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Pharmacology of Aging: *Drosophila* as a Tool to Validate Drug Targets for Healthy Lifespan

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Abstract

Finding effective therapies to manage age-related conditions is an emerging public health challenge. Although disease-targeted treatments are important, a preventive approach focused on aging can be more efficient. Pharmacological targeting of aging-related processes can extend lifespan and improve health in animal models. However, drug development and translation are particularly challenging in geroscience. Preclinical studies have survival as a major endpoint for drug screening, which requires years of research in mammalian models. Shorter-lived invertebrates can be exploited to accelerate this process. In particular, the fruit fly *Drosophila melanogaster* allows the validation of new drug targets using precise genetic tools and proof-of-concept experiments on drugs impacting conserved aging processes. Screening for clinically approved drugs that act on aging-related targets may further accelerate translation and create new tools for aging research. To date, 31 drugs used in clinical practice have been shown to extend the lifespan of flies. Here, we describe recent advances in the pharmacology of aging, focusing on *Drosophila* as a tool to repurpose these drugs and study age-related processes.

Conflict of Interest

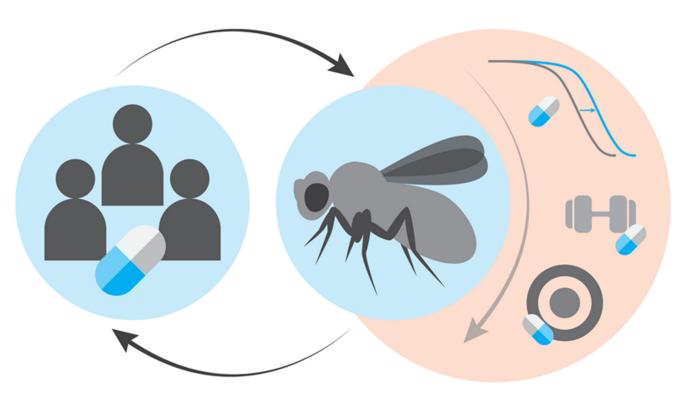
The authors declare that they have no conflict of interest.

Data Selection

Drugs were selected from the DrugAge database⁵⁰ (https://genomics.senescence.info/drugs), based on the following inclusion criteria: 1) median/mean lifespan extension of *Drosophila melanogaster* by at least 1%, 2) statistically significant effect, and 3) clinically authorized according to the European Medicines Agency, Article 57(2) of Regulation (EC) No. 726/2004 EMA/518502/2018 (https://www.ema.europa.eu/en/human-regulatory/post-authorisation/data-medicines-iso-idmpstandards/public-data-article-57-database). Information was accessed on 20 June 2022, revised on 3 May 2024.

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Introduction—Preventive Medicine Applied to Age-Related Disease

Aging is the main risk factor for chronic diseases and geriatric syndromes^{1–3}. The increasing prevalence of these conditions is a serious public health challenge^{4,5}. Age-related disorders are responsible for major healthcare costs and a growing burden on healthcare provision and caregiving, which threaten to become unsustainable from an economic and societal standpoint^{6,7}.

Major advances in modern medicine, with significant improvements in human health and life expectancy, arose from preventive approaches, such as immunization against infectious diseases and public health campaigns⁸. In geriatric medicine, personalized prevention already occurs in clinical practice. For instance, death from cardiovascular events is prevented pharmacologically by addressing risk factors such as hypercholesterolemia and hypertension. Similarly, the development of other age-related diseases, such as type-2 diabetes and certain cancer types, is delayed through dietary programs and the cessation of smoking^{9–11}.

Geroscience proposes the targeting of a more comprehensive risk factor: aging (Fig. 1). Treating or preventing an individual disease, if prevalent and deadly, can increase human life expectancy. However, the added years do not necessarily correlate with an improved quality of life¹², as aging will affect the whole organism and promote the development of other diseases and geriatric syndromes. Geriatric syndromes are linked to increased morbidity and reduced quality of life in the elderly² as well as higher mortality¹³. For instance, falls are a main cause of hospital admissions in the elderly¹⁴. Similar to other

geriatric syndromes, falls result from multiple age-related factors, such as deteriorating muscle strength, loss of balance, and impaired vision, which illustrates the challenge of managing these conditions with an organ- or disease-focused approach. Instead, targeting aging has the potential to decrease the incidence of both age-related diseases and geriatric syndromes, extending human life with added years of health. Indeed, interventions that target aging-related processes improve healthspan in a range of animals ^{15,16}.

Pharmacology of Aging: An Ancient Quest for a Modern Problem

The search for a compound able to target aging and extend life predates modern scientific history. Until just a few decades ago, this pursuit was still compared with medieval alchemy, even by prominent figures within the then-emerging field of aging research¹⁷. Initial studies in biogerontology were faced with scepticism at the idea of controlling longevity and concerns that a longer lifespan may not translate into a healthier one¹⁸. However, lifespan extension is often a side effect of health-promoting interventions, such as dietary restriction (DR). DR, defined as reduced food intake without malnutrition, is one of the most robust evolutionary conserved strategies to extend healthy lifespan¹⁵. Since the initial reports of DR in the early 20th century, researchers have strived to identify drugs that could act as DR mimetics and recapitulate its effects^{19,20}. In addition, other dietary regimes, such as intermittent fasting, and genetic interventions within nutrient-sensing signaling cascades have similar benefits^{21,22}. Accordingly, most current aging-related pharmacological interventions act—at least partly—via nutrient-sensing pathways²³.

In addition to dysregulated nutrient sensing, aging is associated with other processes that impact homeostasis with age, such as the accumulation of senescent cells or mitochondrial dysfunction²³⁻²⁶. Therapies targeting these processes also improve health in animal models²⁷⁻³¹.

Drug development is a slow and lengthy process, with most drugs failing to reach clinical trials³². This is particularly challenging in geroscience, because survival is a major endpoint of preclinical studies. Although new drug discovery is needed, a range of drugs already approved for clinical use in humans have exhibited prolongevity effects in model organisms. Studying the potential of these repurposed drugs to prevent human age-related disease may be an important first step in bringing the pharmacology of aging to the clinic, because they can be more rapidly and safely translated and may serve as proof-of-principle that targeting aging processes can improve population health. In addition, dissecting their mechanisms in animal models can help increase treatment efficacy by finding more specific longevity-modulating targets. This goal underpins the Intervention Testing Program by the U.S. National Institute on Aging^{33,34} and some recent biotech companies^{35,36}, most of which investigate longevity in mice.

Drosophila as a Tool to Repurpose Drugs for Healthy Aging

Mammals such as mice are valuable model systems from a translational perspective due to their closer evolutionary proximity to humans, in particular to study drug metabolism, excretion, and toxicity. However, invertebrate models are important complementary tools

in pharmacology by allowing quicker, cheaper, and larger-scale studies, with fewer bureaucratic restrictions^{37,38}. Short-lived invertebrates are particularly useful in aging research to validate the targets of known drugs as longevity modulators, and to screen for effective drugs before further pre-clinical assessment of toxicity and pharmacokinetics in mammals (Fig. 2). When repurposing approved drugs, invertebrates may accelerate translation even more by providing proof-of-concept studies in the context of longevity and disease prevention³⁹.

Two invertebrate systems, the nematode worm *Caenorhabditis elegans* and the fruit fly *Drosophila melanogaster*, are extensively used in aging research. As one of the most prevalent occurrences across life, aging is expected to happen through analogous processes and manifest in invertebrates via shared pathways. Indeed, many pioneering discoveries in the aging field were initially made in the worm and the fly, and subsequently shown to be evolutionarily conserved in mammals^{40–42}.

Drosophila has a relatively short lifespan (typically ~2–3 months in the laboratory compared with ~2–3 years for mice) and exhibits strong evolutionary conservation of metabolic and signaling pathways associated with aging. Flies also allow the use of powerful genetic tools that can validate drug targets, enabling the mechanistic analysis of pharmacological interventions and the assessment of systemic physiological consequences⁴⁰.

The *Drosophila* model is historically important in the discovery and characterization of compounds that modulate longevity. Diiodomethane was the first nonnutrient shown to increase fly life-span in the $1970s^{43}$. Since then, other examples with comparatively strong effects on fly survival include the metabolites phenylbutyrate⁴⁴ and α -ketoglutarate⁴⁵ as well as compounds such as trichostatin $A^{46,47}$, torin 1^{48} , and dihydromyricetin⁴⁹.

According to the DrugAge database, there are >600 compounds that significantly extend the lifespan of different animal models⁵⁰. Among these, >150 compounds have been tested in *Drosophila*. Here, we specifically focus on a subset of 31 drugs that are clinically approved by the European Medicines Agency and significantly extend median/mean lifespan in flies (Table 1). The following eight drugs recently identified by a high-throughput screen will not be described in detail, as they require further validation and characterization in the context of fly survival: bupivacaine, fluspirilene, fluvoxamine, haloperidol, mianserin, promethazine, tetracycline, and thioridazine⁵¹. In this review, we discuss the remaining 22 clinically approved drugs present in the DrugAge database, along with an additional compound (zoledronate) from a recent study⁵², which meets our inclusion criteria, and we highlight how the *Drosophila* model has contributed to advancing our understanding of their prolongevity benefits.

Most of these drugs act on nutrient-sensing pathways. To adapt to the energy supply and demands of the extracellular environment, cells sense nutrients essentially via two key pathways: the insulin/insulin-like growth factor signaling (IIS) pathway and the mechanistic target-of-rapamycin (TOR) pathway, which are activated under conditions of high glucose and amino acids, respectively. In contrast, under low cellular energy status, AMP-activated protein kinase (AMPK) promotes ATP production via the increased expression of genes and

activity of proteins involved in catabolism, and energy conservation by inhibiting anabolic processes. Pharmacological modulation of components within these key metabolic cascades can impact survival (Fig. 3). The mechanisms whereby other clinically relevant drugs (i.e., not classically implicated in nutrient sensing) extend *Drosophila* lifespan are currently less explored and understood, as discussed in the respective sections.

Rapamycin and derivatives: Inhibiting the TOR pathway

Rapamycin, also known as sirolimus, is a drug that blocks the activation of the TOR kinase, more specifically acting on TOR complex 1 (TORC1). In humans, this results in the inhibition of T-cell activation and proliferation, leading to immunosuppression. Rapamycin is clinically used in immunosuppressive regimens, including the prophylaxis of kidney transplant rejection⁵³.

As the TOR pathway is also a key player in longevity modulation, rapamycin has been thoroughly studied in this context⁵⁴. Rapamycin was shown to increase the survival of multiple organisms, such as worms, flies, and mice^{55–58}. Importantly, rapamycin extends mammalian survival even when administered chronically later in life^{34,55,59,60} and improves health by preventing aging-related phenotypes, such as liver degeneration, tendon stiffness, periodontal bone loss and inflammation, and loss of hematopoietic stem cell capacity^{59,61,62}.

In *Drosophila*, rapamycin extends lifespan independently of sex and fly genetic background⁵⁶, although not consistently in males⁶³. Its derivative everolimus was independently found to extend mean lifespan in males⁶⁴. Transient rapamycin treatment during fly development and early adult life is sufficient to extend lifespan as well as chronic treatment from middle and old age^{65,66}. Consistent with other organisms, this effect is related to the inhibition of TORC1⁵⁶. By inhibiting TORC1, rapamycin induces a cellular perception of amino acid deprivation, resulting in the induction of autophagy and reduced protein synthesis via S6K inhibition^{56,67}. The lifespan extension is not dependent on the mRNA translation inhibitor 4E-BP, another target of TORC1, even though 4E-BP is necessary for the protective role of rapamycin in fly neurodegeneration models^{68,69}.

Tissue-specific effects of TOR pathway inhibition have systemic consequences sufficient to extend fly lifespan. For example, inhibitory modulation of TOR pathway gene expression in the fat body increases fly survival⁷⁰. In the gut, rapamycin delays age-related barrier dysfunction and decreases the rate of intestinal stem cell (ISC) proliferation^{58,67,71}. These effects are mediated by the indirect inhibition of polymerase III, an enzyme downstream of TORC1 that generates ribosomal RNA involved in protein synthesis⁵⁸, and the overexpression of histones H3 and H4^{63,71}. The counter-intuitive synthesis of these histone proteins in enterocytes occurs via a noncanonical mechanism mediated by translation initiation factor 3 (IF-3), resulting in changes to chromatin organization in enterocyte nuclei and altered expression of autophagy-related genes. These genes include *bchs*, a cargo adaptor for selective degradation of ubiquitinated protein aggregates, which is required for rapamycin-associated lifespan extension^{63,71}. Similarly, the lifespan extension after early life transient rapamycin treatment is mediated by sustained elevated autophagy in intestinal enterocytes⁶⁶. Overall, these studies suggest that rapamycin extends fly lifespan by promoting intestinal health. Male flies have preserved intestinal barrier function with age

and lower ISC proliferation rates⁷² as well as high basal levels of enterocyte autophagy that are not further increased by rapamycin, which can explain the sexually dimorphic effect of rapamycin on lifespan⁶³.

A side effect of rapamycin in female flies is decreased fecundity^{56,58,73}, although rapamycin treatment can still extend survival in sterile females⁵⁶. More recently, rapamycin was shown to only extend the lifespan of females under sterol-limited conditions⁷³. Because egg production depletes sterol availability and flies are incapable of *de novo* cholesterol synthesis, rapamycin may extend survival by inhibiting sterol-consuming processes such as egg production. Indeed, cholesterol supplementation alone extends fly lifespan, which is not further increased by the addition of rapamycin⁷³. This diet–drug interaction seems to be independent of the microbiota, because the effects of rapamycin on *Drosophila* tissue aging and lifespan are unaffected under germ-free conditions⁷⁴.

Another effect of rapamycin treatment in flies is systemic fat accumulation in the form of triglycerides (TAG), which recapitulates the hypertriglyceridemia seen in humans taking rapamycin⁵³. This may be a consequence of *fork head (fkh)* upregulation. FKH is a transcription factor whose overexpression increases nutrient transporter genes resulting in raised TAG levels in the fly, and it is required for rapamycin-related survival extension⁷⁵. This effect may explain why rapamycin promotes resistance to starvation independently of S6K inhibition or autophagy upregulation⁵⁶. In humans, rapamycin also reduces insulin sensitivity and glucose tolerance⁷⁶, but this adverse effect was shown to be mediated by chronic TORC2 inhibition and did not affect survival in a mammalian model⁷⁷. In fact, insulin resistance *per se* can be uncoupled from survival in both mice and flies^{77,78}.

In addition to lower doses and shorter treatment duration⁶⁰, more specific TORC1 inhibitors may prevent the side effects related to chronic TORC2 inhibition and improve treatment efficacy. Recent reports from clinical trials of TORC1 inhibitors show promising results in terms of safety and efficacy for reducing infection rates in the elderly^{79–81}.

