scientific reports



OPEN Characterization of semi-arid Chadian sweet sorghum accessions as potential sources for sugar and ethanol production

Gapili Naoura¹, Yves Emendack^{2⊠}, Nébié Baloua³, Kirsten vom Brocke^{4,7}, Mahamat Alhabib Hassan¹, Nerbewende Sawadogo⁵, Amos Doyam Nodjasse¹, Reoungal Djinodji¹, Gilles Trouche^{4,7} & Haydee Echevarria Laza⁶

Sweet sorghum (Sorghum bicolor (L.) Moench) is an important crop in Chad that plays an economic role in the countryside were stalks are produced mainly for human consumption without any processing. Unfortunately, very little information exists on its genetic diversity and brix content. Studies performed in 2014 and 2015 showed that there were significant variations (p < 0.001) for all assessed quantitative traits. Potential grain yield (0.12-1.67 t ha⁻¹), days to 50% flowering (68.3-126.3 days), and plant height (128.9–298.3 cm) were among traits that exhibited broader variability. Brix content range from 5.5 to 16.7% across accessions, was positively correlated to stalk diameter and plant height, but negatively correlated to moisture content in fresh stalk and potential grain yield. Fresh stalk yield range from 16.8 to 115.7 Mg ha⁻¹, with a mean value of 58.3 Mg ha⁻¹ across accession. Moisture content in fresh stalk range from 33.7 to 74.4% but was negatively correlated to fresh stalk yield. Potential sugar yield range from 0.5 to 5.3 Mg ha⁻¹ across accession with an average of 2.2 Mg ha⁻¹. Theoretical ethanol yield range from 279.5 to 3,101.2 L ha⁻¹ across accession with an average of 1,266.3 L ha⁻¹ which is significantly higher than values reported under similar semiarid conditions. Overall, grain yields were comparatively low. However, two accessions had grain yield of more than 1.5 t ha⁻¹; which is greater than the average 1.0 t ha⁻¹ for local grain sorghum varieties in Chad. These could have multi-purpose uses; grains, sugar and bioenergy production.

Sweet sorghum [Sorghum bicolor (L.) Moench] is an annual, seed-propagated C4 grass that derived its name from the high concentration of soluble sugars (a mixture of sucrose, glucose, and fructose) contained in its tall, juicy stalks^{22,26,38}. In Chad, sweet sorghum also commonly known as sugar sorghum, is cultivated by several farmers on small areas or sprinkled in the grain sorghum fields. It is also cultivated around village huts for consumption by children while their parents are away to their farms out of the villages¹³. The national agricultural statistics of the ministry in charge of agricultural production in Chad does not take sweet sorghum into account, thus information on the national production, the extent of its cultivation and genetic diversity are not available. The cultivation of sweet sorghum had been abandoned by farmers, threatening its genetic resources. Recent regained interest in the sale and consumption of sweet sorghum stalk in local markets has led to a boost in cultivation by small scale farmers¹³.

Most of the world's ethanol production is obtained from two major crops: corn and sugarcane⁶. Sweet sorghum offers one of the best plant-based bioethanol productions from its sugary stalk and is considered a potential bioenergy crop throughout most of the tropical and temperate zones of the world, and it is also a leading contender for biofuel production in the southern United States⁴⁶. Numerous studies have been performed to assess the agronomic performance and yields of juice, sugar and ethanol of sweet sorghum^{2,3,6,10,11,12,18,20,32,44}.

¹Institut Tchadien de Recherche Agronomique Pour le Développement (ITRAD), B.P. 5400, N'Djaména, Chad. ²Cropping Systems Research Laboratory, USDA-ARS, Lubbock, TX 79415, USA. ³International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Bamako, Mali. ⁴AGAP, Univ Montpellier, CIRAD, INRA, Montpellier SupAgro, University of Montpellier, 34090 Montpellier, France. ⁵Laboratoire Biosciences, Équipe Génétique et Amélioration des Plantes, Université Joseph KI-ZERBO, 03 BP 7021, Ouagadougou 03, Burkina Faso. ⁶Department of Plant and Soil Sciences, Texas Tech University, Lubbock, TX 79409, USA. ⁷CIRAD, UMR AGAP, 34398 Montpellier, France. [™]email: Yves.Emendack@ars.usda.gov

				Annual rainfall (mm)		
Regions	Departments	Number of villages	Number of accessions	2015	2014	
	Lac Houé	1	1	951	1,100	
Logone Occ	Dodjé	2	6	1,278	1,077	
Logotte Occ	Ngourkosso	2	2	905	934	
	Total	5	9			
	Mont de Lam	3	11	1639	1,281	
	Kou Est	1	4	1,432	1,298	
I accoma Oni	Kou Ouest	2	4	1,229	920	
Logone Ori	Nya-Pendé	3	10	1,428	1,159	
	Pendé	2	10	1,240	951	
	Total	11	39			
Mandoul	Mandoul Occ	2	4	914	881	
	Mandoul Ori	4	8	1,014	851	
	Total	6	12			
Mayo-Kebbi E	Mayo Boneye	2	7	874	712	
	Mayo Dallah	5	11	1,006	860	
Mayo-Kebbi W	Lac Léré	3	3	778	896	
	Total	8	14			
Moyen Chari	Barh Kôh	1	5	1,157	1,062	
	Tandjilé W	1	3	1,017	896	
m lui/	Tandjilé E	8	16	1,069	925	
	Total	9	19			
Tandjilé	CHAD	41	105			
	ICRISAT		5			
	TOTAL		110			

Table 1. Rainfall, regional distribution, and origins of sweet sorghum accessions used in study.

Compared to other bioenergy crops such as corn, wheat, sugarcane, sugar beet, cassava, sweet potato; sweet sorghum is drought tolerant, requires lower quantity of water (e.g. 1/3 of sugarcane, 1/2 of corn) and fertilization inputs, has tolerance to salinity, i.e. can be grown in marginal regions that are not commonly used for crop production and it also has lower greenhouse gas emissions on a life-cycle basis^{1,2,9}.

Sweet sorghum stalk is used for the production of food grade syrup, alcohol, and even chewed fresh in Brazil and India¹⁷. With its high sucrose content, the stalk is fermented for the production of bioethanol^{7,29} and can yield up to 8,000 L ha⁻¹ of ethanol which is approximately twice the ethanol yield of corn and 30% greater than the average produced from sugarcane²⁴. In 2019, the global fuel ethanol production was 110.1 billion liters, the two largest producers being the U.S. and Brazil with 59.8 and 32.6 billion litters respectively⁴⁵.

There are about 4,000 sweet sorghum cultivars distributed through the world³⁵. In Chad, sugar sorghum is grown for human consumption without any prior processing. Stalks are sold on the roadsides in villages or transported by trucks to large urban centers. In some rural areas in southern Chad, sugar sorghum is the main source of revenue and economic sustainability¹³. Despite the importance of sweet sorghum in Chad's human nutrition, few researches have evaluated the genetic diversity of the Chadian accessions. A study conducted as part of the sweet sorghum collection survey revealed a large diversity in the cultivars grown by farmers¹³.

