

Phase 2a Pharmacokinetic, Safety, and Exploratory Efficacy Evaluation of Oral Gepotidacin (GSK2140944) in Female Participants with Uncomplicated Urinary Tract Infection (Acute Uncomplicated Cystitis)

J. Scott Overcash,^a Courtney A. Tiffany,^b Nicole E. Scangarella-Oman,^b Caroline R. Perry,^b Yu Tao,^b Mohammad Hossain,^{b*} Aline Barth,^b Etienne F. Dumont^b

^aeStudySite, La Mesa, California, USA ^bGlaxoSmithKline, Collegeville, Pennsylvania, USA

AMERICAN SOCIETY FOR Antimicrobial Agents

MICROBIOLOGY and Chemotherapy®

ABSTRACT Gepotidacin, a triazaacenaphthylene bacterial type II topoisomerase inhibitor, is in development for treatment of uncomplicated urinary tract infection (uUTI). This phase 2a study in female participants with uUTI evaluated the pharmacokinetics (primary objective), safety, and exploratory efficacy of gepotidacin. Eligible participants (n = 22) were confined to the clinic at baseline, received oral gepotidacin at 1,500 mg twice daily for 5 days (on-therapy period; days 1 to 5), and returned to the clinic for test-of-cure (days 10 to 13) and follow-up (day 28 ± 3) visits. Pharmacokinetic, safety, clinical, and microbiological assessments were performed. Maximum plasma concentrations were observed approximately 1.5 to 2 h postdose. Steady state was attained by day 3. Urinary exposure over the dosing interval increased from 3,742 µg·h/ml (day 1) to 5,973 µg·h/ml (day 4), with trough concentrations of 322 to $352 \mu g/ml$ from day 3 onward. Gepotidacin had an acceptable safety-risk profile with no treatment-limiting adverse events and no clinically relevant safety trends. Clinical success was achieved in 19 (86%) and 18 (82%) of 22 participants at test-of-cure and follow-up visits, respectively. Eight participants had a qualifying baseline uropathogen (growth; $\geq 10^5$ CFU/ml). A therapeutic (combined clinical and microbiological [no growth; <10³ CFU/ml]) successful response was achieved in 6 (75%) and 5 (63%) of 8 participants at test-of-cure and follow-up visits, respectively. Plasma area under the free-drug concentration-time curve over 24 h at steady state divided by the MIC (fAUC₀₋₂₄/MIC) and urine AUC₀₋₂₄/MIC ranged from 6.99 to 90.5 and 1,292 to 121,698, respectively. Further evaluation of gepotidacin in uUTI is warranted. (This study has been registered in ClinicalTrials.gov under identifier NCT03568942.)

KEYWORDS gepotidacin, uncomplicated urinary tract infection, acute uncomplicated cystitis, pharmacokinetics, safety

Predominant uropathogens in uncomplicated urinary tract infections (uUTIs; acute uncomplicated cystitis) are *Escherichia coli* (75% to 90%), *Staphylococcus saprophyticus* (5% to 15%), and *Klebsiella, Enterobacter, Proteus*, and enterococcus uropathogens (5% to 10%) (1–3). Multidrug-resistant (MDR) uropathogens, commonly associated with nosocomial infections, have emerged at the community level, and treatment for uUTIs has become more difficult (4–6). Health authorities recognize extended-spectrum β -lactamase (ESBL)-producing *Enterobacteriaceae* as a serious threat (7) and drug-resistant *Enterobacteriaceae* as a critical priority pathogen (8). The MDR *E. coli* sequence type 131 clone has emerged as a cause of urinary tract infections and bacteremia

Citation Overcash JS, Tiffany CA, Scangarella-Oman NE, Perry CR, Tao Y, Hossain M, Barth A, Dumont EF. 2020. Phase 2a pharmacokinetic, safety, and exploratory efficacy evaluation of oral gepotidacin (GSK2140944) in female participants with uncomplicated urinary tract infection (acute uncomplicated cystitis). Antimicrob Agents Chemother 64:e00199-20. https://doi.org/10.1128/AAC.00199-20.

Copyright © 2020 Overcash et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Caroline R. Perry, Caroline.R.Perry@gsk.com, or Etienne F. Dumont, Etienne.F.Dumont@gsk.com.

* Present address: Mohammad Hossain, Agios Pharmaceuticals, Cambridge, Massachusetts, USA.

Received 30 January 2020 Returned for modification 2 February 2020 Accepted 4 April 2020

Accepted manuscript posted online 13 April 2020 Published 23 June 2020





worldwide (9–11). The availability of oral antimicrobials effective against ESBLs is limited and, for some outpatient infections, no oral options remain.

Gepotidacin (GSK2140944) is a triazaacenaphthylene bacterial type II topoisomerase inhibitor with a novel mode of action and with *in vitro* activity against most target pathogens resistant to established antibacterials (12–15). Phase 2 studies have demonstrated the efficacy of gepotidacin in acute bacterial skin and skin structure infections and uncomplicated urogenital gonorrhea (16–18). The microbiological activity of gepotidacin includes *E. coli*, the key causative uropathogen of uUTI, and *S. saprophyticus*, and *Enterococcus faecalis*. In addition, the efficacy of gepotidacin against *E. coli* was evaluated in a rat pyelonephritis model, which indicated potential efficacy in uUTI and supported clinical dose selection (19). The pharmacokinetics (PK) of gepotidacin have been well defined in healthy participants and demonstrated urine exposures that may support uUTI treatment (20–23). A phase 2a evaluation of oral gepotidacin in female participants with uUTI was conducted with the main objectives of evaluating plasma and urinary gepotidacin exposures and safety in this population. In addition, exploratory efficacy and PK/pharmacodynamic (PD) endpoints were assessed.

RESULTS

Participant disposition. A total of 22 female participants with uUTI were enrolled and evaluated for PK, safety, and clinical efficacy in this phase 2a, single-center, single-arm, open-label study in the Unites States from July 2018 to January 2019 (see Fig. S1 in the supplemental material). Participants were confined to the clinic from baseline (days –1 to 1 predose) through the on-therapy (days 1 to 5) period and returned as outpatients for test-of-cure (TOC; days 10 to 13) and follow-up (day 28 ± 3) visits. Participants received oral gepotidacin at 1,500 mg twice daily (BID) for 5 days. Two participants (9%) withdrew from the study due to loss to follow-up and family reasons; there were no discontinuations due to adverse events (AEs) (Fig. 1).

