

Ameliorative effect of Phytocee™ Cool against carbon tetrachloride-induced oxidative stress

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ABSTRACT

Background: Antioxidants from natural sources have a major role in reversing the effects of oxidative stress and promoting health, growth and productivity in animals. **Objective:** This study was undertaken to investigate the possible antioxidant activity and hepatoprotective effects of Phytocee™ Cool on carbon tetrachloride (CCl₄) induced oxidative stress and liver damage in rats. **Materials and Methods:** Animals were pretreated with Phytocee™ Cool for 10 days and were challenged with CCl₄ (1:1 v/v) in olive oil on the 10th day. After 24 h of CCl₄ administration blood was collected and markers of hepatocellular damage aspartate aminotransferase (AST), alanine aminotransferase (ALT) were evaluated. Rats were sacrificed and oxidative stress in liver was estimated using malondialdehyde (MDA), reduced glutathione (GSH), superoxide dismutase (SOD) and catalase. **Results:** CCl₄ caused a significant increase in serum AST, ALT, hepatic MDA and GSH levels, whereas the SOD and catalase activities were decreased. Phytocee™ Cool pretreatment attenuated the MDA, AST ALT levels and increased the activities of SOD and catalase. **Conclusion:** Phytocee™ Cool demonstrated antioxidant potential and hepatoprotective effects and plausibly be used in the amelioration of oxidative stress.

Key words: Antioxidant, carbon tetrachloride, oxidative stress, Phytocee™ Cool

INTRODUCTION

Free radicals generation is an integral feature of normal cellular function or metabolism. The innate enzymatic and nonenzymatic antioxidant defense systems act against the free radicals generated and protect the organisms from radical toxicity.^[1] However, despite the cellular antioxidant defense systems, endogenous or exogenous sources/stressors cause radical related damage due to the imbalance between free radicals and radical scavenging mechanisms. The imbalance so termed as oxidative stress has been implicated in development of various diseases. Although clinical manifestation of chronic pathologies related to oxidative stress in poultry is limited due to its short life span, it is considered as one of the potential causes that lead to deleterious effects on the cell structures, including lipids and membranes, proteins and DNA leading to poor

performance and growth.^[2,3] As the birds are exposed to stressful conditions such as high environmental temperatures, infections, immunization etc., oxidative stress results as an outcome, consequently resulting in reduced performance and productivity in broilers and layers.^[4,5] Therefore, to alleviate the oxidative stress and improve the performance and productivity in poultry, supplementation of synthetic antioxidants has become a common practice.^[6] Alternatively, to improve the growth performance of broiler chickens, productivity in layers, decrease stress challenges, addition of natural antioxidants to the feed may be the most welcome addition.

Numerous medicinal plants for decades have been investigated for their antioxidant potentials and the available literature provides evidence that herbs have gained considerable importance as natural antioxidants in improving the overall health aspects of the poultry industry.^[6,7] The indigenous medicinal plants like *Emblica officinalis*, *Ocimum sanctum* and *Withania somnifera* have already been demonstrated for their antioxidant potentials.^[8] *E. officinalis* (*Euphorbiaceae*) also known as *Phyllanthus emblica*,

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Amla or Indian gooseberry^[9] is a rich source of tannoid principles emblicanin A, emblicanin B, punigluconin and pedunculagin, vitamin C and flavones. These phytochemical entities of amla are reported to have potential anti-oxidant activity.^[10-12] *O. sanctum* (*Labiatae*) in Indian traditional systems of medicines has been used for adaptogenic/antistress, antioxidant and immune stimulating activities.^[13] Furthermore, *O. sanctum* was found to be effective in the management of general stress symptoms in a clinical trial probably due to its antioxidant, antistress and adaptogenic activity.^[14]

Withania somnifera popularly known as Ashwaganda is a perennial plant belonging to the family *Solanaceae* and has been widely reported for its antioxidant activity.^[10,15] Based on the above considerations, a unique polyherbal preparation, Phytocee™ Cool intended for poultry containing *E. officinalis*, *O. sanctum* and *W. somnifera* has been formulated and in addition electrolytes were added. As there is no scientific evidence existing for the antioxidant activity of Phytocee™ Cool, this study was undertaken to evaluate its antioxidant potential in rats.

