

Knockdown of CypA inhibits interleukin-8 (IL-8) and IL-8-mediated proliferation and tumor growth of glioblastoma cells through down-regulated NF- κ B

Shan Sun · Qiuwei Wang · An Giang · Cong Cheng · Chia Soo ·
Cun-Yu Wang · Linda M. Liau · Robert Chiu

Received: 28 January 2010 / Accepted: 28 April 2010 / Published online: 9 May 2010
© The Author(s) 2010. This article is published with open access at Springerlink.com

Abstract Although cyclophilin A (CypA) has been reported to be over-expressed in cancer cells and solid tumors, its expression and role in glioblastomas have not been studied. Herein, we show that expression of CypA in human glioblastoma cell lines and tissues is significantly higher than in normal human astrocytes and normal counterparts of brain tissue. To determine the role of over-expressed CypA in glioblastoma, stable RNA interference (RNAi)-mediated knockdown of CypA (CypA KD) was performed in glioblastoma cell line U87vIII (U87MG · Δ EGFR). CypA KD stable single clones decrease

proliferation, infiltration, migration, and anchorage-independent growth in vitro and with slower growth in vivo as xenografts in immunodeficient nude mice. We have also observed that knockdown of CypA inhibits expression of interleukin-8 (IL-8), a tumorigenic and proangiogenic cytokine. Conversely, enforced expression of CypA in the CypA KD cell line, Ud-12, markedly enhanced IL-8 transcripts and restored Ud-12 proliferation, suggesting that CypA-mediated IL-8 production provides a growth advantage to glioblastoma cells. CypA knockdown-mediated inhibition of IL-8 is due to reduced activity of NF- κ B, which is one of the major transcription factors regulating IL-8 expression. These results not only establish the relevance of CypA to glioblastoma growth in vitro and in vivo, but also suggest that small interfering RNA-based CypA knockdown could be an effective therapeutic approach against glioblastomas.

S. Sun · Q. Wang · A. Giang · R. Chiu (✉)
Dental Research Institute, UCLA School of Dentistry,
Los Angeles, CA 90095, USA
e-mail: rchiu@dentistry.ucla.edu

C. Cheng
Department of Biological Sciences and Biotechnology, School
of Medicine, Tsinghua University, Beijing, China

C. Soo
Department of Orthopaedic Surgery, David Geffen School
of Medicine, UCLA, Los Angeles, CA 90095, USA

C.-Y. Wang
Division of Oral Biology and Medicine, UCLA School
of Dentistry, Los Angeles, CA 90095, USA

L. M. Liau
Department of Neurosurgery, David Geffen School of Medicine,
UCLA, Los Angeles, CA 90095, USA

R. Chiu
Department of Surgery/Oncology, David Geffen School
of Medicine, UCLA, Los Angeles, CA 90095, USA

R. Chiu
Jonsson Comprehensive Cancer Center, UCLA, Los Angeles,
CA 90095, USA

Keywords Cyclophilin A · IL-8 · Glioblastoma ·
RNA interference · Tumor growth

Introduction

Malignant gliomas are among the most devastating of cancers, leading to death in most cases [1]. They present unique challenges for therapy due to their difficult access locations, aggressive biological behavior, and diffuse infiltrative growth. Cyclosporin A (CsA) has been reported to have a chemotherapeutic effect in a variety of cancer cells, including human gliomas [2–4]. In a human glioblastoma cell line and glioblastoma cells from brain slices, CsA was shown to affect their growth/survival and migration/invasion [3, 4]. The mechanisms of these effects may relate to the biochemical properties of CsA such as inhibition of cyclophilin isomerase and calcineurin

phosphatase activities that mediate immunosuppressive effects that can alter several signal pathways.

CypA is found in normal cells, comprises ~0.6% of total cytosolic protein, and is widely conserved among prokaryotes and eukaryotes [5]. It is a member of the peptidyl-prolyl isomerase (PPI) family, a group of proteins that catalyze *cis-trans* isomerization of peptidyl-prolyl bonds during protein folding and/or conformational changes [6, 7]. CypA was first identified as the primary intracellular target of the immunosuppressive drug, CsA [8]. The immunosuppressive activity of CsA is thought to result from engagement of calcineurin by the CsA-CypA complex [9], an observation supported by the finding that CypA knock-out mice are resistant to immunosuppression by CsA [10].

Several lines of research have revealed that PPIs such as CypA may function as molecular signaling “switches” that can act as novel molecular timers to help control the amplitude and duration of a cellular process [11]. Moreover, the role of CypA nuclear translocation or CypA’s participation in activation of other factors or their nuclear translocation also impacts various cellular functions [12–15]. One recent report demonstrated that knockdown of *CypA* inhibited signal transducer and activator of transcription 3 (Stat3) interleukin-6-induced tyrosine phosphorylation and nuclear translocation, resulting in altered gene expression in myeloma cell lines [16]. The CypA inhibitor, CsA, exhibited similar effects. Additional data also suggest that CypA may contribute to the pathology of certain human malignancies. Several experiments have demonstrated that CypA is substantially up-regulated in various types of cancers and cancer cell lines [17–20]. These over-expressed CypAs mediate multiple cellular processes that can confer growth advantage to tumor cells in the neoplastic microenvironment. This was substantiated by CypA over-expression in the SILEK-transformed small airway epithelial cell line, which showed that increased CypA expression correlated with dramatically faster tumor formation *in vivo* [21]. Furthermore, CypA over-expression in cancer cells conferred resistance to anticancer drugs and hypoxia-induced cell death [22, 23]. Therefore, targeted CypA and inhibition of its PPI activity have been suggested as novel therapeutic strategies for treatment of human cancers.

Over-expression of CypA has also been implicated in other pathological processes such as rheumatoid arthritis [24, 25]. In rheumatoid arthritis patients, over-expressed CypA stimulates production of inflammatory cytokines such as tumor necrosis factor alpha (TNF- α), interleukin-1 β (IL-1 β), IL-8, monocyte chemoattractant protein-1 (MCP-1), and matrix metalloproteinase 9 (MMP-9) [25]. These CypA-stimulated products could arise through a pathway that is dependent on NF- κ B activation. However, the means by which CypA mediates signaling to activate NF- κ B remains to be investigated. Similarly, stably

expressed CypA in the SK-Hep1 cell line revealed that CypA up-regulates the expression of many cytokine-related genes such as IL-8, IL-1 β , IL-6, CXCL1, CXCL2, and CXCL3 [22]. These up-regulated cytokines and chemokines may confer tumor cell growth advantage in the neoplastic microenvironment. Therefore, a strategy to reverse these effects might offer attractive options for novel therapeutics.

In human gliomas, IL-8 is expressed and secreted at high levels both *in vitro* and *in vivo*, and recent experiments suggest it is critical to glial tumor neovascularity and progression [26]. IL-8 could act as an inflammatory chemoattractant as part of the host response to neoplasia. Moreover, it also acts as a more general proinflammatory factor released in response to tissue stress and necrosis, a proangiogenic factor that promotes new vessel growth, or an autocrine growth factor secreted by tumor cells to promote their own growth. Secretion of IL-8 could also arise from any other cellular compartments such as microglia, macrophages, and neutrophils. IL-8 gene expression and peptide secretion by glioblastoma cells can be dramatically enhanced by TNF- α and IL-1 [27, 28]. The IL-8 promoter contains binding sites for transcription factors NF- κ B, AP-1, and C-EBP/NF-IL-6, among others, which are mediated to respond to hypoxia, Fas ligation, death receptor activation, cytosolic calcium, TNF- α , IL-1, other cytokines, and various cellular stresses increase IL-8 expression [29–31].

Expression of CypA and its relationship to highly expressed IL-8 in glioblastoma has not been studied. Herein we report that CypA was strongly expressed in a majority of human glioblastomas and cell lines we tested. This ubiquitous over-expression of CypA in glioblastoma suggests that CypA could be an important component of neoplastic transformation. To determine the role of over-expressed CypA in glioblastoma, stable RNA interference (RNAi)-mediated knockdown of CypA (CypA KD) in a glioblastoma cell line was performed. The CypA KD stable single clones showed decreased proliferation, infiltration, migration, and anchorage-independent growth *in vitro*, as well as slower tumor growth *in vivo*. The reversal of the tumor phenotype, and in particular, decreased cell proliferation and slow tumor growth, in CypA KD clones was associated with reduced NF- κ B activity and down-regulated IL-8 expression. Thus, targeted knockdown of CypA could be an effective therapeutic approach for glioblastomas.

