### **RESEARCH ARTICLE**

Revised: 6 June 2021



### Genetic analysis for sow stayability at different parities in purebred Landrace and Large White pigs

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Funding information Ministry of Agriculture, Forestry, and Fisheries of Japan

### Abstract

Genetic parameters for sow stayability were estimated from farrowing records of 10,295 Landrace sows and 8192 Large White sows. The record for sow stayability from parity k to parity k + 1 (k = 1, ..., 6) was 0 when a sow had a farrowing record at parity k but not at parity k + 1, and 1 when a sow had both records. Heritability was estimated by using single-trait linear and threshold animal models. Genetic correlations among parities were estimated by using two-trait linear-linear and single-trait random regression linear animal models. Genetic correlations with litter traits at birth were estimated by using a two-trait linear-linear animal model. Heritability estimates by linear model analysis were low (0.065–0.119 in Landrace & 0.061-0.157 in Large White); those by threshold model analysis were higher (0.136-0.200 & 0.110-0.283). Genetic correlations among parities differed between breeds and models. Genetic correlation between sow stayability and number born alive was positive in many cases, implying that selection for number born alive does not reduce sow stayability. The results seem to be affected by decisions on culling made by farmers.

#### **KEYWORDS**

binary trait, genetic parameter estimation, Landrace, Large White, sow stayability

#### INTRODUCTION 1

Sow lifetime productivity is an economically important complex trait for efficient pork production (e.g., Rohrer et al., 2017; Serenius & Stalder, 2006; Stalder et al., 2003). As it is determined by the number of piglets weaned, or an indicator trait such as number born alive (NBA), and the overall parity number (e.g., Kang et al., 2018; Sasaki & Koketsu, 2008; Serenius & Stalder, 2004), it is crucial to elucidate

[Correction added on 23 August 2021, after first online publication: The copyright line was changed.]

their associations. The number of piglets weaned and NBA have low estimated heritabilities (e.g., Ogawa et al., 2019a, 2019c; Putz et al., 2015). Overall parity can be measured first after culling. The distribution of the measurements is often obscure: factors influencing the reason for culling can change with time, and it is difficult to distinguish between voluntary and involuntary culling (e.g., Serenius & Stalder, 2006; Sölkner & Ducrocq, 1999; Visscher et al., 1999). Furthermore, heritability has been estimated to be low in many cases (e.g., Le et al., 2016; Paixão et al., 2019; Sobczyńska et al., 2013), and therefore, it is thought to be difficult to efficiently improve sow lifetime productivity by direct selection for overall parity number

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(Le et al., 2016; Serenius & Stalder, 2006). These facts are the same for total lifetime litter size and the length of sow productive life (e.g., Le et al., 2016; Paixão et al., 2019; Sobczyńska et al., 2013).

Genetic parameter estimation for stayability, which is defined as a binary trait relating to survivability in a given environment and is treated as an indicator trait for longevity, has been widely conducted in pigs (Aasmundstad et al., 2014; Abell et al., 2016; Hong et al., 2019; Le et al., 2016; López-Serrano et al., 2000; Poulsen et al., 2020; Serenius et al., 2006; Tholen et al., 1996), dairy cattle (e.g., Abe et al., 2020; Bijma & Jensen, 1996; VanRaden et al., 2016), beef cattle (e.g., Fennewald et al., 2018; Martinez et al., 2005; Silva et al., 2003), horses (e.g., Albertsdóttir et al., 2012; Bugislaus et al., 2005; Langlois & Vrijenhoek, 2004), sheep (Lee et al., 2015; McIntyre et al., 2012), and goats (Scholtens et al., 2018). In pigs, genetic parameters for sow stavability from first parity to a particular time, especially second parity, have been estimated, and threshold model analyses have been performed (Abell et al., 2016; Hong et al., 2019; Le et al., 2016). Genetic correlations of sow stayability with production traits, exterior and leg conformations, lameness, farrowing interval, days open, and NBA at first parity have been also studied (Hong et al., 2019: Le et al., 2016; López-Serrano et al., 2000; Tholen et al., 1996). On the other hand, no study has estimated genetic correlations with number stillborn (NSB), survival rate of piglets at birth (SVB), total litter weight at birth (LWB), and mean litter weight at birth (MWB). Several studies have performed random regression analysis of stayabilities in cattle (e.g., Jamrozik et al., 2008; Jamrozik et al., 2013; Veerkamp et al., 2001) and in Thai Landrace and Large White commercial populations (Plaengkaeo et al., 2020). Factors affecting sow stayability and their effect size could change over time, but there appears to be little published information on the genetic associations of sow stayabilities among different parities.