MATERIALS AND METHODS

Animals

Male Wistar rats (150-200 g), bred and reared at central animal facility, Natural Remedies (Bangalore, India) were used in this study. The animals were housed under standard conditions of illumination cycle set to 12 h light and 12 h dark, at a temperature of 20-24° C and 30-70% relative humidity. Standard pelleted rodent feed (M/s. Amrut Laboratory Animal Feeds, Maharashtra, India) and ultraviolet purified and filtered water was provided *ad libitum*.

Drugs and chemicals

Carbon tetrachloride (CCl₄) (Rankem Fine Chemicals., India), vitamin C purified/ascorbic acid (Merck Specialties, India) and Refined olive oil (SOS Cuetara, S. A, Spain) were obtained. Other chemicals used were 2-thiobarbituric acid (TBA) and 5,5'-Dithio bis 2-nitro-benzoic acid (Sigma-Aldrich Co., USA), pyrogallol and potassium dihydrogen orthophosphate (Qualigens Fine Chemicals, India), tris buffer, hydrogen peroxide (H₂O₂) solution 30%, di-sodium hydrogen orthophosphate and trichloro acetic acid, (Ranbaxy fine chemicals, India). All other chemicals and reagents used were of analytical grade.

Plant materials

Phytocee™ Cool is a novel polyherbal formulation containing *E. officinalis* fruits, *O. sanctum* whole plant and *W. somnifera* roots and electrolytes.

Experimental groups and protocol

Male Wistar rats were randomly allocated to six groups, each containing 5-6 animals. Normal control (Group I) and negative/CCl₄ control (Group II) rats received vehicle (demineralized water 10 ml/kg) orally for 10 days whereas, Groups III to VI received vitamin C (20 mg/kg), or Phytocee™ Cool at three dose levels of 50, 100, 200 mg/kg respectively for 10 days by oral administration. On the 10th day, rats from Group II-VI were challenged with CCl₄ (1:1 v/v in olive oil) orally to induce hepatotoxicity.^[16] The animals were anesthetized 24 h after CCl₄ administration, blood was collected and the serum was separated. Subsequently, animals were euthanized; liver was dissected out, blotted and processed for the biochemical estimations.

Estimation of marker enzymes and antioxidant indices

Malondialdehyde (MDA) levels in liver homogenates were estimated as per the method described by Knight *et al.*^[17] The colored MDA-TBA adduct formed as a result of reaction of lipid peroxidation product, MDA with TBA was quantified spectrophotometrically at 532 nm and was considered as an index of lipid peroxidation. The results were expressed as nmol of MDA/g tissue using molar extinction coefficient of the chromophore ($1.56 \times 10^5/M^{-1}/cm^{-1}$).

Serum alanine aminotransferase (ALT), aspartate aminotransferase (AST) activities were determined by Reitman and Frankel^[18] method. Measurement is based on the detection of the increase in absorbance due to the formation of 2,4-dinitrophenyl-hydrazones in the reaction. Superoxide dismutase (SOD) activity was estimated by the method of Marklund and Marklund^[19] in the liver homogenates. This method employs the superoxide-driven auto-oxidation of pyrogallol. One unit of SOD activity was defined as the amount of the enzyme, which inhibits the pyrogallol autoxidation by 50% and results were normalized on the basis of total protein content to express the activity as SOD units/mg protein.

The method described by Aebi^[20] was followed for the estimation of catalase activity in the liver homogenate. The rate of decomposition of H₂O₂ is proportional to the decrease in absorbance at 240 nm. The difference in absorbance per unit time was considered as a measure of catalase activity and was expressed as catalase units/mg protein.

Reduced glutathione (GSH) was estimated by using Ellman's reagent following Sedlak and Lindsay^[21] method. The measurement of SH groups was based on the formation of yellow color product resulting from the reaction of 5,5'-dithiobis (2-nitrobenzoic acid) (DTNB) and GSH. The

yellow color formed was measured spectrophotometrically at 412 nm and was expressed in terms of $\mu\text{moles/g}$ tissue using the molar extinction coefficient of DTNB-GSH conjugate ($13.6 \times 10^3/\text{M}^{-1}/\text{cm}^{-1}$).