Materials and methods

Cell culture and cell proliferation assay

Human glioblastoma cell lines U87vIII, U87MG, T98G, Ln229, U138, U378 and immortalized normal human

astrocytes were grown in DMEM with low glucose and supplemented with 10% fetal bovine serum. Panc-1 and A549 cells were purchased from ATCC. Panc-1 and A549 cells were cultured in DMEM and RPMI 1640 (Gibco), respectively. Cells were maintained at 37°C in a humidified chamber containing 5% CO₂. Cell proliferation was assessed by the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) method (MTT cell proliferation assay, ATCC). Cells (1500/well) were seeded in triplicate into 96-well plates and cultured for various periods. At each time point, 10 µl of MTT reagent was added to each well and incubated for 2 h, followed by addition of 100 µl of detergent reagent to each well for additional 3 h incubation in the dark. At the end of the reaction, the absorbance in each well was measured at 595 nm in a microtiter plate reader (BIO-TEK). Control wells of medium alone were used as blanks. For the rescue experiments, cells were transfected with vector alone or rescue construct, using lipofectamine 2000 (Invitrogen). Cells were then seeded into 96-well plates 18 h post-transfection. Time zero (0 day) was 6 h after seeding.

Immunohistochemistry staining

A GL802 tissue microarray was purchased from U.S. Biomax, Inc. (Rockville, MD). It contains 35 samples of glioblastoma (grade IV), two samples of adjacent normal tissue, and three samples of normal brain tissue from autopsies, with a duplicate core per sample format. The tissues were formalin-fixed and paraffin-embedded. Array sections were mounted on the positively charged glass slide. Tissue sections were deparaffinized, hydrated, and hybridized with rabbit anti-cyclophilin A antibody (Abcam), followed by incubation of sections with ImmPress anti-rabbit Ig (Vector Lab). After washing the slides three times in buffer 5 min each time, the sections were incubated in peroxidase substrate DAB solution until desired stain intensity developed. Counterstaining with hematoxylin was used for blue coloring of negative nuclei.

Xenograft tumor formation

All animal work described herein was approved by the Chancellor's Animal Research Committee at UCLA. Following determination of absence of mycoplasma contamination by nucleic acid hybridization, one million cells each of two CypA KD single clones, Ud-12 and Ud-3, scrambled clone (SC), and parental U87vIII were injected in 100 µl DMEM subcutaneously into the flank of 4-week-old, female, NCRU-M immunodeficient mice (Taconic Farms Inc, Hudson, New York); three animals per cell line were used. Tumor dimensions were measured every

2–3 days with calipers and volume was calculated by the formula $V = 0.5 \times L \times W^2$, where L is long diameter and W is short diameter.

Reverse transcription and quantitative real-time PCR analysis

Total RNA was isolated from cells, using TRIzol reagent (Invitrogen) according to the manufacturer's protocols. One microgram of total RNA was used for cDNA synthesis, using Superscript III (Invitrogen) with an oligo(dT)₁₅ primer. Quantitative real-time PCR (QRT-PCR) was performed using the Power SYBR Green PCR Master Mix (Applied Biosystem) and detected by the 7300 Real Time PCR System (Applied Biosystem). The relative expression of *IL-8* mRNA normalized to the internal reference *18S rRNA* was analyzed using the $2^{-\Delta\Delta C_T}$ method [32]. The amount of *IL-8* transcripts in U87vIII cells, or U87vIII cells transfected with vector alone, was designated as 1.0. Bars indicate the standard error from three individual experiments. The primers used for real-time PCR were: *IL-8*, 5'-GGGTTGTGGAGAA-GTTTTTG-3' (forward), 5'-GTTTCACTGGCATCTTCACTG-3' (reverse); *18S rRNA*, 5'-A-GTTGGTGGAGCGATTT-GTC-3' (forward), 5'-TATTGCTCAATCTCGGGTGG-3' (reverse).

Scratch-induced migration assay

U87vIII, Ud-3, and scrambled clone were separately seeded onto six-well plates at a density of 5×10^5 per well. After 24 h, confluent monolayers were scratched with a 1 ml plastic pipet tip to create a uniform, cell-free area. Then the cells were incubated at 37°C and photographed at time points 0, 16, and 24 h on an Olympus IX71 inverted microscope using an UPlanFL 4×/0.13 N.A. PhL objective (Olympus). Digital images were acquired with a MicroFire digital camera driven by the PictureFrame imaging software. The distance between the opposing edges was measured using the Adobe PhotoShop software.

Matrigel infiltration assay

An infiltration assay was performed using a six-well plate transwell insert with 8-µm pore size filters (Fisher Scientific) coated with Matrigel (BD Biosciences). Cells (5×10^5) from cell lines U87vIII, Ud-3, Ud-12, and scrambled clone containing 1.5 ml of serum-free DMEM medium, were separately placed into each transwell insert. Each well in the lower chambers was filled with 2.5 ml of DMEM medium containing 10% fetal bovine serum (FBS). After 24 h of incubation at 37°C in a cell culture chamber, the cells on the top surface of the filter were wiped off with cotton swabs. Infiltrated cells on the lower surface of the

filter were fixed and stained with crystal violet (Fisher Scientific). Random fields were counted under a light microscope. Olympus microsuite™ B3SV software was used for photographing.

Anchorage-independent growth in soft agar

To evaluate the ability of individual cell lines to grow in an anchorage-independent manner, 3000 cells from each cell line (U87vIII, Ud-3, Ud-12, and scrambled clone) were each mixed with 0.5 ml of UltraPure™ Agarose (Invitrogen) in DMEM medium with 10% FBS (final concentration 0.3%), then overlaid with 1 ml of 0.6% agarose per well of six-well plates (in duplicate). The cells were given 0.5 ml of 10% FBS DMEM per well every 5 days, and incubated for 21 days, then stained by crystal violet. Colonies were counted using a microscope.

IL-8 ELISA

IL-8 ELISA was performed using the Endogen® Human IL-8 ELISA Kit (Thermo Scientific, Inc.) In brief, cells were seeded into 24-well plates 24 h prior to assay, then treated with different chemicals, as described in the Results. Supernatants from cells were collected and analyzed for IL-8 levels according to the manufacturer's instructions. Absorbance was then measured on an ELISA plate reader set at 450 nm and 550 nm. To correct for optical imperfections on the microplate, 550-nm values were subtracted from 450-nm values. The test samples multiply the interpolated value obtained from the standard curve by a dilution factor to calculate pg/ml of human IL-8 in the sample.

Electrophoretic mobility shift assay (EMSA)

Nuclear extracts were prepared using NE-PER nuclear and cytoplasmic extraction reagents (Pierce Biotechnology). EMSA was performed using the Light Shift Chemiluminescent EMSA Kit (Pierce Biotechnology) according to the manufacturer's directions. In brief, the biotin end-labeled probe (100 fmol) was incubated with nuclear extract (15 µg) and electrophoresed on a native polyacrylamide gel. The DNA was then transferred to a nylon membrane, UV cross-linked, probed with the streptavidin-horseradish peroxidase conjugate, and chemiluminescent substrate was used for detection. The EMSA probes with 5' biotin labels were purchased from Sigma-Aldrich. Nucleotide sequences were as follows: NF-κB oligo, 5'-AGTTGAGGGGAC-TT TCCCAGGC-3'; AP1 oligo, 5'-CGCTTGATGAGTCAGC CGGAA-3'; C/EBP oligo, 5'-GCCATCAGTTGCAAATC GTGGC-3'. For competition experiments, 100-fold molar excess of unlabeled oligos were incubated with the protein

at 25°C for 10 min before addition of labeled probes, and then further incubated for 20 min.

Luciferase reporter assays

The luciferase reporter plasmids driven by -162 to +44 of human *IL-8* promoter (p162-IL-8) and its mutants (p162mtAP1, p162mtC/EBP, and p162mtNF-κB) were kindly provided by James J. Pestka (Michigan State University). Cells were plated into 48-well plates and cultured for 1 day, then transfected with 0.3 µg/well of reporter plasmids and 0.006 µg/well *Renilla* luciferase plasmid, using lipofectamine 2000 (Invitrogen). Firefly and *Renilla* luciferase activity were measured with the dual-luciferase reporter assay system (Promega). Results are presented as relative reporter activity after normalization to the *Renilla* luciferase activity. For promoter activity at basal level, cells were assayed 24 h post-transfection. For TNFα and cyclosporin A (CsA) (Sigma-Aldrich) treatment, cells were incubated with 1 µg/ml CsA for 5 h at 24 h post-transfection, then stimulated with 10 ng/ml TNF-α for another 4 h. Cells were then harvested and promoter activity determined.

Statistical analysis

Results are expressed as means ± SE from at least three independent experiments. Statistical analysis was performed using student's *t* test. Unless otherwise indicated, *P* < 0.05 was deemed significant.

Results

Elevated cyclophilin A (CypA) in human glioblastoma cell lines and tissue specimens

CypA is substantially up-regulated in various types of tumors and cancer cell lines [17–21]. To investigate whether elevated CypA at the protein level is also present in glioblastoma cell line, U87MG and its EGFR deletion mutant, U87vIII, and other glioblastoma cell lines, we performed Western blot analysis of cell lysates prepared from these cell lines and compared them to those isolated from normal human astrocytes (NHA). Our data demonstrate that CypA protein expression in glioblastoma cell lines is significantly higher than the control (Fig. 1a, lanes 1–3). This elevated CypA expression is comparable to that seen in the human pancreatic cancer cell line, Panc-1 [17], and the human lung epithelial carcinoma cell line, A549 [21] (Fig. 1a, lanes 4–14). These data clearly demonstrate that CypA is over-expressed in all glioblastoma cell lines we tested.

