

Evaluation of micronutrient status in serum and saliva of oral submucous fibrosis patients: A clinicopathological study

Shishir Ram Shetty,
Subhas Babu, Suchetha
Kumari¹, Pushparaja Shetty²,
R. Vijay³, Arvind Karikal⁴

Department of Oral Medicine and Radiology, AB Shetty Memorial Institute of Dental Science, ¹Department of Biochemistry, KS Hegde Medical Academy, ²Department of Oral Pathology, AB Shetty Memorial Institute of Dental Sciences, ³Central Research Laboratory, Nitte University, ⁴Department of Oral Surgery, AB Shetty Memorial Institute of Dental Science Mangalore, Karnataka, India

Address for correspondence:

Dr. Shishir Ram Shetty,
Department of Oral Medicine and Radiology, AB Shetty Memorial Institute of Dental Sciences, Nitte University, Mangalore, Karnataka, India.
E-mail: drshishirshettyomr@yahoo.com

ABSTRACT

Background: Oral submucous fibrosis (OSMF) is one of the most commonly occurring potentially malignant disorders in India and south East Asian countries where betel chewing is common practice. Iron and ascorbic acid are important agents for collagen synthesis. **Aims:** The aims of this study were to estimate the levels of iron and ascorbic acid in serum and saliva in patients with OSMF and to correlate change in levels of iron and ascorbic acid with the histopathological grading of OSMF. **Materials and Methods:** The study group comprised of 65 clinically diagnosed and histopathologically confirmed cases of OSMF; 21 age- and sex-matched controls were also enrolled in the study. Serum and salivary ascorbic acid were analyzed by the dinitrophenyl hydrazine method whereas serum and salivary iron were analyzed by the dipyriddy method. **Results:** The serum and salivary ascorbic acid levels consistently decreased with the progression of histopathological grading of OSMF. Serum and salivary iron levels were also decreased in OSMF patients, but this was not significant. **Conclusion:** Ascorbic acid and iron may have been used for the excessive collagen synthesis occurring during progression of OSMF. Hence, serum and salivary monitoring may play a crucial role in the early diagnosis and prognosis of OSMF.

Key words: Ascorbic acid, collagen, iron, oral submucous fibrosis, saliva

INTRODUCTION

Iron is one of the most abundant and necessary transition metals in the body, which is an essential component in DNA synthesis and in respiratory and oxidative metabolism.^[1] Recent studies have revealed depleted iron levels in patients with oral pre-cancerous lesions.^[2,3]

Ascorbic acid, or vitamin C, has the potential to protect both cytosolic and membrane components of cells from oxidant damage. In the cytosol, ascorbate acts as a primary

antioxidant to scavenge free radical species that are generated as by-products of cellular metabolism.^[4] Studies have revealed decrease in the ascorbic acid levels in oral submucous fibrosis (OSMF).^[5]

OSMF is a fibrotic condition of the oral cavity and is always associated with chronic epithelial inflammation and progressive deposition of collagenous extracellular matrix (ECM) proteins in the subepithelial layer of the buccal mucosa.^[6] The disease is seen in those from Indian subcontinent and from many parts of South-East Asia such as Taiwan.^[7] We carried out a study with an objective of assessing the status of serum and saliva in different histopathological grades of OSMF.

MATERIALS AND METHODS

Eighty-six patients between the age range of 20 and 40 years, reporting to the Department of Oral Medicine

Access this article online

Quick Response Code:



Website:
www.ijmpo.org

DOI:
10.4103/0971-5851.107087

and Radiology in a dental college in south India, were enrolled into the study. The study subjects included 65 histopathologically confirmed cases of OSMF and 21 age- and sex-matched healthy controls. A detailed case history that included habit index was taken from each subject in the study. Subjects with any other long-term systemic illness and long-term medication were excluded from the study. Five milliliters of unstimulated saliva was obtained by the spit method after following a standard pre-collection protocol. Five milliliters of venous blood was obtained from the antecubital vein, centrifuged and stored. Serum and salivary ascorbic acid were analyzed by the dinitrophenyl hydrazine method whereas serum and salivary iron were analyzed by the dipyriddy method. The data obtained was subjected to statistical analysis using the SPSS version 17 software.

RESULTS

The OSMF subjects were graded histopathologically into three categories. Of the 65 subjects in the control group, 22, 20 and 23 were categorized under grade 1, 2 and 3, respectively [Figure 1a–c] after histopathological examination. Mean serum and salivary ascorbic acid levels were significantly decreased in cases when compared with controls ($P<0.001$). However, serum and salivary iron levels showed no significant difference in case and controls ($P=0.073$ and 0.097 , respectively) [Table 1]. Both serum and salivary ascorbic levels consistently decreased as the histopathological grading progressed; there was a very highly significant difference among the three groups. Serum and salivary iron levels decreased as the grades progressed, but this was not significant [Table 2]. Serum and salivary levels showed significant correlation among cases ($r=0.315$ and $P=0.011$), but not among controls. Serum and salivary ascorbic acid levels showed no correlation among cases and controls [Tables 3 and 4]. There was no significant difference in the frequency of chewing habits and the levels of serum and salivary ascorbic acid or iron levels.