Participant baseline characteristics. The majority of participants were White, ranged in age from 19 to 60 years, and had a body mass index ranging from 20.9 to 37.9 kg/m^2 (Table 1). The number of past uUTI episodes ranged from 0 to 10 over the past 12 months across participants; the majority of participants reported ≤ 2 episodes.

The mean total clinical symptom score at baseline was 7.9 (range, 4 to 12). All participants reported frequency and urgency, and all but 1 participant (5%) reported dysuria.

In the 22 intent-to-treat (ITT) participants, 19 baseline uropathogens were recovered, and 8 uropathogens from 8 participants (36%) met the qualifying uropathogen definition and inclusion in the microbiological intent-to-treat (micro-ITT) population (Fig. 1).

TABLE 1	Baseline	demographics	(ITT	population)
---------	----------	--------------	------	-------------

Demographic parameter ^a	Value for the parameter $(n = 22)$			
Age (yr)	37.1 (12.26)			
Reproductive status (no. [%])	2 (14)			
Sterile (of childbearing age)	1 (5)			
Potentially able to bear children	18 (82)			
Body mass index (kg/m²) Height (cm) Weight (kg)	26.96 (5.366) 163.22 (6.033) 72.01 (16.015)			
Ethnicity (no. [%]) Hispanic or Latino Not Hispanic or Latino	6 (27) 16 (73)			
Race (no. [%]) Black or African American White (White/Caucasian/European heritage)	4 (18) 18 (82)			

aUnless otherwise indicated, values are means (standard deviations).

This subset of 8 participants underwent both clinical and microbiological efficacy assessments.

Pharmacokinetics. Median gepotidacin plasma concentrations peaked rapidly, with median maximum concentrations (T_{max}) observed at 1.50 and 1.92 h postdose on days 1 and 4, respectively (Fig. 2). Concentrations declined in a multiphasic manner. Plasma exposure (maximum observed concentration [C_{max}] and area under the concentration-time curve from time zero to the 12-h dosing interval [AUC_{0-\tau}]) was approximately 1.4-fold higher on day 4 than on day 1 (Table 2). The accumulation was consistent with an effective elimination half-life of 6.6 h. The between-participant variability in plasma exposures was moderate, with a higher coefficient-of-variation range for C_{max} (38% to 47%) than for the AUC_{0-\tau} (29% to 32%) across days 1 and 4. Based on observed plasma predose concentrations (C_{τ}) and statistical analysis, steady state was achieved by day 3 (Fig. S2 and Table S1).



FIG 2 Median gepotidacin plasma concentration-time plot (pharmacokinetic population). The lower limit of quantification, represented by the dashed line, was 0.10 μ g/ml. Day 1 plasma pharmacokinetic data after the 0.5-h collection for two participants were excluded due to vomiting. The 12-h pharmacokinetic data for one participant on day 1 and one participant on day 4 were excluded because the samples were collected after the second daily dose.

TABLE 2 Summary	of gepotidacin	plasma PK	parameters	(PK parameter	population)
$(N = 22)^{a}$					

PK parameter and	Value for the parameter ^c	Value for the parameter ^c				
time point ^b	Geometric mean (% CVb)	Min–max				
C _{max} (μg/ml) Day 1 Day 2	5.89 (47.3)	1.82–12.8				
Day 3 Day 4 Day 5	8.44 (38.0)	3.82-16.8				
T _{max} (h) Day 1 Day 2	1.50 ^{<i>d</i>}	0.470-3.07				
Day 3 Day 4 Day 5	1.92 ^d	0.450-4.12				
AUC _{0-τ} (μg·h/ml) Day 1 Day 2	20.2 (28.6)	11.0-31.0				
Day 3 Day 4 Day 5	29.3 (31.8)	15.2–49.5				
CL _{ss} /F (l/h) Day 1 Day 2 Day 3 Day 4 Day 5	51.2 (31.8)	30.3-98.7				
R _o Day 1 Day 2 Day 3 Day 4 Day 5	1.40 (20.4)	1.09-2.20				
C _τ (μg/ml) Day 1 Day 2 Day 3 Day 4 Day 5	0.621 (62.3) 0.789 (37.4) 0.851 (41.4) 0.819 (46.4)	0.122–1.84 0.371–1.60 0.460–1.99 0.327–1.93				

^aN, number of participants in the treatment.

 ${}^{b}C_{maxi}$ maximum observed concentration; T_{maxi} time of occurrence of C_{maxi} AUC₀₋₇ area under the concentration-time curve from time zero to the 12-h dosing interval; CL_{ss}/F , apparent steady-state clearance; R_{o} , accumulation ratio based on the AUC₀₋₇; C_{τ} predose concentration. The numbers of participants with evaluable PK parameter data were 20 for day 1 and 21 for days 2 to 5, with the exception that day 4 R_{o} data are for 19 participants.

^cCVb, between-participant geometric coefficient of variation; max, maximum; min, minimum.

^dValues are medians.

Median urine gepotidacin concentrations were generally higher on day 4 than on day 1 (Fig. 3). On day 1, approximately 20% of the dose was excreted in urine over the dosing interval, increasing to 31% on day 4 (Table 3). Overall exposure in urine (AUC_{0- τ}) also increased from day 1 to day 4, with urine C_{τ} values ranging from 322 to 352 μ g/ml from day 3 onward. The renal clearance values were similar on days 1 and 4. Approximately 460 mg of unchanged gepotidacin was excreted in urine over the steady-state dosing interval, with a minimum steady-state AUC_{0- τ} of 2,256 μ g-h/ml.

Gepotidacin was measurable in cervical, rectal, and pharyngeal swabs on day 4, with the highest concentrations in rectal swabs (Table S2).

Safety. Twenty-one participants (95%) experienced AEs; gastrointestinal-related disorders had the highest prevalence (Table 4). Gastrointestinal AEs reported in >10%



FIG 3 Median gepotidacin urine concentration-time plot (pharmacokinetic population). The lower limit of quantification, represented by the dashed line, was 1.00 μ g/ml. Data are plotted by the planned relative midpoint time for each interval.

of participants were diarrhea, nausea, and vomiting and were the most prevalent drug-related events. Other drug-related AEs were vulvovaginal mycotic infection (2 participants, 9%), headache (1 participant, 5%), and chest discomfort (1 participant, 5%; noncardiac in nature).