Lithium: Inhibiting the IIS pathway

Lithium is a mood stabilizer. Although the key mechanisms underlying its mode of action are still unclear, lithium is approved for the treatment of mood conditions, such as bipolar disorder⁸². In the UK Biobank cohort, patients treated with lithium show longer survival compared with individuals taking other antipsychotics⁸³. Interestingly, lithium extends the lifespan of multiple organisms, including flies^{84–86}.

In *Drosophila*, low doses of lithium extend lifespan, even when administered from middle age (32 days of adulthood). This effect is greatest under fully fed conditions (i.e., on a yeast-rich diet), suggesting that lithium may partly act via mechanisms mimicking DR⁸⁶, consistent with previous evidence from *C. elegans* studies⁸⁷. However, lithium extends lifespan beyond DR, indicating additional independent or synergistic effects⁸⁶. Lithium also increases the survival of flies fed a sucrose-rich diet, which is linked to changes in lipid metabolism^{86,88}.

The lifespan extension by lithium in flies relies on its ubiquitous inhibition of Shaggy (Sgg), the fly ortholog of glycogen synthase kinase-3 (GSK-3), and the activation of cap'n'collar C (CncC), the fly ortholog of nuclear factor erythroid 2-related factor (NRF-2). The prolongevity effect seems to be hormetic, because strong inhibition of Sgg/GSK-3 is detrimental for survival⁸⁶. Increased lithium bioavailability due to different food composition might thus explain the shortening of female fly lifespan in an independent study, where flies of similar genetic background were fed equivalent doses of the drug⁸⁹. This consideration prompts the need for further research exploring lithium—diet interactions, which is important given the particularly narrow therapeutic window⁸⁶.

GSK-3 is upregulated in many disease states, such as neurodegeneration, type-2 diabetes, inflammatory conditions, and some types of cancer⁹⁰. More selective GSK-3 inhibitors are in development and being assessed in clinical trials for Alzheimer's disease and progressive supranuclear palsy⁹¹. These drugs could also be repurposed for longevity modulation and may avoid the influence of lithium on mood, interactions with other molecular targets, and possible long-term side effects, such as renal damage^{88,92}.

Trametinib: Inhibiting the Ras-Erk pathway

Trametinib is an inhibitor of the Ras pathway by targeting the mitogen-activated protein kinase kinases (MEK1/2) and used as a chemotherapeutic agent for tumors in patients with the BRAF V600E activating mutation, commonly found in melanomas⁹³.

In *Drosophila*, trametinib extends lifespan by preventing the activation of the downstream MEK target, extracellular signal-regulated kinase (Erk), both when supplemented from early adulthood (2 days old) or midlife (30 days old). This inhibition is not accompanied by a compensatory overactivation of the upstream Ras pathway or the PI3K/Akt pathway⁹⁴.

Trametinib lowers fecundity, which is expected because loss-of-function female Ras mutants are sterile⁹⁴. Similar to rapamycin, reduced egg laying may improve survival by avoiding the depletion of essential micronutrients, an example of trade-off between fertility and longevity⁹⁵. However, trametinib extends lifespan when administered from a late age when fecundity is already decreased, which makes this scenario less likely. Furthermore, despite affecting fecundity, trametinib does not alter feeding⁹⁴, suggesting that its beneficial effects on lifespan are not via induction of DR.

Similar to rapamycin, trametinib may extend lifespan by promoting gut health. Although an initial study suggested that trametinib does not protect from age-related gut barrier dysfunction or alter proliferation rates of ISCs at either 15 or 65 days of age in female flies⁹⁴, subsequent studies using the same fly background, sex, and drug dose showed gut barrier protection by trametinib at 60 days and decreased proliferation of ISCs at 35 days⁷². However, the prolongevity effects of trametinib are inconsistent in males and seemingly unrelated to gut homeostasis⁹⁶. Similar to rapamycin, trametinib indirectly decreases RNA polymerase III activity and its prolongevity effect is partially mediated by the polymerase III repressor Maf1⁹⁶. These studies suggest a convergent mechanism through which life-extending drugs act on specific RNA synthesis pathways to preserve female gut health.

Combination therapy: Rapamycin, lithium, and trametinib

Because the three drugs described previously—rapamycin, lithium, and trametinib—target different interconnected pathways (Fig. 3), understanding whether they act separately or via the same mechanisms within a network is crucial. This question was addressed in a study showing that the triple drug combination extends lifespan by almost 50%, and results in prolongevity benefits beyond those of additive effects, suggesting synergy⁹⁷. This synergistic effect is consistent with observations in *C. elegans*, where concomitant interventions in the TOR and IIS pathways had higher effects on survival than the sum of their individual effects⁹⁸.

Pharmacologically, this synergy may result from improvements in different health determinants that favor each other, or the neutralization of respective negative drug side effects. For instance, lithium treatment reverses the accumulation of TAG associated with rapamycin, accordingly decreasing rapamycin-induced resistance to starvation. In addition, lithium further extends the lifespan of long-lived IIS mutant flies lacking the *Drosophila* insulin-like peptides (dILPs) 2, 3 and 5, counteracting the potential activation of Sgg as a result of lowered IIS signaling, which would partly limit survival⁹⁷. Trametinib improves insulin resistance in obese mice⁹⁹ and may prevent this side effect associated with chronic rapamycin treatment in flies.

Feeding behavior is unaltered by the triple drug combination, and reduced fecundity is mainly a consequence of trametinib, as no further change was observed when the other two drugs were included⁹⁷, which makes the trade-off scenario unlikely to explain the effects of the combination. Systemic levels of each drug were unaltered by the triple combination⁹⁷, suggesting the absence of pharmacokinetic effects in flies causing increased bioavailability of a certain drug. This study supports the principle of an intervention that combines multiple active substances targeting different aging-related processes to extend healthspan and lifespan.

Metformin: Modulating the metabolism of microbiota

Metformin is the first-line treatment for type-2 diabetes. It is a cheap and well-tolerated drug that increases insulin sensitivity in peripheral tissues and decreases liver glucose production, thereby decreasing hyperglycemia with a good safety profile, i.e., with a low risk of hypoglycaemia. Beyond its antidiabetic properties, epidemiological and animal model studies indicate that metformin can improve health markers and survival, including in late-onset interventions in mammals ^{100–102}. A series of clinical trials are planned to assess if metformin also prevents the incidence of age-related diseases in humans ¹⁰³.

Metformin has multiple modes of action, with part of its metabolic effects reported to occur via the inhibition of complex I in the mitochondrial respiratory chain and the indirect activation of AMPK ^{104,105}. Despite evidence of prolongevity benefits in other model organisms, such as worms and rodents, metformin was initially reported not to extend lifespan in *Drosophila*, even though *in vivo* AMPK activation and decreased TAG levels were observed as expected ¹⁰⁶. However, a subsequent study showed that metformin can extend fly lifespan under defined nutritional and microbial conditions ¹⁰⁷. Consistent

with earlier findings in *C. elegans*¹⁰⁸, the microbiota was found to be essential for the prolongevity effects of metformin in flies. Metformin leads to the production of the bacterial metabolite agmatine, which is essential for survival benefits in both the worm and fly host¹⁰⁷, although how bacteria-derived agmatine leads to lifespan extension in the host remains to be elucidated. A further independent study reported lifespan extension by metformin in flies of a different strain¹⁰⁹. This effect was related to the prevention of ubiquitinated protein aggregates in adult muscle, linked to muscle autophagy induction¹⁰⁹.

Overall, the case of metformin exemplifies the importance of the gut microbiota in modulating different aspects of nutritional physiology and pharmacological metabolism to impact on host longevity^{104,105,110}, which adds another layer of complexity to the study of drug interventions in aging.

NSAIDs in longevity: Aspirin, salicylamide, ibuprofen, and celecoxib

Nonsteroidal anti-inflammatory drugs (NSAIDs) are first-line treatment options in a wide range of medical conditions, mostly related to their anti-inflammatory benefits. They include non-selective cyclooxygenase (COX) inhibitors, such as aspirin and ibuprofen, as well as the selective COX-2 inhibitor celecoxib¹¹¹.

Although the existence of COX enzymes in *Drosophila* remains unclear, the NSAIDs aspirin, ibuprofen, and celecoxib have all been shown to increase fly survival^{112–116}, which makes the fly system particularly useful to assess the action of these drugs on other targets and characterize the effect of those targets on life-span modulation.

Aspirin has been shown to extend lifespan in different model organisms, including mammals¹¹⁷. In humans, a recent report indicated that aspirin does not prolong disability-free survival of healthy individuals over the age of 70¹¹⁸. However, aspirin was previously shown to improve aspects of numerous age-related diseases, such as certain types of cancer, type-2 diabetes, atherosclerosis, and neurodegenerative diseases¹¹³, and it may prevent disease if administered from earlier in life. Aspirin extends lifespan in *Drosophila* and decreases fecundity without altering food intake^{112,113,119}, indicating that DR does not play a role in the longevity effect. In fact, similar to rapamycin, aspirin-treated flies have higher TAG content and increased starvation resistance. Aspirin was recently shown to prevent the dysbiosis of commensal microbiota in the fly gut as well as age-related gut leakage and ISCs over-proliferation, partly through the downregulation of the inflammatory Imd pathway¹¹⁹. Salicylamide, another salicylic acid derivative, was also found to extend *Drosophila* lifespan, but the mechanisms remain unexplored¹²⁰.

The effects of ibuprofen on fly survival are controversial. Initially, ibubrofen was reported to increase lifespan moderately in both sexes when supplemented to the diet at 0.5 or 1 μM^{114} . In another independent study, 1 μM ibuprofen extended the survival of females, but not males, when given for 10 days during middle age (from the age of 30 days) but not from early adulthood 115 . In fact, ibuprofen treatment in males was found to impair physical activity, a parameter of healthspan, by an unknown mechanism 115 . Therefore, the role of ibuprofen in modulating fly survival and the context of its prolongevity effects still require further clarification.

Celecoxib, in the form of 2,5-dimethyl-celecoxib (DMC), can extend *Drosophila* lifespan in a sex-independent way, even when administered from later timepoints. The ability of DMC to modulate lifespan was shown to require both IIS and TOR signaling and to be dependent on Akt inhibition ¹¹⁶.

Nonselective COX inhibition by aspirin and ibuprofen can have serious side effects in the long term, such as gastrointestinal bleeding and nephrotoxicity, to which the elderly are particularly susceptible ¹¹¹. Therefore, drugs specifically aimed at COX-independent targets that promote longevity and health must be developed to improve their potential for translation. Long-term COX-2 inhibition is also associated with life-threatening complications. The celecoxib-derivative DMC lacks COX inhibitory function, which may allow the use of this drug in the context of longevity ¹¹⁶.

Simvastatin and zoledronate: Mevalonate pathway inhibition

Simvastatin belongs to the statin class of drugs and is a competitive inhibitor of 3-hydroxy-3-methyl-glutaryl-coenzyme A (HMG-CoA) reductase, part of the mevalonate pathway responsible for cholesterol biosynthesis. In the clinic, simvastatin is used to treat pathologically elevated cholesterol levels, providing protection against cardiovascular disease. Zoledronate is a bisphosphonate that inhibits the mevalonate pathway down-stream of HMG-CoA by targeting the enzyme farnesyl pyrophosphate synthase (FPPS). This results in the inhibition of osteoclastic bone resorption in humans, making zoledronate suitable for diseases such as osteoporosis.

Both simvastatin and zoledronate extend male *Drosophila* median lifespan when administered continuously from early life^{52,121}. Zoledronate also extends both male and female survival when given from a later timepoint⁵². Both compounds improve health markers with age: simvastatin reduces age-related cardiac arrhythmias¹²¹ and zoledronate improves climbing activity and the maintenance of intestinal epithelium function with age⁵².

Unlike mammals, flies lack several enzymes required for *de novo* cholesterol biosynthesis, and therefore the beneficial effects of simvastatin on *Drosophila* health and survival must be cholesterol independent. The mevalonate pathway is also important for protein prenylation and as a precursor for the generation of sterol hormones and isoprenoids. By targeting HMG-CoA reductase, simvastatin could potentially decrease juvenile hormone (JH) signaling or levels of coenzyme Q, which have both been implicated in longevity ^{122,123}.

The potential role of protein prenylation in determining fly survival has been demonstrated, with pharmacological inhibition of isoprenyl transferases extending *Drosophila* lifespan¹²¹. Although the detection of prenylated proteins in fly samples was not technically possible, decreased prenylation of Ras family small GTPases was observed in the liver of mice after simvastatin treatment (188 mg per kg of food)¹²¹. However, in an independent study, simvastatin at low and high doses (20 and 120 mg per kg of food, respectively) did not exhibit a prolongevity effect in either male or female mice³⁴. These discordant results suggest that the beneficial effects of statins on survival may be dose-, disease-, and/or species-specific and require further investigation.

Sevelamer

Sevelamer is an anion exchange resin that binds phosphate and was developed to reduce elevated serum phosphate in individuals with chronic kidney disease. In flies, sevelamer treatment (1% w/v in the diet) results in lifespan extension, which is abolished when food is supplemented with excess (30 mM) phosphate ¹²⁴. Intriguingly, sevelamer does not change the concentration of phosphate circulating in the fly hemolymph¹²⁴. As the study did not include lifespan data under low dietary phosphate conditions, it is difficult to conclude that reduced phosphate uptake is responsible for the sevelamer-mediated increased longevity. Further investigations are warranted to explore other potential actions of this drug as well as the effects of phosphate independently on physiology and survival. For instance, sevelamer in humans is able to reduce blood uric acid levels and inflammatory factors such as IL-6¹²⁵, which have been implicated in longevity modulation^{23,78}.

N-acetylcysteine

N-acetylcysteine (NAC) is administered in humans to treat paracetamol overdose and as a mucolytic for conditions such as cystic fibrosis. Several studies have assessed the effects of NAC treatment on fly survival, with mixed outcomes depending on the sex and genetic background $^{126-128}$. NAC was initially reported to extend the lifespan of $Oregon\ R$ males when either 1 or 10 mg/mL were supplemented to the food 126 . Subsequently, NAC was shown to only extend the survival of $Canton\ S$ male flies but not females 128 . Conversely, NAC was recently found to extend lifespan in females but not males of the w^{1118} strain at a dose of 1 mg/mL, while a higher concentration of 10 mg/mL was toxic 127 . The reason for this variability between sexes and genetic backgrounds is unclear and may be influenced by other factors such as differences in the basal diet composition between the studies (see Table 1).