The main purposes of this study were to: (1) determine the agro-morphological and phenological diversity in traits associated with yields of juice and ethanol of sweet sorghum accessions grown in the Sudanese zone of Chad, (2) identify high-performing cultivars which could be used for genetic improvement of grain sorghum accessions grown and sold in Chad and (3) identify high performing accessions with respect to ethanol production which could be beneficial to the global bioenergy production research especially in similar ecological regions.

Materials and methods

Plant materials. The plant material consisted of 105 landraces from the Bébédjia research station. These were local varieties from a prospecting collection carried out in the Sudanese zone (comprising Logone Occidental, Logone Oriental, Mandoul, Mayo Kebbi West, Mayo Kebbi East, Moyen Chari, and Tandjilé) of Chad in 2012¹³ (Table 1). Five improved sweet sorghum varieties from ICRISAT Mali, were used as checks (see Supplementary Table 1 for accessions numbers and names of varieties).

Experimental design. Field experiments were conducted at the ITRAD Research Centre in Bébédjia (9° 55' N Latitude North and 15° 8' Longitude East), during the growing season from April to October for two consecutive years; 2014 and 2015, on a poorly desaturated sandy clay soil type. The experimental design was an α -lattice with three replications, each of which was subdivided into twelve plots of ten lines. Seeds from each

accession were sown in holes on 10 m long lines, with 0.7 m spacing between lines and 0.3 m between seed holes. 100 kg ha^{-1} of NPK fertilizer was applied 15 days after planting.

Data collection. Seasonal rainfall distribution was measured every 10 days through the growing season in both years. Plant data were collected over the entire period of plant development, from emergence to maturity and after harvesting. As qualitative characters, the color and vigor of seedling were noted at emergence; seedling vigor rated from 1 to 9 based on stem size, leaf thickness and length of seedlings. 1 indicated poor vigor, 3 weak vigor, 5 fair vigor, 7 good vigor, and 9 excellent vigor. Panicle compactness, color of leaf midrib, grain color, presence of dimple, glume color, and glume hairiness were determined at maturity. The botanical race was determined in the field according to ¹⁵ and confirmed in the laboratory. Virtuosity was estimated according to the BONO scale (0–4): 0 for totally floury grains, 1 for rather floury grains, 2 for vitreous grains (at 50% vitreous), 3 for rather vitreous grains (more than 50% vitreous), and 4 for entirely vitreous grains.

Quantitative characteristics such as number of days to heading (NDH) and number of days to flowering (NDF; 50% flowering in accession) were determined from planting. At the hard dough stage, the perultimate leaf length (PLL; base of leaf to leaf tip), perultimate leaf width (PLW; at widest part of leaf), plant height (PHT, base of plant to tip of panicle), number of internodes (NIN), stem diameter (SDI; at fourth internode under panicle), panicle length (PAL; base to tip of panicle), panicle width (PAW; at broadest part of panicle), and internode length (INL; average of third and fifth internode under panicle) were determined from 10 randomly selected plants per accession. At maturity, main panicles of the 10 plants were harvested, air dried to constant weight for at least 10 days, and weighed to determine the main panicle (PWT). Panicles were thrashed and grains weighed to determine the weight of grains of the main panicle (PGW). The potential yield (PYI) was obtained by multiplying the average PGW from the 10 main panicles by the number of seedlings per hectare for each accession. Thousand grains weight (TGW) was also determined.

Brix content was measured from internodes using a hand-held refractometer (Master, Atago, Japan). Brix content is influenced significantly by the positions of the internodes³⁹. For this study the brix content was measured from the fourth and sixth internodes below the panicle. To determine the fresh and dry weight of stalk, the 10 randomly selected plants were (harvested at 5 cm above the soil surface) were stripped of leaves and panicles, and the stalks were weighed for fresh stalk weight (FSW). The stalks were air dried in the shade for at least 10 days until they reached constant weight for three consecutive days, at which the field dried stalk weight (DSW) was measured. Moisture content was determined as the percent difference between FSW and DSW. The fresh stalk yield (FSY) and dry stalk yield (DSY) were calculated as average of FSW and DSW respectively multiplied by number of plants per hectare^{10,39}.

Juice and sugar yields were calculated according to⁴³:

$$CSY = (FSY - DSY) \times Brix \times 0.75$$

$$JCY (80\% extracted) = [FSY - (DSY - CSY)] \times 0.8$$

$$SGY = JCY \times Brix \times 0.75$$

where CSY is conservative sugar yield (Mg ha⁻¹), FSY is fresh stalk yield (Mg ha⁻¹), DSY is dry stalk yield (Mg ha⁻¹), JCY is juice yield (Mg ha⁻¹), and SGY is sugar yield (Mg ha⁻¹).

Sugar concentration of juice (SCJ) was determined as 75% of brix expressed in g kg⁻¹ sugar juice³⁵:

$$SCJ(gKg^{-1}) = 0.75 \times Brix$$

Theoretical ethanol yield (TEY, L ha^{-1}) from extracted juice was calculated as sugar yield (kg ha^{-1}) multiplied by a conversion factor of 0.581 L kg^{-1} sugar⁴¹:

TEY
$$(L ha^{-1}) = CSY \times 0.581$$

Total soluble sugar (y, %) was estimated using equation by²³:

$$y(\%) = 0.8111x - 0.37285$$

where x is the Brix of stalk juice.

Data analysis. Analysis of variance (ANOVA) and Newman-Keuls test were performed to determine if the average of the quantitative characteristics varied significantly (at 5.0, 1.0, and 0.1% probability thresholds) between accessions. Some parametric distributions were calculated to see the dispersion of the values of the characteristics based on the average, the minimum, the maximum and the coefficient of variation. For all characteristics, genetic variabilities were estimated from the components of the analysis of variance, and broad heritability (H²) was calculated using the formulas by¹¹6. Bivariate analysis was carried out, using the Pearson correlation coefficient to see the link between two characteristics. Multivariate analyses were performed through Principal Component Analysis (PCA) to highlight uncorrelated characters, which were used to build the dendrogram from the Hierarchical Ascending Classification (HAC). Then the Discriminant Factor Analysis (DFA) was carried out to characterize the group from the HAC.

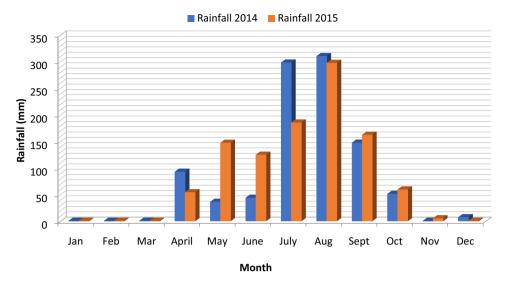


Figure 1. Season rainfall distribution measured during the 2014 and 2015 sweet sorghum growing season at the ITRAD Research Centre in Bébédjia, Chad.