Statistical analysis

Data were expressed as mean and standard error of the mean and were analyzed using one-way ANOVA followed by Bonferroni method as *post-hoc* test. In case of heterogeneous data after transformation, Dunnett T3 method was used. Statistical significance was set at $P \leq 0.05$.

RESULTS

The hepatic levels of MDA were increased significantly ($P \leq 0.05$) in CCl_4 treated animals when compared to the normal control, while pretreatment with Phytocee™ Cool significantly decreased the CCl_4 induced increase in hepatic MDA levels in a dose dependent manner [Figure 1].

The mean serum ALT and AST levels, markers for hepatic tissue damage are depicted in Figure 2. Administration of CCl_4 to rats significantly ($P \leq 0.05$) increased the activity of serum hepatic marker enzymes when compared to normal control, whereas serum ALT and AST levels decreased significantly ($P \leq 0.05$) in groups pretreated with Phytocee™ Cool when compared to CCl_4 group.

The free radical induced hepatotoxicity of CCl_4 showed a significant decrease in SOD levels, nonsignificant decrease in catalase levels and significant increase in the GSH levels when compared to normal control group. However, Phytocee™ Cool pretreatment significantly increased SOD activity at all dose levels and catalase activity at a dose

of 100 mg/kg, conversely hepatic levels of GSH were decreased as compared to CCl_4 treated group [Table 1].

DISCUSSION

Stress is of major concern for poultry production systems, as birds are exposed routinely to an array of stressors such as immunization, high and low environmental temperatures, preslaughter holding etc., As a consequence of stress, release of oxidants, imbalance between oxidants and *in vivo* antioxidants occurs at the cellular level leading to modification of cellular macromolecules, cell death by apoptosis or necrosis and structural tissue damage. Eventually the cellular level damage leads to reduced performance, health as well as serious economic losses. Control of oxidative stress related damage has led to a substantial increase in the use of antioxidant feed supplements in poultry diet^[22-24] since high antioxidant status is observed as one of the most important factors positively affecting bird's performance in the poultry industry.^[25,26] With the growing interest in natural feed supplements, poultry farming has been seeking sources of

Table 1: Effect of Phytocee™ Cool on CCl_4 induced changes in reduced GSH, SOD and catalase of rat liver

Treatment	GSH ($\mu\text{moles/g}$ liver)	SOD (U/mg protein)	Catalase (U/mg protein)
Control (DM water; 10 ml/kg)	5.99±0.37	4.62±0.38	17.98±1.04
CCl_4 (CCl_4 and olive oil)	8.59±0.42*	3.04±0.14*	12.75±0.54
Vitamin C (20 mg/kg)	7.97±0.56	5.09±0.32#	17.36±1.37
Phytocee™ Cool (50 mg/kg)	8.03±0.24	5.31±0.48#	18.17±1.34
Phytocee™ Cool (100 mg/kg)	8.14±0.30	5.37±0.29#	23.40±2.64#
Phytocee™ Cool (200 mg/kg)	8.21±0.29	4.96±0.29#	20.21±2.71

CCl_4 : Carbon tetrachloride, GSH: Glutathione, SOD: Superoxide dismutase, SEM: Standard error of the mean. Values are expressed as mean±SEM; n=5-6. * $P < 0.05$ control versus CCl_4 , # $P < 0.05$ CCl_4 versus treatment

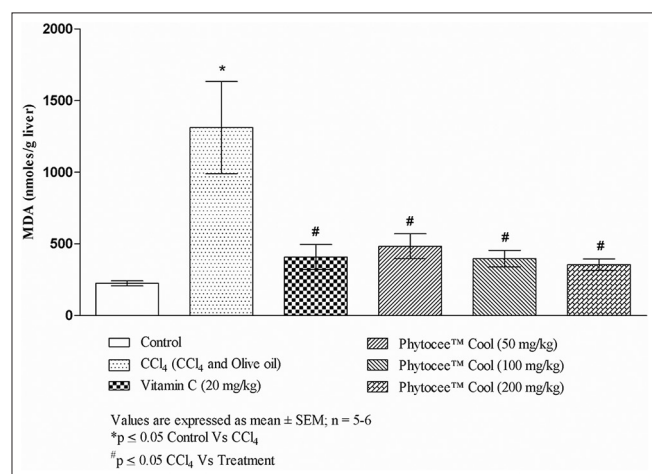


Figure 1: Effect of Phytocee™ Cool on carbon tetrachloride induced lipid peroxidation in rat liver