All three reports claim that the modulation of survival by NAC is through its antioxidant activity, both directly by detoxifying reactive oxygen species (ROS) and indirectly by upregulating antioxidant systems. Recent findings suggest that NAC does not alter H₂O₂ flux but rather suppresses complex I-linked respiration in female flies, while typically maintaining a reduced glutathione pool at lower doses (1 mg/mL)¹²⁹. NAC was shown to elevate the transcript levels of catalase and phospholipid-hydroperoxide glutathione peroxidase in the whole body, and more specifically in the head and abdomen of females flies, although these results again seem to be strain-specific^{127,128}. The activity of these enzymes, known to be important ROS scavengers, was also increased in response to NAC and correlated with enhanced resistance to paraquat in one report¹²⁷. However, a recent study in *C. elegans* shows dose-dependent shortened survival following NAC administration associated with the inhibition of the worm NRF-2 ortholog and suggests that scavenging naturally occurring ROS may be harmful by inhibiting healthy redox signaling¹³⁰. Indeed, NAC can induce reductive stress at high concentrations¹²⁹.

Overall, considering the discrepancy of these results, further studies should dissect the interacting factors or conditions that allow NAC to extend lifespan and explore the role of antioxidant factors and specific redox signaling pathways, which may uncover new and more consistent targets for pharmacological intervention in this context¹³¹.

Corticosteroids

Corticosteroids are synthetic analogs of the steroid hormones produced in the cortex of adrenal glands. They can have either glucocorticoid or mineralocorticoid properties, or both to varying degrees. Corticosteroids are prescribed to ameliorate a wide range of medical conditions. Glucocorticoids can be used for their immunosuppressive and anti-inflammatory properties in autoimmune or inflammatory disorders involving effectively any organ. Mineralocorticoids regulate electrolytes and water balance and are used in combination with glucocorticoids as replacement hormonal therapy, for example, in the case of adrenal insufficiency ¹³².

Both types of corticosteroid are reported to extend fly life-span¹²⁰. The glucocorticoid triamcinolone extends the survival of both males and females. Hydrocortisone and its pro-drug cortisone, which possess both glucocorticoid and mineralocorticoid activities, similarly extend fly lifespan independently of sex¹²⁰. Other glucocorticoids, fluprednisolone and dexamethasone, were reported to have no impact or even to shorten fly lifespan¹²⁰, despite appearing as drugs that significantly extend lifespan in the DrugAge database⁵⁰. Conversely, the mineralocorticoids fludrocortisone and desoxycorticosterone are absent from the database but are reported to significantly extend lifespan¹²⁰. Although these drugs were proposed to maintain membrane stability and thereby slow aging, no mechanistic evidence was provided¹²⁰.

Flies were recently found to have a receptor that responds to cortisone, encoded by the estrogen-related receptor (ERR) gene¹³³. Both cortisone and dexamethasone may act in the fly by conserved immunosuppressive mechanisms to increase susceptibility to infection^{133,134}. Interestingly, an endogenous ligand for the fly ERR has yet to be identified and, therefore, further research is needed to understand the role of steroid hormones in fly physiology¹³³.

The anti-inflammatory effects of glucocorticoids may decrease the age-related low-grade inflammation known to occur in mammals, in part related to the overproduction of proinflammatory factors and aged innate immunity cells¹³⁵. The decreased corticoid production with age in mice correlates with the surge of this systemic pro-inflammatory profile and was proposed to be one of its drivers¹³⁵. However, in humans, daily glucocorticoid production varies with age in a U-shaped pattern¹³⁶. At older ages, total levels actually increase, and are related to circadian changes, with the peak diurnal rhythm shifted to later in the day, and higher concentrations in the late evening and early night 137. This circadian shift may be detrimental for health, for example, by affecting quality of sleep¹³⁷. Together with known adverse effects of chronic glucocorticoid intake, such as osteoporosis and dyslipidaemia, this suggests that glucocorticoids may be of limited use to improve human health in old age. Similarly, chronic intake of mineralocorticoids in the absence of adrenal insufficiency will likely be detrimental due to the retention of sodium and water, and increased risk of hypertension, a major risk factor for cardiovascular events. Despite this, further research into maintaining a normal circadian production of these hormones over time and their beneficial specific targets for fly survival may uncover new strategies to preserve human health in old age.

Mifepristone

Mifepristone (or RU486) is another synthetic steroid that acts instead as an antagonist to human progesterone and type II glucocorticoid receptors and is clinically used to terminate pregnancy¹³⁸

In fly research, mifepristone is used as the inducing agent for the GeneSwitch conditional gene expression system^{139–141}. This inducible method allows mifepristone-dependent temporal changes in gene expression in flies with the exact same genetic background and has been widely applied in longevity studies. More recently, this system has been reported to be leaky under some conditions (i.e., to induce expression of the transgene in the absence of the drug), dependent on the upstream activating sequence and driver lines used^{142,143}.

In addition to potential interference by endogenous ligands at the GeneSwitch receptor in the absence of the drug, mifepristone itself has been reported to affect fly metabolism and lifespan 138,142–145. This effect was first described in mated female flies but not in nonmated females or males, and it was found to be genotype-dependent 143. In an independent study, mifepristone was shown to decrease food intake in low-yeast conditions (0.1% w/v) and to exacerbate lifespan-shortening undernutrition, but had little to no effect on either food intake or lifespan of flies at higher yeast content (5% w/v), independent of their sex 142. The studies where mifepristone showed a positive effect on survival used food with an intermediate yeast content (~2.5% w/v), without change in food intake 138,143,144. Although later reported to also moderately extend nonmated female lifespan 138,144, mifepristone was shown to mainly extend survival of mated females to nonmated levels by counteracting the negative effects of mating, which are related to innate immune system activation by bacterial factors. Mifepristone is proposed to prevent these effects by antagonizing JH signaling downstream of male sex peptide in the midgut 138,144,145.

Overall, these studies suggest that when using the GeneSwitch system in the context of longevity, experiments should be adequately controlled or interpreted taking into account the potential indirect effects of mifepristone¹⁴¹. The action of mifepristone also appears to depend on the nutritional context and varies according to the composition of the fly food. Considering the specific conditions under which mifepristone itself affects survival, it is unclear whether this drug acts on aging or aging-related disease mechanisms that can be translated to the clinic.

Rifampicin

Rifampicin is clinically used as an antibiotic to treat infections caused by a broad range of bacteria. It binds specifically to the β subunit of bacterial RNA polymerases to inhibit transcription, while having little to no activity against human polymerases ¹⁴⁶.

In *Drosophila*, rifampicin alone was shown to only moderately extend male lifespan but displayed a strong synergistic effect with rapamycin and even more in triple combination with allantoin to extend mean survival by up to 77% compared with the short-lived controls¹⁴⁷. The mechanism of action was not explored in the fly model, but in *C. elegans*, rifampicin was dependent on *daf-16*, the worm ortholog of the forkhead box-O transcription factor (FOXO), for the survival outcome¹⁴⁷. In a recent study, rifampicin is proposed

to prevent the activation of the innate immune system by gut microbiota¹⁴⁸. Therefore, further research into innate immunity activation and survival of flies may reveal new pharmacological targets for improved health in old age.

Minocycline

Minocycline, a second-generation tetracycline, is a broad-spectrum antibiotic prescribed for conditions such as acne vulgaris. Its bacteriostatic action is related to the inhibition of protein synthesis in bacteria by binding to the bacterial 30S ribosomal subunit¹⁴⁹. Beyond its antibiotic effect, minocycline is shown to improve health in animal models of several neurodegenerative and inflammatory diseases. Multiple mechanisms have been proposed related to different molecular targets interacting with the drug. These include its antioxidant properties, calcium chelation, and the ability to inhibit proinflammatory enzymes¹⁴⁹.

In *Drosophila*, minocycline extends the survival of both males and females from different fly strains. This effect correlates with a delay in the age-related loss of motor activity 150-¹⁵³. Interestingly, minocycline also extends the lifespan of germ-free flies¹⁵¹, suggesting its prolongevity action is at least partially independent of the microbiota. This drug does not alter feeding rates or fecundity, indicating that its effects on survival are not caused by DR or a reproductive trade-off¹⁵¹. Oxidative stress resistance is enhanced by minocycline in different species 149. However, as discussed with NAC, its ROS scavenging effect might be detrimental under homeostatic conditions. The transcription factor FOXO seems to be necessary for minocycline-associated oxidative stress resistance and lifespan extension, because these effects are lost in foxo-null mutant flies. Consistent with this observation, minocycline increases FOXO expression in thoracic muscle and the fat body 151 and prevents the accumulation of ubiquitinated protein aggregates in flight muscles by upregulating autophagy in a FOXO-Hsp70-dependent manner¹⁵⁴. FOXO overexpression in skeletal muscle¹⁵⁵ and fat body^{156,157} increases fly lifespan. However, minocycline further extends the lifespan of flies overexpressing FOXO ubiquitously 151, prompting additional studies not only to characterize the interaction of minocycline with this transcription factor but also to dissect other potential molecular targets impacting longevity.

Lamotrigine

Lamotrigine is used in the clinic as an anticonvulsant to treat conditions such as epilepsy as well as a mood stabilizer for individuals with bipolar disorder⁸². In *Drosophila*, lamotrigine is reported to increase survival in both males and females¹⁵⁸. However, lamotrigine also reduces fly locomotor activity, suggesting that the lifespan extension may not be accompanied by healthspan benefits¹⁵⁸, which weakens its potential for translation. Nevertheless, future studies providing mechanistic insight into how lamotrigine promotes fly survival may help find more specific targets that do not compromise parameters of health.

Morphine

Morphine is an opiate commonly used in the clinic as pain-relief (antalgic) therapy. In *Drosophila*, morphine is reported to extend lifespan, primarily in males, even when administered later in life¹⁵⁹, although no mechanistic insight was provided. The adverse effects of morphine in humans, such as respiratory depression, constipation, gain of

tolerance, and physical dependence, undermine its potential to be repurposed for health benefits or disease prevention. However, understanding how morphine promotes survival in flies may uncover novel aging-related targets to benefit human health.

Future Directions and Challenges

Drug repurposing can facilitate clinical translation ¹⁶⁰, but screening compounds for agerelated disease prevention is still challenging. These issues are not specific to *Drosophila*, but addressing them will make the fly a more reliable model for pre-clinical studies and drug translation in the context of aging. Here, we discuss the following challenges: 1) the use of survival as a proxy for aging, 2) the lack of comprehensive health scores, 3) false hits and the conservation of drug targets, 4) the diet dependency of survival effects, and 5) the downside of combination treatments.

1 The use of survival as a proxy for aging

In the absence of universal and robust biomarkers, population survival is the most widely used parameter as a proxy for aging, which is an important limitation to consider. While targeting aging should extend population survival, this is not sufficient to classify an intervention as geroprotective or antiaging. Life-extending interventions may promote fly health and survival independently of aging¹⁶¹. Semantically, interventions that extend lifespan must be termed prolongevity or prosurvival until further evidence. Rejuvenating antiaging therapies will be more potent and should be the ultimate goal of biogerontology, but these will only emerge from a better understanding of aging itself¹⁶².

Among other factors, survival within a population depends on individual genetic predisposition to develop disease. Therefore, it is important to use genetically heterogeneous populations when testing longevity interventions ¹⁶³. In a genetically heterogeneous population, individuals may still share the same predisposition to develop a disease with age that limits the life expectancy of the population. Treating these aging-unrelated factors to protect against the development of that disease can theoretically extend median lifespan. However, other age-related conditions will eventually develop resulting in smaller changes to the maximum lifespan of the population. Directly treating aging itself should instead lower the risk for all age-related diseases by definition, and thus extend both median and maximum lifespan (Fig. 1), which is necessary to classify a drug as a geroprotector.

2 The lack of comprehensive health scores

To further assess the efficacy of longevity interventions in both preclinical and clinical studies, drugs should improve comprehensive health scores ¹⁶⁴. If aging is associated with a loss of organ function and homeostasis over time, measuring functional and systemic homeostasis markers is necessary to screen for antiaging compounds. These scores could replace survival as the primary endpoint of early clinical studies, which may accelerate the translation of interventions focused on aging. Health scores to assess aging interventions should combine functional, imaging, and biochemical parameters. Some are already applied in geriatric medicine: for example, frailty indexes measure performance in activities of daily living, while mental state scores examine cognitive function ^{165,166}. These scores can be

expanded to include more detailed physical and psychological performance tests combined with imaging signs of tissue aging and molecular biomarkers of normal organ function to create a global view of individual health.

In this context, it is equally important to standardize healthspan and frailty measures in invertebrate animal model studies, as attempted in mammals ¹⁶⁷. For instance, the creation of a *Drosophila* health and frailty index (e.g., based on physiological and behavioral assays including physical activity, movement and feeding, as well as markers of organ function) would be valuable to assess the effects of interventions on health the same way across studies. This would complement lifespan information when screening for drugs that target aging and could be used as exclusion criteria when screening for geroprotectors.

3 False hits and the conservation of drug targets

Drug screening in animal models can result in false negative and false positive hits. In addition to applying comprehensive health scores, a way to increase confidence in positive hits is to show conservation of the molecular target in functionally equivalent tissues.

A false negative occurs when a drug fails to extend the lifespan or improve the health of a model organism, while able to prevent the consequences of aging in humans. Aging is associated with systemic loss of homeostasis, which is maintained differently across species depending on respective organ composition and physiology. Consequently, aging can manifest differently between species ¹⁶⁸. Particular organs can even be key drivers for age-related disease in a given species. For instance, preventing human thymic involution may ameliorate age-related immune dysfunction and improve human health in old age. However, fundamental aging processes can be the same across different organs. Building on the previous example, thymus involution occurs partially through stem cell exhaustion, which is a consequence of aging seen across multiple organs and species^{23,169}.

Even though flies lack certain organs present in humans, they do have many functionally similar organs with comparable anatomical distribution and physiology. Flies are useful to uncover interventions that act on conserved aging processes, which are conceptually more likely to affect multiple organs and therefore be more potent targets. Overall, false negatives are hard to predict and avoid, as prior knowledge of these human-specific aging processes is required, but they also have fewer consequences for drug development compared with false positives in terms of time and financial investment.

A false positive happens when a drug extends the lifespan of a model organism via mechanisms that prevent survival-limiting pathophysiological processes specific to that model ¹⁶¹. To prevent false positives, it is important to show that the drug target is conserved and that potential organ-specific effects occur in tissues that have at least a functional equivalent in humans. *Drosophila* research is equipped with powerful genetic tools to easily and precisely validate beneficial drugs and targets in a tissue-specific manner. This can be accomplished by: 1) engineering knock-in mutants where the proposed target is unresponsive to the drug, cancelling its effects; 2) knocking down or overexpressing the proposed target to simulate action of the drug on the endpoint, without further

pharmacological additive effects; and 3) using tissue specific or inducible drivers to apply the above genetic constructs in a localized or temporally defined manner.

Measuring drug bioavailability in the fly hemolymph—the circulating extracellular fluid analogous to blood—is technically possible⁹⁷ and should be considered in future studies to ensure that a drug has the ability to reach target tissues. For example, FOXO and AMPK increase fly lifespan when genetic interventions are restricted to the gut^{170,171}. Drugs that act on these pathways may similarly have gut-specific effects with systemic consequences for health. However, if drug absorption is not shown, additional systemic effects might be missed.

