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Comparison of six commercial kits for the diagnosis of rotavirus infection in man and calves

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Seventy-two human and 72 bovine faecal specimens were tested for rotavirus by polyacrylamide gel electrophoresis (PAGE), four commercial enzyme-linked immunosorbent assay (ELISA) kits (RotaScreen, Wellcozyme, Rotazyme II and IDEIA) and two latex agglutination (LA) kits (RotaScreen and Wellcome).

Specimens which were negative by PAGE but positive by one or more of the kits were further examined by direct and immuno-electron microscopy (DEM and IEM). If also negative by DEM and IEM the kit result was considered to be a false positive. Three kits (RotaScreen and IDEIA ELISAs and RotaScreen LA) had specificity and sensitivity greater than 90% on the human specimens but only two (RotaScreen ELISA and LA) had specificity and sensitivity over 80% on the bovine specimens. These kits can therefore be used with reasonable confidence for rotavirus diagnosis, but none of them has any advantage over PAGE other than speed.

Keywords: rotavirus diagnosis, PAGE, electron microscopy, ELISA, latex agglutination.

Introduction

Rotaviruses are the main cause of infantile diarrhoea worldwide¹. They can also cause mild to severe diarrhoea in adults² and they play a major role in neonatal diarrhoea in many mammalian and avian species³.

Electron microscopy (EM) is used for rotavirus diagnosis and remains the standard by which other assays are judged. However, the requirement for expensive equipment with skilled operation limits its use. Further, with this method, it is also difficult to handle large numbers of specimens.

The demonstration of rotavirus double-stranded RNA in silver-stained polyacrylamide gels⁴ is another well-recognized method for diagnosing rotavirus infections. This technique also requires special equipment but this is inexpensive, simple to use, requires less technical expertise than EM and is suitable for processing many specimens. Polyacrylamide gel electrophoresis (PAGE) is the method used in our laboratories for the diagnosis of rotavirus infections.

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Latex agglutination (LA) and enzyme-linked immunosorbent assay (ELISA) kits for rotavirus diagnosis are widely available and often used. This present study was therefore initiated to assess the comparative specificity and sensitivity of ELISA and LA kits.

Materials and methods

Faecal specimens

Seventy-two human and 72 bovine faecal specimens submitted for routine examination were tested. Some of these contained viruses other than rotaviruses (adenovirus, enterovirus, coronavirus). Specimens were stored at -70° C for up to six months prior to this study and were tested at room temperature.

PAGE

The technique was similar to that of Herring *et al.*⁴ Any differences were for convenience and did not affect results.

Electron microscopy

Direct EM (DEM): Faecal specimens were mixed with an equal volume of distilled water and $3.5 \,\mu$ l were added to a carbon coated grid. After 30 seconds the grid was drained briefly by touching to filter paper. Phosphotungstic acid (1%), pH 7.0, was then added for 30 seconds before the grid was drained as above and allowed to dry. Stained grids were examined on Siemens Elmiskop 1 and 102 electron microscopes at a magnification of 40,000. Unless rotavirus was detected, each sample was examined for 20 minutes.

Immuno-electron microscopy (IEM): With minor modifications the method of Roberts and Harrison⁵ was followed. Grids were coated with an IgG fraction of a bovine anti-rotavirus serum for one hour at 37°C. Faecal specimens were prepared as 20% suspensions in distilled water, ground with carborundum powder and clarified at 10,000 \times g for 90 seconds. Supernatant fluid was added to the antibody-coated grid and allowed to adsorb for one hour at 37°C. Grids were then stained and examined as above.

Commercial kits

The ELISA kits used were: RotaScreen EIA (Mercia Diagnostics Ltd., Guildford, Surrey, U.K.), Wellcozyme Rotavirus (Wellcome Diagnostics Ltd., Dartford, U.K.), Rotazyme II (Abbott Diagnostics Division, Maidenhead, Berkshire, U.K.), IDEIA Rotavirus Test (Boots-Celltech Diagnostics Ltd., Slough, U.K.).

The LA kits used were: RotaScreen (Mercia Diagnostics Ltd.), The Wellcome Rotavirus Latex Test (Wellcome Diagnostics Ltd.).

All the tests were done in one laboratory and strictly according to the manufacturers' instructions. All ELISA results were read initially by eye, and then immediately by spectrophotometer. A result within 10% of the cut-off value in the Rotazyme II ELISA is suspect and repetition of the test is advised. However, in this study, such specimens were further examined by DEM and IEM.

Comparison with reference techniques

All specimens were examined by all the kits and PAGE. Only specimens which were negative by PAGE, but positive by one or more of the kits, were examined by DEM and

IEM. A specimen was considered positive if rotavirus was detected by any of PAGE. DEM or IEM.