DISCUSSION

The incidence of OSMF is increasing like an epidemic among youngsters in the Indian and South East Asian population. The etiology for OSMF is still obscure and a varied number of factors have been proposed, acrcanut chewing being the most important.^[8] Several recent studies have been carried out on micronutrient status in OSMF; the copper and zinc studies being the most highlighted.^[8–10] Ascorbic acid levels have been investigated in several cancer-related studies^[11]; some studies have reported that ascorbic acid enhances destruction of cancer cells.^[12] We observed a reduction in iron levels in OSMF cases, although it was statistically not significant. This decrease in iron levels could be due to the requirement of iron for collagen synthesis by enzymes in hydroxylation of proline and lysine. This hydroxylation of proline and lysine is catalyzed by proline hydroxylase and peptidyl lysine hydroxylase, respectively. Peptidyl proline hydroxylase requires as co-factors molecular oxygen, ferrous iron, alfa keto-glutarate and ascorbic acid.^[13] We found a significant decrease in the levels of ascorbic acid in both serum and saliva. This could be attributed to its role in collagen synthesis. In a recent study involving 36 OSMF patients, tissue collagen and serum iron levels were assessed.^[5] They found elevated tissue collagen levels and depleted ascorbic acid and iron reserves in OSMF patients. We used the dinitrophenyl hydrazine method to assess serum and salivary ascorbic

Table 1: Comparison between iron and ascorbic acid levels in cases and controls

Parameters	n	Mean	Std. deviation	t
Se Fe cases	65	126.7312	26.82294	1.81600
Controls	21	140.1248	36.39456	$P=0.073$ ns
Sa Fe cases	65	107.4475	28.93174	1.67800
Controls	21	119.8981	31.51926	$P=0.097$ ns
Se Asc cases	65	1.1992	0.27486	5.13500
Controls	21	1.5671	0.31704	$P<0.001$ vhs
Sa Asc cases	65	0.6651	0.28221	3.84800
Controls	21	0.9357	0.27376	$P<0.001$ vhs

Students t test

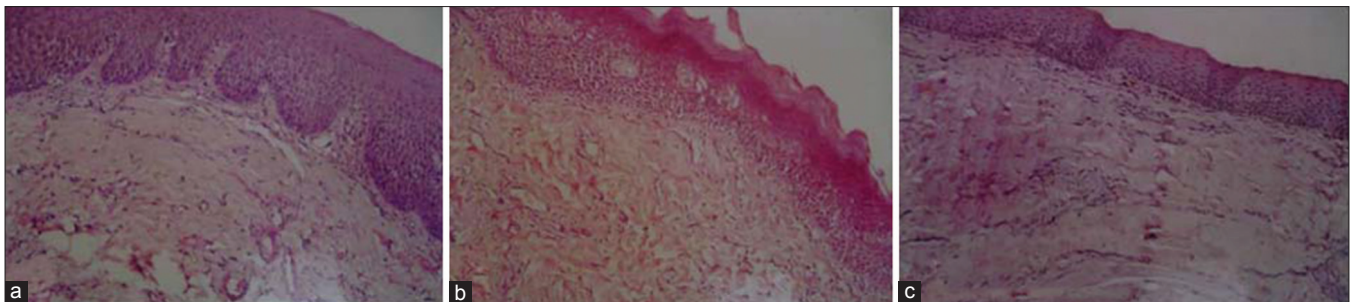


Figure 1: (a) Photomicrograph (40×) showing loose thin and thick fibers; (b) photomicrograph (40×) showing loose thin or thick fibers with partial hyalinization; (c) photomicrograph (40×) showing complete hyalinization

Table 2: Intercomparison of iron and ascorbic acid levels among histopathological grades of OSMF and controls

Parameter	n	Mean	Std deviation	F	P
Se Fe					
Grade 1	22	136.6795	21.55596	2.656	0.054
Grade 2	20	125.4335	23.27194		
Grade 3	23	118.3439	31.74315		
Controls	21	140.1248	36.39456		
Sa Fe					
Grade 1	22	111.9223	26.67577	1.284	0.285
Grade 2	20	107.8955	31.38907		
Grade 3	23	102.7778	28.46029		
Controls	21	119.8981	31.51926		
Se Asc					
Grade 1	22	1.2273	0.25757	8.830	<0.001 (vhs)
Grade 2	20	1.1610	0.24794		
Grade 3	23	1.2057	0.31812		
Controls	21	1.5671	0.31704		
Sa Asc					
Grade 1	22	0.5977	0.25370	6.065	<0.001 (vhs)
Grade 2	20	0.6490	0.29973		
Grade 3	23	0.7435	0.28543		
Controls	21	0.9357	0.27376		