4 The diet dependency of survival effects

Drugs can interact with the diet to impact longevity in several ways. Food composition can affect the outcome of an intervention by influencing drug bioavailability ¹⁷². At a biological level, the presence/absence of specific nutrients can determine whether a drug is mechanistically able to extend lifespan ^{73,108}. Furthermore, as exemplified by metformin and rapamycin, diet and drugs can jointly modulate microbiota and host cell metabolism with systemic effects ¹⁰⁵. Therefore, it is critical to report comprehensive details of media recipes for all drug studies and possibly standardize diets across laboratories to make findings more comparable and reproducible ^{172–174}.

Diet alone is a key factor in chronic disease prevention¹⁷⁵. Integrating dietary information with drug interventions targeting aging and their metabolic consequences may uncover synergies that promote survival more potently. In *Drosophila*, this can be accomplished by using chemically defined media, where the constituents of the fly diet can be altered individually^{172,174} and by performing multilevel screens integrating nutritional and pharmacological information with metabolic and microbiome data¹⁰⁷.

While fine-tuning the nutritional context to improve drug efficacy and favor longevity is important, an ideal antiaging drug should be effective under a broad range of dietary conditions. In fact, interventions that target nutrient-sensing pathways may promote health and survival of animal models independently of aging, by ameliorating the side effects of overnutrition when reared in the laboratory compared with the wild. Similarly, DR mimetics may improve human health solely by preventing the consequences of overfeeding. Drugs targeting aging must show additional health and longevity benefits even under DR conditions.

5 The downside of combination treatments

Combining therapies that target each of the currently described aging-related processes may be a potent strategy to deal with such a pervasive problem as aging²³. Within the same hallmark of aging, targeting different components can similarly be beneficial. For example, the triple drug combination study⁹⁷ shows that multiple nutrient-sensing-related drugs acting on different processes can synergise, possibly by counteracting respective side effects. Another example of potential synergy in age-related drugs is the combination of metformin with rapamycin in mammals, even though further conclusive data are required¹⁷⁶.

An issue with combination therapies is that polypharmacy itself is a geriatric syndrome. Drug interactions, side effects, and dosage are particularly important problems in the clinical management of elderly patients¹⁷⁷. Combining different active substances into one pill, as proposed previously¹⁷⁸, may work in an optimized experimental cohort, but translation to humans, especially to the elderly, will face challenges related to altered metabolic rates, enzyme activities, or kidney function. Individuals may also be taking other disease-targeted drugs that can interfere with their respective bioavailability, leading to the abolition of benefits or to toxicity. A poly-pill strategy may avoid some of these challenges if applied early in life and/or in a personalized manner, but substantial progress is needed to ensure effectiveness and safety (Table 2).

Another approach is to find divergent or convergent aging targets. There is evidence that different processes of aging interact, which means that a single target may have multiple beneficial effects. For example, pharmacologically targeting components of nutrient-sensing pathways can ameliorate age-related loss of proteostasis⁷⁴, cellular senescence¹⁷⁹, and altered intercellular communication¹⁸⁰.

Conclusion

Longevity can be modulated pharmacologically. As a short-lived metazoan, *Drosophila* is a useful tool to dissect how drugs increase lifespan and study their biological targets *in vivo*. However, there is still a long road ahead for the pharmacology of aging to deliver on its promise. Further studies are needed to discover new fundamental mechanisms of aging and validate interventions as reliable promoters of healthy aging before clinical translation. Drug repurposing can be an important initial step in bringing the pharmacology of aging to the clinic. *Drosophila* enables rapid preclinical proof-of-concept experiments in this context. These studies will shape the future of the field, potentially paving the way for rejuvenating antiaging drug discovery.

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References

- 1. Rae MJ, Butler RN, Campisi J, De Grey ADNJ, Finch CE, Gough M, Martin GM, et al. Logan BJ. The demographic and biomedical case for late-life interventions in aging. Sci Transl Med. 2010; 2 (40) 40cm21 doi: 10.1126/scitranslmed.3000822 [PubMed: 20630854]
- 2. Inouye SK, Studenski S, Tinetti ME, Kuchel GA. Geriatric syndromes: Clinical, research, and policy implications of a core geriatric concept. J Am Geriatr Soc. 2007; 55 (5) 780–791. DOI: 10.1111/j.1532-5415.2007.01156.x [PubMed: 17493201]
- 3. Niccoli T, Partridge L. Ageing as a risk factor for disease. Curr Biol. 2012; 22 (17) R741–R752. [PubMed: 22975005]
- 4. Kontis V, Bennett JE, Mathers CD, Li G, Foreman K, Ezzati M. Future life expectancy in 35 industrialised countries: Projections with a Bayesian model ensemble. Lancet. 2017; 389 (10076) 1323–1335. DOI: 10.1016/S0140-6736(16)32381-9 [PubMed: 28236464]

5. Ferrari AJ, Santomauro DF, Aali A, Abate YH, Abbafati C, Abbastabar H, et al. GBD 2021 Diseases and Injuries Collaborators. Global incidence, prevalence, years lived with disability (YLDs), disability-adjusted life-years (DALYs), and healthy life expectancy (HALE) for 371 diseases and injuries in 204 countries and territories and 811 subnational locations, 1990–2021: A systema. Lancet. 2024; 403 (10440) 2133–2161. DOI: 10.1016/S0140-6736(24)00757-8 [PubMed: 38642570]

- 6. European Commission Directorate-General for Economic and Financial Affairs. The 2018 ageing report Economic & budgetary projections for the 28 EU Member States (2016-2070). 2018.
- 7. Scott AJ, Ellison M, Sinclair DA. The economic value of targeting aging. Nat Aging. 2021; 1 (7) 616–623. DOI: 10.1038/s43587-021-00080-0 [PubMed: 37117804]
- 8. Partridge L, Deelen J, Slagboom PE. Facing up to the global challenges of ageing. Nature. 2018; 561 (7721) 45–56. [PubMed: 30185958]
- Everitt A, Hilmer SN, Brand-Miller JC, Jamieson HA, Truswell AS, Sharma AP, et al. Le Couteur DG. Dietary approaches that delay age-related diseases. Clin Interv Aging. 2006; 1 (1) 11–31. DOI: 10.2147/ciia.2006.1.1.11 [PubMed: 18047254]
- Burns DM. Cigarette smoking among the elderly: Disease consequences and the benefits of cessation. Am J Heal Promot. 2000; 14 (6) 357–361. [PubMed: 11067570]
- 11. Stewart J, Manmathan G, Wilkinson P. Primary prevention of cardiovascular disease: A review of contemporary guidance and literature. JRSM Cardiovasc Dis. 2017; 6 204800401668721 doi: 10.1177/2048004016687211 [PubMed: 28286646]
- 12. Wang H, Abbas KM, Abbasifard M, Abbasi-Kangevari M, Abbastabar H, Abd-Allah F, et al. GBD 2019 Demographics Collaborators. Global age-sex-specific fertility, mortality, healthy life expectancy (HALE), and population estimates in 204 countries and territories, 1950–2019: A comprehensive demographic analysis for the Global Burden of Disease Study 2019. Lancet. 2020; 396 (10258) 1160–1203. DOI: 10.1016/S0140-6736(20)30977-6 [PubMed: 33069325]
- 13. Keeble E, Roberts HC, Williams CD, Van Oppen J, Conroy SP. Outcomes of hospital admissions among frail older people: A 2-year cohort study. Br J Gen Pract. 2019; 69 (685) E555–E560. DOI: 10.3399/bjgp19X704621 [PubMed: 31308000]
- 14. Galet C, Zhou Y, Eyck PT, Romanowski KS. Fall injuries, associated deaths, and 30-day readmission for subsequent falls are increasing in the elderly us population: A query of the WHO mortality database and national readmission database from 2010 to 2014. Clin Epidemiol. 2018; 10: 1627–1637. DOI: 10.2147/CLEP.S181138 [PubMed: 30519111]
- 15. Fontana L, Partridge L, Longo VD. Extending healthy life span-from yeast to humans. Science. 2010; 328 (5976) 321–326. DOI: 10.1126/science.1172539 [PubMed: 20395504]
- Partridge L, Fuentealba M, Kennedy BK. The quest to slow ageing through drug discovery. Nat Rev Drug Discov. 2020; 19 (8) 513–532. [PubMed: 32467649]
- 17. Hayflick L. How and why we age. Exp Gerontol. 1998; 33 (7–8) 639–653. [PubMed: 9951612]
- Zainabadi K. A brief history of modern aging research. Exp Gerontol. 2018; 104: 35–42. [PubMed: 29355705]
- 19. Lane MA, Ingram DK, Roth GS. 2-Deoxy-D-glucose feeding in rats mimics physiologic effects of calorie restriction. J Anti Aging Med. 1998; 1 (4) 327–337. DOI: 10.1089/rej.1.1998.1.327
- 20. Castillo-Quan JI, Kinghorn KJ, Bjedov I. Genetics and pharmacology of longevity. Advances in Genetics. 2015; 90: 1–101. [PubMed: 26296933]
- 21. Catterson JH, Khericha M, Dyson MC, Vincent AJ, Callard R, Haveron SM, et al. Partridge L. Short-term, intermittent fasting induces long-lasting gut health and TOR-independent lifespan extension. Curr Biol. 2018; 28 (11) 1714–1724. e4 doi: 10.1016/j.cub.2018.04.015 [PubMed: 29779873]
- 22. López-Otín C, Galluzzi L, Freije JMP, Madeo F, Kroemer G. Metabolic control of longevity. Cell. 2016; 166 (4) 802–821. [PubMed: 27518560]
- 23. López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. Cell. 2013; 153 (6) 1194–1217. DOI: 10.1016/j.cell.2013.05.039 [PubMed: 23746838]
- 24. Kennedy BK, Berger SL, Brunet A, Campisi J, Cuervo AM, Epel ES, et al. Sierra F. Geroscience: Linking aging to chronic disease. Cell. 2014; 159 (4) 709–713. DOI: 10.1016/j.cell.2014.10.039 [PubMed: 25417146]

25. López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. Hallmarks of aging: An expanding universe. Cell. 2023; 186 (2) 243–278. [PubMed: 36599349]

- De Grey ADNJ, Ames BN, Andersen JK, Bartke A, Campisi J, Heward CB, et al. Stock G. Time to talk SENS: Critiquing the immutability of human aging. Ann N Y Acad Sci. 2002; 959 (1) 452–462. [PubMed: 11976218]
- 27. Amor C, Feucht J, Leibold J, Ho Y-J, Zhu C, Alonso-Curbelo D, et al. Lowe SW. Senolytic CAR T cells reverse senescence-associated pathologies. Nature. 2020; 583 (7814) 127–132. DOI: 10.1038/s41586-020-2403-9 [PubMed: 32555459]
- 28. Johmura Y, Yamanaka T, Omori S, Wang T-W, Sugiura Y, Matsumoto M, et al. Nakanishi M. Senolysis by glutaminolysis inhibition ameliorates various age-associated disorders. Science. 2021; 371 (6526) 265–270. [PubMed: 33446552]
- 29. Xu M, Pirtskhalava T, Farr JN, Weigand BM, Palmer AK, Weivoda MM, et al. Kirkland JL. Senolytics improve physical function and increase lifespan in old age. Nat Med. 2018; 24 (8) 1246–1256. DOI: 10.1038/s41591-018-0092-9 [PubMed: 29988130]
- 30. Mills KF, Yoshida S, Stein LR, Grozio A, Kubota S, Sasaki Y, et al. Imai SI. Long-term administration of nicotinamide mononucleotide mitigates age-associated physiological decline in mice. Cell Metab. 2016; 24 (6) 795–806. DOI: 10.1016/j.cmet.2016.09.013 [PubMed: 28068222]
- 31. Yousefzadeh MJ, Zhu Y, McGowan SJ, Angelini L, Fuhrmann-Stroissnigg H, Xu M, et al. Niedernhofer LJ. Fisetin is a senotherapeutic that extends health and lifespan. EBioMedicine. 2018; 36: 18–28. DOI: 10.1016/j.ebiom.2018.09.015 [PubMed: 30279143]
- 32. Mohs RC, Greig NH. Drug discovery and development: Role of basic biological research.

 Alzheimer's Dement Transl Res Clin Interv. 2017; 3 (4) 651–657. DOI: 10.1016/j.trci.2017.10.005
 [PubMed: 29255791]
- 33. Nadon NL, Strong R, Miller RA, Nelson J, Javors M, Sharp ZD, et al. Harrison DE. Design of aging intervention studies: The NIA interventions testing program. Age (Dordr). 2008; 30 (4) 187–199. DOI: 10.1007/s11357-008-9048-1 [PubMed: 19424842]
- 34. Miller RA, Harrison DE, Astle CM, Baur JA, Boyd AR, de Cabo R, et al. Strong R. Rapamycin, but not resveratrol or simvastatin, extends life span of genetically heterogeneous mice. Gerontol Ser A. 2011; 66A (2) 191–201. DOI: 10.1093/gerona/glq178 [PubMed: 20974732]
- 35. de Magalhães JP, Stevens M, Thornton D. The business of anti-aging science. Trends Biotechnol. 2017; 35 (11) 1062–1073. [PubMed: 28778607]
- 36. de Magalhães JP. Longevity pharmacology comes of age. Drug Discov Today. 2021; 26 (7) 1559–1562. [PubMed: 33617794]
- 37. Fernández-Hernández I, Scheenaard E, Pollarolo G, Gonzalez C. The translational relevance of *Drosophila* in drug discovery. EMBO Rep. 2016; 17 (4) 471–472. DOI: 10.15252/embr.201642080 [PubMed: 26882560]
- Papanikolopoulou K, Mudher A, Skoulakis E. An assessment of the translational relevance of Drosophila in drug discovery. Expert Opin Drug Discov. 2019; 14 (3) 303–313. [PubMed: 30664368]
- 39. Ziehm M, Kaur S, Ivanov DK, Ballester PJ, Marcus D, Partridge L, Thornton JM. Drug repurposing for aging research using model organisms. Aging Cell. 2017; 16 (5) 1006–1015. DOI: 10.1111/acel.12626 [PubMed: 28620943]
- 40. Piper MDW, Partridge L. *Drosophila* as a model for ageing. Biochim Biophys Acta Mol Basis Dis. 2018; 1864 (9) 2707–2717. [PubMed: 28964875]
- 41. Tissenbaum HA, Guarente L. Model organisms as a guide to mammalian aging. Dev Cell. 2002; 2 (1) 9–19. [PubMed: 11782310]
- 42. Olsen A, Vantipalli MC, Lithgow GJ. Using *Caenorhabditis elegans* as a model for aging and age-related diseases. Ann N Y Acad Sci. 2006; 1067 (1) 120–128. [PubMed: 16803977]
- 43. Massie HR, Williams TR. Increased longevity of *Drosophila melanogaster* with lactic and gluconic acids. Exp Gerontol. 1979; 14 (3) 109–115. [PubMed: 110607]
- 44. Kang H-L, Benzer S, Min K-T. Life extension in *Drosophila* by feeding a drug. Proc Natl Acad Sci U S A. 2002; 99 (2) 838–843. DOI: 10.1073/pnas.022631999 [PubMed: 11792861]

