

Review Article

Volatile uptake, transport, perception, and signaling shape a plant's nose

 Lei Wang and  Matthias Erb

Institute of Plant Sciences, University of Bern, Altenbergrain 21, 3013 Bern, Switzerland

Correspondence: Lei Wang (lei.wang@ips.unibe.ch) or Matthias Erb (matthias.erb@ips.unibe.ch)



Herbivore-induced plant volatiles regulate defenses in undamaged neighboring plants. Understanding the mechanisms by which plant volatiles are taken up, perceived, and translated into canonical defense signaling pathways is an important frontier of knowledge. Volatiles can enter plants through stomata and the cuticle. They are likely perceived by membrane-associated receptors as well as intracellular receptors. The latter likely involves metabolization and transport across cell membranes by volatile transporters. Translation of volatiles into defense priming and induction typically involves mitogen-activated protein kinases (MAPKs), WRKY transcription factors, and jasmonates. We propose that the broad range of molecular processes involved in volatile signaling will likely result in substantial spatiotemporal and ontogenetic variation in plant responsiveness to volatiles, with important consequences for plant–environment interactions.

Introduction

Plants produce and emit volatile organic compounds to mediate interactions with other organisms [1]. Upon insect herbivory, plants emit a complex blend of herbivore-induced plant volatiles [2,3]. These plant volatiles typically include green leaf volatiles (GLVs), terpenoids, and phenylpropanoid/benzenoid volatiles. Based on the plant species and the herbivores that trigger the emission, the volatile blends differ in composition, quantity, and timing. Herbivore-induced plant volatiles play critical roles shaping the interactions between plant–insect herbivores, directly or indirectly. They function directly by triggering or mediating defense response in plants, or by acting as toxins or repellents against herbivores. They also function indirectly by attracting the natural enemies of insect herbivores [2–4].

A particularly interesting function of herbivore-induced plant volatiles is their ability to mediate defense in the systemic undamaged tissues or neighboring undamaged plants. [5–7]. Many volatiles have been shown capable of mediating interactions between plants (see a recent review for a comprehensive summary) [8]. Other than the phytohormone precursors MeJA and MeSA and the volatile hormone ethylene, GLVs are likely the most conserved volatile signals mediating plant–plant interactions. They trigger an array of defense responses in many plants, such as *Arabidopsis*, tomato, lima bean, and maize [9]. GLVs treatment in maize triggers the expression of defense-related genes and biosynthesis of defense-related metabolites, including many plant volatiles [10,11]. The emission of these volatiles further contributes to indirect defense [2]. In sweet potato, the homoterpene (E)-4,8-dimethyl-1,3,7-nonatriene (DMNT) can induce expression of proteinase inhibitor genes and increase herbivore resistance [12]. Volatiles such as indole and linalool typically do not induce defense directly, but they can prime several plants for stronger defense upon insect herbivory [11,13,14].

Recent years have seen substantial progress in understanding the biosynthesis, emission, and ecological function of plant volatiles. The perception of plant volatiles is now also being unraveled [1–3,15]. To fully understand how volatiles mediate plant–plant interactions, it is important to address how volatiles enter plant tissues and get recognized thereafter. A few recent reviews have summarized the latest discoveries in volatile biosynthesis, emission, and bioactivity [1,8,16]. Here, we discuss the possible paths