Here, to obtain information necessary to efficiently improve sow lifetime productivity, we estimated the heritabilities of sow stayabilities at different parities, defined as binary traits, and the genetic correlations of sow stayabilities among parities and with NBA, NSB, SVB, LWB, and MWB in purebred Landrace and Large White pigs using large-scale datasets obtained from a single Japanese pig breeding company.

### 2 | MATERIALS AND METHODS

### 2.1 | Ethics statement

Approval of the Animal Care and Use Committee was not required for this study because the data were acquired from an existing database.

### 2.2 | Phenotypic information and pedigree data

CIMCO Corporation (Tokyo, Japan) provided 72,606 farrowing records for 13,747 Landrace sows and 59,116 records for 11,345 Large White sows, obtained during 2000–2018, and pedigree data

covering 83,333 Landrace pigs and 72,168 Large White pigs. CIMCO operates two great-grandparent and several grandparent farms on a Specific-Pathogen-Free system across Japan (Ogawa et al., 2019c).

We excluded farrowing records with a negative gestation length, NBA = 0, or MWB outside the mean  $\pm$  3 standard deviation (SD); we assumed a mean MWB of 1.48 and SD = 0.23 in both breeds (Damgaard et al., 2003), because the values calculated from our data were greatly affected by obvious outliers (Konta et al., 2019, 2020; Ogawa et al., 2019a, 2019b, 2019c). Next, we extracted sows having all farrowing records from first through final or seventh parities. This gave 43,215 records for 10,295 Landrace sows and 33,513 records for 8192 Large White sows.

Six binary traits for sow stayability—from first to second (STAY12), second to third (STAY23), third to fourth (STAY34), fourth to fifth (STAY45), fifth to sixth (STAY56), and sixth to seventh parities (STAY67)—were defined. The sow stayability from parity k to parity k + 1 (k = 1, ..., 6) was recorded as 0 when a sow had a calving record at parity k but not at parity k + 1, 1 when a sow had both records, and missing when a sow did not have a record at parity k (Figure 1). Table 1 summarizes phenotypic measurements of the traits studied.

### 2.3 | Numerical analyses

Two kinds of single-trait animal models—linear and threshold—were used to estimate the heritability of sow stayability. The following single-trait linear animal model (denoted as Model 1) was used:

$$y = Xb + Za + e$$
,

where **y** is the vector of phenotypic records; **b** is the vector of macroenvironment effects (farrowing year, 2000–2018; farrowing season, spring [March to May], summer [June to August], autumn [September to November], winter [December to February]; farm, seven levels for Landrace, eight levels for Large White; mating sire breed, Landrace, Large White, Duroc); **a** is the vector of breeding values; **e** is the vector

Sow	Has a farrowing record at each parity?											
	1st	2nd	3rd 4	th 5th	6 <b>th</b>	7th						
1	Yes	Yes	Yes Y	Yes Yes	Yes	Yes						
2	Yes	Yes	Yes Y	Yes No	No	No						
3	Yes	No	No 1	No No	No	No						
Sow STAY12 STAY23 STAY34 STAY45 STAY56 STAY67												
	SIAII2	51A125	51A134	51A145	51A150	51A107						
1	1	1	1	1	1	1						
2	1	1	1	0	Missing	Missing						
3	0	Missing	Missing	Missing	Missing	Missing						



TABLE 1 Descriptive statistics of phenotypic measurements of the traits studied

		Landrace				Large White					
Trait	Abbreviation	N	Mean	SD	Min	Max	N	Mean	SD	Min	Max
Number born alive	NBA	43,215	10.23	2.72	1	22	33,513	10.21	2.57	1	22
Number stillborn	NSB	43,215	0.66	1.02	0	10	33,513	0.74	1.08	0	10
Survival rate of piglets at birth	SVB	43,215	0.94	0.08	0.33	1.00	33,513	0.94	0.09	0.36	1.00
Total litter weight at birth (kg)	LWB	43,215	15.34	3.77	1.0	31.0	33,513	15.65	3.61	1.1	28.0
Mean litter weight at birth (kg)	MWB	43,215	1.53	0.24	0.79	2.17	33,513	1.56	0.24	0.80	2.17
Stayability from first to second parity	STAY12	10,295	0.80	0.40	0	1	8192	0.80	0.40	0	1
Stayability from second to third parity	STAY23	8276	0.85	0.36	0	1	6563	0.84	0.37	0	1
Stayability from third to fourth parity	STAY34	7008	0.85	0.36	0	1	5507	0.84	0.37	0	1
Stayability from fourth to fifth parity	STAY45	5956	0.83	0.37	0	1	4606	0.81	0.39	0	1
Stayability from fifth to sixth parity	STAY56	4956	0.79	0.41	0	1	3716	0.77	0.42	0	1
Stayability from sixth to seventh parity	STAY67	3903	0.72	0.45	0	1	2860	0.72	0.45	0	1