All AEs were mild (4 participants, 18%) or moderate (16 participants, 73%), except for a nonfatal serious AE of major depression with voluntary psychiatric hospitalization in 1 participant (5%) that occurred 9 days after the last dose and was considered by the investigator not to be related to gepotidacin. No participant experienced a drug-related AE of an intensity greater than moderate.

No clinically relevant laboratory changes were observed. Baseline and repeat urine dipstick results were consistent with the uUTI under study.

There were no clinically significant electrocardiogram (ECG) findings or changes from baseline. No participants had a QT interval corrected for heart rate according to Fridericia (QTcF) of \geq 480 ms or an increase of >30 ms (Fig. 4). Mean QTcF (minimum, maximum) change from baseline to day 4 at 2 h postdose was 3.4 (-89, 27) ms. No clinically relevant changes in vital signs were observed.

Exploratory efficacy. (i) Clinical. In the ITT population, at TOC, clinical success was observed for 19 of 22 participants (86%; Clopper-Pearson 95% confidence interval [CI], 65% to 97%), and clinical failure was observed for 3 of 22 participants (14%; Clopper-Pearson 95% CI, 3% to 35%) (Table 5). Clinical success was achieved in 12 of 14 participants (86%) who did not have a qualifying baseline uropathogen, indicating complete resolution of clinical symptoms in these participants. Clinical response results were similar between TOC and follow-up visits (Table S3); however, per sponsor-determined clinical response, there was an additional clinical failure at follow-up.

The mean baseline total clinical symptom score (7.9) decreased to 0.1 and 0 at TOC and follow-up, respectively, with a similar trend for each symptom category (Fig. 5). All participants who completed the full course of gepotidacin and presented at TOC (n = 19) had complete symptom resolution; 1 participant who received only 6 doses of gepotidacin had a score of 2. All 20 participants with clinical scores reported at follow-up had a score of 0.

Clinical cure in most uropathogen groups was observed by approximately day 4 with an increase through TOC and follow-up.

(ii) Microbiological. The 8 qualifying baseline isolates consisted of 5 *E. coli* isolates and 1 isolate each of *Citrobacter koseri*, *Klebsiella pneumoniae*, and *S. saprophyticus* (Table 6). Of the 5 qualifying *E. coli* uropathogens, 2 were MDR, and 1 of those was both MDR and quinolone resistant.

TABLE 3 Summary of	gepotidacin	urine PK	parameters	(PK parameter	population)
$(N = 22)^{a}$					

PK parameter and	Value for the parameter ^c					
time point ^b	Geometric mean (% CVb)	Min-max				
Ae ₁₂ (mg)						
Day 1	299 (107.6)	9.55–578				
Day 2						
Day 3	460 (66.0)	125 1 100				
Day 4	400 (55.8)	135-1,100				
Day 5						
AUC _{0-τ} (μg·h/ml)						
Day 1	3,742 (93.9) ^d	1,034–24,858				
Day 2						
Day 3	5 0 7 2 (0 7 2) 2	2 256 20 425				
Day 4	5,973 (87.2) ^e	2,256-30,425				
Day 5						
AUC _{0−24} (μg·h/ml)						
Day 1						
Day 2						
Day 3						
Day 4	11,945 (87.2) ^e	4,512–60,849				
Day 5						
fe% (%)						
Day 1	19.9 (107.6)	0.637–38.5				
Day 2						
Day 3						
Day 4	30.7 (55.8)	9.03–73.5				
Day 5						
CLr (l/h)						
Day 1	14.8 (118.2)	0.420-41.5				
Day 2						
Day 3						
Day 4	15.7 (45.2)	8.91–41.6				
Day 5						
C_{τ} (μ g/ml)						
Day 1						
Day 2	279 (154.7)	26.8-1,800				
Day 3	322 (138.8)	42.1–3,670				
Day 4	327 (248.7)	32.8-4,540				
Day 5	352 (146.5)	68.2–4,010				

^aN, number of participants in the treatment.

^bAe₁₂, total unchanged drug excreted over 12 h; AUC₀₋₇, area under the concentration-time curve from time zero to the 12-h dosing interval; AUC₀₋₂₄, area under the concentration-time curve from 0 to 24 h; fe%, percentage of the given dose of drug excreted in urine; CLr, renal clearance; C₇₇ predose concentration. The numbers of participants with evaluable PK parameter data, except as otherwise noted, were 20 for days 1 and 2 and 21 for days 3 to 5. Day 1 urine PK parameter data for 2 participants were excluded from the summary statistics analysis due to vomiting.

^cCVb, between-participant geometric coefficient of variation; max, maximum; min, minimum; *n*, number of participants with evaluable values.

 $^{d}n = 16.$

^en = 18.

Most qualifying baseline uropathogens were resistant to ampicillin (5 of 8 isolates, 63%); none were resistant to nitrofurantoin, meropenem, or fosfomycin. No phenotypic ESBL-producing uropathogens were recovered. Against the 8 qualifying baseline uropathogens in the micro-ITT population, gepotidacin MIC values ranged from 0.06 to 4 μ g/ml (Table S4).

For 7 of 8 participants (88%) in the micro-ITT population, no growth (eradication) was observed starting on day 2. At TOC, microbiological success was achieved in 7 of 8 participants (88%), including a participant with *K. pneumoniae* infection (Table 7 and Table S5). At TOC, there was 1 microbiological failure (13%) due to an indeterminant

TABLE 4 Summary of	of adverse	events	(safety	population)
--------------------	------------	--------	---------	-------------

Event system organ class and preferred term	No. of events (%) $(n = 22)$
Any adverse event	21 (95)
Gastrointestinal disorders	21 (95)
Diarrhea	18 (82)
Nausea	17 (77)
Vomiting	5 (23)
Anal pruritus	1 (5)
Colitis	1 (5)
Dyspensia	1 (5)
Fructation	1 (5)
Feres soft	1 (5)
Flatulence	1 (5)
Infections and infestations	6 (27)
Viral upper respiratory tract infection	2 (9)
Vulvovaginal mycotic infection	2 (9)
Gastroenteritis	1 (5)
Upper respiratory tract	1 (5)
Nervous system disorders	5 (23)
Headache	5 (23)
Musculoskeletal and connective tissue disorders	3 (14)
Back pain	2 (9)
Muscle spasms	1 (5)
Myalgia	1 (5)
General disorders and administration site conditions	2 (9)
Chest discomfort	2 (9)
Psychiatric disorders	1 (5)
Major depression	1 (5)
Respiratory, thoracic, and mediastinal disorders	1 (5)
Oropharyngeal pain	1 (5)

laboratory result (i.e., out-of-stability specimen); however, the participant was a clinical success. Microbiological response results were similar between TOC (Table S5) and follow-up (Table S6) visits except that there was an additional microbiological failure at follow-up. One participant had regrowth of *C. koseri* with no change in gepotidacin MIC or susceptibility.