Received: 21 February 2022
Revised: 04 July 2022
Accepted: 20 July 2022

Version of Record published:
30 September 2022

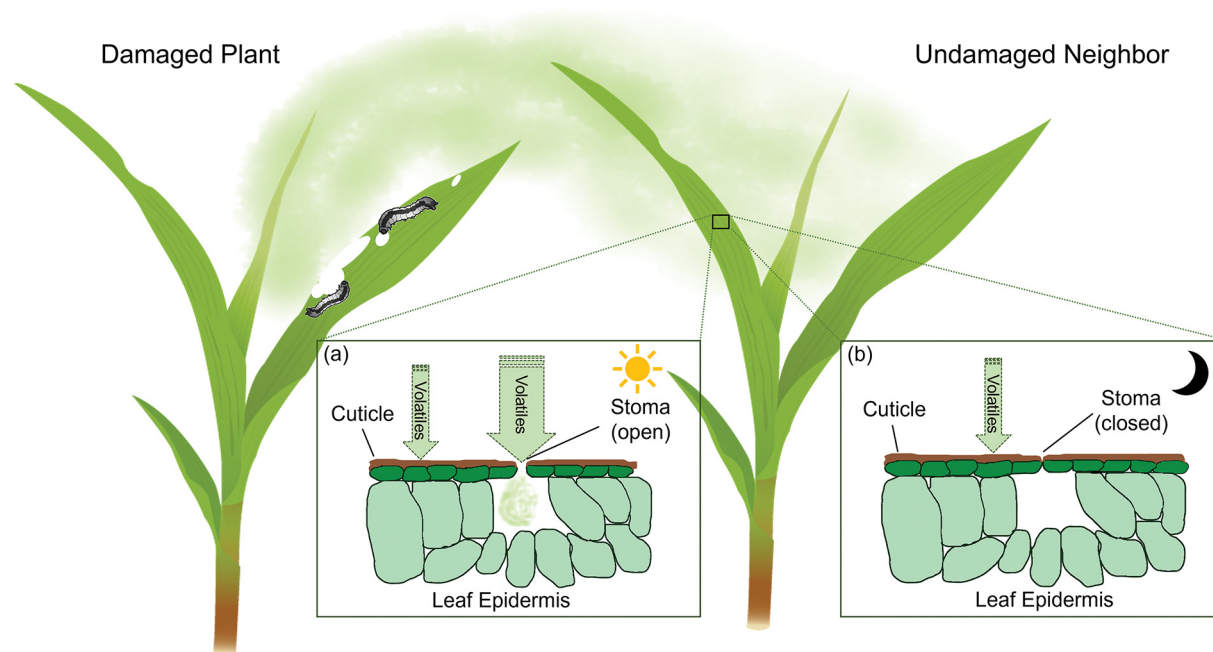


Figure 1. Schematic view of possible volatile entry routes into leaf tissues

Herbivore-induced volatiles emitted from damaged plants may enter the plant tissue of undamaged neighbors through stomata and/or diffuse through the cuticle. (a) During daytime, stomata may be the main entry sites due to low resistance. (b) At night, when the stomata are closed, diffusion through the cuticle may become more important.

that volatiles may take to enter leaf tissues. We review the current evidence regarding the role of receptors/receptor complexes in volatile perception, with a focus on herbivore-induced plant volatiles. We further summarize the downstream signaling events that are triggered by herbivore-induced plant volatiles, and explore the hypothesis that spatiotemporal variation in volatile uptake, transport, perception, and signaling shape a plant's 'nose', i.e. the tissues that are involved in perceiving volatiles as environmental cues. Pathogen-induced volatiles are covered in a separate paper in the same issue by Vlot and colleagues [17].

Entry of volatiles into plant leaves

To trigger defense response in the cells, volatiles need to access the plasma membrane or intracellular compartments. Volatiles may enter the leaves either through stomata or pass through the cuticular wax layer (Figure 1).

Entry through stomata

Stomata are the breathing pores of plant leaves, balancing photosynthetic carbon dioxide uptake and evaporative water loss [18]. These natural openings provide entry low resistance points for environmental agents such as microbes [19]. Recent studies show that stomata also shape plant–insect interactions by controlling volatile emission [20–22]. In maize, stomatal closure, induced by darkness or abscisic acid (ABA) treatment, constrains the emission of elicitor-triggered sesquiterpenes. Maize plants induced by elicitors in the dark show a burst of sesquiterpenes emission when light is switched on, indicating stomata as the gate for these volatiles to be released into the atmosphere [20]. Similarly, tomato and soybean leaves emit less volatiles when the stomata are partially closed by glucose oxidase, a salivary protein from the caterpillar *Helicoverpa zea* [21].

Given the considerations above, it is reasonable to assume that stomata also serve as low-resistance entry points for volatiles. Volatiles from attacked plants may thus enter the leaves of neighboring plants through these openings. If stomata are the main entry path for plant volatiles, several aspects need to be considered. First, as plants close their stomata at night [23], this would imply that plants are not able to perceive volatiles during nighttime, despite the fact that herbivore-induced plant volatiles such as GLVs are released as danger cues at night [24]. Another important aspect to consider is the developmental stage of stomata in leaves. In grasses, for instance, the stomatal complex is under differentiation and formation in the developmental zone in the young leaves [18]. Thus, if stomata are important for

volatile uptake, the ability of young leaves to respond to volatiles will differ from that of the old leaves, where stomata are fully developed. Experiments that investigate diurnal and developmental variation in volatile perception could thus provide first indications on the potential role of stomata in volatile perception.

Entry through the cuticle

The plant cuticle is the final barrier for volatiles to be released into atmosphere from nonvegetative organs [16]. In petunia flowers, the cuticle acts both as a resistance barrier and a sink for VOCs. Its thickness thus affects the dynamics of volatile emission [25]. This effect also depends on the physiochemical properties of VOCs, with volatiles having lower ambient vapor pressure facing higher resistance [16,26]. Several plants use their leaves to adsorb/take up volatiles from neighboring plants for enhanced herbivore resistance [27]. A recent study shows that plant leaf cuticular waxes can sequester exogenous volatiles [28]. Thus, it can be hypothesized that plant volatiles may pass the leave cuticle and diffuse across the more permeable cell wall to reach the plasma membrane, effectively bypassing stomata.