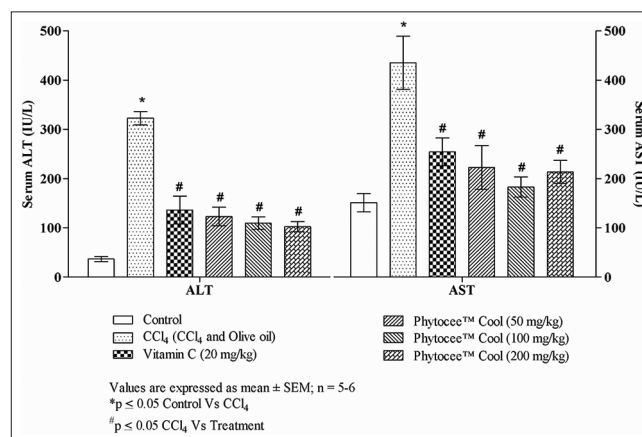


Figure 2: Effect of Phytocee™ Cool on carbon tetrachloride induced rise in serum alanine aminotransferase and aspartate aminotransferase in rats

natural antioxidants. Considering the aforementioned facts, the present study investigated the antioxidant potential of a unique polyherbal formulation Phytocee™ Cool in rats.

In this study, oxidative stress and liver injury was induced in rats by the administration of CCl_4 which is one of the well-recognized and widely used animal models to investigate the antioxidant and hepatoprotective effects of substances.^[27-29] CCl_4 induced oxidative stress is dependent on the bioactivation of CCl_4 to trichloromethyl radical (CCl_3) and trichloromethylperoxyl radicals. The initial step of biotransformation is the cytochrome P450 dependent reductive dechlorination of CCl_4 to CCl_3 that may form adducts with lipids and proteins or is trapped by molecular oxygen to form trichloromethylperoxyl radical (CCl_3OO).^[30,31] CCl_3OO is more reactive and interact with lipids, causing lipid peroxidation along with the production of 4-hydroxyalkenals.^[32,33] Earlier studies on natural products that showed protective effect against CCl_4 induced hepatotoxicity attributed the protective mechanism to antioxidant property of natural products.^[34,35] Hence, attenuation of CCl_4 induced oxidative stress by Phytocee™ Cool directly connotes its antioxidant effect.

The active radical metabolic intermediates of CCl_4 covalently bind to macromolecules such as lipids, produce hepatic lipid peroxidation. These free radicals attack polyunsaturated fatty acids (PUFA) to generate lipid peroxides, which are highly instable resulting in breakdown products such as reactive aldehydes (MDA), 4-hydroxyneonal or acrolein.^[36] MDA, one of the end products of PUFA peroxidation is considered as a marker for lipid peroxidation. The level of MDA reflects extent of lipid peroxidation in hepatocytes.^[37] In the present study, MDA levels in the liver homogenates of rats challenged with CCl_4 were significantly increased. However, the increase in the MDA levels were significantly prevented in the rats pretreated with Phytocee™ Cool indicating the ability of Phytocee™ Cool to break the chain reaction of lipid peroxidation. Furthermore, the lipid peroxidation in CCl_4 challenged rats lead to substantial increase in the serum ALT and AST concentrations. The increase in ALT and AST indicate considerable hepatocellular damage as these enzymes are normally localized in the cytoplasm and are released into the circulation after occurrence of cellular damage.^[38,39] The serum enzyme levels in the groups pretreated with Phytocee™ Cool were significantly low indicating that Phytocee™ Cool was able to prevent the leakage of enzymes in the cytosol to circulation by protecting the hepatic cell membranes from CCl_4 induced oxidative damage. Thus, CCl_4 induced significant hepatocellular damage as evident from hepatic MDA levels coupled with marked

elevation of AST and ALT enzymes, while Phytocee™ Cool pretreatment demonstrated substantial protection against oxidative stress and hepatocellular damage.