Results

Seasonal rainfall distribution. Total season rainfall (Fig. 1) from planting to harvest was lower in 2014 (908.2 mm) compared to 2015 (1,098.5 mm). However, seasonal rainfall distribution showed crops receiving similar amount of rainfall (609.6 vs. 605.7) during the peak of crop growth and development in 2014 and 2015 respectively.

Variability in agro-morphological traits. Analysis of qualitative traits (Table 2) showed a great diversity among sweet sorghum accessions from Chad. Generally, 64% of seedlings were green and 36% violet, and these were accession specific. Seedling vigor ratings showed frequencies of 31.4, 32.9, and 21.2% for low, fair, and good vigor respectively. The color of the leaf midrib (a phenotypic characteristic of sweet sorghum) was green for 69.5% and white for 29.5% of the accessions. Most accessions had loose panicles (51.9%) with straw colored glumes (80.9%), red seed (84.5%), often very floury in vitreousity (69.7%) and mainly from the caudatum (64.55%) or bicolor (35.45%) race.

The analysis of variance (Table 3) showed highly significant differences (p < 0.001) between accessions in all assessed quantitative agronomic characteristics. The coefficients of variation (CV) for most characters was above 30%, reflecting variability amongst accessions. Heritability (H^2) range from 58.6 to 98.8%. The strongest agronomic heritability ($H^2 > 80\%$) were recorded for number of days to flowering (95.6), number of days to heading (95.5), plant height (94.9), number of internodes (94.5), internode length (90.2), panicle length (90.2), stalk diameter (83.9), dry stalk weight (82.1), and perultimate leaf width (81.0). The lowest heritability was scored for potential yield (64.8), panicle grain weight (64.8), panicle weight (61.5) and panicle width (58.6).

Other traits (see Supplementary Table 1) showed significant variability with some Chadian accessions performing better than the ICRISAT checks. Compared to the average of ICRISAT checks, 22 Chadian accessions were taller, 63 had shorter days to heading, 54 had shorter days to flowering, 46 had higher number of internodes, 20 had longer internode lengths, 16 had longer perultimate leaf, 7 had wider perultimate leaf, 19 had longer panicle, 48 with wider panicle, 41 with higher fresh stalk biomass, 74 with higher dry stalk biomass, and 33 with broader stem diameter. Five of the 105 accessions screened had potential grain yields equal to or higher than the average 1.0 t ha⁻¹ of grain sorghum production in semi-arid Chad.

All assessed quantitative characteristics linked to sugar production showed significant variability (p < 0.001) within and when compared to ICRISAT checks (Table 4). The coefficient of variation was high (CV > 30%) for most characteristics except for Brix (19.8%), total soluble sugar (20.6%) and the sugar concentration on juice (20.1%). The heritability was very high for all assessed characteristics except for the Juice yield (59.6%) and moisture content of fresh stalk weight (57.6%). Heritability was strongest for brix (98.8%), sugar concentration of juice (98.8%) and total soluble sugar (98.8%), followed by dry stalk yield (82.1%) and then fresh stalk yield (79.8%).

The fresh stalk yield range from 16.75 to 115.7 Mg ha⁻¹ and the dry stalk yield from 6.54 to 75.5 Mg ha⁻¹, giving a moisture content range of 33.7 to 74.4%, with an average of 48.7%. The juice yield for 80% of the sugar extracted (represented the sugar for the stalk) range from 7.34 to 43 Mg ha⁻¹ with an average of 23.6 Mg ha⁻¹. The brix content range from 5.5 to 16.7% and the total soluble sugar range from 4.1 to 13.1 Mg ha⁻¹ with an average of 9.3 Mg ha⁻¹. The potential sugar yield range from 0.5 to 5.3 Mg ha⁻¹ with an average of 2.2 Mg ha⁻¹ and the theoretical ethanol yield range from 279.5 to 3,101.2 L ha⁻¹ with average of 1,266 L ha⁻¹.

Compared (see Supplementary Table 1) to the ICRISAT checks (F60, IS23525, IS23536, IS23541, and IS23574), 37 Chadian accessions had more brix, 39 more fresh stalk yield, 31 more dry stalk yield, 35 more juice yield, 32 more conservative sugar yield, 19 more juice yield, 12 more soluble sugars, and 22 more theoretical ethanol yield;

Characteristics	Modality	Frequency (%)	
0.1 6 11:	Green	64.2	
Color of seedling	Violet	35.7	
	Excellent	7.3	
	Good	21.2	
Vigor of seedling	Fair	32.9	
	Weak	31.4	
	Poor	7.3	
	Green	69.7	
Color of leaf midrib	White	29.5	
Color of leaf midrib	Black	11.8	
	Yellow	0.2	
Glume color	Straw	80.9	
Giunie color	Red	7.3	
Seed color	Red	84.5	
Seed color	White	15.5	
Vitreousity	Floury	69.7	
Vitreousity	Vitreous	30.3	
	Loose	51.9	
Panicle	Semi-compact	32.6	
	Compact	15.5	
Botanical race	Caudatum	64.6	
Botamcai race	Bicolor	35.4	
	Minimal	55.8	
Glume hairiness	Partial	36.4	
	Hairy	7.8	
Grain dimple	Absent	52.7	
Grain dinipie	Present	47.3	

Table 2. Modeling for the qualitative characteristics of sweet sorghum in semi-arid Chad.

Characteristics	Minima	Maxima	Average	CV (%)	F-value	H ² (%)
PHT (cm)	128.9	298.3	232.2 ± 47.9	20.6	17.1***	94.9
SDI (cm)	1.0	2.2	1.73 ± 0.3	17.9	5.7***	83.9
PLW (cm)	5.4	11.4	8.2 ± 1.1	13.3	4.4***	81.0
PAW (cm)	4.7	13.0	7.4 ± 1.4	31.7	2.3***	58.6
INL (cm)	14.4	34.9	21.6±3.8	36.2	9.2***	90.2
PLL (cm)	46.0	83.3	68.3 ± 7.1	55.9	3.3***	74.4
PAL (cm)	10.9	35.9	24.8 ± 4.6	45.4	8.9***	90.2
NIN	4.9	16.2	10.3 ± 3.2	43.5	15.1***	94.5
NHD (days)	68.3	126.3	94.2 ± 15.2	16.0	19.8***	95.5
NFW (days)	68.3	126.3	95.1 ± 15.1	16.0	20.3***	95.6
PWT (g)	160.0	1,266.7	502.7 ± 215.8	42.9	2.4***	61.5
PGW (g)	60.0	833.3	272.4 ± 136.7	50.2	2.7***	64.8
PYI (t ha ⁻¹)	0.1	1.67	0.54 ± 0.27	50.2	2.7***	64.8
TGW (g)	12.7	33.3	21.39 ± 4.31	20.2	-	-
FSW (g)	116.7	783.3	403.8 ± 150	37.1	4.6***	79.8
DSW (g)	44.3	511.1	217.3 ± 101.9	46.9	5.0***	82.1

Table 3. Variance of agronomic characteristics of Chadian sweet sorghum accessions. ***Indicates significance at p < 0.001, CV; coefficient of variation, H²; heritability, PHT; plant height, SDI; stalk diameter, PLW; perultimate leaf width, PAW; panicle width, INL; internode length, PLL; perultimate leaf length, PAL; panicle length, NIN; number of internodes, NHD; number of days to heading, NFW; number of days to flowering, PWT; panicle weight, PGW; panicle grain weight, PYI; potential yield, TGW; 1,000-grain weight, FSW; fresh stalk weight, DSW; field dried stalk weight.