Plant cuticular waxes comprise mainly very long-chain fatty acids and their derivatives. Both the wax composition and structure change greatly during leaf development. In wheat, leaf surface wax keeps accumulating until the leaf blade finishes expanding. Meanwhile, the carbon chain length of the wax constituents increases drastically, and the wax crystals form different structure [29]. These changes likely cause different volatile permeability in young leaves and old leaves. Different physiochemical properties of volatiles will further increase the variance. So far, responses in volatile perception are either analyzed on the whole plant level or on a specific leaf. Experiments with cuticle mutants will help to uncover the role of cuticle as entry sites for volatiles relative to stomata.

Volatile perception at the cell membrane

Once accessing the plasma membrane, VOCs may be perceived by receptor/receptor complexes to trigger cellular response or taken up for further metabolic processing. Finding plant volatile receptors has been a long-standing question, but a breakthrough has yet to be achieved [3,15]. Ample progress has been made on identifying receptors for microbe-associated molecular patterns (MAMPs), herbivore-associated molecular patterns (HAMPs), and damage-associated patterns (DAMPs) [30,31]. These discoveries may inspire the discovery of a plant volatile receptors.

GLVs, C6 aldehydes, alcohols, and esters are enzymatically generated from membrane lipids upon disruption of membrane integrity in leaves, upon mechanical wounding or insect feeding [9]. Based on the plant origin and their ability to induce defense, these fatty acid-derived molecules can also be termed as DAMPs [32]. In *Arabidopsis*, the lectin receptor kinase LORE recognizes bacterial medium-chain 3-hydroxy fatty acid as a MAMP to trigger immunity [33]. Another *Arabidopsis* lectin receptor kinase LecRK-I.8 is critical for the defense triggered by phosphatidylcholines primarily with C16- to C18-fatty acyl chains [34]. The maize ZmFACS protein, a leucine-rich repeat (LRR) receptor kinase, mediates defense triggered by the fatty acid–amino acid conjugates (FACs) [35]. GLVs, as fatty acid derivatives, share certain biochemical properties with the molecular patterns mentioned above. Thus, they may be perceived by plasma membrane-localized receptor kinases or receptor proteins as well. Typically, these receptors need to form a protein complex with coreceptors to achieve full immune responses. These coreceptors are often from the somatic embryogenesis receptor kinase (SERKs) family [31]. Thus, screening lectin/LRR receptor kinase/receptor protein mutants and SERKs mutants for abolished or reduced GLVs response may help identifying components of the hypothesized GLVs receptor complex.

Volatile uptake into cells

MAMPs and DAMPs receptors are plasma membrane-localized receptors [31]. Perception of danger-related molecules can also happen inside of plant cells, such as the perception of effectors by nucleotide-binding domain leucine-rich repeat-containing (NLR) proteins [36]. It is possible that perception of some VOCs happens intracellularly. Additionally, plants take up volatiles directly for further metabolic processing [27]. In either case, these volatiles need to pass through the plasma membrane. Direct diffusion may happen for membrane lipid derived volatiles such as GLVs. This is unlikely for most other volatiles [16]. In petunia, transport of VOCs across the plasma membrane relies on an adenosine triphosphate-binding cassette (ABC) transporter [37]. Similar volatile transporters may mediate the channeling of volatiles from the extracellular into the intracellular space. Transporters or ion channels may act as receptors as well. For example, the *Arabidopsis* anion channel SLAC1 plays an important role in sensing CO₂/bicarbonate in the guard cells [38]. Within the cells, volatiles may bind specific proteins to initiate cellular