In order to further elucidate the antioxidant defense mechanisms of Phytocee™ Cool, hepatic enzymatic and nonenzymatic antioxidant defense mechanisms were evaluated. Enzymatic antioxidant defenses studied were SOD and catalase. SOD catalyzes conversion of superoxide to less toxic H_2O_2 , while catalase converts the H_2O_2 into nontoxic H_2O .^[40] CCl_4 induced generation of peroxy radicals leads to inactivation of SOD and catalase^[41] a finding in corroboration with CCl_4 challenged group in the present study. However, Phytocee™ Cool administration demonstrated significant increase in the activities of SOD and catalase indicating that it might have restored/activated the enzyme activities. These findings provide a supportive and convincing evidence for the antioxidant potential of Phytocee™ Cool.

Another antioxidant defense mechanism includes nonenzymatic antioxidants such as GSH. GSH in CCl_4 administered rats increased and the mechanism underlying this increase is unknown, albeit such increase in hepatic GSH on CCl_4 administration was also observed in several studies by Di Simplicio and Mannervik,^[42] Nakagawa^[43] and Lai *et al.*,^[44] Nonetheless, Phytocee™ Cool modulated the GSH levels.

Antioxidants play a major role in delaying or inhibiting the oxidation of easily oxidizable macromolecules like lipids by protecting these molecules from actions of free radicals or reactive oxygen species^[24] and accordingly lower plasma levels of antioxidants have been inversely correlated with the oxidative damage.^[45,46] Phytocee™ Cool demonstrated its antioxidant activity by protecting the hepatic cells from CCl_4 induced oxidative stress and by modulating the antioxidant defense systems. This protective effect of polyherbal formulation of Phytocee™ Cool can be attributed to the cumulative effect of its individual herbs, as the individual herbs (*E. officinalis*, *O. sanctum* and *W. somnifera*) are earlier reported for their antioxidant potential. Extracts of *E. officinalis* decreased the liver lipid peroxides in CCl_4 treated rats and increased activities of SOD and catalase enzymes *in vivo*.^[47-49] Another study revealed hepatoprotection by *E. officinalis* as evidenced by its ability to decrease AST and ALT levels in rats treated with ethanol.^[50] Meanwhile, *O. sanctum* extract demonstrated significant decrease in LPO levels coupled with significant increase in SOD and catalase in liver homogenates of rats exposed to oxidative stress.^[51,52] Further, earlier reports by Rajakumar and Rao (1993) have demonstrated that isoeugenol from *O. sanctum* with a double bond in its structure is a potent free radical scavenger.^[53] Furthermore,

flavonoids present in *Ocimum* extract protected the radiation induced cellular damage by scavenging the free radicals suggest strong evidence of antioxidant potential of *Ocimum*.^[54] *W. somnifera* restored the LPO levels and enhanced the activities of catalase in stress induced rats. Moreover, *Withania* treatment significantly reduced the total barbituric acid reactive substances, AST and ALT when compared to ammonium chloride treated group.^[55] Further, the phytoactives of *W. somnifera* showed antioxidant effects by modulating enzymatic and nonenzymatic defense systems.^[56] The available literature on the individual ingredients of Phytocee™ Cool suggests their potent antioxidant activity and the present study also revealed similar antioxidant and protective effects that is indicative of the plausible cumulative effect of the herbs in the formulation.

Phytocee™ Cool contains several phytoactives viz., flavonoids, phenolic compounds, ascorbic acid, tannoid principles etc., Flavonoids, phenolic compounds and ascorbic acid are well-reported in earlier studies for scavenging/chelating free radicals and may be involved in chain breaking activity. Furthermore, the tannoid principles largely would have contributed for the rise in antioxidant levels.^[10,49] These aforementioned mechanisms might plausibly be responsible for antioxidant activity of Phytocee™ Cool. Since, it is already established that ascorbic acid from natural sources is more potent antioxidant compared to synthetic vitamin C,^[57,58] Phytocee™ Cool can be used as a better substitute for synthetic vitamin C. Overall, Phytocee™ Cool is a unique combination of herbs and electrolytes that may contribute to alleviate the heat stress in birds; although the effect of electrolytes on the heat induced stress is yet to be explored. Thus, Phytocee™ Cool revealed its antioxidant effects by ameliorating oxidative stress induced changes and by modulating the activities of antioxidant defense systems.

CONCLUSION

Our study demonstrated CCl_4 induced marked oxidative stress was attenuated by Phytocee™ Cool. This protective effect of Phytocee™ Cool can be correlated to its potent antioxidant property.

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