Growth was observed for 2 isolates posttreatment (1 *E. coli* isolate on day 3 and 1 *C. koseri* isolate at follow-up); both were resistant to ampicillin at baseline and the posttreatment time point. There was no change in the gepotidacin MIC (Table S4).

No participants had a baseline uropathogen that demonstrated a reduction in susceptibility to gepotidacin (i.e., \geq 4-fold increase in gepotidacin MIC for baseline uropathogens versus postbaseline uropathogens of the same species and from the same participant) at any time point in the ITT population.

(iii) **Therapeutic response.** The overall participant-level therapeutic responses in the micro-ITT population were success for 6 of 8 participants (75%) and failure for 2 of 8 participants (25%) at TOC (Table 7 and Table S7). Details on clinical and microbiological failures leading to therapeutic response failures are described in the previous paragraphs (Table 7). Therapeutic response results were comparable between TOC and follow-up visits (Table 7 and Table S8); however, there was an additional microbiological failure at follow-up. No clinical or microbiological failures required an alternative antibiotic for treatment of uUTI throughout the study.

Pharmacokinetic/pharmacodynamic. Four out of the five participants with a qualifying *E. coli* uropathogen at baseline had evaluable PK/PD parameters (Table 7). Other qualifying baseline uropathogens were observed, but data were limited. Similar

Box Plots of Change from Baseline in QTcF over Time



FIG 4 Box plot of change from baseline in QTcF over time (safety population). The triangle (inside the box) represents the mean value; the circle represents individual change from baseline; the top, middle, and bottom lines of the box represent the 75th, 50th (median), and 25th percentiles, respectively. The interquartile range is the distance between the 25th and 75th percentiles. The top and bottom whiskers represent maximum and minimum values, which are within $1.5 \times$ the interquartile range from the edge of the box, respectively. Any points outside the whiskers are deemed outliers. QTcF, QT interval corrected for heart rate according to Fridericia.

to the *E. coli* data, the PK parameter/MIC ratios were higher for the urine parameters than for the free-plasma parameters.

For the 4 participants with qualifying *Enterobacteriaceae* uropathogens who were also microbiological successes at TOC, the area under the free-drug concentration-time curve over 24 h at steady state divided by the plasma MIC ($fAUC_{0-24}$ /MIC) ranged from 6.99 to 90.5, and urine AUC_{0-24} /MICs ranged from 1,292 to 121,698 (Table 7). The participant with the lowest plasma $fAUC_{0-24}$ /MIC (6.99) and urine AUC_{0-24} /MIC (1,292) had a *K. pneumoniae* isolate with a gepotidacin MIC of 4 µg/ml and was a microbiological success.

TABLE 5 Summary of investigator-determined and sponsor-determined clinical outcomes and responses at test-of-cure by qualifying uropathogen isolated at baseline

	No. of participants by population and method (% [95% Cl]) ^a						
Uropathogen group, clinical response (success or failure), and clinical	Intent-to-treat population	(<i>n</i> = 22)	Microbiological intent-to-treat population $(n = 8)$				
outcome category	Investigator-determined	Sponsor-determined	Investigator-determined	Sponsor-determined			
All qualifying uropathogens ($n = 8$)							
Success	7 (88 [47 to >99])	7 (88 [47 to >99])	7 (88 [47 to >99])	7 (88 [47 to >99])			
Clinical success	7 (88)	7 (88)	7 (88)	7 (88)			
Failure	1 (13 [<1 to 53])	1 (13 [<1 to 53])	1 (13 [<1 to 53])	1 (13 [<1 to 53])			
Clinical failure	1 (13)	1 (13)	1 (13)	1 (13)			
Unable to determine	0	0	0	0			
No qualifying uropathogen ($n = 14$)							
Success	12 (86 [57 to 98])	12 (86 [57 to 98])					
Clinical success	12 (86)	12 (86)					
Failure	2 (14 [2 to 43])	2 (14 [2 to 43])					
Clinical failure	0	0					
Unable to determine	2 (14)	2 (14)					
Total for groups (all participants)							
Success	19 (86 [65 to 97])	19 (86 [65 to 97])	7 (88 [47 to >99])	7 (88 [47 to >99])			
Failure ^b	3 (14 [3 to 35])	3 (14 [3 to 35])	1 (13 [<1 to 53])	1 (13 [<1 to 53])			

^aA participant was counted more than once under a uropathogen category if multiple qualifying uropathogens within that uropathogen category were isolated at baseline for the participant. Other Gram-negative bacilli consisted of *Citrobacter koseri* (1) and *Klebsiella pneumoniae* (1). Cl, confidence interval (Clopper-Pearson). ^bNo failures required an alternative antibiotic for treatment of uncomplicated urinary tract infection throughout the study.



FIG 5 Individual clinical symptom score and box plot of total score over time (ITT population) (n = 22). The box represents the 25th to 75th percentiles. Within the box, the horizontal line represents the median, and the square indicates the mean. The upper and lower whiskers represent $1.5 \times$ the interquartile range. The open circles represent individual participant outlier scores.

DISCUSSION

These results suggest that gepotidacin may provide a new oral treatment option for uUTI (acute uncomplicated cystitis) with further evaluation. The novel mechanism of action of gepotidacin could help meet the current need for oral antibacterial agents with activity against drug-resistant uropathogens (12–15).