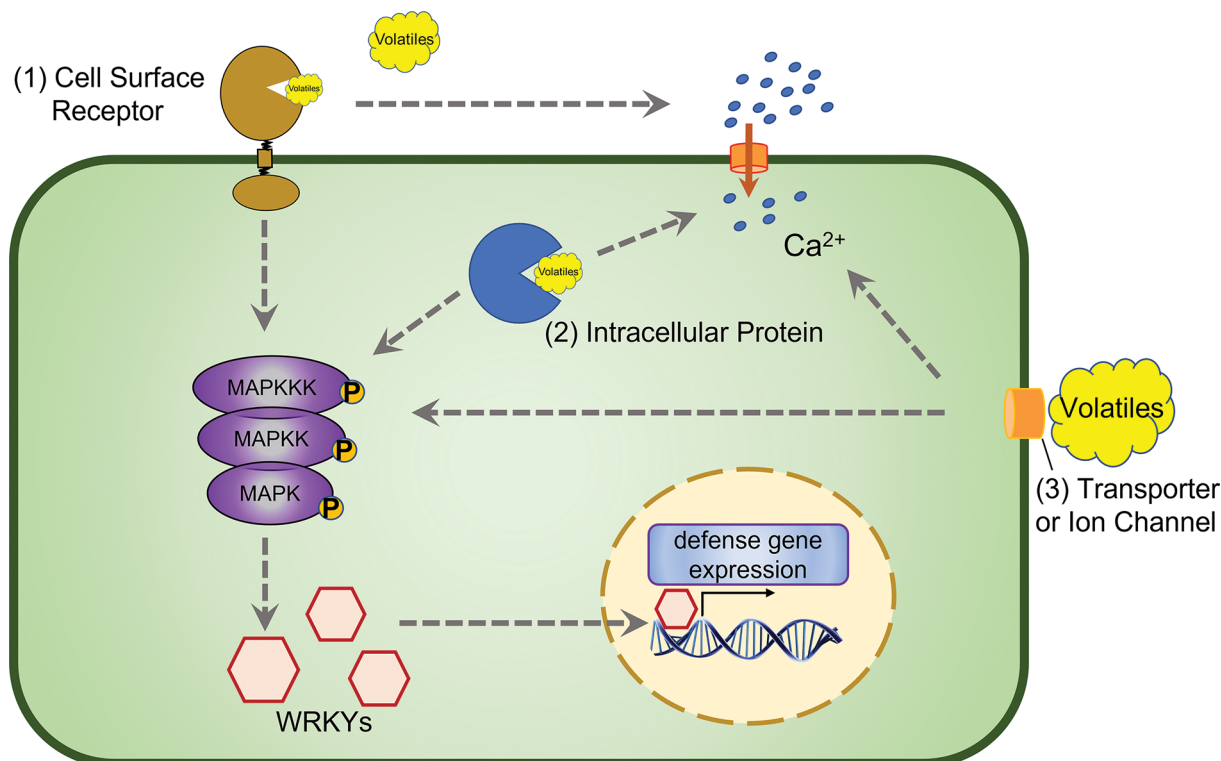


Figure 2. Schematic representation of plant volatile perception and signaling

Plant volatiles are likely perceived in three different ways. The respective responsible proteins are: (1) cell surface receptors, (2) intracellular proteins, and (3) volatile transporters or ion channels. Volatile perception leads to a cascade of defense responses, including calcium influx, MAPK activation, and WRKY transcription factor-regulated expression of defense genes. Dashed arrows indicate unclear molecular mechanisms of the signaling cascade.

responses. Recently, the transcription regulators TOPLESS-like proteins (TPLs) were found to bind the sesquiterpene caryophyllene [39]. However, it is unclear whether TPLs alone are sufficient to act as receptors to transduce caryophyllene-triggered responses. So far, hypothetical plant odorant-binding proteins (OBPs) have also been proposed to act as plant volatile receptors, based on their similarity with animal OBPs. A recent study used *in silico* molecular docking to prove plant OBPs can bind monoterpenes [40]. The specificity and *in vivo* activity of OBPs remains to be uncovered.

Translation into defense signaling

Volatiles are well established to trigger defense pathways that are typically associated with MAMPs and DAMPs [8,30] (Figure 2).

Upon exposure to GLVs, *Arabidopsis* and tomato plants show a cytosolic calcium influx. In *Arabidopsis*, this transient influx happens rapidly upon GLVs exposure and peaks at 5–10 min [41]. The calcium influx kinetic is unclear in tomato likely due to the lack of a calcium reporter line [42]. Additionally, ocimene, myrcene, pinene, and DMNT can also trigger transient cytosolic calcium influx in *Arabidopsis*, similar as the one triggered by GLVs [41]. Calcium influx is a typical early response, following the perception of MAMPs and DAMPs. NLR activation after effector recognition also leads to calcium influx, *albeit* with different dynamics [36]. Collectively, these studies indicate calcium influx is a conserved immune response upon danger perception, including volatiles.

Activation of mitogen-activated protein kinases (MAPKs) is another classical early immune response upon pattern recognition [43]. Similarly, GLV exposure leads to rapid activation of MAPKs in the grass *Lolium temulentum* [44]. The sesquiterpene (E)-Nerolidol increases both the transcript and protein of MAPK in tea plants [45]. Indole exposure does not activate MAPKs directly but increases MAPK gene expression and activation following simulated herbivory in rice. Knocking down *MPK3* and *MPK6* leads to abolished or greatly reduced defense priming effect by indole [46]. MAPK cascades are also points of convergence between different signaling pathways [47]. This role may explain the

phenomenon that dual exposure of the GLV (Z)-3-hexenyl-acetate (HAC) and indole in maize generates stronger defense than HAC exposure alone [11].