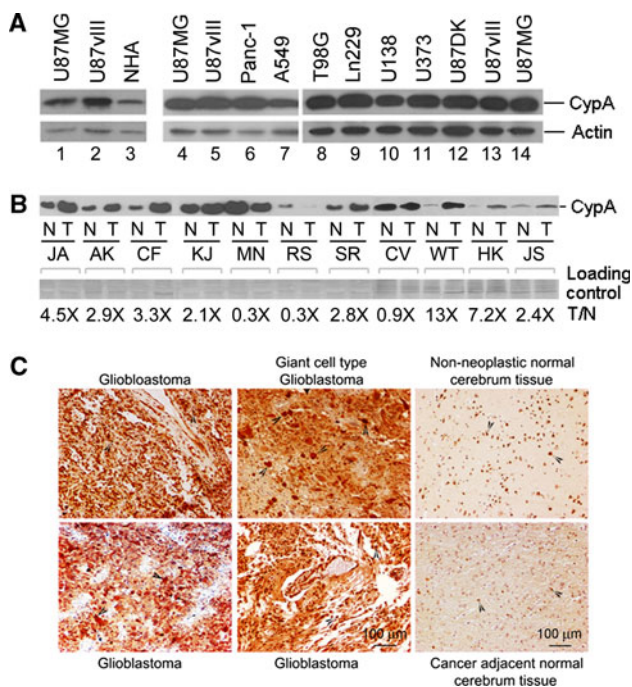


Fig. 1 Over-expression of CypA in glioblastoma cell lines and human tissue specimens. **a** Elevated CypA at the protein level in glioblastoma cell lines. Five micrograms of cell lysate prepared from U87MG and U87MG. Δ EGFR (U87vIII) and normal human astrocyte (NHA) were subjected to Western blot analysis to detect CypA expression levels (lanes 1–3). Actin was used as a loading control. Over-expression of CypA levels in glioblastoma cell lines is comparable with those in pancreatic and non-small cell lung carcinoma cell lines (lanes 6 and 7). Lanes 8–11 are other glioblastoma cell lines, and U87DK (lane 12) represents K721 mutation and kinase inactive at EGFR. **b** Relative CypA levels in human glioblastoma and normal tissues. Ten micrograms of tissue lysates were subjected to Western blot analysis. Coomassie brilliant blue R250-stained signals on the nitrocellulose membrane served as internal controls. (N, normal brain tissue; T, tumor tissue). **c** CypA expression in glioblastoma multiforms (GBMs). Deparaffinized and hydrated tissue sections were hybridized with rabbit anti-cyclophilin A antibody (Abcam), followed by incubating sections with ImmPress anti-rabbit Ig (Vector Lab) and DAB (brown) staining. Arrows indicate typical positive staining for CypA. Non-neoplastic normal cerebrum tissue and cancer-adjacent normal cerebrum tissue were used as controls

To correlate CypA over-expression in glioblastoma cell lines with glioblastoma specimens, eleven human glioblastoma specimens and surrounding normal brain tissue in each specimen were obtained from the laboratory of Dr. Linda Liao (UCLA). Coomassie brilliant blue R250-stained signals on the nitrocellulose membrane served as internal controls prior to immunoblot analysis. As shown in Fig. 1b, the human glioblastoma specimens had 2.1- to 13-fold increases in CypA protein levels compared to adjacent normal tissue. About 73% of tumor tissues examined contained over-expressed CypA, suggesting that CypA could be involved in glioblastoma pathogenesis. In addition, we obtained a GL802 tissue microarray from U.S.

Biomax, Inc. (Rockville, MD). Deparaffinized and hydrated tissue sections were hybridized with rabbit anti-cyclophilin A antibody (AbCame), followed by incubation of the sections with ImmPress anti-rabbit Ig (Vector Lab) and DAB (brown) staining. Counterstaining with hematoxylin was used for blue coloring of negative nuclei. CypA immunohistochemical quantitation in glioblastoma multiforms (GBMs) is shown in Table 1. About 86% of tumor tissues contained over-expressed CypA. A representative CypA stain is shown in Fig. 1c.

CypA knockdown correlated with decreased cell proliferation in vitro and slower tumor growth in vivo

Infection of U87vIII cells with the pSilencer-CypA RNAi retrovirus vector yielded five stable single clones with 70–90% CypA knockdown (CypA KD), as quantitated by chemiluminescence of Western blots (Fig. 2a). Among these single clones, Ud-12 had the greatest CypA KD, and the four other single clones (Ud-3, Ud-8, Ud-13 and Ud-15) had lesser amounts. Single clones selected with pSilencer-scrambled RNAi showed no differences in CypA levels relative to parental U87vIII cells (Fig. 2a). To determine whether the CypA KD single clones had an intrinsic effect on cell proliferation, we performed MTT assays to measure cell proliferation in the CypA KD single clones, Ud-3 and Ud-12, and their parental and scrambled control cells. As shown in Fig. 2b, a difference was noted between knock-downs and parental cells and scrambled control cells. Thus, we conclude that the CypA KD single clones Ud-3 and Ud-12 led to decreased proliferation compared to parental U87vIII and SC.

To determine whether CypA KD single clones have a similar effect of slowing tumor growth in vivo, we grew CypA KD clones as xenografts in immunodeficient nude mice. CypA KD cells Ud-3 and Ud-12 yielded slower-growing tumors than control cells for the U87vIII cells and SC (Fig. 2c). At 21 days, tumors from CypA KD Ud-12 and Ud-3 cells were, on average, 96 and 74% smaller, respectively, than tumors from control cells. These results indicate that CypA KD had a significant effect on tumor growth in vivo.

CypA KD decreases anchorage-independent growth, cell migration and infiltration in vitro

To determine whether the CypA KD single clones can reverse the transformed phenotype of glioblastoma, we examined anchorage-independent growth of CypA KD, scrambled and parental control cells, using a soft agar assay. After 21 days of incubation, the cells were stained with crystal violet and imaged. Colonies larger than 0.2 mm² were counted. Results of a typical experiment

Table 1 CypA expression in GBMs

Bank case number	Cypa (set 1)		Cypa (set2)	
	C	T	C	T
1	–		+	
2	–		+	
3	–		–	
4	–		–	
A1		+++		+++
A2		++		+++
A3		++		++
A4		+++		++++
A5		+++		+++
B1		++		+++
B2		+++		+++
B3		+++		+++
B4		+		++++
B5		++		+
C1		++		+++
C2		+++		++
C3		+++		+++
C4		–		+++
C5		++		++
D1		+++		+++
D2		++		++
D3		++++		++++
D4		+		+
D5		+++		+++
E1		+		++
E2		++++		++++
E3		++++		++++
E4		+++		+++
E5		++++		++++
F1		–		–
F2		+		++
F3		++		+
F4		++		++
F5		++++		++++
G1		++++		++++
G2		++++		++++
G3		+++		+++
G4		+++		++++
G5		+++		+++

CypA immunohistochemical quantitation in GBMs. *C* normal cerebrum tissue; *T* tumor. Scoring: –, no staining; +, staining of less 10% of cells; ++, staining of 20–30% of cells; +++, staining of 30–60% of cells; +++++, staining of greater than 60% of cells

showed that U87vIII cells and SC formed significant numbers of colonies in soft agar. The Ud-12 clone caused significant decreases in colony numbers (Fig. 3a); notably, Ud-12 also decreased colony sizes. Taken together, these

results indicate that CypA KD inhibits anchorage-independent growth of U87vIII cells. Thus, we concluded that CypA KD attenuates the transformed phenotype of U87vIII cells.

To investigate the effects of CypA KD on a second manifestation of the transformed phenotype, cell motility of glioblastoma cells, we performed a scratch-induced migration assay. Subconfluent monolayers of CypA KD single clones, parental U87vIII cells, and SC were wounded at time 0 with a plastic scraper to create a cell-free space 2 mm wide, and were monitored at 16 and 24 h after wounding by photomicrograph. The U87vIII cells and SC migrated into the cell-free region, and the wounded area was nearly completely recovered after 24 h (Fig. 3b). However, recovery of the wound area in the CypA KD Ud-3 cells was only 50% at the same time point (Fig. 3b). These results demonstrated that gap closure was strongly attenuated in the CypA KD single clone, Ud-3, which seemed to progress more slowly than the control cells.

Cell infiltration is similar to cell migration, but it requires a cell to migrate through an extracellular matrix or a basement membrane extract barrier, and then become established in a new location. Transwell plates from Corning Life Sciences provide a relatively simple approach to performing cell infiltration assays in vitro. The effect of CypA KD on cell infiltration was evaluated using Matrigel as an extracellular matrix component. Chambers were incubated for 24 h under normal cell culture conditions. Invading cells on the lower side of the filter were stained with 0.1% crystal violet (Fig. 3c) and quantified after dissolving the cell-bound crystal violet in 10% acetic acid (v/v), followed by measurement of optical density at 540 nm (Fig. 3c). Results are presented as the relative OD 540 nm reading.