The gepotidacin dose regimen in this population provided >600-fold-higher concentrations in urine than in free plasma at steady state, which is the target site of action for the treatment of uUTIs. The minimum gepotidacin urine concentrations remained above the gepotidacin MIC value of 4 μ g/ml throughout the dosing interval. Of note, fluid intake was not standardized during the study; thus, any impact hydration status had on gepotidacin urinary exposures is unknown. The gepotidacin renal excretion for female participants with uUTI was higher than that of healthy participants with normal renal function or mild renal impairment (20% versus 7.5% of the dose), and day 1

TABLE 6 Uropathogens recovered	at	baseline	(ITT	popula	tion)
---------------------------------------	----	----------	------	--------	-------

Group and uropathogen recovered ^a	No. (%) (<i>n</i> = 22) ^{<i>b</i>}
Total group	19
Acinetobacter pittii	1 (5)
Citrobacter freundii complex	1 (5)
Citrobacter koseri	1 (5)
Escherichia coli	14 (74)
MDR Escherichia coli	2 (11)
Quinolone-resistant Escherichia coli	1 (5) ^c
Klebsiella pneumoniae	1 (5)
Staphylococcus saprophyticus	1 (5)
Qualifying group ($\geq 10^5$ CFU/ml)	8
Citrobacter koseri	1 (13)
Escherichia coli	5 (63)
MDR Escherichia coli	2 (25)
Quinolone-resistant Escherichia coli	1 (13) ^c
Klebsiella pneumoniae	1 (13)
Staphylococcus saprophyticus	1 (13)

^aMultidrug resistant (MDR) refers to a uropathogen that was resistant to \geq 3 relevant antibiotic classes. ^bThe denominator for the percentage calculations was the number of pathogens.

^cOf the *E. coli* uropathogens, two were MDR and one of those was both MDR and quinolone resistant.

	Oualifying baseline	Gepotidacin	Plasma	Urine	Microbiological response		Clinical response		Therapeutic response	
Participant no.	uropathogen	MIC (μ g/ml)	fAUC ₀₋₂₄ /MIC	AUC ₀₋₂₄ /MIC	тос	Follow-up	тос	Follow-up	тос	Follow-up
1	C. koseri	0.5	79.6	NA	S	F	S	S	S	F
2	E. coli	2	22.1	7,379	S	S	S	S	S	S
3	E. coli	0.5	90.5	121,698	S	S	S	S	S	S
4	E. coli	2	30.6	9,011	S	S	S	S	S	S
5	E. coli ^b	1	30.6	7,926	Fc	F^d	S	S	F	F
6	E. coli ^e	2	NA	NA	S	S	F^{f}	F ^f	F	F
7	K. pneumoniae	4	6.99	1,292	S	S	S	S	S	S
8	S. saprophyticus	0.06	1,040	543,252	S	S	S	S	S	S

TABLE 7 Summary of plasma and urine PK/PD, microbiological response, clinical response, and therapeutic response at TOC and followup by qualifying uropathogen isolated at baseline (micro-ITT population)^a

^aAUC₀₋₂₄, area under the concentration-time curve from time zero to 24 h; fAUC, area under the free-drug concentration-time curve; NA, not available (steady-state pharmacokinetic data not available); TOC, test-of-cure; S, success; F, failure.

^bIsolate was multidrug resistant (e.g., resistant to \geq 3 relevant antibiotic classes) and quinolone resistant.

^cMicrobiological failure due to an out-of-stability urine specimen.

^dMicrobiological failures at TOC were also considered microbiological failures at follow-up.

^elsolate was multidrug resistant.

Participant received only 6 doses of gepotidacin due to withdrawal by the participant. The participant had a baseline total clinical score of 10 that decreased to 2 at TOC; however, that was not complete symptom resolution, and the clinical response was clinical failure. At follow-up, the total clinical symptom score was 0, which was a sponsor-determined clinical outcome of delayed clinical success; however, the clinical response remained a clinical failure per the analysis plan.

plasma exposures in this study appeared to be higher than in previous phase 1 studies (mean C_{max} of 5.89 μ g/ml versus 3.20 μ g/ml) (unpublished data). Furthermore, gepotidacin concentrations were measurable in cervical, rectal, and pharyngeal swabs, supporting the evaluation of gepotidacin for gonorrhea (clinical trial NCT04010539).

An acceptable safety-risk profile was demonstrated after gepotidacin administration with no treatment-limiting AEs and no discontinuations due to AEs. The safety profile of gepotidacin was similar to that observed in previous studies. A high prevalence of gastrointestinal AEs was expected (16, 17, 20–23); however, the prevalence of gastro-intestinal AEs in the current study (95%) was higher than that observed previously. This was the first study in all-female participants and in uUTI. The investigator observed that most nausea AEs had an acute onset within the first few doses and that tolerance was observed with repeat dosing.

Based on a previous gepotidacin corrected QT (QTc) evaluation (24), this study strategically included an on-treatment ECG assessment at the maximum steady-state gepotidacin exposures (i.e., day 4 at 2 h postdose); however, there were no cardiac AEs reported and no clinically significant ECG findings. In addition, there were no clinically relevant trends in the safety parameters.

In the ITT population (n = 22), symptom resolution (i.e., clinical score of 0) was achieved in 19 participants at TOC and in 20 participants at follow-up. The only participant without symptom resolution at TOC withdrew from study treatment and did not receive the full 5-day course of gepotidacin but underwent TOC and follow-up assessments. All participants had clinical efficacy observations consistent with expectations for a uUTI antibacterial.

Nonclinical models have shown that the PK/PD index most predictive of gepotidacin efficacy is fAUC/MIC (25). When the minimum exposure of gepotidacin in human urine at steady state measured in this study (minimum urine AUC_{0-τ} = 2,256 µg·h/ml; thus, minimum AUC₀₋₂₄ = 4,512 µg·h/ml) and a gepotidacin MIC value of 4 µg/ml are applied, the minimum human urine AUC₀₋₂₄/MIC achieved for the oral gepotidacin 1,500-mg BID dose exceeds the fAUC/MIC resistance suppression target of 275, as determined from an *in vitro* PK/PD hollow-fiber infection model (26), by approximately 4-fold, and 100% target attainment is expected in participants infected with uropathogens with gepotidacin MICs of \leq 4 µg/ml.