Other commonly reported volatile responses include the increased expression of genes-encoding transcription factors, defense hormone biosynthesis enzymes, and defense metabolite biosynthesis enzymes [8]. Members of the WRKY transcription factor family are often induced by various biotic and abiotic stress and regulate hormone biosynthesis in turn [48,49]. In *Arabidopsis*, the GLV (E)-2-hexenal induces the expression of several WRKY genes, including *AtWRKY6*, *AtWRKY40*, and *AtWRKY53*. Knocking out these genes leads to increased expression of (E)-2-hexenal-specific responsive genes, indicating that these transcription factors as negative regulators of GLVs signaling in *Arabidopsis* [48]. The GLV (Z)-3-hexenol increases *ZmWRKY12* transcripts in maize, but the importance of this phenomenon is unclear [50]. Similarly, *CsWRKY3* expression is up-regulated by (E)-Nerolidol but its role in (E)-Nerolidol signaling is unknown [45].

Plant VOCs may also suppress defense in some cases [14,51]. Whitefly-infested tomato plants emit a unique blend of volatiles to suppress JA-dependent defense but prime SA-dependent defense. The dual role in defense is triggered by two terpenes: β -myrcene or β -caryophyllene. On the contrary, linalool, a monoterpene elicited by the *Spodoptera exigua* caterpillar attacked tomato plants, primes the expression of two JA-pathway proteinase inhibitor genes, *PI-I* and *PI-II* [14]. The molecular mechanisms behind these differences are unknown. Future work on how volatile-triggered defense is differentially regulated will greatly help to uncover novel volatile signaling components and pathways.

Spatiotemporal patterns of volatile perception

Since the discovery of ‘talking trees’, plant volatile-mediated plant–plant interactions have fascinated many scientists and led to substantial research efforts [5]. We now have a detailed understanding of plant volatile biosynthesis and transport [1,16]. Based on this knowledge, we can infer that plant volatile perception likely involves a number of physical structures such as stomata and cuticles as well as molecular elements such as transporters, receptors, and signal integration proteins. Given the substantial variation in the expression of these elements in different plant parts and developmental stages, we predict that plant volatile perception will not be uniform, but will show significant variation within a given plant. Certain leaves are likely to be much more sensitive to others, and could thus be viewed as a plants ‘nose’. Understanding these patterns and linking them to our increased understanding of the mechanisms of plant volatile perception will be important to unravel the ecological dynamics that are elicited by volatiles and to exploit plant volatiles as crop-reprogramming signals. As comprehensively summarized in a recent review, the potential applications of plant volatiles for sustainable agricultural practices will include breeding crops with enhanced volatile emissions and inducing volatile release in a targeted manner [2]. A good understanding of the mechanisms and spatiotemporal variation in volatile perception will also facilitate the selection and breeding of plants that are sensitive to plant volatiles in the right place at the right time. Such work will help to unlock the potential of plant volatiles as crop-reprogramming agents.

Summary

- Plant volatiles may enter the inner space of plant leaves through stomata or cuticle. The relative importance of these entry sites is likely to vary with development and environmental conditions.
- Plant volatile perception is likely mediated by cell surface receptors, plasma membrane-localized transporters or ion channels, and intracellular proteins.
- Plant volatiles regulate canonical defense signaling pathways, with MAPKs and WRKY transcription factors playing important roles as signal integration hubs.
- Spatiotemporal variation in volatile uptake and perception elements will likely determine where and when plants respond to volatiles.

Competing Interests

The authors declare that there are no competing interests associated with the manuscript.

Funding

L.W. is funded by the Horizon 2020 Marie Skłodowska-Curie Actions (886651). M.E. is funded by the European Research Council (ERC) under the European Union's Horizon 2020 Research and Innovation Programme (ERC-2016-STG 714239), the Swiss National Science Foundation (Grant Nr. 200355), The Velux Stiftung (Grant Nr. 1231) and the University of Bern.

Author Contributions

L.W. conceived the manuscript, wrote the first draft and edited the final version. M.E. conceived the manuscript, helped write the first draft and edited the final version.

Acknowledgements

The authors would like to thank two anonymous reviewers for their helpful comments on an earlier version of the manuscript.

Abbreviations

ABA, abscisic acid; ABC, adenosine triphosphate-binding cassette; DAMP, damage-associated molecular pattern; DMNT, (E)-4,8-dimethyl-1,3,7-nonatriene; FAC, fatty acid–amino acid conjugate; GLV, green leaf volatile; HAC, (Z)-3-hexenyl-acetate; HAMP, herbivore-associated molecular pattern; JA, jasmonic acid; LRR, leucine-rich repeat; MAMP, microbe-associated molecular pattern; MAPK, mitogen-activated protein kinase; NLR, nucleotide-binding, leucine-rich repeat; OBP, odorant-binding protein; SA, salicylic acid; SERK, somatic embryogenesis receptor kinase; TPL, TOPLESS-like proteins; VOC, volatile organic compound.