CypA KD increases apoptosis under the condition of serum withdrawal

It has been reported that over-expression of CypA protects cells from apoptosis [23]. To determine whether CypA KD would sensitize cells to apoptosis under stress conditions, we used a well-established in vitro model, serum withdrawal, to induce apoptosis. The CypA KD single clones, Ud-3 and Ud-12, and parental U87vIII control cells were grown in 96-well plates in media without serum for 6 days. At different time points, cells were subjected to MTT assays to measure cell viability. After 6 days, greater cell death was observed in the KD cells than in the control cells (Fig. 4). As shown in Fig. 4, viable cells of the CypA KD clones, Ud-3 and Ud-12, are significantly reduced on day 6, suggesting that KD cells are more sensitive to serum withdrawal than the control cells.

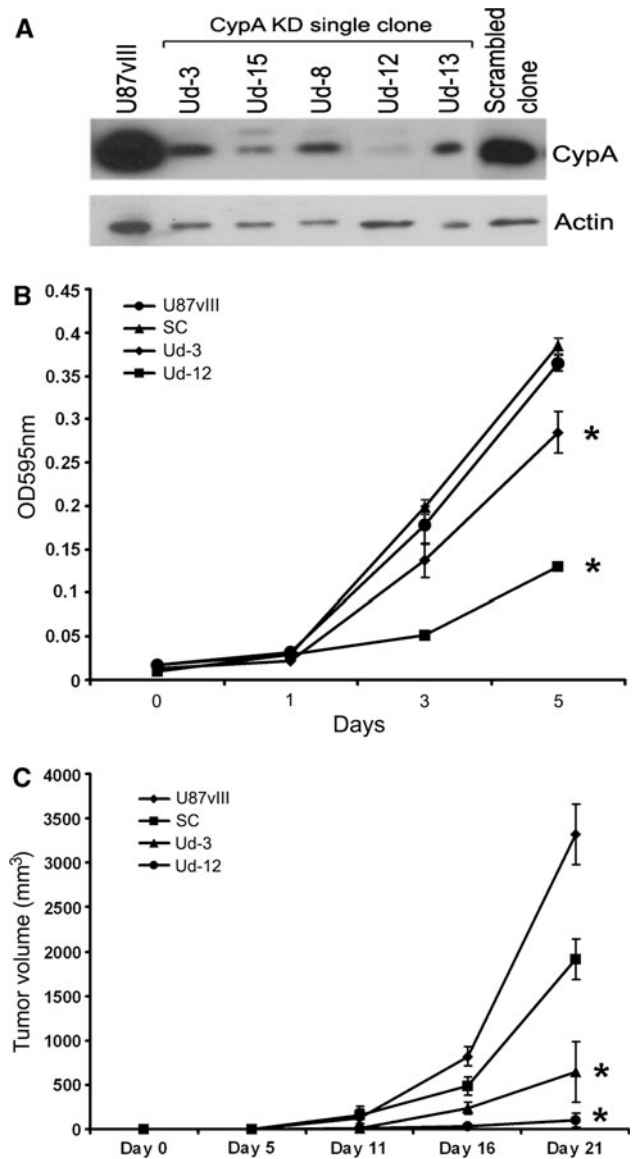
Fig. 2 CypA KD single clones result in decreased cell proliferation and tumor growth. **a** CypA KD stable single clones generated from U87vIII cells. U87vIII cells were seeded into 24-well plates at 5×10^4 cells per well 1 day before infection with enriched viral media prepared from the CypA-specific siRNA insert, scrambled siRNA vector, and linearized retrovirus vector (supplied with the pSilencer H1-Retro kit purchased from Ambion). Cells were transferred into 6-well plates 3 days post-infection and selected with 2 μ g/ml puromycin for 2 weeks. Puromycin-resistant clones were harvested and lysed with RIPA buffer supplemented with a protease inhibitor cocktail (Sigma). Five micrograms of total protein was used for Western blot analysis. Membranes probed with actin antibody were used as loading controls. **b** CypA KD results in decreased cell proliferation. Ud-3 and Ud-12 were compared to parental U87vIII cells and scrambled clone (SC), using MTT assays according to the manufacturer's protocol. Values of OD 595 absorbance at each time point were the average of triplicate individual readings (bars, SE; *, $P < 0.05$ compared with parental wild-type cells). **c** CypA KD decreases tumor growth. Taconic Balb/c/NIH(s) female nu/nu mice were used for xenografts. One million cells were injected. At each time point, tumor dimensions were measured with calipers, and volume was calculated as $V = 0.5 \times L \times W^2$. Three mice were used at each time point and each cell type (bars, SE; *, $P < 0.05$ compared with parental wild-type cells)

CypA KD in the U87vIII glioblastoma cell line significantly reduces IL-8 expression

To determine whether CypA KD can affect expression of cytokines such IL-8 in the glioblastoma cell line, U87vIII, real-time RT-PCR was performed to assess cytokine expression in the CypA KD single clones, Ud-12 and Ud-3, as compared to its parental cells and scrambled clone. As shown in Fig. 5a, expression of *IL-8* in Ud-12 and Ud-3 cells was significantly decreased by 90 and 80%, respectively, as compared with parental U87vIII control cells. To determine whether down-regulated *IL-8* occurs at the protein level, we performed ELISA assays using the Endogen® Human IL-8 ELISA Kit (Thermo Scientific Inc). As shown in Fig. 5b, the levels of IL-8 expression in the CypA KD clones, Ud-12 and Ud-3, were 98 and 90% lower than parental U87vIII cells. Thus, this observation is very important regarding reversal of tumor phenotype.

Rescue of the knockdown phenotype restores IL-8 expression and cell proliferation

To ensure that the observed knockdown phenotype described above is the result of silencing *CypA*, the intended target, we introduced siRNA-resistant wild-type *CypA* with myc-tagged plasmid into Ud-12 cells to reverse siRNA effects. Western blot analysis indicated that exogenous myc-tagged CypA was expressed in Ud-12 cells at levels similar to endogenous CypA in wild-type cells (Fig. 6a). We then analyzed their effects on IL-8 expression and cell proliferation. Results shown in Fig. 6b and c demonstrate that re-introduction of wild-type CypA



restored the decreased IL-8 levels and cell proliferation caused by CypA RNAi, indicating that these defects were due to a reduction of CypA. As reported previously, CypA may contribute to proliferation, the inflammatory process, and angiogenesis mediated through activation of cytokine IL-8 [22, 25]. The mechanism for CypA activation of IL-8 and CypA KD reversal remains to be elucidated.

Identification of CypA-associated transcription factor(s) in regulation of IL-8 expression in U87vIII cells

CypA KD inhibits IL-8 expression at both the mRNA and protein levels, suggesting that inhibition of IL-8 expression occurs at the transcriptional level. Although CypA is not a transcription factor and has no DNA-binding motif in the promoter of the *IL-8* gene, CypA-associated *IL-8*