Characteristics	Minima	Maxima	Average	CV (%)	F-value	H ² (%)	
MFS (%)	33.7	74.4	48.7 ± 8.7	30.3	2.3***	57.6	
FSY (Mg ha ⁻¹)	16.7	115.7	58.3 ± 22.1	38.0	4.6***	79.8	
DSY (Mg ha ⁻¹)	6.5	75.5	31.3 ± 14.7	47.1	5.0***	82.1	
JCY (Mg ha ⁻¹)	7.3	43.0	23.6±7.7	32.7	2.4***	59.6	
CSY (Mg ha ⁻¹)	0.6	5.9	2.5 ± 1.1	43.4	4.0***	75.7	
SGY (Mg ha ⁻¹)	0.5	5.3	2.2 ± 1.0	44.5	4.2***	76.7	
SCJ (g kg ⁻¹)	41.3	125.0	89.7 ± 18	20.1	86.8***	98.8	
y (%)	4.1	13.1	9.3 ± 1.9	20.6	86.8***	98.8	
TEY (L ha ⁻¹)	279.5	3,101.2	1,266.3 ± 563	44.5	4.2***	76.7	
Brix (%)	5.5	16.7	11.9 ± 2.4	19.8	86.8***	98.8	

Table 4. Analysis of variance of twelve characteristics determining sugar and ethanol production in 105 accessions of semi-arid Chadian sweet sorghum. ***Indicates significance at p < 0.001, CV; Coefficient of variation, MFS; moisture content of fresh stalk, FSY; fresh stalk yield, DSY; field dried stalk yield, CSY; conservative sugar yield, JCY; juice yield, SGY; sugar yield, SCJ; sugar concentration of juice, TEY; Theoretical ethanol yield, y; total soluble sugar. \pm standard error.

Traits	PHT	PLL	PAL	NFW	SDI	NIN	Brix	FSY	DSY	JCY	SGY	TEY	MFS	TGW
PLL	0.48***													
PAL	0.72***	0.44***												
NFW	0.70***	0.50***	0.32											
SDI	0.65***	0.59***	0.41***	0.81***										
NIN	0.82***	0.49***	0.45***	0.92***	0.86***									
Brix	0.50***	0.14	0.32***	0.37***	0.37***	0.43***								
FSY	0.76***	0.50***	0.45***	0.79***	0.81***	0.81***	0.52***							
DSY	0.78***	0.52***	0.42***	0.82***	0.82***	0.85***	0.54***	0.97***						
JCY	0.64***	0.40***	0.44***	0.63***	0.69***	0.65***	0.46***	0.92***	0.80***					
SGY	0.65***	0.31	0.43***	0.60***	0.66***	0.64***	0.75***	0.88***	0.80***	0.92***				
TEY	0.65***	0.31	0.43***	0.60***	0.66***	0.64***	0.75***	0.88***	0.80***	0.92***	1.00***			
MFS	-0.73***	-0.54***	-0.37***	-0.73***	-0.64***	-0.77***	-0.47***	-0.68***	-0.80***	-0.39***	-0.45***	-0.45***		
TGW	-0.34***	-0.14	-0.12	-0.31**	-0.21*	-0.31**	-0.20*	-0.34***	-0.33***	-0.29**	-0.27**	-0.27**	0.32**	
PYI	-0.24*	0.09	-0.23*	-0.03	0.06	-0.06	-0.26*	-0.20*	-0.17	-0.23*	-0.24*	-0.24*	0.19	0.30**

Table 5. Correlation coefficient for agro-morphological and phenotypic characteristics of 105 semi-arid Chadian sweet sorghum accessions. *,**,*** Significance at p < 0.05, p < 0.01, p < 0.001; PHT: plant height; PLL: perultimate leaf length; PAL: panicle length; NFW: number of days to flowering; SDI: stem diameter; NIN: number of internodes; FSY: fresh stalk yield; DSY: field dried stalk yield; JY: juice yield; SY: sugar yield; TEY: theoretical ethanol yield, MFS: moisture content in fresh stalk; TGW: 1,000-grain weight; PYI: potential yield.

with 10 accessions (ecotypes numbers 21, 46, 64, 66, 72, 80, 81, 82, 130, and 137) having significantly higher theoretical ethanol yield than the best ICRISAT variety (IS23541; TEY of 1695 L ha^{-1}).

Correlation analysis amongst characteristics. The Pearson correlation matrix (Table 5) shows the strength of the relationship between quantitative traits based on 5% (p < 0.05), 1% (p < 0.01) and 0.1% (p < 0.001) probability levels. Brix content was significantly and positively correlated with theoretical ethanol yield (r = 0.75, p < 0.001), potential sugar yield (r = 0.75, p < 0.001), fresh stalk yield (r = 0.52, p < 0.001). Moisture content in fresh stalk was negatively correlated to brix content (r = -0.47, p < 0.001). Brix content was negatively correlated to thousand grain weight (r = -0.20, p < 0.05) and potential grain yield (r = -0.26, p < 0.05).

Greater theoretical ethanol yields were correlated to taller plants (r = 0.65, p < 0.001), thicker stems (r = 0.66, p < 0.001), number of internodes internodes (r = 0.64, p < 0.001), longer panicles, fresh stalk (r = 0.88, p < 0.001) and dry stalk (r = 0.80, p < 0.001) yields, number of days to flowering (r = 0.60, p < 0.001). Moisture content in fresh stalk had a negative correlation to all assessed characteristics except thousand grain weight (r = 0.32, p < 0.01).

Late maturity correlated positively to larger stem diameter, number of internodes, fresh and dry stalk yields, juice and sugar yields but negatively to thousand grain weight, with no significant correlation to potential grain yield. Taller plants with longer panicles, high brix content, high fresh and field dried stalk yields, and high juice and sugar yields were negatively related to potential grain yield.