This was a single-center evaluation in the United States, which led to very few drug-resistant isolates for evaluation. There was also a low prevalence of baseline uropathogens meeting growth criteria for the micro-ITT population (i.e., small sample

size for microbiological assessment). The study was open label and did not include a comparator antibacterial. Global, multicenter, noninferiority gepotidacin phase 3 studies (clinical trials NCT04020341 and NCT04187144) should address these limitations.

The gepotidacin PK parameters were well defined in this female uUTI population. Oral gepotidacin at 1,500 mg BID for 5 days demonstrated an acceptable safety-risk profile (i.e., no discontinuations due to AEs) and provided positive exploratory efficacy findings, with no resistance development. These data support additional evaluation of gepotidacin in uUTI.

MATERIALS AND METHODS

Study population. The study recruited nonpregnant females who were \geq 18 and \leq 65 years of age. Participants were required to have two or more of the following clinical signs and symptoms with onset \leq 72 h at screening: dysuria, frequency, urgency, or lower abdominal pain. In addition, they were required to have pyuria (\geq 10 white blood cells/mm³ or the presence of leukocyte esterase) and/or nitrite from a pretreatment urine sample. Participants who had any preexisting condition that may have impacted gepotidacin absorption, distribution, metabolism, or excretion were excluded. Full inclusion and exclusion criteria are provided in the supplemental material.

This study was conducted in accordance with the Declaration of Helsinki and Good Clinical Practice guidelines. Protocol and procedures were reviewed and approved by an institutional review board. Written informed consent was obtained from participants before any study procedures were performed.

Study design. This was a phase 2a, single-center, single-arm, open-label study. Participants were confined to the clinic from baseline (days –1 to 1 predose) through the on-therapy period (days 1 to 5). Participants returned for outpatient visits at TOC (days 10 to 13) and follow-up (day 28 \pm 3). Gepotidacin (1,500 mg; two 750-mg tablets) was administered orally BID for 5 days under site supervision. The target sample size was approximately 20 participants based on PK requirements.

Pharmacokinetic assessments. Serial blood and urine samples were collected from predose to 12 h postdose on day 1 (first dose) and day 4 (time matched to the first dose on day 1) (collection time points are given in Fig. S1 in the supplemental material). For steady-state assessment, predose blood (single collection) and urine (0- to 2-h interval) samples were collected before each time-matched dose on days 1 through 5.

For each plasma PK sample, 3 ml of whole blood was collected into tubes containing EDTA anticoagulant via an indwelling catheter and/or direct venipuncture. Each tube was inverted approximately 5 to 10 times immediately after the sample was drawn. The whole-blood sample may have been stored at room temperature for up to 60 min prior to centrifugation. The sample was centrifuged under refrigerated conditions (2°C to 8°C) at approximately 650 to $1,450 \times g$. Approximately 1.5 ml of plasma was transferred via a pipette into a 2-ml cryovial tube and kept frozen until analysis. Batched samples were shipped on dry ice to the bioanalytical laboratory for validated analysis.

For predose urine PK samples, a urine cup was used for collection. For all postdose urine PK samples, a urine jug was used for each collection interval. For each interval, approximately 1 ml of urine was transferred via a pipette into a 2-ml cryovial tube and kept frozen until analysis. Batched samples were shipped on dry ice to the bioanalytical laboratory for validated analysis.

Exploratory PK assessment included the collection of cervical, rectal, and pharyngeal swab specimens on day 4 (predose and 2 h postdose).

All PK samples were analyzed using validated ultra- or high-performance liquid chromatography with tandem mass spectroscopy methods by PPD Bioanalytical Laboratory (Middleton, WI).

Safety assessments. Adverse event monitoring, vital sign measurements, clinical laboratory evaluations, and ECGs, including on-treatment ECGs on days 1 and 4 matched with the 2-h PK collection, were performed.

Exploratory efficacy assessments. (i) Clinical. Clinical signs and symptoms of uUTI were recorded based on participant interview at baseline (pretreatment), days 2 through 5, TOC, and follow-up using a 0- to 3-point scale (0, none; 1, mild; 2, moderate; 3, severe) for the categories of dysuria, frequency, urgency, and lower abdominal or suprapubic pain (Fig. S3). At each on-therapy assessment, clinical success included both resolution of or improvement in signs and symptoms. Clinical success at TOC and follow-up was defined as resolution of signs and symptoms present at baseline (and no new signs and symptoms) and no use of other antimicrobial therapy for the current uUTI. At TOC, a score of zero was required for a participant to be deemed a clinical success. At follow-up, the participant must have had a score of zero at TOC that persisted from TOC to follow-up for a response of clinical success.

(ii) Microbiological. A urine sample was collected at baseline (pretreatment), predose days 2 through 5, TOC, and follow-up for Gram stain, quantitative bacteriology culture, and *in vitro* antimicrobial susceptibility testing using standard methods at a central laboratory (PPD Laboratories Central Lab, Highland Heights, KY). Susceptibility testing was conducted for all uropathogens by broth microdilution and gradient diffusion (fosfomycin only) according to guidelines of the Clinical and Laboratory Standards Institute (27, 28). Inclusion in the micro-ITT population required growth of a qualifying baseline uropathogen ($\geq 10^5$ CFU/ml) (29, 30) (Fig. S4). Microbiological success was defined as culture-confirmed eradication (no growth; <10³ CFU/ml) of the qualifying baseline uropathogen. Multidrug resistance was defined as a baseline uropathogen that was resistant to ≥ 3 relevant antibiotic classes.

Statistical analysis. (i) Analysis populations. Analysis populations are defined in Table S9.

(ii) Pharmacokinetic. Noncompartmental PK analyses were performed using Phoenix WinNonlin, version 6.4 (Certara USA, Inc., Princeton, NJ), and SAS, version 9.3 (SAS Institute, Inc, Cary, NC), with actual sampling times. As total gepotidacin concentrations were measured in plasma, unbound values were derived by multiplying total concentrations by 0.67 to correct for the plasma protein binding of gepotidacin (33%) (unpublished data). Steady-state achievement was assessed using a linear mixed model with the Helmert transformation.

(iii) **Safety.** Adverse events, change from baseline values for clinical chemistry, hematology, vital signs, and ECG findings were summarized using SAS, version 9.3. Posthoc QTcF plots were generated using SAS, version 9.4.

(iv) Exploratory efficacy. Efficacy data were summarized by qualifying baseline uropathogen using counts and percentages, with the 95% Clopper-Pearson CI presented at TOC and follow-up using SAS, version 9.3.