45. Su Y, Wang T, Wu N, Li D, Fan X, Xu Z, Mishra SK, Yang M. Alpha-ketoglutarate extends *Drosophila* lifespan by inhibiting mTOR and activating AMPK. Aging (Albany NY). 2019; 11 (12) 4183–4197. DOI: 10.18632/aging.102045 [PubMed: 31242135]

- 46. Tao D, Lu J, Sun H, Zhao Y-M, Yuan Z-G, Li X-X, Huang B-Q. Trichostatin A extends the lifespan of *Drosophila melanogaster* by elevating hsp22 expression. Acta Biochim Biophys Sin (Shanghai). 2004; 36 (9) 618–622. [PubMed: 15346199]
- 47. Zhao Y, Sun H, Lu J, Li X, Chen X, Tao D, et al. Huang B. Lifespan extension and elevated *hsp* gene expression in *Drosophila* caused by histone deacetylase inhibitors. J Exp Biol. 2005; 208 (4) 697–705. [PubMed: 15695762]
- 48. Mason JS, Wileman T, Chapman T. Lifespan extension without fertility reduction following dietary addition of the autophagy activator Torin1 in *Drosophila melanogaster*. PLoS One. 2018; 13 (1) e0190105 doi: 10.1371/journal.pone.0190105 [PubMed: 29329306]
- 49. Fan X, Zeng Y, Fan Z, Cui L, Song W, Wu Q, et al. Yang M. Dihydromyricetin promotes longevity and activates the transcription factors FOXO and AOP in *Drosophila*. Aging (Albany NY). 2021; 13 (1) 460–476. DOI: 10.18632/aging.202156 [PubMed: 33291074]
- 50. Barardo D, Thornton D, Thoppil H, Walsh M, Sharifi S, Ferreira S, et al. de Magalhães JP. The DrugAge database of aging-related drugs. Aging Cell. 2017; 16 (3) 594–597. DOI: 10.1111/acel.12585 [PubMed: 28299908]
- 51. Lombard DB, Kohler WJ, Guo AH, Gendron C, Han M, Ding W, et al. Miller RA. High-throughput small molecule screening reveals Nrf2-dependent and -independent pathways of cellular stress resistance. Sci Adv. 2020; 6 (40) eaaz7628 doi: 10.1126/sciadv.aaz7628 [PubMed: 33008901]
- Chen Z, Cordero J, Alqarni AM, Slack C, Zeidler MP, Bellantuono I. Zoledronate extends healthspan and survival via the mevalonate pathway in a FOXO-dependent manner. Gerontol Ser A. 2021; 77 (8) 1494–1502. DOI: 10.1093/gerona/glab172 [PubMed: 34137822]
- 53. Klawitter J, Nashan B, Christians U. Everolimus and sirolimus in transplantation-related but different. Expert Opin Drug Saf. 2015; 14 (7) 1055–1070. DOI: 10.1517/14740338.2015.1040388 [PubMed: 25912929]
- 54. Liu GY, Sabatini DM. mTOR at the nexus of nutrition, growth, ageing and disease. Nat Rev Mol Cell Biol. 2020; 21 (4) 183–203. DOI: 10.1038/s41580-019-0199-y [PubMed: 31937935]
- 55. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, et al. Miller RA. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. Nature. 2009; 460 (7253) 392–395. DOI: 10.1038/nature08221 [PubMed: 19587680]
- 56. Bjedov I, Toivonen JM, Kerr F, Slack C, Jacobson J, Foley A, Partridge L. Mechanisms of life span extension by rapamycin in the fruit fly *Drosophila melanogaster*. Cell Metab. 2010; 11 (1) 35–46. DOI: 10.1016/j.cmet.2009.11.010 [PubMed: 20074526]
- 57. Robida-Stubbs S, Glover-Cutter K, Lamming DW, Mizunuma M, Narasimhan SD, Neumann-Haefelin E, et al. Blackwell TK. TOR signaling and rapamycin influence longevity by regulating SKN-1/Nrf and DAF-16/FoxO. Cell Metab. 2012; 15 (5) 713–724. DOI: 10.1016/j.cmet.2012.04.007 [PubMed: 22560223]
- 58. Filer D, Thompson MA, Takhaveev V, Dobson AJ, Kotronaki I, Green JWM, et al. Alic N. RNA polymerase III limits longevity downstream of TORC1. Nature. 2017; 552 (7684) 263–267. DOI: 10.1038/nature25007 [PubMed: 29186112]
- 59. Chen C, Liu Y, Liu Y, Zheng P. mTOR regulation and therapeutic rejuvenation of aging hematopoietic stem cells. Sci Signal. 2009; 2 (98) ra75 doi: 10.1126/scisignal.2000559 [PubMed: 19934433]
- 60. Bitto A, Ito TK, Pineda V, LeTexier NJ, Huang HZ, Sutlief E, et al. Kaeberlein M. Transient rapamycin treatment can increase lifespan and healthspan in middle-aged mice. eLife. 2016; 5 e16351 doi: 10.7554/eLife.16351 [PubMed: 27549339]
- 61. An JY, Kerns KA, Ouellette A, Robinson L, Morris HD, Kaczorowski C, et al. Kaeberlein M. Rapamycin rejuvenates oral health in aging mice. eLife. 2020; 9 e54318 doi: 10.7554/eLife.54318 [PubMed: 32342860]

62. Wilkinson JE, Burmeister L, Brooks S, Chan C-C, Friedline S, Harrison DE, et al. Miller RA. Rapamycin slows aging in mice. Aging Cell. 2012; 11 (4) 675–682. DOI: 10.1111/j.1474-9726.2012.00832.x [PubMed: 22587563]

- 63. Regan JC, Lu Y-X, Ureña E, Meilenbrock RL, Catterson JH, Kißler D, et al. Partridge L. Sexual identity of enterocytes regulates autophagy to determine intestinal health, lifespan and responses to rapamycin. Nat Aging. 2022; 2 (12) 1145–1158. DOI: 10.1038/s43587-022-00308-7 [PubMed: 37118538]
- 64. Spindler SR, Li R, Dhahbi JM, Yamakawa A, Sauer F. Novel protein kinase signaling systems regulating lifespan identified by small molecule library screening using *Drosophila*. PLoS One. 2012; 7 (2) e29782 doi: 10.1371/journal.pone.0029782 [PubMed: 22363408]
- 65. Aiello G, Sabino C, Pernici D, Audano M, Antonica F, Gianesello M, et al. Tiberi L. Transient rapamycin treatment during developmental stage extends lifespan in *Mus musculus* and *Drosophila melanogaster*. EMBO Rep. 2022; 23 (9) e55299 doi: 10.15252/embr.202255299 [PubMed: 35796299]
- 66. Juricic P, Lu Y-X, Leech T, Drews LF, Paulitz J, Lu J, et al. Partridge L. Long-lasting geroprotection from brief rapamycin treatment in early adulthood by persistently increased intestinal autophagy. Nat Aging. 2022; 2 (9) 824–836. DOI: 10.1038/s43587-022-00278-w [PubMed: 37118497]
- 67. Fan X, Liang Q, Lian T, Wu Q, Gaur U, Li D, et al. Yang M. Rapamycin preserves gut homeostasis during *Drosophila* aging. Oncotarget. 2015; 6 (34) 35274–35283. DOI: 10.18632/oncotarget.5895 [PubMed: 26431326]
- 68. Partridge L, Alic N, Bjedov I, Piper MDW. Ageing in *Drosophila*: The role of the insulin/Igf and TOR signalling network. Exp Gerontol. 2011; 46 (5) 376–381. DOI: 10.1016/j.exger.2010.09.003 [PubMed: 20849947]
- 69. Tain LS, Mortiboys H, Tao RN, Ziviani E, Bandmann O, Whitworth AJ. Rapamycin activation of 4E-BP prevents Parkinsonian dopaminergic neuron loss. Nat Neurosci. 2009; 12 (9) 1129–1135. DOI: 10.1038/nn.2372 [PubMed: 19684592]
- Kapahi P, Zid BM, Harper T, Koslover D, Sapin V, Benzer S. Regulation of lifespan in *Drosophila* by modulation of genes in the TOR signaling pathway. Curr Biol. 2004; 14 (10) 885–890. DOI: 10.1016/j.cub.2004.03.059 [PubMed: 15186745]
- 71. Lu Y-X, Regan JC, Eßer J, Drews LF, Weinseis T, Stinn J, et al. Partridge L. A TORC1-histone axis regulates chromatin organisation and non-canonical induction of autophagy to ameliorate ageing. eLife. 2021; 10 e62233 doi: 10.7554/eLife.62233 [PubMed: 33988501]
- 72. Ureña E, Xu B, Regan J, Atilano ML, Minkley LJ, Filer D, et al. Partridge L. Trametinib ameliorates aging-associated gut pathology in *Drosophila* females by reducing Pol III activity in intestinal stem cells. Proc Natl Acad Sci U S A. 2024; 121 (4) e2311313121 doi: 10.1073/pnas.2311313121 [PubMed: 38241436]
- 73. Zanco B, Mirth CK, Sgrò CM, Piper MDW. A dietary sterol trade-off determines lifespan responses to dietary restriction in *Drosophila melanogaster* females. eLife. 2021; 10 e62335 doi: 10.7554/eLife.62335 [PubMed: 33494859]
- 74. Schinaman JM, Rana A, Ja WW, Clark RI, Walker DW. Rapamycin modulates tissue aging and lifespan independently of the gut microbiota in *Drosophila*. Sci Rep. 2019; 9 (1) 7824 doi: 10.1038/s41598-019-44106-5 [PubMed: 31127145]
- 75. Bolukbasi E, Khericha M, Regan JC, Ivanov DK, Adcott J, Dyson MC, et al. Partridge L. Intestinal fork head regulates nutrient absorption and promotes longevity. Cell Rep. 2017; 21 (3) 641–653. DOI: 10.1016/j.celrep.2017.09.042 [PubMed: 29045833]
- 76. Blagosklonny MV. Fasting and rapamycin: Diabetes versus benevolent glucose intolerance. Cell Death Dis. 2019; 10 (8) 607. doi: 10.1038/s41419-019-1822-8 [PubMed: 31406105]
- 77. Lamming DW, Ye L, Katajisto P, Goncalves MD, Saitoh M, Stevens DM, et al. Baur JA. Rapamycin-induced insulin resistance is mediated by mTORC2 loss and uncoupled from longevity. Science. 2012; 335 (6076) 1638–1643. DOI: 10.1126/science.1215135 [PubMed: 22461615]

78. van Dam E, van Leeuwen LAG, dos Santos E, James J, Best L, Lennicke C, et al. Cochemé HM. Sugar-induced obesity and insulin resistance are uncoupled from shortened survival in *Drosophila*. Cell Metab. 2020; 31 (4) 710–725. e7 doi: 10.1016/j.cmet.2020.02.016 [PubMed: 32197072]

- Mannick JB, Morris M, Hockey H-UPU, Roma G, Beibel M, Kulmatycki K, et al. Klickstein LB. TORC1 inhibition enhances immune function and reduces infections in the elderly. Sci Transl Med. 2018; 10 (449) eaaq1564 [PubMed: 29997249]
- 80. Mannick JB, Del Giudice G, Lattanzi M, Valiante NM, Praestgaard J, Huang B, et al. Klickstein LB. mTOR inhibition improves immune function in the elderly. Sci Transl Med. 2014; 6 (268) 268ra179 [PubMed: 25540326]
- 81. Mannick JB, Teo G, Bernardo P, Quinn D, Russell K, Klickstein L, et al. Shergill S. Targeting the biology of ageing with mTOR inhibitors to improve immune function in older adults: Phase 2b and phase 3 randomised trials. Lancet Heal Longev. 2021; 2 (5) e250–e262. DOI: 10.1016/S2666-7568(21)00062-3 [PubMed: 33977284]
- 82. Li X, Frye MA, Shelton RC. Review of pharmacological treatment in mood disorders and future directions for drug development. Neuropsychopharmacology. 2012; 37 (1) 77–101. DOI: 10.1038/npp.2011.198 [PubMed: 21900884]
- 83. Araldi E, Jutzeler CR, Ristow M. Lithium treatment extends human lifespan: Findings from the UK Biobank. Aging (Albany NY). 2023; 15 (2) 421–440. DOI: 10.18632/aging.204476 [PubMed: 36640269]
- 84. Matsagas K, Lim DB, Horwitz M, Rizza CL, Mueller LD, Villeponteau B, Rose MR. Long-term functional side-effects of stimulants and sedatives in *Drosophila melanogaster*. PLoS One. 2009; 4 (8) e6578 doi: 10.1371/journal.pone.0006578 [PubMed: 19668379]
- 85. Zarse K, Terao T, Tian J, Iwata N, Ishii N, Ristow M. Low-dose lithium uptake promotes longevity in humans and metazoans. Eur J Nutr. 2011; 50 (5) 387–389. DOI: 10.1007/s00394-011-0171-x [PubMed: 21301855]
- 86. Castillo-Quan JI, Li L, Kinghorn KJ, Ivanov DK, Tain LS, Slack C, et al. Partridge L. Lithium promotes longevity through GSK3/NRF2-dependent hormesis. Cell Rep. 2016; 15 (3) 638–650. DOI: 10.1016/j.celrep.2016.03.041 [PubMed: 27068460]
- 87. McColl G, Killilea DW, Hubbard AE, Vantipalli MC, Melov S, Lithgow GJ. Pharmacogenetic analysis of lithium-induced delayed aging in *Caenorhabditis elegans*. J Biol Chem. 2008; 283 (1) 350–357. DOI: 10.1074/jbc.M705028200 [PubMed: 17959600]
- 88. Roux M, Dosseto A. From direct to indirect lithium targets: A comprehensive review of omics data. Metallomics. 2017; 9 (10) 1326–1351. [PubMed: 28885630]
- 89. Zhu F, Li Q, Zhang F, Sun X, Cai G, Zhang W, Chen X. Chronic lithium treatment diminishes the female advantage in lifespan in *Drosophila melanogaster*. Clin Exp Pharmacol Physiol. 2015; 42 (6) 617–621. [PubMed: 25810251]
- 90. Takahashi-Yanaga F. Activator or inhibitor? GSK-3 as a new drug target. Biochem Pharmacol. 2013; 86 (2) 191–199. [PubMed: 23643839]
- 91. Eldar-Finkelman H, Martinez A. GSK-3 inhibitors: Preclinical and clinical focus on CNS. Front Mol Neurosci. 2011; 4: 32. doi: 10.3389/fnmol.2011.00032 [PubMed: 22065134]
- 92. Gitlin M. Lithium side effects and toxicity: Prevalence and management strategies. Int J Bipolar Disord. 2016; 4 (1) 27. doi: 10.1186/s40345-016-0068-y [PubMed: 27900734]
- 93. Kakadia S, Yarlagadda N, Awad R, Kundranda M, Niu J, Naraev B, et al. Mahmoud F. Mechanisms of resistance to BRAF and MEK inhibitors and clinical update of US Food and Drug Administration-approved targeted therapy in advanced melanoma. Onco Targets Ther. 2018; 11: 7095–7107. DOI: 10.2147/OTT.S182721 [PubMed: 30410366]
- 94. Slack C, Alic N, Foley A, Cabecinha M, Hoddinott MP, Partridge L. The Ras-Erk-ETS-signaling pathway is a drug target for longevity. Cell. 2015; 162 (1) 72–83. DOI: 10.1016/j.cell.2015.06.023 [PubMed: 26119340]
- 95. Miller PB, Obrik-Uloho OT, Phan MH, Medrano CL, Renier JS, Thayer JL, et al. Bloch Qazi MC. The song of the old mother: Reproductive senescence in female *Drosophila*. Fly (Austin). 2014; 8 (3) 127–139. DOI: 10.4161/19336934.2014.969144 [PubMed: 25523082]
- 96. Ureña E, Xu B, Regan JC, Atilano ML, Minkley LJ, Filer D, et al. Partridge L. Trametinib ameliorates aging-associated gut pathology in *Drosophila* females by reducing Pol III activity