References

- Dudareva, N., Klempien, A., Muhlemann, J.K. and Kaplan, I. (2013) Biosynthesis, function and metabolic engineering of plant volatile organic compounds. *New Phytol.* **198**, 16–32, <https://doi.org/10.1111/nph.12145>
- Turlings, T.C.J. and Erb, M. (2018) Tritrophic interactions mediated by herbivore-induced plant volatiles: mechanisms, ecological relevance, and application potential. *Annu. Rev. Entomol.* **63**, 433–452, <https://doi.org/10.1146/annurev-ento-020117-043507>
- Heil, M. (2014) Herbivore-induced plant volatiles: targets, perception and unanswered questions. *New Phytol.* **204**, 297–306, <https://doi.org/10.1111/nph.12977>
- Ton, J., D'Alessandro, M., Jourdie, V., Jakab, G., Karlen, D., Held, M. et al. (2007) Priming by airborne signals boosts direct and indirect resistance in maize. *Plant J.* **49**, 16–26, <https://doi.org/10.1111/j.1365-3113X.2006.02935.x>
- Baldwin, I.T., Halitschke, R., Paschold, A., von Dahl, C.C. and Preston, C.A. (2006) Volatile signaling in plant-plant interactions: “talking trees” in the genomics era. *Science* **311**, 812–815, <https://doi.org/10.1126/science.1118446>
- Moreira, X. and Abdala-Roberts, L. (2019) Specificity and context-dependency of plant-plant communication in response to insect herbivory. *Curr. Opin. Insect. Sci.* **32**, 15–21, <https://doi.org/10.1016/j.cois.2018.09.003>
- Karban, R. (2021) Plant communication. *Annu. Rev. Ecol. Evol. Syst.* **52**, 1–24, <https://doi.org/10.1146/annurev-ecolsys-010421-020045>
- Brosset, A. and Blande, J.D. (2022) Volatile-mediated plant-plant interactions: volatile organic compounds as modulators of receiver plant defence, growth, and reproduction. *J. Exp. Bot.* **73**, 511–528, <https://doi.org/10.1093/jxb/erab487>
- Ameje, M., Allmann, S., Verwaeren, J., Smaghe, G., Haesaert, G., Schuurink, R.C. et al. (2018) Green leaf volatile production by plants: a meta-analysis. *New Phytol.* **220**, 666–683, <https://doi.org/10.1111/nph.14671>
- Engelberth, J., Alborn, H.T., Schmelz, E.A. and Tumlinson, J.H. (2004) Airborne signals prime plants against insect herbivore attack. *Proc. Natl. Acad. Sci. U.S.A.* **101**, 1781–1785, <https://doi.org/10.1073/pnas.0308037100>
- Hu, L., Ye, M. and Erb, M. (2019) Integration of two herbivore-induced plant volatiles results in synergistic effects on plant defence and resistance. *Plant Cell Environ.* **42**, 959–971, <https://doi.org/10.1111/pce.13443>
- Meents, A.K., Chen, S.-P., Reichelt, M., Lu, H.-H., Bartram, S., Yeh, K.-W. et al. (2019) Volatile DMNT systemically induces jasmonate-independent direct anti-herbivore defense in leaves of sweet potato (*Ipomoea batatas*) plants. *Sci. Rep.* **9**, 17431, <https://doi.org/10.1038/s41598-019-53946-0>
- Erb, M., Veyrat, N., Robert, C.A.M., Xu, H., Frey, M., Ton, J. et al. (2015) Indole is an essential herbivore-induced volatile priming signal in maize. *Nat. Commun.* **6**, 6273, <https://doi.org/10.1038/ncomms7273>
- Zhang, P.-J., Wei, J.-N., Zhao, C., Zhang, Y.-F., Li, C.-Y., Liu, S.-S. et al. (2019) Airborne host-plant manipulation by whiteflies via an inducible blend of plant volatiles. *Proc. Natl. Acad. Sci. U.S.A.* **116**, 7387–7396, <https://doi.org/10.1073/pnas.1818599116>
- Loreto, F. and D'Auria, S. (2022) How do plants sense volatiles sent by other plants? *Trends Plant Sci.* **27**, 29–38, <https://doi.org/10.1016/j.tplants.2021.08.009>
- Widhalm, J.R., Jaini, R., Morgan, J.A. and Dudareva, N. (2015) Rethinking how volatiles are released from plant cells. *Trends Plant Sci.* **20**, 545–550, <https://doi.org/10.1016/j.tplants.2015.06.009>
- Eccleston, L., Brambilla, A. and Vlot, A.C. (2022) New molecules in plant defence against pathogens. *Essays Biochem.*, <https://doi.org/10.1042/EBC20210076>
- Nunes, T.D.G., Zhang, D. and Raissig, M.T. (2020) Form, development and function of grass stomata. *Plant J.* **101**, 780–799, <https://doi.org/10.1111/tpj.14552>