Clinical outcome (investigator- and sponsor-determined) and response were summarized for the ITT and micro-ITT populations. Mean clinical symptom scores were summarized for the ITT population. Clinical cure was summarized for qualifying baseline uropathogens.

Microbiological outcome and response were summarized by predefined uropathogen groups or species. Urine quantitative bacteriology culture results were summarized. Results and interpretations of susceptibility testing for all uropathogens against gepotidacin and other antimicrobials were summarized.

Therapeutic response (success/failure), determined by statistical programming, was a measure of the overall efficacy response. Therapeutic success required both clinical success and microbiological success, or the participant was a deemed therapeutic failure. Therapeutic success was summarized by perparticipant microbiological response and clinical response.

(v) Pharmacokinetic/pharmacodynamic. The plasma $fAUC_{0-24}$ /MIC and the urine AUC_{0-24}/MIC ratio were determined using the day 4 PK parameters and the qualifying baseline uropathogen (day 1) MIC.

SUPPLEMENTAL MATERIAL

Supplemental material is available online only. **SUPPLEMENTAL FILE 1**, PDF file, 0.6 MB.

ACKNOWLEDGMENTS

We thank the study participants, clinical staff, and investigator at eStudySite for their participation. We also thank the PPD laboratories and PK and biostatistics team for their analysis and study contributions, and Jodi Stahlman (PPD) for editorial and writing assistance. Anonymized individual participant data and study documents can be requested for further research from www.clinicalstudydatarequest.com.

This study was supported by GlaxoSmithKline (GSK; Collegeville, PA, USA). This work was also supported in whole or in part with Federal funds from the Office of the Assistant Secretary for Preparedness and Response, Biomedical Advanced Research and Development Authority (HHSO100201300011C). Medical writing assistance was funded by GSK.

C.A.T., N.E.S-O., C.R.P., A.B., and E.F.D. are employees of and own stock in GSK. Y.T. is an employee of GSK. M.H. is a former employee of GSK and holds company stock. Authors were not paid for their manuscript contributions.

J.S.O. was the principal investigator for the study. All authors, with the exception of A.B., contributed to the study concept/design. Only J.S.O. contributed to the acquisition of study data. All authors, with the exception of J.S.O., contributed to the data analysis/interpretation. All authors provided manuscript review and approval.

REFERENCES

- Stamm WE, Hooton TM. 1993. Management of urinary tract infections in adults. N Engl J Med 329:1328–1334. https://doi.org/10.1056/ NEJM199310283291808.
- Talan DA, Stamm WE, Hooton TM, Moran GJ, Burke T, Iravani A, Reuning-Scherer J, Church DA. 2000. Comparison of ciprofloxacin (7 days) and trimethoprim-sulfamethoxazole (14 days) for acute uncomplicated pyelonephritis in women: a randomized trial. JAMA 283:1583–1590. https:// doi.org/10.1001/jama.283.12.1583.
- Foxman B, Barlow R, D'Arcy H, Gillespie B, Sobel JD. 2000. Urinary tract infection: self-reported incidence and associated costs. Ann Epidemiol 10:509–515. https://doi.org/10.1016/S1047-2797(00)00072-7.
- Hooton TM. 2012. Clinical practice. Uncomplicated urinary tract infection. N Engl J Med 366:1028–1037. https://doi.org/10.1056/NEJMcp1104429.
- Flamm RK, Sader HS, Farrell DJ, Jones RN. 2014. Ceftazidime-avibactam and comparator agents tested against urinary tract isolates from a global surveillance program (2011). Diagn Microbiol Infect Dis 80:233–238. https://doi.org/10.1016/j.diagmicrobio.2014.07.005.
- Sanchez GV, Babiker A, Master RN, Luu T, Mathur A, Bordon J. 2016. Antibiotic resistance among urinary isolates from female outpatients in the United States in 2003 and 2012. Antimicrob Agents Chemother 60:2680–2683. https://doi.org/10.1128/AAC.02897-15.
- Centers for Disease Control and Prevention. 2019. Antibiotic resistance threats in the United States. https://www.cdc.gov/drugresistance/pdf/ threats-report/2019-ar-threats-report-508.pdf.
- 8. World Health Organization. 2017. Global priority list of antibiotic resistant bacteria to guide research, discovery, and development of new

antibiotics. http://www.who.int/medicines/publications/WHO-PPL-Short _Summary_25Feb-ET_NM_WHO.pdf?ua=1.