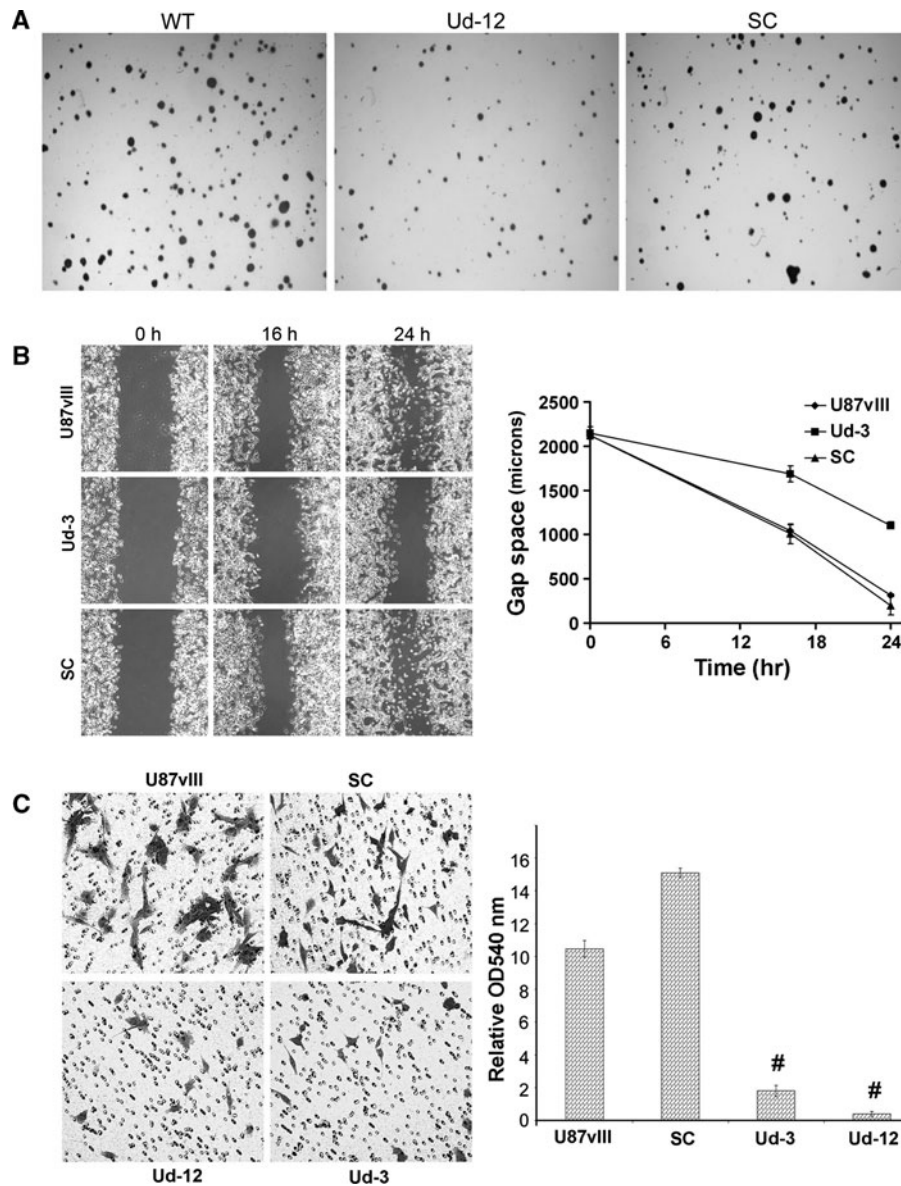


Fig. 3 The CypA KD U87vIII single clone decreases anchorage-independent growth, and reduces cell migration and infiltration ability in vitro. **a** Anchorage-independent growth in vitro. Ud-12 and parental U87vIII cells and SC were plated in duplicate in six-well plates in 0.5% soft agarose on a 1.4% agarose bed agar medium. After 21 days of incubation, the cells were stained with crystal violet and imaged. Colonies larger than 0.2 mm² were counted. Ud-12/U87vIII=47.6%, $P < 0.05$. **b** Knock down of CypA in U87vIII reduces cell migration. Ud-3 cells were seeded into 6-well plates. When cells reached subconfluent monolayers, a scratch wound was inflicted, and the resulting gap was imaged at the time of the wound and after 16 and 24 h. Images are from a typical experiment. **c** Infiltration assay in

vitro. Transwell plates from Corning Life Sciences were used for invasion assays. The effect of CypA KD on cell infiltration was evaluated using Matrigel as an extracellular matrix component. Chambers were incubated for 24 h under normal cell culture conditions. Invading cells on the lower side of the filter were stained with 0.1% crystal violet (left panel) and quantified after dissolving the cell-bound crystal violet in 10% acetic acid (v/v), followed by measurement of optical density at 540 nm. (Columns, mean of the relative OD 540 nm readings from three independent experiments; bars, SE (right panel); #, $P < 0.05$ compared with parental wild-type cells)

expression may be regulated by transcription factors such as NF- κ B, C/EBP, and AP1, which are important in the regulation of *IL-8* expression. We used a wild-type and a series of mutant *IL-8* promoter luciferase constructs to assess the regulation of *IL-8* expression in U87vIII cells mediated by the above transcription factor-binding sites.

U87vIII cells were transiently transfected with a -162-bp *IL-8* promoter construct and mutation constructs containing AP-1 (-126 to -120), C/EBP (-94 to -81), or NF- κ B (-80 to -70) site mutations for 48 h, and then assayed for luciferase activity (Fig. 7a). As shown in Fig. 7b, mutation of the NF- κ B, C/EBP, or AP1 sites resulted in a significant

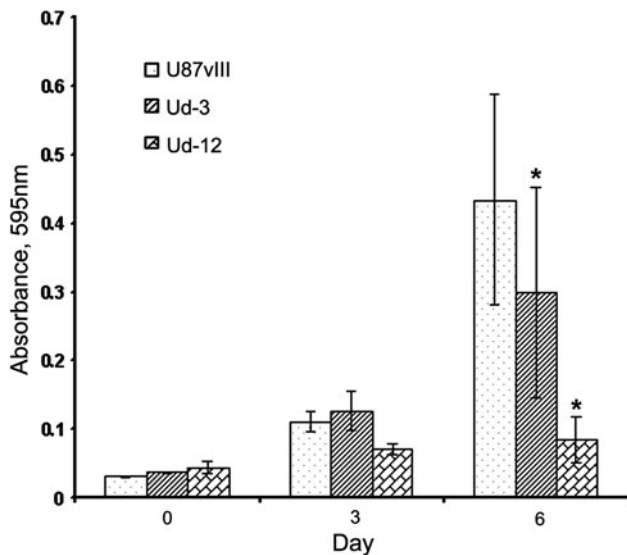


Fig. 4 CypA KD correlates with increased apoptosis under the condition of serum withdrawal. The CypA KD single clones, Ud-3 and Ud-12, and parental U87vIII control cells were grown in 96-well plates in media without serum for 6 days. Cells (10,000/well) were plated in replicates of three. Viable cells were measured at the indicated time points by MTT assays. *Columns*, mean of the relative OD 595 nm readings from three independent experiments; *bars*, SE; *, $P < 0.05$ compared with parental wild-type cells

decrease of luciferase activity, indicating that all of these sites are necessary for regulation of *IL-8* expression in U87vIII cells.

An attempt to identify CypA-associated transcription factors involved in regulation of the *IL-8* promoter in the CypA KD cell line was unsuccessful, due to very low basal levels of *IL-8* production. Therefore, we examined whether inhibition of CypA isomerase activity is able to suppress TNF α -induced *IL-8* expression in the wild-type U87vIII cells. Cells were treated with CsA, which inhibits CypA isomerase and calcineurin phosphatase activity. Cells were also treated with rapamycin as a control, since rapamycin binds to FKBP-12 but not a calcineurin inhibitor. As shown in Fig. 7C, when the cells transfected with the reporter plasmid driven by the wild type *IL-8* promoter were treated with TNF α , luciferase activity increased markedly. This increase was inhibited by CsA but not rapamycin, indicating that CypA isomerase activity is associated with regulation of *IL-8* expression in U87vIII cells.

NF- κ B activity decreases in CypA KD cells

To further identify the transcription factors responsible for down-regulation of *IL-8* expression; we performed EMSAs on nuclear extracts from the CypA KD clones, Ud-3 and Ud-12, to compare to those isolated from parental U87vIII cells and SC. When the NF- κ B oligo was used as probe, the density of the protein/DNA complex in CypA KD clones

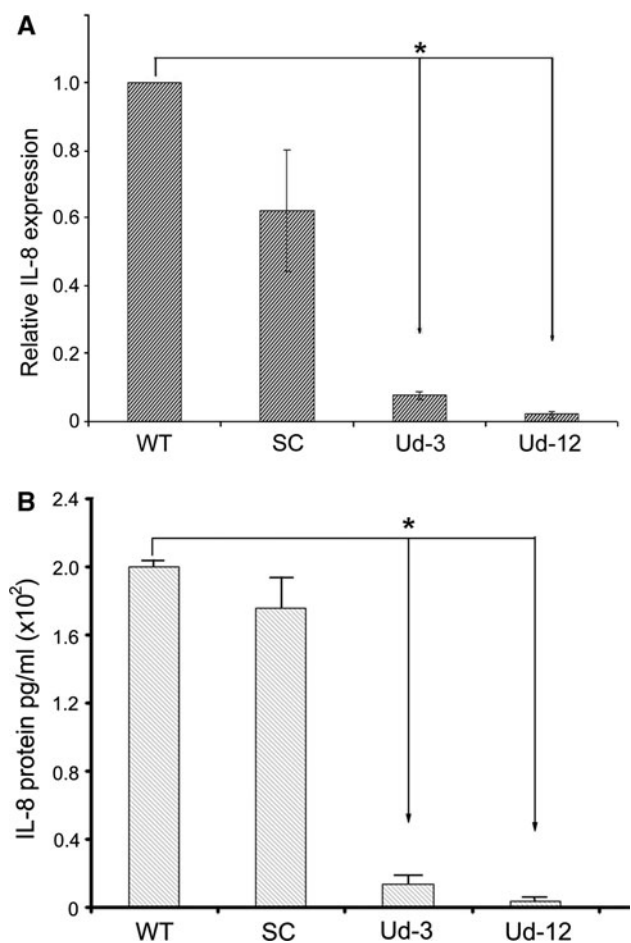


Fig. 5 CypA KD-mediated down-regulation of *IL-8* expression. **a** *IL-8* expression levels were determined by a quantitative real-time PCR analysis. One microgram of total RNA was used for cDNA synthesis, using Superscript III (Invitrogen) with the oligo(dT)₁₅ primer. Quantitative real-time PCR (QRT-PCR) was performed using the Power SYBR Green PCR Master Mix (Applied Biosystem) detected by the 7300 Real Time PCR System (Applied Biosystems). The relative expression of *IL-8* mRNA normalized to the internal reference *18S rRNA* was analyzed using the $2^{-\Delta\Delta CT}$ method (*columns*, mean of results from three independent experiments; *bars*, SE; *, $P < 0.05$ compared with parental wild-type cells). **b** *IL-8* ELISA. Cells were seeded into 24-well plates for 24 h. Cell media was collected for measurement of *IL-8* expression according the manufacturer’s protocol (*columns*, mean of results from three independent experiments; *bars*, SE; *, $P < 0.05$ compared with parental wild-type cells)

was much lower than in the parental U87vIII cells and SC (Fig. 8a). In contrast, when C/EBP and AP1 oligos were used as probes, no alteration was observed when subjecting these cells to EMSA (Fig. 8b and c). We next examined whether CypA KD affects expression and phosphorylation of I κ B α , using Western blot analysis. As shown in Fig. 8d (upper panel), the levels of phosphorylated I κ B α are lower in CypA KD clones than in wild-type cells and SC, whereas the levels of I κ B α in the wild-type cells and SC are lower than in the CypA KD clones. In Fig. 8d (lower