Calculations: The formulae used for evaluating each kit were:

Sensitivity =
$$\frac{\text{True positive results detected}}{\text{Total true positive results}} \times 100$$

Specificity
$$= \frac{\text{True negative results detected}}{\text{Total true negative results}} \times 100$$

Results

Human specimens

There were 28 positive and 44 negative specimens. There was agreement with all tests for 18 of the 28 (64%) positive and 25 of the 44 (57%) negative specimens (Table 1). All the positive specimens were detected (i.e. 100% sensitivity) by all but one of the kits (Table 2). However, false positive results were obtained with 3 of the kits, and specificity ranged from 75–100%. ELISA results in these Tables refer to spectrophotometer readings. A virus other than rotavirus was cultured in 10 of the 44 negative specimens, but did not appear to affect the specificity. Small round viruses were seen in three rotavirus negative specimens.

Bovine specimens

There were 35 positive and 37 negative specimens. There was agreement with all tests for 13 of the 35 (37%) positive and 12 of the 37 (32%) negative specimens (Table 3). Both false positive and false negative results occurred more commonly than with the human specimens, with sensitivities of 40–91% and specificities of 49–97% (Table 4). ELISA results in these Tables refer to spectrophotometer readings. Coronavirus was seen in two and small round viruses in one of the negative specimens.

Method of reading ELISAs

The results of the ELISAs read by eye and spectrophotometer were compared (Table 5). There were 55 discrepancies between the visual and spectrophotometer results. Reading by eye tended to reduce slightly the sensitivity with both human and bovine specimens (Table 6). However, reading by eye tended to enhance considerably the specificity to the range 93 100% for human specimens and to 86% with bovine specimens.

Discussion

Many factors such as cost, speed and ease of use influence the choice of diagnostic tests, but the outstanding criteria must be satisfactory specificity and sensitivity.

When used on human facces, all the kits except Wellcome LA had sensitivies of 100%. The 64% sensitivity attained by Wellcome LA is too low a level of accuracy for results from this test to be interpreted with any confidence. Of the remaining five kits, three (RotaScreen and IDEIA ELISAs and RotaScreen LA) had specificities greater than 90%. These kits seem sufficiently accurate to be useful for human rotavirus diagnosis.

Reference		ELISA (spectrop	photometer result)		LA		Number of
techniques	RotaScreen	Wellcozyme	Rotazyme II	IDEIA	RotaScreen	Wellcome	specimens
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Table 1 Human enciments: Results of FUISA and 1 A kits compared with reference techniques. PAGF or FM

Key to symbols: + Positive results.
- Negative results.
* With Rotazyme II, one specimen in both of these groups gave a suspect result.
† One specimen contained small round viruses.

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Kit				Result			
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	I	Pos	Neg	Sensitivity (%)	Neg	Pos	Specificity (%)
ELISA:	RotaScreen	28	0	100	41	3	93
	Wellcozyme	28	0	100	33	Ξ	75
	Rotazyme II	28	0	001	35	6	80
	IDEIĂ	28	0	100	44	0	100
LA:	RotaScreen	28	0	100	44	0	100
	Wellcome	18	10	64	44	0	100
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Diagnosis of rotavirus infection

Reference		ELISA (spectrop	photometer result	()	L/	4	Number of
recnniques	RotaScreen	Wellcozyme	Rotazyme II	IDEIA	RotaScreen	Wellcome	specimens
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							Total 72
Kev to symbols – + Positive result - Negative result		* One specimen g † One specimen o ‡ One specimen o	ave a suspect readi ontained small rou ontained coronavir	ing with Rotaz nd viruses. 'us.	yme II.		
		§ Atypical rotavir	.ns.				

Table 3. Bovine specimens: Results of ELISA and LA kits compared with reference techniques. PAGE or EM

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Kit				Result			
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		Pos	Neg	Sensitivity (%)	Neg	Pos	Specificity (%)
ELISA:	RotaScreen	31	4	89	31	Q	84
	Wellcozymc	32	~,	91	28	ų	76
	Rotazymc II	31	4	89	28	6	76
	IDEIA	32	٣,	16	18	19	49
LA:	RotaScreen	59	ç	83	32	Ś	86
	Wellcome	14	21	40	36	_	97