(Fischer's test)

Table 3: Correlation between serum and salivary iron levels

Parameters	Sa Fe
Cases	
r	0.315
P	0.0119 (significant)
N	65
Controls	
r	0.322
P	0.155
N	21

Table 4: Correlation between serum and salivary ascorbic acid levels

Parameters	Sa Asc
Cases	
r	0.023
P	0.853
N	65
Controls	
r	0.286
P	0.209
N	65

acid. This method was first used to assess salivary ascorbic acid in 1969.^[14] Few studies however have stated that there is lack of correlation between serum and salivary ascorbic

acid levels.^[15] In our study, serum and salivary ascorbic acid levels showed good correlation. The ascorbic acid levels consistently increased as the histopathological grades of OSMF progressed. This indicated that iron and ascorbic acid may have been used for excessive collagen production and cross-linking occurring in OSMF. Hence, serum and salivary iron and ascorbic acid levels could be an important indicator for the progression of OSMF.

REFERENCES

- Weinberg ED. Roles of iron in neoplasia. Promotion, prevention, and therapy. *Biol Trace Elem Res* 1992;34:123-40.
- Gupta PC, Hebert JR, Bhonsle RB, Murti PR, Mehta H, Mehta FS. Influence of dietary factors on oral precancerous lesions in a population-based case-control study in Kerala, India. *Cancer* 1999;85:1885-93.
- Rajendran R, Vasudevan DM, Vijayakumar T. Serum levels of iron and proteins in oral submucous fibrosis (OSMF). *Ann Dent* 1990;49:23-5.
- May JM. Is ascorbic acid an antioxidant for the plasma membrane?. *FASEB J* 1999;13:995-1006.
- Anuradha CD, Devi CS. Serum protein, ascorbic acid and iron and tissue collagen in oral submucous fibrosis—a preliminary study. *Indian J Med Res* 1993;98:147-51.
- Murthi PR, Bhonsle RB, Gupta PC, Daftafray DK, Pindborg JJ, Mehta FS. Etiology of oral submucous fibrosis with special references to the role of arecanut chewing. *J Oral Pathol Med* 1995;24:145-52.
- Kiran Kumar K, Saraswathi TR, Ranganathan K, Uma Devi M, Joshua E. Oral Submucous Fibrosis: A clinico histopathological study in Chennai. *Indian J Dent Res* 2007;18:106-11.
- Khanna SS, Karjodkar FR. Circulating Immune Complexes and trace elements (Copper, Iron and Selenium) as markers in oral precancer and cancer: A randomised, controlled clinical trial. *Head Face Med* 2006;2:33.
- Gupta RP, Rai K, Hemani DD, Gupta AK. Study of trace elements (copper and zinc) in oral submucous fibrosis. *Indian J Otolaryngol Head Neck Surg* 1987;39:104-6.
- Rajendran. Serum levels of some trace and bulk element in OSMF. *J Indian Dent Ass* 1992;631:251-4.
- Chan SW, Reade PC. The role of ascorbic acid in oral cancer and carcinogenesis. *Oral Dis* 1998;4:120-9.
- Chen Q, Espey MG, Krishna MC, Mitchell JB, Corpe CP, Buettner GR, *et al.* Pharmacologic ascorbic acid concentrations selectively kill cancer cells: Action as a pro-drug to deliver hydrogen peroxide to tissues. *Proc Natl Acad Sci U S A* 2005;102:13605-9.
- Huang S, Ling T, Wu H. Experimental study on aqueous areca nut extracts inducing OSMF in rats. effect of mast cells on collagen metabolism. *Hua Xi Kou Qiang Yi, Xve Za Zhi*, 1997;15:94-6.
- Mäkilä E, Kirveskari P. A study of ascorbic acid in human saliva. *Arch Oral Biol* 1969;14:1285-92.
- Leggott PJ, Robertson PB, Rothman DL, Murray PA, Jacob RA. Response of Lingual Ascorbic Acid Test and Salivary Ascorbate Levels to Changes in Ascorbic acid intake. *J Dent Res* 1986;65:131-4.

How to cite this article: Shetty SR, Babu S, Kumari S, Shetty P, Vijay R, Karikal A. Evaluation of micronutrient status in serum and saliva of oral submucous fibrosis patients: A clinicopathological study. *Indian J Med Paediatr Oncol* 2012;33:224-6.
Source of Support: Nil, **Conflict of Interest:** None declared.