- in intestinal stem cells. Proc Natl Acad Sci U S A. 2024; 121 (4) e2311313121 doi: 10.1073/pnas.2311313121 [PubMed: 38241436]
- 97. Castillo-Quan JI, Tain LS, Kinghorn KJ, Li L, Grönke S, Hinze Y, Partridge L. A triple drug combination targeting components of the nutrient-sensing network maximizes longevity. Proc Natl Acad Sci U S A. 2019; 116 (42) 20817–20819. DOI: 10.1073/pnas.1913212116 [PubMed: 31570569]
- 98. Lan J, Rollins JA, Zang X, Wu D, Zou L, Wang Z, et al. Chen D. Translational regulation of non-autonomous mitochondrial stress response promotes longevity. Cell Rep. 2019; 28 (4) 1050–1062. e6 doi: 10.1016/j.celrep.2019.06.078 [PubMed: 31340143]
- Banks AS, McAllister FE, Camporez JPG, Zushin P-JH, Jurczak MJ, Laznik-Bogoslavski D, et al. Spiegelman BM. An ERK/Cdk5 axis controls the diabetogenic actions of PPARγ. Nature. 2015; 517 (7534) 391–395. DOI: 10.1038/nature13887 [PubMed: 25409143]
- 100. Pryor R, Cabreiro F. Repurposing metformin: An Old drug with new tricks in its binding pockets. Biochem J. 2015; 471 (3) 307–322. DOI: 10.1042/BJ20150497 [PubMed: 26475449]
- 101. Martin-Montalvo A, Mercken EM, Mitchell SJ, Palacios HH, Mote PL, Scheibye-Knudsen M, et al. de Cabo R. Metformin improves healthspan and lifespan in mice. Nat Commun. 2013; 4 (1) 2192 doi: 10.1038/ncomms3192 [PubMed: 23900241]
- 102. Alfaras I, Mitchell SJ, Mora H, Lugo DR, Warren A, Navas-Enamorado I, et al. de Cabo R. Health benefits of late-onset metformin treatment every other week in mice. NPJ Aging Mech Dis. 2017; 3: 16. doi: 10.1038/s41514-017-0018-7 [PubMed: 29167747]
- 103. Barzilai N, Crandall JP, Kritchevsky SB, Espeland MA. Metformin as a tool to target aging. Cell Metab. 2016; 23 (6) 1060–1065. DOI: 10.1016/j.cmet.2016.05.011 [PubMed: 27304507]
- 104. Clark RI, Walker DW. Role of gut microbiota in aging-related health decline: Insights from invertebrate models. Cell Mol Life Sci. 2018; 75 (1) 93–101. DOI: 10.1007/s00018-017-2671-1 [PubMed: 29026921]
- 105. Pryor R, Martinez-Martinez D, Quintaneiro L, Cabreiro F. The role of the microbiome in drug response. Annu Rev Pharmacol Toxicol. 2020; 60 (1) 417–435. [PubMed: 31386593]
- 106. Slack C, Foley A, Partridge L. Activation of AMPK by the putative dietary restriction mimetic metformin is insufficient to extend lifespan in *Drosophila*. PLoS One. 2012; 7 (10) e47699 doi: 10.1371/journal.pone.0047699 [PubMed: 23077661]
- 107. Pryor R, Norvaisas P, Marinos G, Best L, Thingholm LB, Quintaneiro LM, et al. Cabreiro F. Host-microbe-drug-nutrient screen identifies bacterial effectors of metformin therapy. Cell. 2019; 178 (6) 1299–1312. e29 doi: 10.1016/j.cell.2019.08.003 [PubMed: 31474368]
- 108. Cabreiro F, Au C, Leung K-Y, Vergara-Irigaray N, Cochemé HM, Noori T, et al. Gems D. Metformin retards aging in *C. elegans* by altering microbial folate and methionine metabolism. Cell. 2013; 153 (1) 228–239. DOI: 10.1016/j.cell.2013.02.035 [PubMed: 23540700]
- 109. Suzuta S, Nishida H, Ozaki M, Kohno N, Le TD, Inoue YH. Metformin suppresses progression of muscle aging via activation of the AMP kinase-mediated pathways in *Drosophila* adults. Eur Rev Med Pharmacol Sci. 2022; 26 (21) 8039–8056. [PubMed: 36394755]
- 110. Norvaisas P, Cabreiro F. Pharmacology in the age of the holobiont. Curr Opin Syst Biol. 2018; 10: 34–42. DOI: 10.1016/j.coisb.2018.05.006
- 111. Wongrakpanich S, Wongrakpanich A, Melhado K, Rangaswami J. A comprehensive review of non-steroidal anti-inflammatory drug use in the elderly. Aging Dis. 2018; 9 (1) 143. doi: 10.14336/AD.2017.0306 [PubMed: 29392089]
- 112. Danilov A, Shaposhnikov M, Shevchenko O, Zemskaya N, Zhavoronkov A, Moskalev A. Influence of non-steroidal anti-inflammatory drugs on *Drosophila melanogaster* longevity. Oncotarget. 2015; 6 (23) 19428–19444. DOI: 10.18632/oncotarget.5118 [PubMed: 26305987]
- 113. Song C, Zhu C, Wu Q, Qi J, Gao Y, Zhang Z, et al. Yang M. Metabolome analysis of effect of aspirin on *Drosophila* lifespan extension. Exp Gerontol. 2017; 95: 54–62. [PubMed: 28457986]
- 114. He C, Tsuchiyama SK, Nguyen QT, Plyusnina EN, Terrill SR, Sahibzada S, et al. Polymenis M. Enhanced longevity by ibuprofen, conserved in multiple species, occurs in yeast through inhibition of tryptophan import. PLoS Genet. 2014; 10 (12) e1004860 doi: 10.1371/journal.pgen.1004860 [PubMed: 25521617]

115. Proshkina E, Lashmanova E, Dobrovolskaya E, Zemskaya N, Kudryavtseva A, Shaposhnikov M, Moskalev A. Geroprotective and radioprotective activity of quercetin, (-)-epicatechin, and ibuprofen in *Drosophila melanogaster*. Front Pharmacol. 2016; 7: 505. doi: 10.3389/fphar.2016.00505 [PubMed: 28066251]

- 116. Wu Q, Lian T, Fan X, Song C, Gaur U, Mao X, et al. Yang M. 2, 5-Dimethyl-celecoxib extends *Drosophila* life span via a mechanism that requires insulin and target of rapamycin signaling. Gerontol Ser A Biol Sci Med Sci. 2016; 72 (10) glw244 [PubMed: 28025308]
- 117. Strong R, Miller RA, Astle CM, Floyd RA, Flurkey K, Hensley KL, et al. Harrison DE. Nordihydroguaiaretic acid and aspirin increase lifespan of genetically heterogeneous male mice. Aging Cell. 2008; 7 (5) 641–650. DOI: 10.1111/j.1474-9726.2008.00414.x [PubMed: 18631321]
- 118. McNeil JJ, Woods RL, Nelson MR, Reid CM, Kirpach B, Wolfe R, et al. ASPREE Investigator Group. Effect of aspirin on disability-free survival in the healthy elderly. N Engl J Med. 2018; 379 (16) 1499–1508. DOI: 10.1056/NEJMoa1800722 [PubMed: 30221596]
- 119. Zhu Y, Cai Q, Zheng X, Liu L, Hua Y, Du B, et al. Ji S. Aspirin positively contributes to *Drosophila* intestinal homeostasis and delays aging through targeting Imd. Aging Dis. 2021; 12 (7) 1821. doi: 10.14336/AD.2020.1008 [PubMed: 34631223]
- 120. Hochschild R. Effect of membrane stabilizing drugs on mortality in *Drosophila melanogaster* . Exp Gerontol. 1971; 6 (2) 133–151. [PubMed: 4397875]
- 121. Spindler SR, Li R, Dhahbi JM, Yamakawa A, Mote P, Bodmer R, et al. Ablao KP. Statin treatment increases lifespan and improves cardiac health in *Drosophila* by decreasing specific protein prenylation. PLoS One. 2012; 7 (6) e39581 doi: 10.1371/journal.pone.0039581 [PubMed: 22737247]
- 122. Liu J, Wu Q, He D, Ma T, Du L, Dui W, et al. Jiao R. *Drosophila* sbo regulates lifespan through its function in the synthesis of coenzyme Q *in vivo*. J Genet Genomics. 2011; 38 (6) 225–234. [PubMed: 21703546]
- 123. Yamamoto R, Bai H, Dolezal AG, Amdam G, Tatar M. Juvenile hormone regulation of Drosophila aging. BMC Biol. 2013; 11 (1) 85. doi: 10.1186/1741-7007-11-85 [PubMed: 23866071]
- 124. Bergwitz C. Dietary phosphate modifies lifespan in *Drosophila*. Nephrol Dial Transplant. 2012; 27 (9) 3399–3406. [PubMed: 22942172]
- 125. Locatelli F, Del Vecchio L. Cardiovascular mortality in chronic kidney disease patients: Potential mechanisms and possibilities of inhibition by resin-based phosphate binders. Expert Rev Cardiovasc Ther. 2015; 13 (5) 489–499. [PubMed: 25804298]
- 126. Brack C, Bechter-Thüring E, Labuhn M. N-Acetylcysteine slows down ageing and increases the life span of *Drosophila melanogaster*. Cell Mol Life Sci. 1997; 53 (11) 960–966. DOI: 10.1007/PL00013199 [PubMed: 9447249]
- 127. Niraula P, Kim MS. N-Acetylcysteine extends lifespan of *Drosophila* via modulating ROS scavenger gene expression. Biogerontology. 2019; 20 (4) 533–543. [PubMed: 31115735]
- 128. Shaposhnikov MV, Zemskaya NV, Koval LA, Schegoleva EV, Zhavoronkov A, Moskalev AA. Effects of N-acetyl-L-cysteine on lifespan, locomotor activity and stress-resistance of 3 *Drosophila* species with different lifespans. Aging (Albany NY). 2018; 10 (9) 2428–2458. DOI: 10.18632/aging.101561 [PubMed: 30243020]
- 129. Camus MF, Rodriguez E, Kotiadis V, Carter H, Lane N. Redox stress shortens lifespan through suppression of respiratory complex I in flies with mitonuclear incompatibilities. Exp Gerontol. 2023; 175 112158 [PubMed: 36965604]
- Gusarov I, Shamovsky I, Pani B, Gautier L, Eremina S, Katkova-Zhukotskaya O, et al. Nudler E. Dietary thiols accelerate aging of *C. elegans*. Nat Commun. 2021; 12 (1) 4336 doi: 10.1038/s41467-021-24634-3 [PubMed: 34267196]
- 131. Lennicke C, Cochemé HM. Redox signalling and ageing: Insights from *Drosophila*. Biochem Soc Trans. 2020; 48 (2) 367–377. DOI: 10.1042/BST20190052 [PubMed: 32196546]
- 132. Liu D, Ahmet A, Ward L, Krishnamoorthy P, Mandelcorn ED, Leigh R, et al. Kim H. A practical guide to the monitoring and management of the complications of systemic corticosteroid therapy. Allergy, Asthma Clin Immunol. 2013; 9 (1) 30. doi: 10.1186/1710-1492-9-30 [PubMed: 23947590]

133. Bartolo G, Gonzalez LO, Alameh S, Valencia CA, Martchenko Shilman M. Identification of glucocorticoid receptor in *Drosophila melanogaster*. BMC Microbiol. 2020; 20 (1) 161. doi: 10.1186/s12866-020-01848-x [PubMed: 32539689]