Diagnosis of rotavirus infection

Table	5. Results of ELISA kits rea	d by eye	or spect	trophot	ometer	compare	d with 1	eferenc	e technic	ques. P∕	AGE or	EM	
							Res	ult					
Kit	Method of reading	R I	72 8 Positiv	Human es	specim 44	ens Negativ	/es	ň	72 5 Positive	Bovine	specime 37	ens Negativ	es
)	ЬA	by AGE or I	EM	PAG	by GE and	EM	ΡA	by GE or E	W	PA(by GE and 1	M
		Pos	Equiv	Neg	Neg	Equiv	Pos	Pos	Equiv	Neg	Neg	Equiv	Pos
RotaScreen	Eve	28	0	0	42	7	0	31	0	4	32	0	s.
	Spectrophotometer	28	0	0	41	0	m	31	0	4	31	0	9
Wellcozyme	Eve	27	0	1	41	m	0	28	0	7	31	_	ŝ
•	Spectrophotometer	28	0	0	33	0	Π	32	0	ŝ	28	0	6
Rotazvme II	Eve	26	7	0	34	7	m	29	2	4	30	0	Ś
	Spectrophotometer	28	0	0	33	2	6	31	0	4	27		6
IDEIA	Eye	28	0	0	4	0	0	29	0	9	32	0	S
	Spectrophotometer	28	0	0	4	0	0	32	0	ę	18	0	61

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K.	Sensitivity (%)		Specificity (%)	:	Sensitivity (%)		Specificity (%)	
	Spectrophotometer	Eye	Spectrophotometer	Eyc	Spectrophotometer	Eye	Spectrophotometer	Eye
RotaScreen	100	100	93	100	89	. 68	84	. 86
Wellcozyme	100	96	75	100	16	80	76	86
Rotazymc II	100	93	80	93	89	83	76	86
IDEIA	100	100	100	100	16	83	49	86

ctrophotometer 1 0...0 vd hoor red when Table 6. Sensitivity and specificity of ELISA kits

Diagnosis of rotavirus infection

Most of the kits performed noticeably less accurately in detecting rotavirus in calf faeces. It should be noted that the kits are not specifically designed for veterinary use, but they might be expected to detect group A bovine rotaviruses since they are serologically related to human group A rotaviruses. No kit had both sensitivity and specificity greater than 90%. Only the two kits (RotaScreen ELISA and LA) with sensitivities and specificities greater than 80% could be used with any confidence in herd or individual diagnosis.

Other studies have reported similar results on human specimens for IDEIA⁶. RotaScreen LA^{7.8}, Rotazyme II⁸ and Wellcome LA⁹. RotaScreen LA has also been found adequate for bovine rotavirus diagnosis¹⁰.

A recent report¹¹ assessing five commercially available rotavirus kits did not compare its results with any reference technique, but assumed a result to be a true positive when two or more of the kits being tested gave a positive result. This study reported false positives with Rotazyme II, Wellcozyme and the Wellcome LA. Our results suggest that this is an invalid assumption, especially with the ELISAs, where most of our false positive results occurred, and that they are probably underestimating the number of false positives. We are also surprised by the high number of false positives they obtained with the Wellcome LA. Our results on human specimens gave high specificity but low sensitivity and this is in agreement with another recent report⁹.

Others have also reported false positive results with Rotazyme $II^{8,9}$, and RotaScreen LA⁷ and false negative results with Rotazyme II^{12} .

All the ELISA kits are recommended to be read by spectrophotometer with visual reading possible, albeit with lower sensitivity, if suitable facilities do not exist. The superior specificities obtained by visual reading with all the ELISAs appear anomalous. This must be due to the ability of the spectrophotometer to detect low optical densities. An increase in the recommended cut-off point for some of these kits might eliminate this problem.

Of the ELISA kits tested, the specimen preparation and test procedure were simplest and quickest with IDEIA, all being completed within $3\frac{1}{2}$ hours. Poor specificities have often been noted in ELISAs¹³ which have been attributed to endogenous enzymes¹⁴ or cross-reacting substances¹⁵. Non-specific reactions may account for those specimens which were positive by most tests but negative by our reference techniques. It has been found necessary with specimens from both man¹³ and calves (unpublished observations) to include a blocking antiserum to eliminate false positive results.

LA kits were significantly quicker (less than 30 minutes) and easy to use on small numbers of specimens, but proved time-consuming when many specimens were examined. LAs are read visually, thus contributing a significant subjective element. It is likely that they involve significant inter-operator differences, although in this study particular care was taken to adhere rigorously to the instructions, with continuous rocking of the test card and reading the result at a maximum of two minutes.

Cost per specimen also varied considerably, from approximately $\pounds 1$ for both LAs, $\pounds 2$ for Wellcozyme, RotaScreen and IDEIA ELISAs to $\pounds 4$ for Rotazyme II ELISA. By contrast, the cost of consumables per specimen for PAGE is approximately $\pounds 0.25$.