- 19 Melotto, M., Zhang, L., Oblessuc, P.R. and He, S.Y. (2017) Stomatal defense a decade later. *Plant Physiol.* **174**, 561–571, <https://doi.org/10.1104/pp.16.01853>
- 20 Seidl-Adams, I., Richter, A., Boomer, K.B., Yoshinaga, N., Degenhardt, J. and Tumlinson, J.H. (2015) Emission of herbivore elicitor-induced sesquiterpenes is regulated by stomatal aperture in maize (*Zea mays*) seedlings. *Plant Cell Environ.* **38**, 23–34, <https://doi.org/10.1111/pce.12347>
- 21 Lin, P.-A., Chen, Y., Chaverro-Rodriguez, D., Heu, C.C., Zainuddin, N.B., Sidhu, J.S. et al. (2021) Silencing the alarm: an insect salivary enzyme closes plant stomata and inhibits volatile release. *New Phytol.* **230**, 793–803, <https://doi.org/10.1111/nph.17214>
- 22 Lin, P.-A., Chen, Y., Ponce, G., Acevedo, F.E., Lynch, J.P., Anderson, C.T. et al. (2021) Stomata-mediated interactions between plants, herbivores, and the environment. *Trends Plant Sci.*
- 23 Caird, M.A., Richards, J.H. and Donovan, L.A. (2007) Nighttime stomatal conductance and transpiration in C3 and C4 plants. *Plant Physiol.* **143**, 4–10, <https://doi.org/10.1104/pp.106.092940>
- 24 de Moraes, C.M., Mescher, M.C. and Tumlinson, J.H. (2001) Caterpillar-induced nocturnal plant volatiles repel conspecific females. *Nature* **410**, 577–580, <https://doi.org/10.1038/35069058>
- 25 Liao, P., Ray, S., Boachon, B., Lynch, J.H., Deshpande, A., McAdam, S. et al. (2021) Cuticle thickness affects dynamics of volatile emission from petunia flowers. *Nat. Chem. Biol.* **17**, 138–145, <https://doi.org/10.1038/s41589-020-00670-w>
- 26 Matsui, K. (2016) A portion of plant airborne communication is endorsed by uptake and metabolism of volatile organic compounds. *Curr. Opin. Plant Biol.* **32**, 24–30, <https://doi.org/10.1016/j.pbi.2016.05.005>
- 27 Sugimoto, K., Matsui, K. and Takabayashi, J. (2016) Uptake and conversion of volatile compounds in plant–plant communication. In *Deciphering Chemical Language of Plant Communication* (Blande, J.D. and Glinwood, R., eds), pp. 305–316, Springer International Publishing, Cham, https://doi.org/10.1007/978-3-319-33498-1_13
- 28 Camacho-Coronel, X., Molina-Torres, J. and Heil, M. (2020) Sequestration of exogenous volatiles by plant cuticular waxes as a mechanism of passive associational resistance: a proof of concept. *Front. Plant Sci.* **11**, 121, <https://doi.org/10.3389/fpls.2020.00121>
- 29 Wang, Y., Wang, J., Chai, G., Li, C., Hu, Y., Chen, X. et al. (2015) Developmental changes in composition and morphology of cuticular waxes on leaves and spikes of glossy and glaucous wheat (*Triticum aestivum* L.). *PLoS ONE* **10**, e0141239, <https://doi.org/10.1371/journal.pone.0141239>
- 30 Erb, M. and Reymond, P. (2019) Molecular interactions between plants and insect herbivores. *Annu. Rev. Plant Biol.* **70**, 527–557, <https://doi.org/10.1146/annurev-arplant-050718-095910>
- 31 Albert, I., Hua, C., Nürnberger, T., Pruitt, R.N. and Zhang, L. (2020) Surface sensor systems in plant immunity. *Plant Physiol.* **182**, 1582–1596, <https://doi.org/10.1104/pp.19.01299>
- 32 Meents, A.K. and Mithöfer, A. (2020) Plant-plant communication: is there a role for volatile damage-associated molecular patterns? *Front. Plant Sci.* **11**, 583275, <https://doi.org/10.3389/fpls.2020.583275>
- 33 Kutschera, A., Dawid, C., Gisch, N., Schmid, C., Raasch, L., Gerster, T. et al. (2019) Bacterial medium-chain 3-hydroxy fatty acid metabolites trigger immunity in Arabidopsis plants. *Science* **364**, 178–181, <https://doi.org/10.1126/science.aau1279>
- 34 Stahl, E., Brillatz, T., Ferreira Queiroz, E., Marcourt, L., Schmiesing, A., Hilfiker, O. et al. (2020) Phosphatidylcholines from *Pieris brassicae* eggs activate an immune response in Arabidopsis. *Elife* **9**, e60293
- 35 Poretsky, E., Ruiz, M., Ahmadian, N., Steinbrenner, A.D., Dressano, K., Schmelz, E.A. et al. (2021) Comparative analyses of responses to exogenous and endogenous antiherbivore elicitors enable a forward genetics approach to identify maize gene candidates mediating sensitivity to herbivore-associated molecular patterns. *Plant J.* **108**, 1295–1316