- Johnson JR, Nicolas-Chanoine MH, DebRoy C, Castanheira M, Robicsek A, Hansen G, Weissman S, Urban C, Platell J, Trott D, Zhanel G, Clabots C, Johnston BD, Kuskowski MA, MASTER Investigators. 2012. Comparison of *Escherichia coli* ST131 pulsotypes, by epidemiologic traits, 1967–2009. Emerg Infect Dis 18:598–607. https://doi.org/10.3201/eid1804.111627.
- Nicolas-Chanoine MH, Bertrand X, Madec JY. 2014. Escherichia coli ST131, an intriguing clonal group. Clin Microbiol Rev 27:543–574. https://doi .org/10.1128/CMR.00125-13.
- Peirano G, Richardson D, Nigrin J, McGeer A, Loo V, Toye B, Alfa M, Pienaar C, Kibsey P, Pitout J. 2010. High prevalence of ST131 isolates producing CTX-M-15 and CTX-M-14 among extended-spectrum-beta-lactamaseproducing *Escherichia coli* isolates from Canada. Antimicrob Agents Chemother 54:1327–1330. https://doi.org/10.1128/AAC.01338-09.
- Bax BD, Chan PF, Eggleston DS, Fosberry A, Gentry DR, Gorrec F, Giordano I, Hann MM, Hennessy A, Hibbs M, Huang J, Jones E, Jones J, Brown KK, Lewis CJ, May EW, Saunders MR, Singh O, Spitzfaden CE, Shen C, Shillings A, Theobald AJ, Wohlkonig A, Pearson ND, Gwynn MN. 2010. Type IIA topoisomerase inhibition by a new class of antibacterial agents. Nature 466:935–940. https://doi.org/10.1038/nature09197.
- Biedenbach DJ, Bouchillon SK, Hackel M, Miller LA, Scangarella-Oman NE, Jakielaszek C, Sahm DF. 2016. *In vitro* activity of gepotidacin, a novel triazaacenaphthylene bacterial topoisomerase inhibitor, against a broad spectrum of bacterial pathogens. Antimicrob Agents Chemother 60: 1918–1923. https://doi.org/10.1128/AAC.02820-15.
- Flamm RK, Farrell DJ, Rhomberg PR, Scangarella-Oman NE, Sader HS. 2017. Gepotidacin (GSK2140944) in vitro activity against gram-positive and gramnegative bacteria. Antimicrob Agents Chemother 61:e00468-17. https://doi .org/10.1128/AAC.00468-17.
- Gibson EG, Bax B, Chan PF, Osheroff N. 2019. Mechanistic and structural basis for the actions of the antibacterial gepotidacin against *Staphylococcus aureus* gyrase. ACS Infect Dis 5:570–581. https://doi.org/10.1021/ acsinfecdis.8b00315.
- O'Riordan W, Tiffany C, Scangarella-Oman N, Perry C, Hossain M, Ashton T, Dumont E. 2017. Efficacy, safety, and tolerability of gepotidacin (GSK2140944) in the treatment of patients with suspected or confirmed gram positive acute bacterial skin and skin structure infections. Antimicrob Agents Chemother 61:e02095-16. https://doi.org/10.1128/AAC .02095-16.
- Taylor SN, Morris DH, Avery AK, Workowski KA, Batteiger BE, Tiffany CA, Perry CR, Raychaudhuri A, Scangarella-Oman NE, Hossain M, Dumont EF. 2018. Gepotidacin for the treatment of uncomplicated urogenital gonorrhea: a phase 2, randomized, dose-ranging, single-oral dose evaluation. Clin Infect Dis 67:504–512. https://doi.org/10.1093/cid/ciy145.
- Scangarella-Oman NE, Hossain M, Dixon PB, Ingraham K, Min S, Tiffany CA, Perry CR, Raychaudhuri A, Dumont EF, Huang J, Hook EWIII., Miller LA. 2018. Microbiological analysis from a phase 2 randomized study in adults evaluating single oral doses of gepotidacin in the treatment of uncomplicated urogenital gonorrhea caused by *Neisseria gonorrhoeae*. Antimicrob Agents Chemother 62:e01221-18. https://doi.org/10.1128/ AAC.01221-18.
- 19. Hoover JL, Singley CM, Elefante P, Rittenhouse S. 2019. Efficacy of human

exposures of gepotidacin (GSK2140944) against *Escherichia coli* in a rat pyelonephritis model. Antimicrob Agents Chemother 63:e00086-19. https://doi.org/10.1128/AAC.00086-19.

- Negash K, Andonian C, Felgate C, Chen C, Goljer I, Squillaci B, Nguyen D, Pirhalla J, Lev M, Schubert E, Tiffany C, Hossain M, Ho M. 2016. The metabolism and disposition of GSK2140944 in healthy human subjects. Xenobiotica 46:683–702. https://doi.org/10.3109/00498254.2015.1112933.
- Tiffany CA, Hossain M, McDonald M, Patel A, Lerman S, Patel P, Widdowson KL, Dumont EF. 2013. Safety and pharmacokinetics of single escalating oral doses of GSK2140944, a novel bacterial topoisomerase inhibitor. Abstr 53rd Intersci Conf Antimicrob Agents Chemother, abstr F-1218.
- 22. Tiffany CA, Hossain M, McDonald M, Lerman S, Dumont EF. 2014. Safety and pharmacokinetics of single escalating IV doses of GSK2140944, a novel bacterial topoisomerase inhibitor. Abstr 54th Intersci Conf Antimicrob Agents Chemotherapy, abstr F-277.
- Tiffany CA, Hossain M, McDonald M, Lerman S, Dumont EF. 2014. Safety and pharmacokinetics of repeat escalating IV doses of GSK2140944, a novel bacterial topoisomerase inhibitor. Abstr 54th Intersci Conf Antimicrob Agents Chemotherapy, abstr F-278.
- Hossain M, Zhou M, Tiffany C, Dumont E, Darpo B. 2017. A phase I, randomized, double-blinded, placebo- and moxifloxacin controlled, fourperiod crossover study to evaluate the effect of gepotidacin on cardiac conduction as assessed by 12-lead electrocardiogram in healthy volunteers. Antimicrob Agents Chemother 61:e02385-16. https://doi.org/10.1128/AAC .02385-16.
- Bulik CC, Okusanya ÓO, Lakota EA, Forrest A, Bhavnani SM, Hoover JL, Andes DR, Ambrose PG. 2017. Pharmacokinetic-pharmacodynamic evaluation of gepotidacin against Gram-positive organisms using data from murine infection models. Antimicrob Agents Chemother 61:e00115-16. https://doi.org/10.1128/AAC.00115-16.
- VanScoy BD, Fikes S, Bhavnani SM, Conde H, Scangarella-Oman NE, Ambrose PG. 2018. A hollow-fiber infection model to evaluate the prevention of on-therapy resistance of Neisseria gonorrhoeae to gepotidacin, poster 314. STD Prevention Conf, Washington, DC, 27 to 30 August 2018. https://www.fibercellsystems.com/wp-content/uploads/2018/09/00479_gepo_gc_hfim_16aug18.pdf.
- Clinical and Laboratory Standards Institute. 2018. Methods for dilution antimicrobial susceptibility tests for bacteria that grow aerobically, 11th ed. Document M07-A11. Clinical and Laboratory Standards Institute, Wayne, PA.
- Clinical and Laboratory Standards Institute. 2019. Performance standards for antimicrobial susceptibility testing, 29th ed. Document M100-S29. Clinical and Laboratory Standards Institute, Wayne, PA.
- 29. Food and Drug Administration, Center for Drug Evaluation and Research. 2019. Uncomplicated urinary tract infections: developing drugs for treatment. https://www.fda.gov/media/129531/download.
- European Medicines Agency. 2018. Guideline on the evaluation of medicinal products indicated for treatment of bacterial infections, rev 3. EMA/844951/2018 rev 3. https://www.ema.europa.eu/en/documents/ scientific-guideline/draft-guideline-evaluation-medicinal-products -indicated-treatment-bacterial-infections-revision-3_en.pdf.