- 134. Chamilos G, Lewis RE, Hu J, Xiao L, Zal T, Gilliet M, et al. Kontoyiannis DP. *Drosophila melanogaster* as a model host to dissect the immunopathogenesis of zygomycosis. Proc Natl Acad Sci U S A. 2008; 105 (27) 9367–9372. DOI: 10.1073/pnas.0709578105 [PubMed: 18583479]
- 135. Valbuena Perez JV, Linnenberger R, Dembek A, Bruscoli S, Riccardi C, Schulz MH, et al. Hoppstädter J. Altered glucocorticoid metabolism represents a feature of macroph-aging. Aging Cell. 2020; 19 (6) e13156 doi: 10.1111/acel.13156 [PubMed: 32463582]
- 136. Moffat SD, An Y, Resnick SM, Diamond MP, Ferrucci L. Longitudinal change in cortisol levels across the adult life span. Gerontol Ser A. 2020; 75 (2) 394–400. DOI: 10.1093/gerona/gly279 [PubMed: 31714574]
- 137. Roelfsema F, van Heemst D, Iranmanesh A, Takahashi P, Yang R, Veldhuis JD. Impact of age, sex and body mass index on cortisol secretion in 143 healthy adults. Endocr Connect. 2017; 6 (7) 500–509. DOI: 10.1530/EC-17-0160 [PubMed: 28760748]
- 138. Landis GN, Hilsabeck TAU, Bell HS, Ronnen-Oron T, Wang L, Doherty D, Tower J. Mifepristone increases life span of virgin female *Drosophila* on regular and high-fat diet without reducing food intake. Front Genet. 2021; 12 751647 doi: 10.3389/fgene.2021.751647 [PubMed: 34659367]
- 139. Osterwalder T, Yoon KS, White BH, Keshishian H. A conditional tissue-specific transgene expression system using inducible GAL4. Proc Natl Acad Sci U S A. 2001; 98 (22) 12596–12601. DOI: 10.1073/pnas.221303298 [PubMed: 11675495]
- 140. Scialo F, Sriram A, Stefanatos R, Sanz A. Practical recommendations for the use of the GeneSwitch Gal4 system to knock-down genes in *Drosophila melanogaster*. PLoS One. 2016; 11 (8) e0161817 doi: 10.1371/journal.pone.0161817 [PubMed: 27570965]
- 141. Robles-Murguia M, Hunt LC, Finkelstein D, Fan Y, Demontis F. Tissue-specific alteration of gene expression and function by RU486 and the GeneSwitch system. NPJ Aging Mech Dis. 2019; 5: 6. doi: 10.1038/s41514-019-0036-8 [PubMed: 31123597]
- 142. Yamada R, Deshpande SA, Keebaugh ES, Ehrlich MR, Soto Obando A, Ja WW. Mifepristone reduces food palatability and affects *Drosophila* feeding and lifespan. J Gerontol Ser A Biol Sci Med Sci. 2017; 72 (2) 173–180. DOI: 10.1093/gerona/glw072 [PubMed: 27093874]
- 143. Landis GN, Salomon MP, Keroles D, Brookes N, Sekimura T, Tower J. The progesterone antagonist mifepristone/RU486 blocks the negative effect on life span caused by mating in female *Drosophila*. Aging (Albany NY). 2015; 7 (1) 53–69. DOI: 10.18632/aging.100721 [PubMed: 25614682]
- 144. Tower J, Landis GN, Shen J, Choi R, Fan Y, Lee D, Song J. Mifepristone/RU486 acts in *Drosophila melanogaster* females to counteract the life span-shortening and proinflammatory effects of male sex peptide. Biogerontology. 2017; 18 (3) 413–427. DOI: 10.1007/s10522-017-9703-y [PubMed: 28451923]
- 145. Landis GN, Doherty D, Yen C-A, Wang L, Fan Y, Wang I, et al. Tower J. Metabolic signatures of life span regulated by mating, sex peptide, and mifepristone/RU486 in female *Drosophila melanogaster*. J Gerontol A Biol Sci Med Sci. 2021; 76 (2) 195–204. DOI: 10.1093/gerona/glaa164 [PubMed: 32648907]
- 146. Rothstein DM. Rifamycins, alone and in combination. Cold Spring Harb Perspect Med. 2016; 6 (7) a027011 doi: 10.1101/cshperspect.a027011 [PubMed: 27270559]
- 147. Admasu TD, Chaithanya Batchu K, Barardo D, Ng LF, Lam VYM, Xiao L, et al. Gruber J. Drug synergy slows aging and improves healthspan through IGF and SREBP lipid signaling. Dev Cell. 2018; 47 (1) 67–79. e5 [PubMed: 30269951]
- 148. Yamashita K, Oi A, Kosakamoto H, Yamauchi T, Kadoguchi H, Kuraishi T, et al. Obata F. Activation of innate immune during development induces unresolved dysbiotic inflammatory gut and shortens lifespan. Dis Model Mech. 2021; 14 (9) 049103 doi: 10.1242/dmm.049103 [PubMed: 34448472]
- 149. Garrido-Mesa N, Zarzuelo A, Gálvez J. Minocycline: Far beyond an antibiotic. Br J Pharmacol. 2013; 169 (2) 337–352. DOI: 10.1111/bph.12139 [PubMed: 23441623]

150. Bonilla E, Contreras R, Medina-Leendertz S, Mora M, Villalobos V, Bravo Y. Minocycline increases the life span and motor activity and decreases lipid peroxidation in manganese treated *Drosophila melanogaster*. Toxicology. 2012; 294 (1) 50–53. [PubMed: 22330257]

- 151. Lee GJ, Lim JJ, Hyun S. Minocycline treatment increases resistance to oxidative stress and extends lifespan in *Drosophila* via FOXO. Oncotarget. 2017; 8 (50) 87878–87890. DOI: 10.18632/oncotarget.21224 [PubMed: 29152127]
- 152. Oxenkrug G, Navrotskaya V, Vorobyova L, Summergrad P. Minocycline effect on life and health span of *Drosophila melanogaster*. Aging Dis. 2012; 3 (3) 352–359. [PubMed: 23185716]
- 153. Mora M, Medina-Leendertz SJ, Bonilla E, Terán RE, Paz MC, Arcaya JL. Minocycline, but not ascorbic acid, increases motor activity and extends the life span of *Drosophila melanogaster*. Invest Clin. 2013; 54 (2) 161–170. [PubMed: 23947005]
- 154. Lim JJ, Hyun S. Minocycline treatment improves proteostasis during *Drosophila* aging via autophagy mediated by FOXO and Hsp70. Biomed Pharmacother. 2022; 149 112803 [PubMed: 35286967]
- 155. Demontis F, Perrimon N. FOXO/4E-BP signaling in *Drosophila* muscles regulates organism-wide proteostasis during aging. Cell. 2010; 143 (5) 813–825. DOI: 10.1016/j.cell.2010.10.007 [PubMed: 21111239]
- 156. Giannakou ME, Goss M, Jünger MA, Hafen E, Leevers SJ, Partridge L. Long-lived *Drosophila* with overexpressed dFOXO in adult fat body. Science. 2004; 305 (5682) 361. [PubMed: 15192154]
- 157. Hwangbo DS, Gersham B, Tu M-P, Palmer M, Tatar M. *Drosophila* dFOXO controls lifespan and regulates insulin signalling in brain and fat body. Nature. 2004; 429 (6991) 562–566. [PubMed: 15175753]
- 158. Avanesian A, Khodayari B, Felgner JS, Jafari M. Lamotrigine extends lifespan but compromises health span in *Drosophila melanogaster*. Biogerontology. 2010; 11 (1) 45–52. DOI: 10.1007/s10522-009-9227-1 [PubMed: 19430925]
- 159. Dubiley TA, Rushkevich YE, Koshel NM, Voitenko VP, Vaiserman AM. Life span extension in *Drosophila melanogaster* induced by morphine. Biogerontology. 2011; 12 (3) 179–184. [PubMed: 21061062]
- 160. Fuentealba M, Dönerta HM, Williams R, Labbadia J, Thornton JM, Partridge L. Using the drug-protein interactome to identify anti-ageing compounds for humans. PLOS Comput Biol. 2019; 15 (1) e1006639 doi: 10.1371/journal.pcbi.1006639 [PubMed: 30625143]
- 161. dos Santos E, Cochemé HM. How does a fly die? Insights into ageing from the pathophysiology of *Drosophila* mortality. GeroScience. 2024; 46: 4003–4015. DOI: 10.1007/s11357-024-01158-4 [PubMed: 38642259]
- 162. de Magalhães JP. The scientific quest for lasting youth: Prospects for curing aging. Rejuvenation Res. 2014; 17 (5) 458–467. DOI: 10.1089/rej.2014.1580 [PubMed: 25132068]
- 163. de Magalhães JP. Why genes extending lifespan in model organisms have not been consistently associated with human longevity and what it means to translation research. Cell Cycle. 2014; 13 (17) 2671–2673. DOI: 10.4161/15384101.2014.950151 [PubMed: 25486354]
- 164. Calimport SRG, Bentley BL, Stewart CE, Pawelec G, Scuteri A, Vinciguerra M, et al. Church G. To help aging populations, classify organismal senescence. Science. 2019; 366 (6465) 576–578. DOI: 10.1126/science.aay7319 [PubMed: 31672885]
- 165. Katz S. Assessing self-maintenance: Activities of daily living, mobility, and instrumental activities of daily living. J Am Geriatr Soc. 1983; 31 (12) 721–727. [PubMed: 6418786]
- 166. Folstein MF, Folstein SE, McHugh PR. Mini-mental state. J Psychiatr Res. 1975; 12 (3) 189–198. [PubMed: 1202204]
- 167. Heinze-Milne SD, Banga S, Howlett SE. Frailty assessment in animal models. Gerontology. 2019; 65 (6) 610–619. [PubMed: 31330523]
- 168. Cohen AA. Aging across the tree of life: The importance of a comparative perspective for the use of animal models in aging. Biochim Biophys Acta Mol Basis Dis. 2018; 1864 (9) 2680–2689. [PubMed: 28690188]

169. Thomas R, Wang W, Su D-M. Contributions of age-related thymic involution to immunosenescence and inflammaging. Immun Ageing. 2020; 17 (1) 2. doi: 10.1186/s12979-020-0173-8 [PubMed: 31988649]

- 170. Alic N, Tullet JM, Niccoli T, Broughton S, Hoddinott MP, Slack C, et al. Partridge L. Cell-nonautonomous effects of dFOXO/DAF-16 in aging. Cell Rep. 2014; 6 (4) 608–616. DOI: 10.1016/j.celrep.2014.01.015 [PubMed: 24508462]
- 171. Stenesen D, Suh JM, Seo J, Yu K, Lee K-S, Kim J-S, et al. Graff JM. Adenosine nucleotide biosynthesis and AMPK regulate adult life span and mediate the longevity benefit of caloric restriction in flies. Cell Metab. 2013; 17 (1) 101–112. DOI: 10.1016/j.cmet.2012.12.006 [PubMed: 23312286]
- 172. Piper MDW, Blanc E, Leitão-Gonçalves R, Yang M, He X, Linford NJ, et al. Partridge L. A holidic medium for *Drosophila melanogaster*. Nat Methods. 2014; 11 (1) 100–105. DOI: 10.1038/nmeth.2731 [PubMed: 24240321]
- 173. Piper MDW, Bartke A. Diet and aging. Cell Metab. 2008; 8 (2) 99–104. [PubMed: 18680711]
- 174. Piper MDW, Soultoukis GA, Blanc E, Mesaros A, Herbert SL, Juricic P, et al. Partridge L. Matching dietary amino acid balance to the in silico-translated exome optimizes growth and reproduction without cost to lifespan. Cell Metab. 2017; 25 (3) 610–621. DOI: 10.1016/j.cmet.2017.02.005 [PubMed: 28273481]
- 175. Schulze MB, Martínez-González MA, Fung TT, Lichtenstein AH, Forouhi NG. Food based dietary patterns and chronic disease prevention. BMJ. 2018; 361 k2396 doi: 10.1136/bmj.k2396 [PubMed: 29898951]
- 176. Strong R, Miller RA, Antebi A, Astle CM, Bogue M, Denzel MS, et al. Harrison DE. Longer lifespan in male mice treated with a weakly estrogenic agonist, an antioxidant, an α-glucosidase inhibitor or a Nrf2-inducer. Aging Cell. 2016; 15 (5) 872–884. DOI: 10.1111/acel.12496 [PubMed: 27312235]
- 177. Maher RL, Hanlon J, Hajjar ER. Clinical consequences of polypharmacy in elderly. Expert Opin Drug Saf. 2014; 13 (1) 57–65. DOI: 10.1517/14740338.2013.827660 [PubMed: 24073682]
- 178. Ingram DK, Zhu M, Mamczarz J, Zou S, Lane MA, Roth GS, DeCabo R. Calorie restriction mimetics: An emerging research field. Aging Cell. 2006; 5 (2) 97–108. [PubMed: 16626389]
- 179. Demidenko ZN, Zubova SG, Bukreeva EI, Pospelov VA, Pospelova T, Blagosklonny MV. Rapamycin decelerates cellular senescence. Cell Cycle. 2009; 8 (12) 1888–1895. [PubMed: 19471117]
- 180. Nassar A, Azab AN. Effects of lithium on inflammation. ACS Chem Neurosci. 2014; 5 (6) 451–458. DOI: 10.1021/cn500038f [PubMed: 24803181]

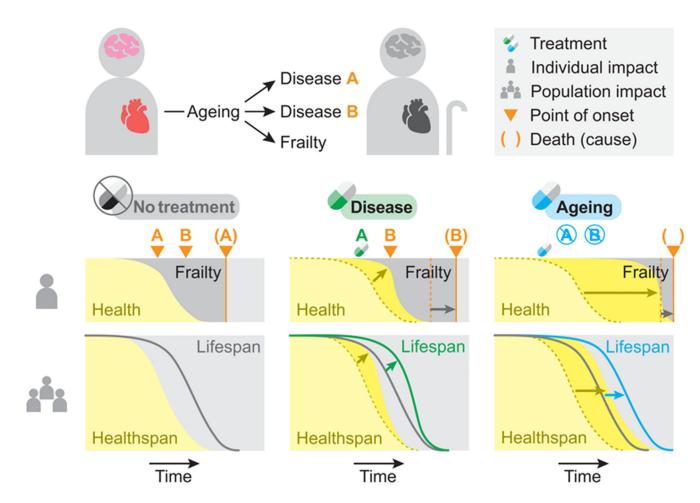


Figure 1. Approaches to health in old age: disease- and aging-centered models.

In this theoretical representation, aging is the major risk factor for prevalent diseases 'A' and 'B' and causes frailty. Left untreated or treated symptomatically, a typical individual in this population will develop diseases 'A' and 'B' and die of complications related to disease 'A' in this hypothetical scenario. Aging combined with these diseases will result in a certain period of frailty. At a population level, the life expectancy or median lifespan will be limited by the prevalent disease 'A,' and the healthspan will be limited by the burden of diseases 'A,' 'B,' and aging. In the disease-centered approach to health in old age, the ideal scenario of curing or preventing disease 'A' will increase both health- and lifespan at an individual and populational level, but the period of frailty is unlikely to change due to the burden of other untreated age-related diseases—here represented by disease 'B'—and aging itself. In the aging-centered approach to health in old age, the ideal scenario of slowing or preventing aging will similarly increase both health and lifespan at an individual and populational level, by preventing or delaying the development of diseases 'A' and 'B' and, necessarily, decrease the period of frailty.

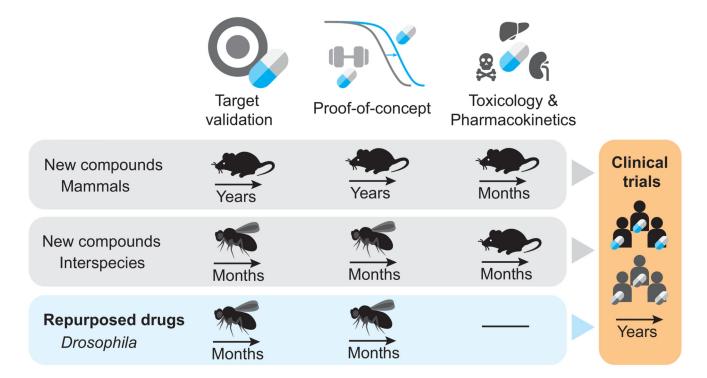


Figure 2. *Drosophila* as a tool to accelerate preclinical drug development in the context of aging. Scheme illustrating different preclinical routes for pharmacological interventions and how this process can be accelerated by a repurposing strategy and the use of *Drosophila* as an *in vivo* model system.