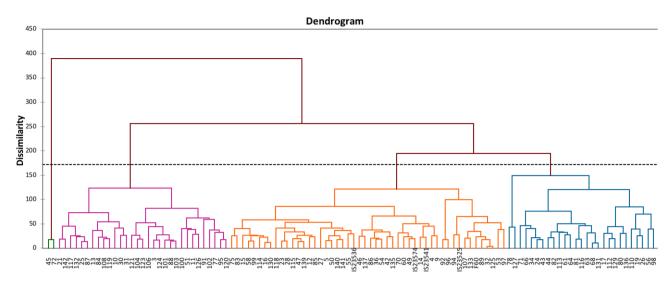


Figure 2. Clustering of 110 sweet sorghum accessions into 4 groups using the standardized squared Euclidean distance of Ward's hierarchical clustering method.

	Group 1	Group 2	Group 3
Group 2	14.0***		
Group 3	49.7***	12.2***	
Group 4	117.5***	52.5***	15.7**

Table 6. Distances of Mahalanobis and statistical significant of Fisher. **,***indicates significance at p < 0.01 and p < 0.001 respectively.

Characteristics	Group 1	Group 2	Group 3	Group 4
PHT (cm)	213.9 ± 46	230.7 ± 47.7	251.5 ± 42.0	295.3 ± 1.6
PLL (cm)	68 ± 6.9	66.9 ± 7.4	70.7 ± 6.1	75.3 ± 1.8
PLW (cm)	7.8 ± 1.0	8.1 ± 1.2	8.7 ± 0.9	8.9 ± 0.7
PAL (cm)	24.5 ± 3.9	23.9 ± 3.9	26.4±5.9	30.5 ± 4
PAW (cm)	6.9 ± 1.5	7.4 ± 1.4	7.9 ± 1.4	7.9 ± 0.5
INL (cm)	22.39 ± 4.1	21.2 ± 3.5	21.4±4.2	21.9 ± 3.2
NFW (days)	88.2 ± 11.2	94.2 ± 15.8	103.4 ± 13.7	109.3 ± 8
FSW (g)	223.6 ± 53.9	403.7 ± 46.0	583.3 ± 63.5	777.8 ± 7.9
Brix (%)	10.7 ± 2.7	11.64±1.8	13.7 ± 1.7	15.2 ± 2.1
Number of accessions	31	50	27	2

Table 7. Comparison of agro-morphological and phenotypic traits of four groups of Chadian sweet sorghum derived from Ward's hierarchical clustering. PHT: plant height; PLL: perultimate leaf length; PLW: perultimate leaf width; PAL: panicle length; PAW: panicle width; INL: internode length; NFW: number of days to flowering; FSW: fresh stalk weight. ± standard error.

Cluster analysis and diversity organization. Hierarchical clustering, using Ward's method and truncation on level 175, grouped the 110 sweet sorghum accessions into four major groups (Fig. 2). Group 1 was composed of 31 accessions and was further subdivided into 2 sub-groups; 1a (13 accessions) and 1b (18 accessions). Group 2 had the largest number of accessions (50) and was subdivided into 2 sub-groups; 2c (38 accessions) and 2d (12 accessions). Group 3 consisted of 27 accessions divided into 2 sub-groups; 3e (2 accessions) and 3f. (25 accessions). Group 4 was composed of 2 accessions. The Mahalanobis distance obtained through the Discriminant Factor Analysis (DFA) showed long distance between all the groups (p < 0.001 and p < 0.01), which signified that the four groups were quite distinct (Table 6).

The mean values of the accessions in each cluster are given in Table 7, obtained through the Discriminant Factor Analysis (DFA). Group 1 accessions were characterized as short, early flowering, with lowest fresh stalk weight and the lowest average brix content (10.7%). Groups 2 and 3 showed mean values of all characteristics studied,

Regions	Group 1	Group 2	Group 3	Group 4	Total
Tandjilé	9	8	2	0	19
Mayo Kebbi Est	3	4	0	0	7
Mayo Kebbi Ouest	4	5	5	0	14
Logone Oriental	10	13	14	2	39
Logone Occidental	3	6	1	0	9
Mandoul	2	6	4	0	12
Moyen Chari	0	4	1	0	5
Accession from ICRISAT	0	5	0	0	5
Total	31	50	27	2	110

Table 8. Regional distribution of the accessions of Chadian sweet sorghum following Ward's hierarchical clustering into four major groups.

Accessions Name	Brix	FSY	DSY	JCY	CSY	SGY	SCJ	y	TEY
(Ecotype #)	(%)	(Mg ha	(Mg ha ⁻¹)					(%)	(L ha ⁻¹)
"Balnda" (66)	16.7	80.0	32.8	42.6	5.9	5.3	166.7	13.1	3,101.2
"Sian Guebeuh" (21)	15.0	104.3	57.8	41.4	5.2	4.7	150.0	11.8	2,712.2
"Var137" (137)	15.7	97.8	54.2	39.0	5.1	4.6	156.7	12.3	2,669.5
"Zimikay" (72)	13.7	115.7	67.0	43.0	5.0	4.4	136.7	10.7	2,570.0
"Chian Woua" (46)	16.7	114.1	75.5	34.7	4.8	4.3	166.7	13.1	2,530.5
"Kadbal" (80)	14.3	96.0	52.9	38.2	4.6	4.1	143.3	11.3	2,393.1
"Kad bel hym" (81)	15.3	82.5	47.2	31.5	4.1	3.6	153.3	12.1	2,111.0
"Kad Nda" (82)	15.3	80.5	46.3	30.5	3.9	3.5	153.3	12.1	2047.2
"Bagnadé" (64)	15.7	77.6	44.3	29.7	3.9	3.5	156.7	12.3	2036.1
"Syan Teigne" (130)	16.0	91.1	58.7	29.1	3.9	3.5	160.0	12.6	2032.6
IS23541	14.1	71.4	40.4	27.4	3.3	2.9	141.1	11.1	1695.0
IS23574	9.7	66.5	38.8	23.8	2.0	1.7	96.7	7.5	1,004.8
F60	11.7	55.4	31.8	20.6	2.1	1.8	116.7	9.1	1,049.3
IS23525	11.9	52.6	30.5	19.3	2.0	1.7	118.9	9.3	997.2
IS23536	13.7	51.7	32.2	17.2	2.0	1.8	136.7	10.7	1,025.0

Table 9. Compositional characteristics of 10 promising genotypes of Chadian sweet sorghum compared to ICRISAT checks (IS: 23,541, 23,574, 23,525, 23,536, and F60). FSY: fresh stalk yield; DSY: field dried stalk yield; JY: is a juice yield; CSY: conservative sugar yield; SY: sugar yield; SCJ: sugar concentration of juice; y: total soluble sugar; TEY: theoretical ethanol yield.

with an average brix value of 11.6% and 13.7% respectively. The two accessions in group 4 were characterized as the tallest plants, late flowering, with the heaviest fresh stalk weight and the highest average brix value (15.2%).