The prevalence of atypical (non-group A) rotaviruses is less than 1% in the human and bovine populations under study¹⁶, and atypical rotavirus infections in man are commonly detected only in China^{17,18}. There is no serological relationship between group A and atypical rotaviruses¹⁹. The positive results with some kits and the atypical bovine rotaviruses in our study may have been fortuitous. However, a specific reaction due to the serendipitous occurrence of antibodies to atypical rotaviruses in the kit antisera is possible.

Conclusions

Several of the kits tested had specificity and sensitivity greater then 90% on the human specimens, and these (RotaScreen and IDEIA ELISAs and RotaScreen LA) are suitable for the diagnosis of human rotavirus infections. With the bovine specimens, only RotaScreen ELISA and LA, with both specificity and sensitivity over 80%, are suitable for the diagnosis of bovine rotavirus infection. The main advantage of diagnosis by the kits is speed, but specificity and sensitivity should not be sacrificed for this. PAGE is sensitive, specific, cheap, simple to perform, detects all groups of rotavirus, and in our opinion, is the method of choice for diagnosing rotavirus infection.

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References

- 1. Flores J. Nakagomi O, Nakagomi T *et al.* The role of rotaviruses in pediatric diarrhoea. Pediatr Infect Dis 1986; 5 (Suppl 1): S53–62.
- 2. Cukor G, Blacklow NR. Human viral gastroenteritis. Microbiol Rev 1984; 48: 157-79.
- 3. McNulty MS. Rotaviruses. J Gen Virol 1978; 40: 1-18.
- Herring AJ, Inglis NF, Ojeh CK, Snodgrass DR, Menzies JD. Rapid diagnosis of rotavirus infection by direct detection of viral nucleic acid in silver-stained polyacrylamide gels. J Clin Microbiol 1982; 16: 473–7.
- 5. Roberts IM, Harrison BD. Detection of potato leafroll and potato mop-top viruses by immunosorbent microscopy. Ann Appl Biol 1979; 93: 289–97.
- 6. Westmoreland D, Ashley CR, Caul EO. Rapid and reliable routine diagnosis of rotavirus using a commercial monoclonal antibody based immunoassay. J Clin Path 1987; 40: 1384–6.
- 7. Bryden AS. An evaluation of commercial latex agglutination kits for rotavirus detection. Serodiag Immunother Infect Dis 1987; 1: 131–9.
- 8. de Vries WP, Houben AW, Stobberingh EE. Comparison of four commercial assays for detection of rotavirus in childhood gastroenteritis. Eur J Clin Microbiol 1986; 5: 542-4.
- 9. Dennehy PH, Gauntlett DR, Tente WE. Comparison of nine commercial immunoassays for the detection of rotavirus in faecal specimens. J Clin Microbiol 1988; 26: 1630–4.
- Herbst W, Lange H, Zschöck M, Krauss H. Nachweis von kälberrotavirus mit dem latextest RotaScreen und dem elektronenmikroskopeine vergleichende untersuchung. DTW 1986; 93: 317-19.
- 11. Jenkins CT. An evaluation of five commercially available kits for the diagnosis of rotavirus infection. Serodiag Immunother Infect Dis 1988; 2: 137-41.
- 12. Martin AL, Follett EAC. An assessment of the sensitivity of three methods for the detection of rotavirus. J Virol Methods 1987; 16: 39-44.
- 13. Brandt CD, Kim HW, Rodriguez WJ et al. Comparison of direct electron microscopy, immune electron microscopy and rotavirus enzyme-linked immunosorbent assay for detection of gastroenteritis viruses in children. J Clin Microbiol 1981; 13: 976–81.
- 14. Beards GM. Endogenous alkaline phosphatase is a cause of non-specific reactions in enzyme-

immunoassays for rotavirus based on alkaline phosphatase conjugates. Med Lab Sci 1988; 45: 97-8.

- 15. Yolken RH, Stopa PJ. Analysis of nonspecific reactions in enzyme-linked immunosorbent assay testing for human rotavirus. J Clin Microbiol 1976; 10: 703-7.
- 16. Snodgrass DR, Herring AJ, Campbell I, Inglis JM, Hargreaves FD. Comparison of atypical rotaviruses from calves, piglets, lambs and man. J Gen Virol 1984; 65: 909–14.
- 17. Hung T, Chen G, Wang C *et al.* Waterborne outbreak of rotavirus diarrhoea in adults in China caused by a novel rotavirus. Lancet 1984; i: 1139–42.
- 18. Chen G, Hung T, Bridger JC, McCrae MA. Chinese adult rotavirus is a group B rotavirus. Lancet 1985; ii: 1123-4.
- 19. McCrae MA. Nucleic acid-based analyses of non-group A rotaviruses. In: Bock G, Whelan J, eds. Novel diarrhoea viruses. Chichester: John Wiley and Sons, 1987: 24–48.

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