to ICU, and that these agents should not be discontinued to prevent the development of severe COVID-19 [160].

Two other retrospective cohort studies from China comparing disease severity and mortality rates between hypertensive patients taking ACE inhibitors or ARBs and those not taking these agents are available [161, 162]. One, a single-center study, showed that the percentage of patients taking ACE inhibitors and ARBs did not differ between those with severe and nonsevere infections or between survivors and nonsurvivors [161]. In the other, a multicenter cohort study, hypertensive patients with COVID-19 who were taking ACE inhibitors or ARBs were compared with those who were taking antihypertensive drugs other than ACE inhibitors and ARBs as well as patients with COVID-19 without hypertension [162]. The risk for 28-day all-cause mortality was lower in the ACE inhibitor/ARB group than in the control group (adjusted HR 0.42, 95% CI 0.19-0.92; P = 0.03) and in matched subgroup analysis (adjusted HR 0.30, 95% CI 0.12–0.70; P = 0.01).

In addition, two clinical studies from northern Italy and from New York provide further evidence on the association between antihypertensive medication and COVID-19 [163, 164]. In the case-control study in Italy, which included 6272 patients with COVID-19 and 30,759 controls matched for age, sex, and municipality of residence, after adjustment for drugs and coexisting conditions, the odds ratios for the use of ACE inhibitors and ARBs were 0.96 (95% CI 0.87-1.07) and 0.95 (95% CI 0.86-1.05), respectively, among all patients and 0.91 (95% CI 0.69-1.21) and 0.83 (95% CI 0.63-1.10), respectively, among patients who had a severe or fatal course of the disease [163]. In the New York study, none of the major classes of antihypertensive drugs, including ACE inhibitors and ARBs, were associated with a positive SARS-CoV-2 test or disease severity [164].

Taken together, the data from these six studies, although retrospective, from different countries [159–164] provide evidence for continuing treatment with ACE inhibitors or ARBs in patients with hypertension during the COVID-19 pandemic. Moreover, a recent meta-analysis showed the potential benefit of ACE inhibitors or ARBs in patients with hypertension [165]. In nine studies comprising 3936 patients with hypertension and COVID-19, ACE inhibitors or ARB treatment was not associated with disease severity but was related to lower mortality from COVID-19 compared with other antihypertensive drugs. Although future well-designed randomized controlled trials are needed, these results suggest that treatment with ACE inhibitors or ARBs should be continued in COVID-19 patients with hypertension [165–167].

Use of other antihypertensive agents in patients with COVID-19

The aforementioned two studies on RAS inhibitors also address the association of COVID-19 with other classes of antihypertensive agents [163, 164]. In one study, the adjusted odds ratios for COVID-19 associated with the use of calcium-channel blockers, beta-blockers, thiazide diuretics, loop diuretics, and MRA were 1.03 (95% CI 0.95-1.12), 0.99 (95% CI 0.91-1.08), 1.03 (95% CI 0.86-1.23), 1.46 (95% CI 1.23-1.73), and 0.90 (95% CI 0.75-1.07), respectively [163]. Thus, none of the antihypertensive agents except loop diuretics were associated with an increased risk of COVID-19 in multivariate analysis. In the study population, loop diuretics were used more frequently in patients with COVID-19 than controls (13.9% versus 7.8%; relative difference, 43.6%) [163], and their use may reflect the existence of severe comorbidities such as heart failure and renal dysfunction, the severities of which were not appropriately quantified [163]. In another study, calcium-channel blockers, beta-blockers, and thiazide diuretics were not associated with an increased likelihood of a positive SARS-CoV-2 test [164].

Statements on the use of antihypertensive agents from hypertensive societies and associations worldwide

The International Society of Hypertension [168], the European Society of Hypertension [169], the European Society of Cardiology [170], and the American Heart Association/ Heart Failure Society of America/American College of Cardiology [171] have already made a statement on the use of RAS inhibitors during the COVID-19 pandemic. the Japanese Society of Hypertension [172] and the Japanese Circulation Society [173, 174] have also provided statements regarding the management of cardiovascular diseases during the COVID-19 pandemic. All these statements indicate that there is no good evidence to change or discontinue ACE inhibitors or ARBs to avoid or manage SARS-CoV-2 infection.

Current conclusion regarding the use of antihypertensive agents during the COVID-19 pandemic

In the early phase of the COVID-19 pandemic, there was considerable confusion regarding whether ACE inhibitors or ARBs may have adverse effects on COVID-19 patient morbidity and mortality, which was based on the speculation that ACE2 can be upregulated by ACE inhibitors or ARBs. However, the evidence of ACE2 upregulation is limited to experimental studies. Furthermore, there have been no clinical studies supporting the hypothesis that ACE inhibitors and ARBs augment susceptibility to infection and worsen all-cause mortality and cardiovascular outcomes in COVID-19 patients. Thus, treatment with ACE inhibitors and ARBs should be continued in high-risk patients who have received guideline-directed medical therapy, and hypertension should be managed in accordance with the Japanese Society of Hypertension Guidelines for the Management of Hypertension (JSH2019) [18].

Areas of uncertainty and future perspectives

In this article, we review recent studies on COVID-19 in the context of hypertension and related diseases. As discussed in detail, there are no reliable reports on whether SARS-CoV-2 infection risk is increased in patients with hypertension. Hypertension is known to be associated with endothelial injury, especially in the elderly [175], and recent evidence suggests that thromboembolism triggered by endothelial injury is one of the important complications that influence disease outcome in COVID-19. At this time, it is unclear whether pre-existing endothelial injury increases the severity of COVID-19; however, hypertensive patients with atherosclerotic diseases may need to be carefully monitored for the occurrence of new-onset CVDs during SARS-CoV-2 infection.

Given that hypertension is the leading contributor to the development of cardiovascular and kidney diseases and given that myocardial injury and advanced CKD are associated with an increased risk of severe disease following SARS-CoV-2 infection, optimal management of hypertension can contribute to a better prognosis of COVID-19 by mitigating the progression of these disorders. Regardless, a major challenge is to achieve target BP control in the "New Normal" lifestyle, in which health care workers may have a reduced opportunity for in-person clinical examination of patients. In the Great East Japan Earthquake in 2011, medical care for patients with hypertension was compromised ("disaster hypertension") [176]. After the recognition of BP increases following the disaster, which may have contributed to the increased cardiovascular events [176, 177], the remote BP monitoring system using information and communication technology (ICT) was introduced and was indeed useful in achieving target BP control [178, 179]. Therefore, in the post-COVID-19 era, medical practice using ICT may need to be widely implemented for the management of hypertension. In addition, the COVID-19 pandemic may increase the risk of mental disordersowing to, for example, anxiety, economic issues, and decreased physical activity—all of which can potentially compromise BP control. It is currently unknown whether the COVID-19 pandemic will affect BP control and the development of CVDs in the long term; nonetheless, it is necessary to carefully monitor each patient's BP.

Last, in addition to the development of effective treatment, vaccination for SARS-CoV-2 will of course be helpful. In the case of influenza infection, several epidemiological studies and randomized control trials have clearly shown that there is a strong inverse relationship between influenza vaccination and the risk of cardiovascular events [180].

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Compliance with ethical standards

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