The structuring of the individuals in each group (Table 8) shows a distribution of accessions based on their regions of origin. Group 2 represented all the regions of origin of the accessions. On the other hand, in group 1, the accessions of the Moyen Chari region were not represented. In group 3 accessions of Mayo Kebbi Ouest were not represented. Finally, in group 4, only the accessions of the Oriental Logone were found. The accessions from ICRISAT (see Supplementary Table 1: F60, IS23525, IS23536, IS23541, and IS23574) were found only in group 2.

Promising genotypes for sweet sorghum improvement program in Chad. This multi-year evaluation of the 105 Chadian sweet sorghum accessions showed higher performance in juice and sugar yields of 10 accession compared to the 5 improved sweet sorghum from ICRISAT considered as high performers (Table 9). The "*Balnda*" accession had 16.7% brix, 42.6 Mg ha⁻¹ juice yield, 5.3 Mg ha⁻¹ potential sugar yield, and a theoretical ethanol yield of 3,101.2 L ha⁻¹. It was followed by the accession "Sian *Guebeuh*" with 2,712.2 L ha⁻¹ of theoretical ethanol yield, and "Var137" with 2,669.5 L ha⁻¹ of theoretical ethanol yield. The "Zimikaye Combole" accession, despite its 13.7% brix, achieved a high sugar yield (4.4 Mg ha⁻¹) and ethanol yield (2,570 L ha⁻¹). The best ICRISAT varieties was IS23541 with 14.1% brix, 27.4 Mg ha⁻¹ juice yield, 2.9 Mg ha⁻¹ potential sugar yield, and 1695.0 L ha⁻¹ theoretical ethanol yield.

Discussion

The agro-morphological diversity study of Chadian sweet sorghum showed significant variability in qualitative and quantitative characteristics. In fact, two main leaf midrib colors were observed; white and green which are also the main colors of this type of sorghum²⁸. According to research by²⁸, accessions with white midrib were not

very juicy. However, our study showed that some white midrib accessions were very juicy. The sweet sorghum accessions from the Sudanese zone were identified to be from the *caudatum* and *bicolor* race, unlike the dryseason sorghum which are from the *durra* race¹⁴. According to²⁸, sweet sorghum belongs to *bicolor*, *caudatum*, *durra* and hybrid *bicolor-guinea* race. Ritter et al.³³ suggested that sweet sorghums are of polyphyletic origin, with relatives among *kafir*, *caudatum*, and other grain sorghum types.

Murray et al.²⁶ identified three separate groups of sweet sorghum which often are classified together. He classified these major types as: syrup types; (historical and some modern) which were from the *caudatum* race, modern sugar and energy types; associated with the *kaffir/bicolor* races, and amber types; mainly *durra* and *bicolor* races.

Generally, the 0.5 t ha⁻¹ average grain yield production in this study was quite low compared to the average 1.0 t ha⁻¹ observed in Chadian grain sorghums or 0.87 t ha⁻¹ for dry-season grain sorghum in the same region¹⁴. According to⁷, sweet sorghum is characterized by reduced grain yield as compared to grain sorghum. However, two of the accessions *Begon* (1.7 t ha⁻¹) and *GWS lache* (1.5 t ha⁻¹) showed higher than average grain yields and could be used to improve grain yield of sweet sorghum. Plant height, stem diameter, number of internodes, internode length and other morphological characteristics showed high variability in the accessions studied.

The brix from the current study differed significantly between accessions and it value ranged from 5.5 to 16.7%, with an average of 11.9%. Brix content was lower than that reported by^{8,28}, who obtained brix value ranging from 8.9 to 21.8% and 11.8 to 22.5%, respectively. The "*Balnda*" and "*Chian Woua*" accessions had 16.7% brix, which was higher than what has been recorded in many sweet sorghum studies. According to³⁰, optimal harvesting stage for sweet sorghum is when the juice contains 15.5–16.5% brix which is one of the most important characteristics necessary to obtain juice of high fermentable quality and thus maximize ethanol yield per hectare.

The moisture content of fresh stalk for the 105 landraces ranged from 33.7 to 74.4%, averaging 48.7%. The average value is lower than moisture contents obtained by⁴⁰ (76.0%; using a single cultivar) and¹⁰ (81.0%; using three cultivars). But the value obtained on this current study were high than that obtained by⁵ (16.5%; using 73 sweet sorghum accessions). The higher end of the observed range of juice yield in this study were similar or higher than values obtained by⁴³, but similar to values obtained by⁴¹ working with 31 sweet sorghum lines in Arizona.

Estimated sweet sorghum sugar yields in current study showed high level of diversity (p < 0.001) amongst the 105 cultivars, the average value (2.2 Mg ha⁻¹) being lower than estimated mean values (4.0 Mg ha⁻¹ and 4.0 to10.7 Mg ha⁻¹) reported by^{38,41}; both evaluating 4–6 sweet sorghum lines at variable planting dates and locations respectively. However, 2.2 Mg ha⁻¹ was higher than values reported by³⁴ (1.8 Mg ha⁻¹, 1 variety with variable NPK fertilization management),⁴⁴ (1.8 Mg ha⁻¹ in 2009, using 1 hybrid at variable N-fertilization rates) and³⁵ (1.7 Mg ha⁻¹ using 5 cultivars across 3 years). The average amount of theoretical ethanol yield (1,266 L ha⁻¹) were lower than value (2,854 L ha⁻¹) obtained by⁴¹, but higher than that obtained by³⁵ (1,025 L ha⁻¹) and²(1,000–1,149 L ha⁻¹, using 12 cultivars). According to¹⁹, up to 13.2 Mg ha⁻¹ of total sugars, equivalent to 7,682.0 L ha⁻¹ of ethanol can be produced by sweet sorghum under favorable conditions.

The correlation matrix showed positive correlations of interest between Brix content with plant height, stem diameter, number of internodes and number of days to flowering. Brix is a measure of dissolved sugar to water mass ratio of a liquid; it was positively influenced by the maturity. According to³⁷, all known sweet sorghums are tall, and prior research identified a positive correlation between height and sugar accumulation. The positive relation between brix and maturity suggest that early maturity may not be a desirable characteristics for sweet sorghum variety development since plants will need more days to accumulates more biomass and store energy in its stalk throughout the growing period^{27,32,36}. Positive relations between plant height and days to flowering suggest that taller plants tended to flower later as observed by^{8,42}. The study showed plant height, number of days to flowering and fresh stalk weight negatively affected the moisture content in fresh stalk. Similar results were obtained by⁵ working with sweet sorghum cultivars from the U.S. National Plant Germplasm System collection.