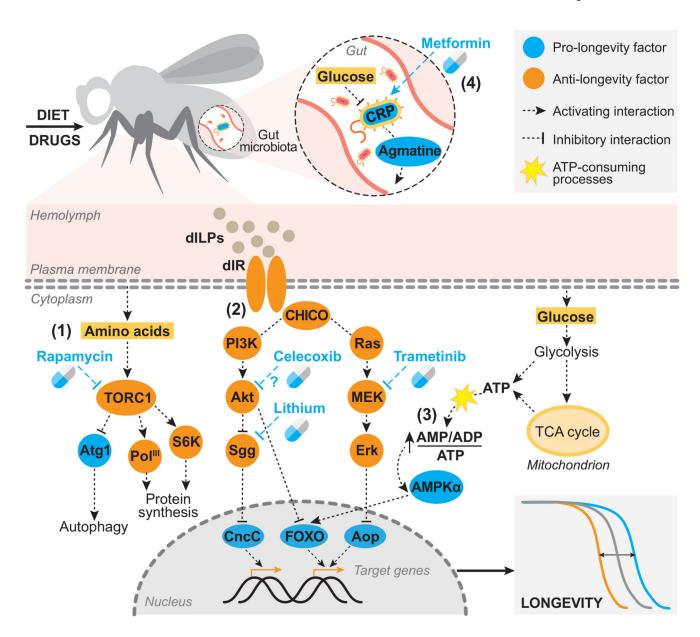


Figure 3. Simplified model of the nutrient-sensing network in *Drosophila* and its pharmacological targets.

(1) The target of rapamycin (TOR) signaling pathway is activated in an amino acid-rich environment. Activated TOR complex 1 (TORC1) phosphorylates S6 kinase (S6K) and inhibits autophagy-related 1 (Atg1), promoting protein translation and inhibiting autophagy, respectively. (2) The insulin/insulin-like growth factor signaling (IIS) pathway is activated when one of the eight *Drosophila* insulin-like peptides (dILPs) binds to their sole receptor, *Drosophila* insulin receptor (dInR). Downstream of the insulin receptor substrate CHICO, activation of the PI3K/protein kinase B (Akt) phosphorylation cascade inhibits the transcription factor dFOXO, as well as the fly GSK3 ortholog *Shaggy* (Sgg), which in turn inhibits CncC, the fly ortholog of the prolongevity transcription factor NRF2. The Ras-Erk pathway is also activated downstream of CHICO, ultimately leading to the inhibition of

the transcription factor Anterior open (Aop). (3) Under low-energy conditions, detected by an increase in the ratio of the adenosine nucleotides AMP and ADP to ATP, the kinase AMPK is activated and phosphorylates dFOXO at a different site. (4) Metformin induces upregulation of a bacterial transcriptional regulator, cAMP receptor protein (CRP), resulting in the production of the metabolite agmatine, which exerts prolongevity effects on the host.

Table 1
Data on clinically approved drugs that extend lifespan in *Drosophila*.

Drug	Target	Treatment Age	Strain and Sex	Food	Concentration	% Mean	% Med.	% Max.	References
			Single	e interventi	ons				
Aspirin	S6K?	From day 2	w^{Dah} Q	SY	0.5 μΜ	-	13%	-	Song et al. (2017) ¹¹³
	Imd?	From day 1	<i>w</i> ¹¹¹⁸ ₽	-	1 mg/L (5.6 μM)	32%	-	-	Zhu et al. (2021) ¹¹⁹
Celecoxib	Akt?	From day 3	$w^{Dah} Q$			-	10%	-	
			$w^{Dah}\sigma''$	Holidic	0.5 μΜ	-	9%	-	Wu et al. (2016) ¹¹⁶
		From day 42	W^{Dah} Q			-	5%	-	
Cortisone	-	From day 1	Canton-S ♀	-	0.188 mg/ml (467 μM)	18%	28%	13%	Hochschild (1971) ¹²⁰
			Canton-S ♂			43%	36%	31%	
Desoxycorti-costerone	-	From day 1	Canton-S ♂	-	$0.005~mg/ml~(15~\mu M)$	21%	29%	21%	Hochschild (1971) ¹²⁰
Everolimus	TORC1	From day -	$Ore^R \sigma$	SYC	3 mM	17%	-	-	Spindler et al. (2012) ⁶⁴
Fludrocortisone	-	From day 1	Canton-S ♀	-	0.0075 mg/ml (18 µM)	11%	23%	9%	Hochschild (1971) ¹²⁰
			Canton-S ♂			-	1%	-6%	
Ibuprofen	-	From day -	Canton-S ♀		0.5 μΜ	-	9%	4%	He et al. (2014) ¹¹⁴
			Canton-S ♂	SYS		-	17%	-8%	
		Day 30-40	Canton-S ♀		1 μΜ	-	23%	-	Proshkina et al.
			Canton-S ♂			-	n/s	-	$(2016)^{115}$
Lamotrigine	-	From day -	IV \circ	BM+YP	12 mg/ml	17%	-	15%	Avanesian et al. (2010) ¹⁵⁸
			$IV\sigma$		(47 mM)	15%	-	18%	
Lithium	GSK-3/ Sgg	From day 2	W^{Dah} Q		1-25 mM	-	16%	18%	Castillo-Quan et al. (2016) ⁸⁶
			$W^{1118}Q$	CV	25 mM	-	13%	18%	
			w^{1118} o [*]	SY	25 mM	-	23%	14%	
			W^{Dah} Q		1 mM	-	15%	10%	Castillo-Quan et al. (2019) ⁹⁷
		From day 32	w ^{Dah} ♀		25 mM	-	9%	8%	Castillo-Quan
Metformin	Complex I E. coli	From day 2	Dah♀		1–10 mM	-	n/s	-	Slack et al. (2012) ¹⁰⁶
	CRP			SY	25 mM	-	-11%	-	
			Dah ♂		1–50 mM	_	n/s	_	

Drug	Target	Treatment Age	Strain and Sex	Food	Concentration	% Mean	% Med.	% Max.	References
					100 mM	-	-31%	-	
		From day 2	W^{Dah} Q	Holidic*	0.5 mM	17%	-	-	Pryor et al. (2019) ¹⁰⁷
		From day 1	w ¹¹¹⁸ ₽	SYC	5 mM	-	17%	-	Suzuta et al. (2022) ¹⁰⁹
Mifepristone (=RU486)	JH	From day 2	<i>w</i> ¹¹¹⁸ ₽			15%	38%	-	Landis et al. (2015) ¹⁴³
			w^{1118} Q*			-1%	0%	-	
			$Ore^R \circ$		0.16 / 1	-10%	-13%	-	
			$Ore^R \mathfrak{P}^*$	SYC	0.16 mg/ml (372 μM)	3%	4%	-	
			Canton-S ♀			5%	12%	-	
			Canton-S Q*			15%	18%	-	
Minocycline	-	From day 1	Ore ^R ♀	SYS	0.87 mM	-	~57%	-	Oxenkrug et al.
			Ore^R σ			-	~71%	-	$(2012)^{152}$
		From day -	$Ore^R \sigma$			-	57%	-	Bonilla et al. (2012) ¹⁵⁰
						-	63%	-	Mora et al. (2013) ¹⁵³
		From day 3	Canton-S ♀	SYC	0.05 mM	-	14%	-	Lee et al. (2017) ¹⁵¹
			Canton-S ♂			-	27%	-	
			w^{1118} Q			-	17%	-	
			W^{1118} o [*]			-	23%	-	
Morphine	-	From day 5	OreR♀	SYC	0.25 mg/ml (665 μM)	9%	-	-	Dubiley et al. (2011) ¹⁵⁹
			Ore^R σ		0.01 mg/ml (27 µM)	16%	-	-	
N-acetyl- _L -cysteine (NAC)	-	From day 1	$Ore^R \sigma$	SYC	10 mg/ml (61 mM)	-	27%	25%	Brack et al. (1997) ¹²⁶
			Canton-S ♀	SY+YP	100 nM	-	-2%	-5%	Shaposhnikov et al. (2018) ¹²⁸
			Canton-S ♂			-	9%	8%	
			W^{118} Q			-	6%	n/s	Niraula & Kim (2019) ¹²⁷
				SYC	1 mg/ml (6 mM)				
			$W^{1118}\sigma$			-	n/s	n/s	
Rapamycin	TORC1		W^{Dah} Q		200 μΜ	-	13%	-	Bjedov et al. (2010) ⁵⁶
		F	w^{Dah} of			-	6%	-	
		From day 2	yw♀	SY		-	4%	-	
			W^{Dah} Q			-	14%	10%	Castillo-Quan et al. (2019) ⁹⁷

Drug	Target	Treatment Age	Strain and Sex	Food	Concentration	% Mean	% Med.	% Max.	References
			Ore ^R ♂	SYC	100 μΜ	38%	-	50%	Admasu et al. (2018) ¹⁴⁷
		L2/3-Pupae	w ^{iso31} ♀			-	n/s	-	Aiello et al. (2022) ⁶⁵
		Day 0-10			200 μΜ	-	10%	-	
		Day 10-13				-	n/s	-	
		Day 10–20 L2/3-Pupae	w^{iso31} of	YSCMS		-	n/s 8%	-	
		Day 0-10				-	14%	-	
		Day 10-13				-	n/s	-	
		Day 10-20				-	n/s	-	
		From day 2	$w^{Dah} Q$			12%	13%	-	Juricic et al. (2022) ⁶⁶
		Day 1-30				11%	13%	-	
		Day 30-100				6%	8%	-	
		Day 45-100		SY		3%	3%	-	
		Day 60-100				n/s	n/s	-	
		Day 1-15				7%	7%	3%	
		Day 15-30			$200 \mu M$	5%	7%	3%	
		From day 2	$W^{Dah} Q$		50 μΜ	-	20%	7%	Regan et al. (2022) ⁶³
			w^{iso31} σ		200 μΜ	-	n/s	13%	
			Dah♀	SY	200 μΜ	-	3%	-	
			Dah ♂			-	4%	-	
			DGRP♀			-	4%	-	
			DGRP ♂			-	n/s	-	
Rifampicin	dFOXO?	From day 1	Ore^R σ	SYA	50 μΜ	11%	-	27%	Admasu et al. (2018) ¹⁴⁷
Salicylamide	-	From day 1	Canton-S ♀	-	6.5 mg/ml	12%	25%	27%	Hochschild (1971) ¹²⁰
			Canton-S ♂		(47 mM)	17%	23%	22%	
Sevelamer	-	From day -	yw♂	YSCM C	1% (54 μM)	-	17%	-	Bergwitz (2012) ¹²⁴
Simvastatin	-	From day 1	Ore ^R ♂	SYC	240 μΜ	14%	-	-	Spindler et al. (2012) ¹²¹
Trametinib	Mek	From day 2	_W Dah ♀	SY	15.6 μΜ	-	12%	-	Slack et al. (2015) ⁹⁴
						-	25%	18%	Castillo-Quan et al. (2019) ⁹⁷
		From day 30				-	4%	-	Slack et al. (2015) ⁹⁴
Triamcinolone	-	From day 1	Canton-S ♀	-	0.04 mg/ml	19%	25%	16%	Hochschild (1971) ¹²⁰
			Canton-S o		(101 µM)	18%	23%	5%	
Zoledronate	-	From day 4	$W^{Dah}Q$	YMC	1 μΜ	-	n/s	-	Chen et al. (2021) ⁵²

Drug	Target	Treatment Age	Strain and Sex	Food	Concentration	% Mean	% Med.	% Max.	References
	,		w ^{Dah} ♂			-	~5%	-	
		From day 40	W^{Dah} Q		10 μΜ	-	17%	-	
			w^{Dah} σ'			-	5%	-	
			Combin	ed interve	ntions				
Rapamycin Rifampicin	TORC1 dFOXO?	From day 1	$Ore^R \sigma$	SY	100 μM 50 μM	63%	-	65%	Admasu et al. (2018) ¹⁴⁷
Rapamycin Lithium	TORC1 Sgg	From day 2	w ^{Dah} ♀	SY	200 μM 1 mM	-	21%	15%	Castillo-Quan et al. (2019) ⁹⁷
Rapamycin Trametinib	TORC1 Mek	From day 2	_W ^{Dah} ♀	SY	200 μM 15.6 μM	-	34%	24%	Castillo-Quan et al. (2019) ⁹⁷
Lithium Trametinib	Sgg Mek	From day 2	_W ^{Dah} ♀	SY	1 mM 15.6 μM	-	27%	20%	Castillo-Quan et al. (2019) ⁹⁷
Rapamycin Lithium Trametinib	TORC1 Sgg Mek	From day 2	_W ^{Dah} ♀	SY	200 μM 1 mM 15.6 μM	-	47%	35%	Castillo-Quan et al. (2019) ⁹⁷

Summary of mean, median (med.), and maximum (max.) % lifespan extension from published studies (n/s, not significant; -, data not available). The type of food is generalized based on the major ingredients, but the relative proportion/concentrations of each component may be different between studies. All media contain agar as a setting agent, with propionic acid and/or nipagin (tegosept) as antimicrobial/fungal preservatives. SY: sugar (sucrose)—yeast-based medium; SYC: SY medium with cornmeal; SYS: SY medium with semolina; SY+YP: SY medium with fresh yeast paste; YMC: yeast—molasses—cornmeal medium; YSCS: yeast—soy flour—cornmeal—corn syrup medium; YSCMC: yeast—soy flour—cornmeal—malt—corn syrup medium; BM+YP: banana—molasses medium with fresh yeast paste; holidic: chemically defined medium that mimics SY; holidic*, modified holidic medium without sucrose; 9*, virgin female.

 $\label{thm:continuous} \textbf{Table 2} \\ \textbf{Proposed criteria for drug screening in } \textit{Drosophila preclinical studies of aging pharmacology.} \\ \\$

Challenges	Proposed Criteria					
Survival as endpoint	• Extends both median and maximum lifespan					
	• Effective in genetically heterogeneous populations					
Health scores	• Improves frailty index scores					
	• Improves imaging signs of tissue aging					
	• Improves markers of organ function					
Target validation	• Target mutation in the binding site cancels drug effect					
	• Target overexpression/knockdown simulates drug effect					
	• Conservation of the target across species					
	• Tissue-specific effects (if any) are seen in analogous organs					
Diet dependency	• Drug effect is observed with different media					
	• Drug effect is not limited by the absence of nutrients in the diet					
	• Effect is present when flies are maintained under dietary restriction					
Polypharmacy	• Identification of common upstream or downstream targets					
	• Improved integration of personalized data					