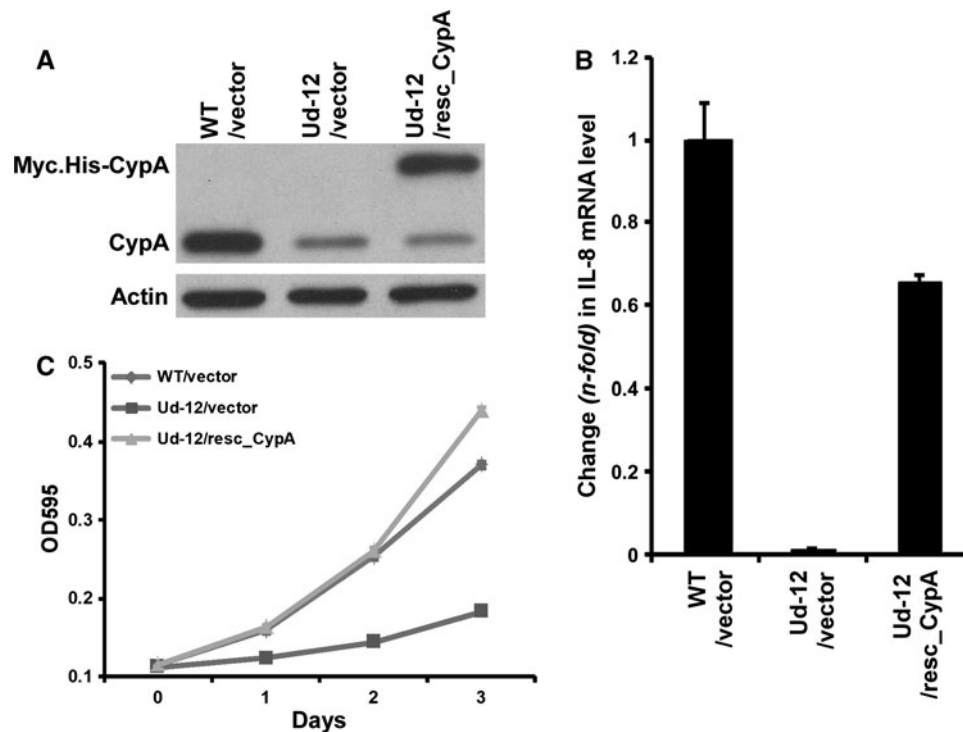


Fig. 6 Expression of a siRNA-resistant wild-type CypA rescue expression plasmid restores *IL-8* expression and cell proliferation defects. **a** Western blot analysis of endogenous CypA and exogenous myc-tagged CypA. U87vIII cells (WT) and CypA KD clone Ud-12 were transfected with the pcDNA3.1/Myc.His vector (vector) or the rescue construct (resc_CypA). Five micrograms of each cell extract were subjected to Western blot analysis probing with anti-CypA and

anti-myc antibodies. **b** Real-time PCR analysis of the *IL-8* mRNA. The values plotted are the mean of three separate experiments. **c** MTT assay of cell proliferation. At various time points, cells were subjected to MTT assays. The absorbance in each well was measured at 595 nm in a microtiter plate reader. Bars indicate the standard error from three individual experiments

panel), nuclear p65 is lower in CypA KD clones than in wild-type cells and SC, which clearly indicated less nuclear translocated NF- κ B in the CypA KD cells and further confirmed our observation above when using EMSA.

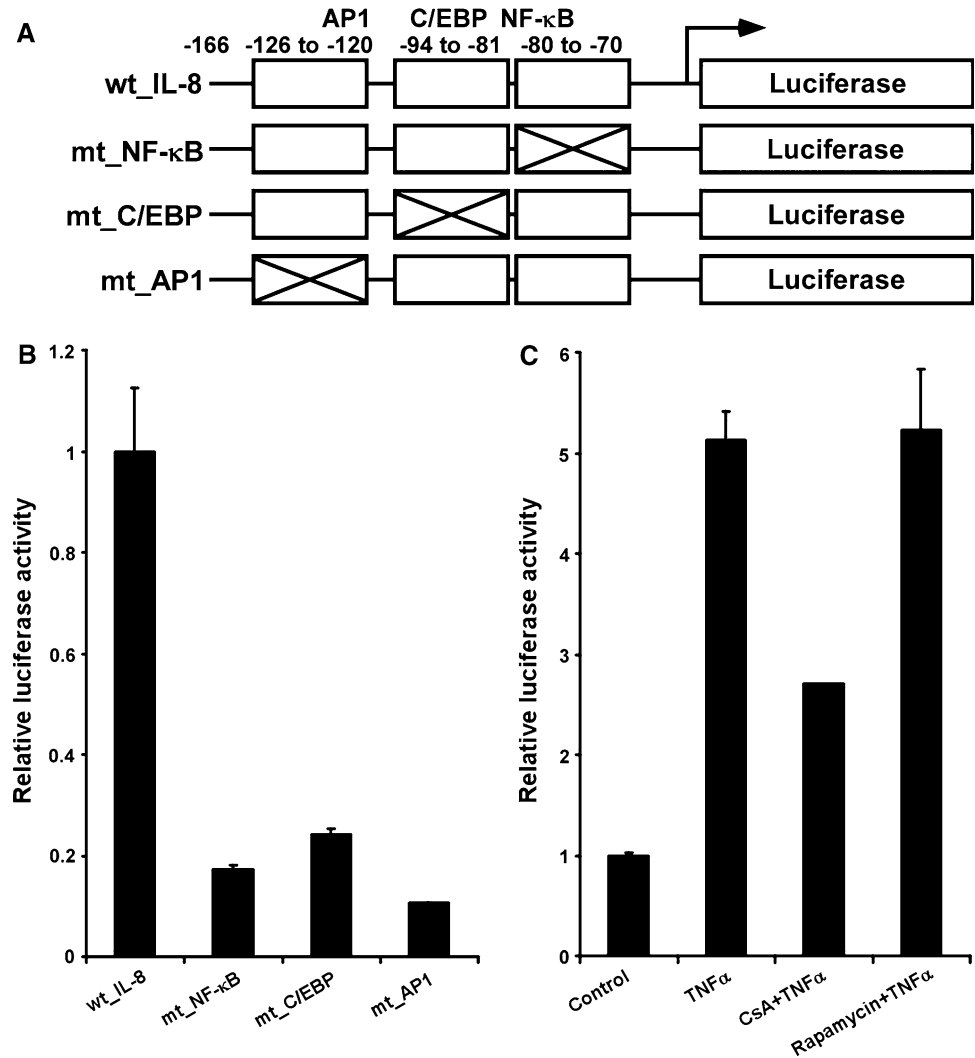
Discussion

It has been suggested that CypA may contribute to the pathology of human malignancies. Indeed, several experiments have demonstrated that CypA is substantially up-regulated in various types of cancers and cancer cell lines, including pancreatic adenocarcinoma [17], non-small cell lung cancer [18], hepatocellular carcinoma [33], endometrial carcinoma [19], and esophageal squamous cell carcinoma [20]. These over-expressed CypA molecules mediate multiple cellular processes and confer growth advantage to tumor cells in the neoplastic microenvironment. Further investigation of the connection between CypA and cancer has led to the conclusion that over-expressed CypA is strongly correlated with malignant lung cancer cells [21, 34]. Our data obtained with glioblastoma cell lines also support this concept. A previous report has demonstrated

that over-expression of CypA in the SILEK-transformed small airway epithelial cell line correlates with dramatically faster tumor formation in vivo [34], this suggests that the effects of CypA deficiency may have important implications for the treatment of aggressive tumor growth. Our data shows strong evidence that CypA KD leads to decreased cell proliferation in vitro and slower tumor growth in vivo (Fig. 2). Similarly, CypA KD increases apoptosis under the condition of serum withdrawal (Fig. 4), this is a reverse effect for over-expression of CypA in cancer cells renders resistance to anticancer drugs and hypoxia-induced cell death [22, 23]. These results not only establish the relevance of CypA to tumorigenesis, but also substantiate the logic of down-regulating CypA as an approach for treatment of cancer patients.