Yields of sugar and theoretical ethanol were significantly (p < 0.05) and negatively influenced by potential grain yield and thousand grain weight. Indeed, sweet sorghum accessions are characterized by the accumulation of carbohydrates in their juicy stems³² to the detriment of the grains which remain rather poorly filled. Clerget et al.⁴ suggested a potential negative interaction between stem development and grain yield in sweet sorghum. This is contrary to grain sorghum accessions where accumulation of carbohydrates is done in favor of grains. Sweet sorghum landraces usually have small panicles and the stem sweetness is commonly attributed to low panicle strength²². However, the interactions and trade-offs between panicle size, grain filling and stem development are complex and can be complicated in photoperiod-sensitive sweet sorghums which tend to show great phenotypic plasticity. While competition for carbohydrates between grain filling and sugar storage in stems has been suggested by other studies^{2,31}, the remobilization of stem reserves towards grain was frequently reported as being small²¹. Thus, varieties in current study which combined high stem reserves with comparatively good grain yield will be great candidates for dual-purpose (food-fuel) sweet sorghum breeding. These varieties are currently under consideration for a potential collaborative project between sorghum scientists in Chad and the United States. Such varieties were reported²⁵ as being cultivated by farmers in semi-arid Mali.

Conclusion

The current study showed that sweet sorghum accessions from Chad were from *caudatum* and *bicolor* race, and most of them had loose panicles with red seeds. Brix values ranged from 5.5 to 16.67% were found and greatly differed (p < 0.001) among accessions. Two accessions "*Balnda*" and "*Chian Woua*" with a high brix value of (16.67%) were identified and could be used as source of sugar to improve grain sorghum in Chad. The yields of potential sugar and theoretical ethanol showed the values ranging from 0.45 to 5.3 Mg ha⁻¹ and 279.5 to 3,101.2 L ha⁻¹ respectively. The study showed high variability (p < 0.001) for all assessed quantitative characters within sweet sorghum accessions from Chad, and identified four major groupings within accessions, each with multiple subgroupings except cluster 4 which had 2 accessions. This study provides valuable findings which could be used to

improve sweet sorghum production in Chad for dual purpose use. Furthermore, the ten accessions with higher brix content than the five improved ICRISAT varieties could be used in biofuel (ethanol) breeding programs in similar geographical production regions. These accessions are currently been considered for improvement in a collaborative project with US based scientists.

Received: 12 February 2020; Accepted: 8 July 2020

Published online: 11 September 2020

References

- 1. Almodores, A. & Hadi, M. R. Production of bioethanol from sweet sorghum: a review. Afr. J. Agric. Res. 4(9), 772-780 (2009).
- 2. Briand, C. H., Geleta, S. B. & Kratochvil, R. J. Sweet sorghum [Sorghum bicolor (L.) Moench] a potential biofuel feedstock: analysis of cultivar performance in the Mid-Atlantic. Renew. Energy 129(2018), 328–333 (2018).
- 3. Channappagoudar, B. B., Biradar, N. R., Patil, J. B. & Hiremath, S. M. Assessment of sweet sorghum genotypes for cane yield, juice characters and sugar levels. *Karnataka J. Agric. Sci.* 20(2), 294–296 (2007).
- 4. Clerget, B., Dingkuhn, M., Goze, E., Rattunde, H. F. W. & Ney, B. Variability of phyllochron, plastochron and rate of increase in height in photoperiod-sensitive sorghum varieties. *Ann. Bot.* **101**, 579–594 (2008).
- 5. Cuevas, H. E., Prom, L. K. & Erpelding, J. E. Tapping the US sweet sorghum collection to identify biofuel germplasm. *Sugar Tech.* 17(4), 428–438 (2015).
- 6. Davila-Gomez, F. J., Chuck-Hernandez, C., Perez-Carrillo, E., Rooney, W. L. & Serna-Saldivar, S. O. Evaluation of bioethanol from five different varieties of sweet and forage sorghums (Sorghum bicolor (L.) Moench). Ind. Crop Prod. 33, 611–616 (2011).
- 7. Disasa, T., Feyissa, T., Admassu, B., Fetene, M. & Venugopal, M. Mapping of QTLs associated with brix and biomass-related traits in sorghum using SSR markers. *Sugar Tech.* **20**(3), 275–285 (2018).
- 8. Disasa, T., Feyissa, T. & Admassu, B. Characterization of Ethiopian sweet sorghum accessions for brix, morphological and grain yield traits. *Sugar Tech.* 19, 72–82 (2017).
- Drapcho, D. A., Nhuan, N. P. & Walker, T. H. Biofuels engineering process technology (McGraw-Hill Publishing Company, Inc., New York, 2008).
- Ekefre, D. E. *et al.* Evaluation of three cultivars of sweet sorghum as feedstocks for ethanol production in the Southeast United States. *Heliyon* 3, e00490 (2017).
- 11. Erickson, J. E. *et al.* Planting date affects biomass and brix of sweet sorghum grown for biofuel across Florida. *Agron. J.* **103**(6),
- Fortmeier, R. & Schubert, S. Storage of non-structural carbohydrates in sweet sorghum [Sorghum bicolor (L.) Moench]: comparison of sterile and fertile lines. J. Agron. Crop Sc. 175, 189–193 (1995).
- Gapili, N., Doyam, N. A., Djinodji, R. & Djondang, K. Prospection et collecte des accessions de sorgho sucré (Sorghum bicolor (L.) Moench) du Tchad. J. Appl. Biosci. 100, 9504–9514 (2016).
- Gapili, N. et al. Assessment of agro-morphological variability of dry-season sorghum cultivars in Chad as novel sources of drought tolerance. Sci. Rep. 9, 19581. https://doi.org/10.1038/s41598-019-56192-6 (2019).
- 15. Harlan, J. R. & de Wet, J. M. J. A simplified classification of cultivated Sorghum. *Crop Sci.* 12(2), 172–176 (1972).
- Hosseini, S. J., Sarvestani, Z. T., Pirdashti, H., Afkhami, A. & Hazrati, S. Estimation of heritability and genetic advance for screening some rice genotypes at salt stress conditions. Int J. Agron. Plant Prod. 3(11), 475–482 (2012).
- 17. House, H. R., Gomez, M., Murty, O. S., Sun, Y. & Verma, B. N. Development of some agricultural industries in several African and Asian countries. In *Sorghum: Origin, history, technology, and production* (eds Smith *et al.*) 131–190 (Wiley, New York, 2000).
- 18. Hunsigi, G., Yekkeli, N. R. & Kongawad, B. Y. Sweet stalk sorghum: an alternative sugar crop for ethanol production. *Sugar Tech.* 12(1), 79–80 (2010).
- 19. Jackson, D. R. et al. Research report on development of sweet sorghum as an energy crop volume 1: Agricultural task (U.S. Department of Commerce, USDOE, Battele, 1980).
- 20. Kim, M. & Day, D. F. Composition of sugar cane, energy cane, and sweet sorghum suitable for ethanol production at Louisiana sugar mills. *J. Ind. Microbiol. Biotechnol.* **38**(7), 803–807 (2010).
- 21. Kiniry, J. R., Tischler, C. R., Rosenthal, W. D. & Gerik, T. J. Nonstructural carbohydrate utilization by sorghum and maize during grain growth. *Crop Sci.* 32, 131–137 (1992).
- 22. Lingle, S. E., Tew, T. L., Rukavina, H. & Boykin, D. L. Post-harvest changes in sweet sorghum I: brix and sugars. *Bioenerg. Res.* 5, 158–167. https://doi.org/10.1007/s12155-011-9164-0 (2012).
- 23. Liu, R., Li, J. & Shen, F. Refining bioethanol from stalk juice of sweet sorghum by immobilized yeast fermentation. *Renew. Energ.* 33, 1130–1135 (2008).
- Luhnow, D., & Samor, G. As Brazil fill up ethanol, it wean-off energy imports. Wall Street Journal, January 2009. https://wsj.com/article/SB113676947533241219.html/. Accessed 10 May 2014 (2006).
- 25. Makanda, I., Tongoona, P. & Derera, L. Quantification of genotypic variability for stem sugar accumulation and associated traits in new sweet sorghum varieties. *Afr. Crop Sci. Conf. Proc.* **9**, 391–398 (2009).
- 26. Murray, S. C., Rooney, W. L., Hamblin, M. T., Mitchell, S. E. & Kresovich, S. Sweet sorghum genetic diversity and association mapping for brix and height. *Plant Gen.* 2, 48 (2009).
- Murray, S. C. et al. Genetic improvement of sorghum as a biofuel feedstock: I QTL for stem sugar and grain nonstructural carbohydrates. Crop Sci. 48, 2165–2179 (2008).
- 28. Nébié, B. et al. Variation de caractères agromorphologiques et du Brix d'une collection de sorghos à tige sucrée du Burkina Faso. Int. J. Biol. Chem. Sci. 7(5), 1919–1928 (2013).
- 29. Nuessly, G. S., Wang, Y., Sandhu, H., Larsen, N. & Cherry, R. T. Entomologic and agronomic evaluations of 18 sweet sorghum cultivars for biofuel in Florida. *Fla Entomol.* **96**(2), 512–528 (2013).
- 30. Prasad, S., Singh, A., Jain, N. & Joshi, H. C. Ethanol production from sweet sorghum syrup for utilization as automotive fuel in India. *Energ. Fuel.* 21, 2415–2420 (2007).
- 31. Rajendran, C., Ramamoorthy, K. & Backiyarani, S. Effect of deheading on juice quality characteristics and sugar yield of sweet sorghum. *J. Agron. Crop Sci.* 185, 23–26 (2000).
- 32. Ritter, K. B. et al. Identification of QTL for sugar-related traits in a sweet × grain sorghum (Sorghum bicolor L. Moench) recombinant inbred population. Mol. Breed. 22, 367–384 (2008).
- 33. Ritter, K. B., McIntyre, C. L., Godwin, I. D., Jordan, D. R. & Chapman, S. C. An assessment of the genetic relationship between sweet and grain sorghums, within Sorghum bicolor ssp. bicolor (L.) Moench, using AFLP markers. Euphytica 157, 161–176 (2007).
- 34. Roy, P. & Barik, S. An agronomic practice for the improvement of sweet sorghum (Sorghum bicolor (L.) Moench) crop: a study at Gangetic plains of West Bengal. Int. J. Appl. Agric. Res. 11(2), 103–113 (2016).
- 35. Rutto, L. K., Xu, Y., Brandt, M., Ren, S. & Kering, M. K. Juice ethanol and grain yield potential of five sweet sorghum bicolor (L.) Moench) cultivars. J. Sustain. Bioenerg. Syst. 3, 113–118 (2013).