- 36 Yuan, M., Ngou, B.P.M., Ding, P. and Xin, X.-F. (2021) PTI-ETI crosstalk: an integrative view of plant immunity. *Curr. Opin. Plant Biol.* **62**, 102030
- 37 Adebessin, F., Widhalm, J.R., Boachon, B., Lefèvre, F., Pierman, B., Lynch, J.H. et al. (2017) Emission of volatile organic compounds from petunia flowers is facilitated by an ABC transporter. *Science* **356**, 1386–1388
- 38 Zhang, J., Wang, N., Miao, Y., Hauser, F., McCammon, J.A., Rappel, W.-J. et al. (2018) Identification of SLAC1 anion channel residues required for CO₂/bicarbonate sensing and regulation of stomatal movements. *Proc. Natl. Acad. Sci. U.S.A.* **115**, 11129–11137
- 39 Nagashima, A., Higaki, T., Koeduka, T., Ishigami, K., Hosokawa, S., Watanabe, H. et al. (2019) Transcriptional regulators involved in responses to volatile organic compounds in plants. *J. Biol. Chem.* **294**, 2256–2266
- 40 Giordano, D., Facchiano, A., D’Auria, S. and Loreto, F. (2021) A hypothesis on the capacity of plant odorant-binding proteins to bind volatile isoprenoids based on in silico evidences. *Elife* **10**, e66741
- 41 Asai, N., Nishioka, T., Takabayashi, J. and Furuichi, T. (2009) Plant volatiles regulate the activities of Ca²⁺-permeable channels and promote cytoplasmic calcium transients in Arabidopsis leaf cells. *Plant Signal. Behav.* **4**, 294–300, <https://doi.org/10.4161/psb.4.4.8275>
- 42 Zebelo, S.A., Matsui, K., Ozawa, R. and Maffei, M.E. (2012) Plasma membrane potential depolarization and cytosolic calcium flux are early events involved in tomato (*Solanum lycopersicon*) plant-to-plant communication. *Plant Sci.* **196**, 93–100, <https://doi.org/10.1016/j.plantsci.2012.08.006>
- 43 Boller, T. and Felix, G. (2009) A renaissance of elicitors: perception of microbe-associated molecular patterns and danger signals by pattern-recognition receptors. *Annu. Rev. Plant Biol.* **60**, 379–406, <https://doi.org/10.1146/annurev.arplant.57.032905.105346>
- 44 Dombrowski, J.E. and Martin, R.C. (2018) Activation of MAP kinases by green leaf volatiles in grasses. *BMC Res. Notes* **11**, 79, <https://doi.org/10.1186/s13104-017-3076-9>
- 45 Chen, S., Zhang, L., Cai, X., Li, X., Bian, L., Luo, Z. et al. (2020) (E)-Nerolidol is a volatile signal that induces defenses against insects and pathogens in tea plants. *Hortic Res.* **7**, 52, <https://doi.org/10.1038/s41438-020-0275-7>
- 46 Ye, M., Glauser, G., Lou, Y., Erb, M. and Hu, L. (2019) Molecular dissection of early defense signaling underlying volatile-mediated defense regulation and herbivore resistance in rice. *Plant Cell.* **31**, 687–698, <https://doi.org/10.1105/tpc.18.00569>
- 47 Zhang, M., Su, J., Zhang, Y., Xu, J. and Zhang, S. (2018) Conveying endogenous and exogenous signals: MAPK cascades in plant growth and defense. *Curr. Opin. Plant Biol.* **45**, 1–10, <https://doi.org/10.1016/j.pbi.2018.04.012>

- 48 Mirabella, R., Rauwerda, H., Allmann, S., Scala, A., Spyropoulou, E.A., de Vries, M. et al. (2015) WRKY40 and WRKY6 act downstream of the green leaf volatile E-2-hexenal in Arabidopsis. *Plant J.* **83**, 1082–1096, <https://doi.org/10.1111/tpj.12953>
- 49 Rushton, P.J., Somssich, I.E., Ringler, P. and Shen, Q.J. (2010) WRKY transcription factors. *Trends Plant Sci.* **15**, 247–258, <https://doi.org/10.1016/j.tplants.2010.02.006>
- 50 Engelberth, J., Contreras, C.F., Dalvi, C., Li, T. and Engelberth, M. (2013) Early transcriptome analyses of Z-3-Hexenol-treated zeas revealed distinct transcriptional networks and anti-herbivore defense potential of green leaf volatiles. *PLoS ONE* **8**, e77465, <https://doi.org/10.1371/journal.pone.0077465>
- 51 Erb, M. (2018) Volatiles as inducers and suppressors of plant defense and immunity—origins, specificity, perception and signaling. *Curr. Opin. Plant Biol.* **44**, 117–121, <https://doi.org/10.1016/j.pbi.2018.03.008>