Over-expression of CypA in rheumatoid arthritis stimulates production of inflammatory cytokines such as TNF- α , IL-1 β , IL-8, MCP-1, and MMP-9 [25]. These CypA-stimulated products could occur through a pathway that is dependent on NF- κ B activation. However, the mechanism by which CypA mediates signaling to activate NF- κ B remains to be investigated. Similarly, stably expressed CypA in the SK-Hep1 cell line revealed that CypA

Fig. 7 Analysis of *IL-8* promoter and regulation of *IL-8* expression by $TNF\alpha$ and CsA. **a** The schematic *IL-8* luciferase reporter constructs used in this study. The luciferase reporter plasmids driven by wild-type *IL-8* promoter and the mutated promoters containing mutated $NF-\kappa B$, C/EBP, or AP1 site are designated as wt_IL-8, mt_NF- κB , mt_C/EBP, and mt_AP1, respectively. **b** U87vIII cells were transfected with indicated reporter plasmids together with *Renilla* luciferase plasmid. Luciferase activity was measured at 24 h post-transfection (columns, mean of results from three independent experiments; bars, SE). **c** U87vIII cells were transfected with the reporter plasmid driven by the wild type *IL-8* promoter together with *Renilla* luciferase plasmid. At 24 h post-transfection, cells were incubated with 1 $\mu g/ml$ CsA or rapamycin for 5 h, then stimulated by 10 ng/ml $TNF-\alpha$ for another 4 h. Cells were then harvested and promoter activity was determined. Results are presented as relative reporter activity after normalization to *Renilla* luciferase activity. Bars indicate the standard error of three replicates



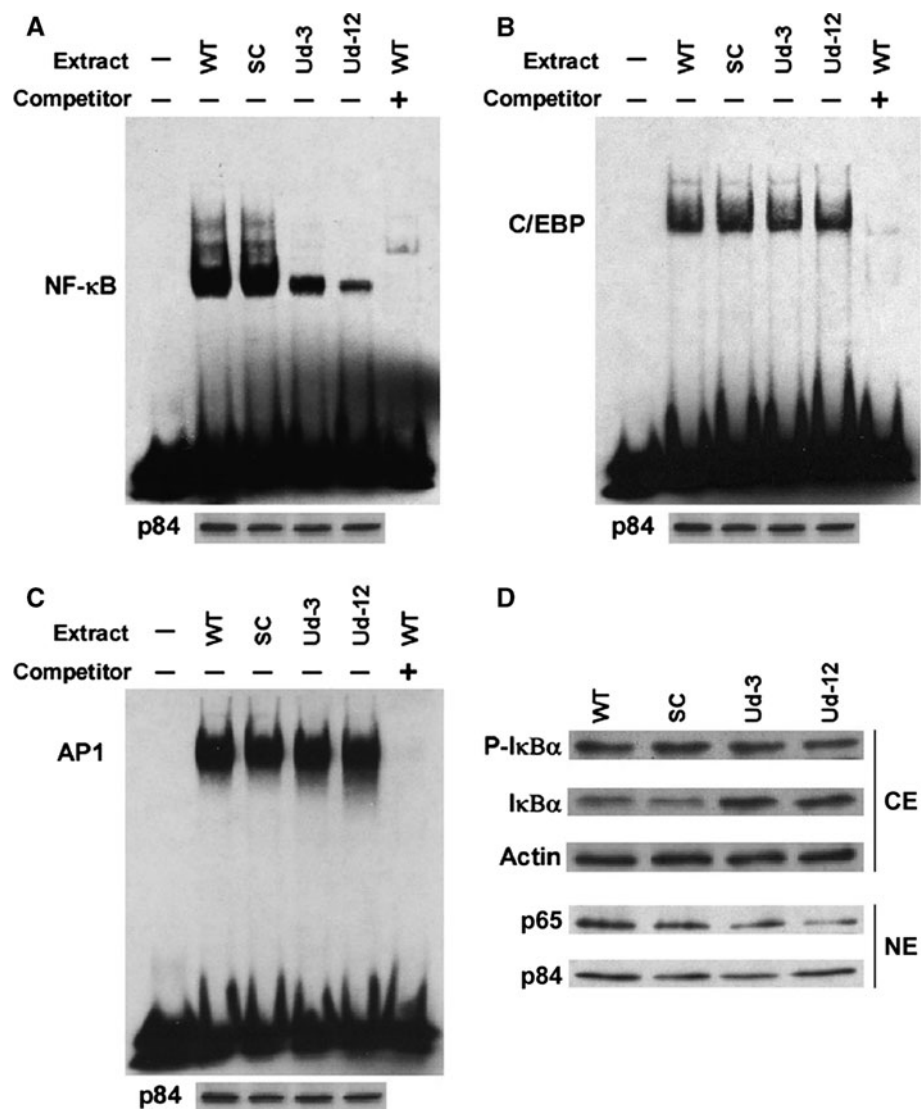
up-regulates the expression of many cytokine-related genes such as *IL-8*, *IL-1 β* , and *IL-6* [22]. These up-regulated cytokines and chemokines may confer tumor cell growth advantage in the neoplastic microenvironment. With these observations in mind, it is conceivable that over-expression of CypA in glioblastomas would lead to highly expressed *IL-8*. Indeed, in human glioblastoma, *IL-8* is expressed and secreted at high levels, and has been suggested to be critical to glial tumor neovascularity and progression [26]. Our data demonstrate that CypA KD in the U87vIII glioblastoma cell line results in profoundly down-regulated *IL-8* expression (Fig. 5), suggesting that a strategy to knock down endogenous over-expressed CypA could prevent tumor progression.

IL-8 is known to possess tumorigenic and proangiogenic properties. Down-regulation of *IL-8* or its regulatory signaling within the tumor microenvironment would have significant therapeutic potential in modulating tumor progression. A recent report indicated that exposure of U251 glioblastoma cells to calcium ionophore and phorbol-

myristate-acetate (PMA) led to dramatic increases in *IL-8* expression, which was suppressed by the calcineurin inhibitor, CsA and FK506 [35]. CsA is a CypA inhibitor; therefore, CsA suppression of *IL-8* expression may contribute to our observation that CypA KD results in down-regulation of *IL-8* expression.

The study conducted in human monocytic leukemia cell line THP-1 demonstrated that CypA induces degradation of $I\kappa B$ and nuclear translocation of $NF-\kappa B$ [25]. Our data concur with their report that CypA KD decreased $NF-\kappa B$ binding activity, decreased phosphorylation and degradation of $I\kappa B\alpha$, reduced nuclear translocation of $NF-\kappa B$ (Fig. 8d), and led to down-regulation of *IL-8* expression. Correspondingly, human recombinant CypA increased phosphorylation of $I\kappa B\alpha$ and stimulated activation of $NF-\kappa B$ in endothelial cells [36]. However, the means by which CypA mediates signaling to activate $NF-\kappa B$ remains to be investigated. A physical and functional map of the human $TNF-\alpha/NF-\kappa B$ signal transduction pathway identified FK506-binding protein 51 (FKBP51), an immunophilin

Fig. 8 Reduced NF- κ B binding activity and decreased p65 associated with CypA KD cells. **a, b,** and **c** EMSA assays were performed using nuclear extracts from U87vIII cells (WT), scrambled control cells (SC), and the CypA KD clones, Ud-3 and Ud-12, with NF- κ B, C/EBP, or AP1 oligos as probes. For competition experiments, protein was pre-incubated with 100-fold molar excess of unlabeled oligos at 25°C for 10 min before addition of labeled probes. p84 expression on Western blots was used as loading control for EMSA. **d** Cytoplasmic extracts (CE) prepared from U87vIII cells (WT), scrambled control cells (SC), and the CypA KD clones, Ud-3 and Ud-12, were immunoblotted for phosphorylated I κ B α (p-I κ B α) and I κ B α , and actin was used as the internal control (*upper panel*). Nuclear extracts (NE) were subjected to Western blot analysis probing with anti-p65 antibody. p84 was used as an internal control (*lower panel*)



with peptidylprolyl isomerase activity, co-purified with IKK α . This interaction was confirmed by co-immunoprecipitation [37]. The NCBI protein data bank shows that IKK α and IKK β contain a potential CypA-binding consensus sequence, FRPFLP/FGPXLPL. Binding of CypA to IKK α and IKK β may affect the phosphorylation at Ser 177 and Ser 181 residues that is necessary to activate kinase activity. The possible interaction between CypA and IKK α or IKK β is currently under investigation.

The present data obtained with both knock down and an ectopically over-expressed rescue plasmid demonstrated that down-regulation and restored *IL-8* expression are CypA-dependent (Fig. 6). Although there is no CypA-binding motif in the *IL-8* promoter to regulate *IL-8* expression, CypA-associated *IL-8* production may be regulated by other transcription factors in an indirect manner. Transcriptional regulation of *IL-8* has been studied extensively at the level of its promoter region, which contains

cis-acting elements for transcription factors C/EBP, AP-1 and NF- κ B. Using a wild-type and a series of mutants of *IL-8* promoter luciferase constructs, we have found that all three binding sites are important positive regulatory elements for regulation of *IL-8*. These data suggest that multiple factors are required to work in concert for optimal *IL-8* expression in glioblastoma cells. Using EMSA, we found decreased binding of NF- κ B to the *IL-8* promoter *cis*-acting element when using nuclear extracts isolated from CypA KD clones (Fig. 8a). These data suggest that CypA is associated with NF- κ B binding activity. Taken together, it has been shown that CypA KD leads to decreased NF- κ B binding, followed by suppression of a synergistically functional interaction between all three transcription factors resulting in reduced *IL-8* expression. Our data demonstrate that knockdown of CypA KD-mediated decreases of cell proliferation, infiltration, and tumor formation may occur through down-regulating *IL-8*. Thus,

targeted knockdown of CypA could be an effective therapeutic approach against devastating glioblastomas.