- Shiringani, A. L., Frisch, M. & Friedt, W. Genetic mapping of QTLs for sugar-related traits in a RIL population of Sorghum bicolor L. Moench. Gen. Res. Crop Evol. 121, 323–336 (2010).
- 37. Shukla, S., Felderhoffa, T. J., Saballos, A. & Vermerris, W. The relationship between plant height and sugar accumulation in the stems of sweet sorghum (Sorghum bicolor (L.) Moench). Field Crops Res. 203, 181–191 (2017).
- 38. Smith, G. A. et al. Evaluation of sweet sorghum for fermentable sugar production potential. Crop Sci. 27, 788-793 (1987).
- 39. Sun, X. Z., Yamana, N., Dohi, M. & Nakata, N. Hardness and brix changes of internodes and extraction characteristics of chopped sweet sorghum. *J. Jpn. Soc. Agric. Mach.* **73**(2), 142–147 (2011) ((in Japanese)).
- 40. Sun, X. Z. & Yamana, N. Determining effects of sowing time and nitrogen fertilizer rate in Brix of sweet sorghum using a gear-type extractor. *Trans. ASABE* 55(4), 1589–1594 (2012).
- Teetor, V. H. et al. Effects of planting date on sugar and ethanol yield of sweet sorghum grown in Arizona. Ind. Crop Prod. 34, 1293–1300 (2011).
- 42. Tesso, T., Tirfessa, A. & Mohamad, H. Association between morphological traits and yield components in the durra sorghums of Ethiopia. *Hereditas* 148, 98–109 (2011).
- 43. Wortmann, C. S. *et al.* Dryland performance of sweet sorghum and grain crops for biofuel in Nebraska. *Agron. J.* **102**(1), 319–326
- 44. Uchino, H. et al. Effects of nitrogen application on sweet sorghum (Sorghum bicolor (L.) Moench) in the semi-arid tropical zone of India. JARQ 47(1), 65–73 (2013).
- 45. https://ethanolrfa.org/statistics/annual-ethanol-production/.
- 46. Viator, H. P., Lu, S. & Aragon, D. Influence of panicles and leafy materials on sweet sorghum juice quality. *J. Am. Soc. Sugarcane Technol.* 35, 21–30 (2015).

Acknowledgements

The authors would like to thank Mr. Djimlelngar Marc, Mr Ganezouné Michel and Mr Djenaissem Alfred, ITRAD technicians for their contribution in field maintenance and data collection.

Author contributions

G.N., M.H., Y.E., and N.S. conceived and designed experiment. G.N., M.H., D.R., N.B., and D.A. collected and provided germplasm for evaluation. G.N., D.A. and D.R., managed field experiments. G.N., Y.E., K.B., T.G., N.B. and H.L. analyzed data. G.N. and Y.E. wrote manuscript. G.N., M.H., Y.E., T.G., K.B., N.S. and H.L. read and reviewed manuscript for final publication.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary information is available for this paper at https://doi.org/10.1038/s41598-020-71506-9.

Correspondence and requests for materials should be addressed to Y.E.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

This is a U.S. Government work and not under copyright protection in the US; foreign copyright protection may apply 2020