Acknowledgments We thank Dr. James J. Pestka (Michigan State University) for IL-8 luciferase reporters, Dr. Kazunari Yokoyama for critical review of the manuscript and discussion, and Rebecca Yang for assistance with experiments.

Open Access This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

References

- Gurney JG, Kadan-Lottick N (2001) Brain and other central nervous system tumors: rates, trends, and epidemiology. *Curr Opin Oncol* 13:160–166
- Ross HJ, Cho J, Osann K et al (1997) Phase I/II trial of low dose cyclosporine A with EP for advanced non-small cell lung cancer. *Lung Cancer* 18:189–198
- Sliwa M, Markovic D, Gabrusiewicz K et al (2007) The invasion promoting effect of microglia on glioblastoma cells is inhibited by cyclosporine A. *Brain* 130:476–489
- Zupanska A, Dziembowska M, Ellert-Miklaszewska A et al (2005) Cyclosporin A induces growth arrest or programmed cell death of human glioma cells. *Neurochem Int* 47:430–441
- Ryffel B, Woerly G, Greiner B, Haendler B, Mihatsch MJ, Foxwell BM (1991) Distribution of the cyclosporine binding protein cyclophilin in human tissues. *Immunology* 72:399–404
- Fischer G, Aumuller T (2003) Regulation of peptide bond cis/trans isomerization by enzyme catalysis and its implication in physiological processes. *Rev Physiol Biochem Pharmacol* 148:105–150
- Kruse M, Brunke M, Escher A, Szalay AA, Tropschug M, Zimmermann R (1995) Enzyme assembly after de novo synthesis in rabbit reticulocyte lysate involves molecular chaperones and immunophilins. *J Biol Chem* 270:2588–2594
- Handschumacher RE, Harding MW, Rice J, Drugge RJ, Speicher DW (1984) Cyclophilin: a specific cytosolic binding protein for cyclosporin A. *Science* 226:544–547
- Bram RJ, Hung DT, Martin PK, Schreiber SL, Crabtree GR (1993) Identification of the immunophilins capable of mediating inhibition of signal transduction by cyclosporine A and FK506: roles of calcineurin binding and cellular location. *Mol Cell Biol* 13:4760–4769
- Colgan J, Asmal M, Yu B, Luban J (2005) Cyclophilin A-deficient mice are resistant to immunosuppression by cyclosporine. *J Immunol* 174:6030–6038
- Lu KP, Finn G, Lee TH, Nicholson LK (2007) Prolyl cis-trans isomerization as a molecular timer. *Nat Chem Biol* 3:619–629
- Colgan J, Asmal M, Neagu M, Yu B, Schneidkraut J, Lee Y, Sokolskaja E, Andreotti A, Luban J (2004) Cyclophilin A regulates TCR signal strength in CD4+ T cells via a praline-directed conformational switch in Itk. *Immunity* 21:189–201
- Song J, Lu Y-C, Yokoyama K, Rossi J, Chiu R (2004) Cyclophilin A is required for retinoic acid-induced neuronal differentiation in p19 cells. *J Biol Chem* 279:24414–24419
- Zhu C, Wang X, Deinum J et al (2007) Cyclophilin A participates in the nuclear translocation of apoptosis-inducing factor in neurons after cerebral hypoxia-ischemia. *J Exp Med* 204:1741–1748
- Pan H, Luo C, Li R et al (2008) Cyclophilin A is required for CXCR4-mediated nuclear export of heterogenous nuclear ribonucleoprotein A2 activation and nuclear translocation of ERK1/2, and chemotactic cell migration. *J Biol Chem* 283:623–637
- Bauer K, Kretzschmar AK, Cvijic H et al (2009) Cyclophilins contribute to Stat3 signaling and survival of multiple myeloma cells. *Oncogene* 28:2784–2795
- Shen J, Person MD, Zhu J, Abbruzzese JL, Li D (2004) Protein expression profiles in pancreatic adenocarcinoma compared with normal pancreatic tissue and tissue affected by pancreatitis as detected by two-dimensional gel electrophoresis and mass spectrometry. *Cancer Res* 64:9018–9026
- Campa MJ, Wang MZ, Howard BA, Fitzgerald MC, Patz EF Jr (2003) Protein expression profiling identifies macrophage migration inhibitory factor and cyclophilin A as potential molecular targets in non-small cell lung cancer. *Cancer Res* 63:1652–1656
- Li Z, Zhao X, Bai S, Wang Z, Chen L, Wei Y, Huang C (2008) Proteomics identification of cyclophilin A as a potential prognostic factor and therapeutic target in endometrial carcinoma. *Mol Cell Proteomics* 7:1810–1823
- Qi YJ, He QY, Ma YF et al (2008) Proteomic identification of malignant transformation-related proteins in esophageal squamous cell carcinoma. *J Cell Biochem* 104:1625–1635
- Howard BA, Furumai R, Campa MJ, Rabbani ZN, Vujaskovic Z, Wang XF, Patz EF Jr (2005) Stable RNA interference-mediated suppression of cyclophilin A diminishes non-small-cell lung tumor growth in vivo. *Cancer Res* 65:8853–8860
- Chen S, Zhang M, Ma H, Saiyin H, Shen S, Xi J, Wan B, Yu L (2008) Oligo-microarray analysis reveals the role of cyclophilin A in drug resistance. *Cancer Chemother Pharmacol* 61:459–469
- Choi KJ, Piao YJ, Lim MJ, Kim JH, Ha J, Choe W, Kim SS (2007) Overexpressed cyclophilin A in cancer cells renders resistance to hypoxia- and cisplatin-induced cell death. *Cancer Res* 67:3654–3662
- Arora K, Gwinn WM, Bower MA et al (2005) Extracellular cyclophilins contribute to the regulation of inflammatory responses. *J Immunol* 175:517–522
- Kim H, Kim WJ, Jeon ST, Koh EM, Cha HS, Ahn KS, Lee WH (2005) Cyclophilin A may contribute to the inflammatory processes in rheumatoid arthritis through induction of matrix degrading enzymes and inflammatory cytokines from macrophages. *Clin Immunol* 116:217–224
- Brat DJ, Bellail AC, Van Meir EG (2005) The role of interleukin-8 and its receptors in gliomagenesis and tumoral angiogenesis. *Neuro Oncol* 7:122–133
- Morita M, Kasahara T, Mukaida N et al (1993) Induction and regulation of IL-8 and MCAF production in human tumor cell lines and brain tumor tissues. *Eur Cytokine Netw* 4:351–358
- Tada M, Suzuki K, Yamakawa Y et al (1993) Human glioblastoma cells produce 77 amino acid interleukin-8. *J Neurooncol* 16:25–34
- Desbaillets I, Diserens AC, de Tribolet N, Hamou MF, Van Meir EG (1999) Regulation of interleukin-8 expression by reduced oxygen pressure in human glioblastoma. *Oncogene* 18:1447–1456
- Hoffmann E, Dittrich-Breiholz O, Holtmann H, Kracht M (2002) Multiple control of interleukin-8 gene expression. *J Leukoc Biol* 72:847–855
- Matsusaki T, Fujikawa K, Nishio Y et al (1993) Transcription factors NF-IL6 and NF-kappa B synergistically activate transcription of the inflammatory cytokines, interleukin 6 and interleukin 8. *Proc Natl Acad Sci USA* 90:10193–10197
- Livak KJ, Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods* 25:402–408

33. Lim SO, Park SJ, Kim W, Park SG et al (2002) Proteome analysis of hepatocellular carcinoma. *Biochem Biophys Res Commun* 291:1031–1037
34. Howard BA, Zheng Z, Campa MC et al (2004) Translating biomarkers into clinical practice: prognostic implications of cyclophilin A and macrophage migratory inhibitory factor identified from protein expression profiles in non-small cell lung cancer. *Lung Cancer* 46:313–323
35. Wakabayashi K, Kambe F, Cao X et al (2004) Inhibitory effects of cyclosporine A on calcium mobilization-dependent interleukin-8 expression and invasion potential of human glioblastoma U251MG cells. *Oncogene* 23:6924–6932
36. Jin ZG, Lungu AO, Xie L, Wang M, Wong C, Berk BC (2004) Cyclophilin A is a proinflammatory cytokine that activates endothelial cells. *Arterioscler Thromb Vasc Biol* 24:1186–1191
37. Bouwmeester T, Bauch A, Ruffner H et al (2004) A physical and functional map of the human TNF- α /NF- κ B signal transduction pathway. *Nat Cell Biol* 6:97–105