Abbreviations: Max, maximum value; Min, minimum value; N, number of phenotypic records; SD, standard deviation.

of random errors; and X and Z are the design matrices relating y to b and a, respectively. The mean and variance-covariance of the vectors a and e were as follows:

$$E\begin{bmatrix}\mathbf{a}\\\mathbf{e}\end{bmatrix} = \begin{bmatrix}\mathbf{0}\\\mathbf{0}\end{bmatrix} \text{ and } \operatorname{var}\begin{bmatrix}\mathbf{a}\\\mathbf{e}\end{bmatrix} = \begin{bmatrix}\mathbf{A}\sigma_a^2 & \mathbf{0}\\\mathbf{0} & \mathbf{I}\sigma_e^2\end{bmatrix},$$

where  $\sigma_a^2$  is the additive genetic variance,  $\sigma_e^2$  is the error variance, **A** is the additive relationship matrix, and **I** is the identity matrix. Variance components were estimated in GIBBS3F90 software (Misztal et al., 2002). A total chain length of 110,000 rounds was run in a single long chain. After the first 10,000 samples were discarded as the burn-in, 1 in every 10 samples was stored. Parameter estimates and their standard errors (SEs) were obtained by calculating the averages and SDs of the 10,000 samples stored, and the 95% confidence intervals (CIs) of were also obtained by using the 10,000 samples.

Estimated heritability,  $\hat{h}^2$ , was converted to a liability scale (e.g., Dempster & Lerner, 1950; Gianola, 1982; Robertson & Lerner, 1949), to compare with the results from threshold model analysis, as follows:

$$\frac{\widehat{h}^2 p(1-p)}{z^2},$$

where *p* is the overall incidence of a score of 1 in a population; *z* is an ordinate of a standard normal density function corresponding to a threshold that divides the probability mass into proportions *p* and 1 - p; and  $\frac{p(1-p)}{r^2}$  takes a minimum value of 1.57 when p = 0.5.

The following single-trait threshold animal model (Model 2) was used:

$$\eta = Xb + Za + e,$$

where  $\eta$  is the vector of liabilities. The mean and variance-covariance of the vectors **a** and **e** were as follows:

$$E\begin{bmatrix}\mathbf{a}\\\mathbf{e}\end{bmatrix} = \begin{bmatrix}\mathbf{0}\\\mathbf{0}\end{bmatrix} \text{ and } \operatorname{var}\begin{bmatrix}\mathbf{a}\\\mathbf{e}\end{bmatrix} = \begin{bmatrix}\mathbf{A}\sigma_a^2 & \mathbf{0}\\\mathbf{0} & \mathbf{I}\sigma_e^2\end{bmatrix}$$

Variance components were estimated in THRGIBBS1F90 software (Tsuruta & Misztal, 2006).

Two kinds of animal models were used to estimate the genetic correlation of sow stayability between parities. First, the following two-trait linear-linear animal model (Model 3) was used:

$$\begin{bmatrix} \mathbf{y}_1 \\ \mathbf{y}_2 \end{bmatrix} = \begin{bmatrix} \mathbf{X}_1 & \mathbf{0} \\ \mathbf{0} & \mathbf{X}_2 \end{bmatrix} \begin{bmatrix} \mathbf{b}_1 \\ \mathbf{b}_2 \end{bmatrix} + \begin{bmatrix} \mathbf{Z}_1 & \mathbf{0} \\ \mathbf{0} & \mathbf{Z}_2 \end{bmatrix} \begin{bmatrix} \mathbf{a}_1 \\ \mathbf{a}_2 \end{bmatrix} + \begin{bmatrix} \mathbf{e}_1 \\ \mathbf{e}_2 \end{bmatrix}$$

The mean and variance-covariance of the vectors  $\mathbf{a}_1$ ,  $\mathbf{a}_2$ ,  $\mathbf{e}_1$ , and  $\mathbf{e}_2$  were as follows:

$$E\begin{bmatrix}a_{1}\\a_{2}\\e_{1}\\e_{2}\end{bmatrix} = \begin{bmatrix}0\\0\\0\\0\end{bmatrix} \text{ and var}\begin{bmatrix}a_{1}\\a_{2}\\e_{1}\\e_{2}\end{bmatrix} = \begin{bmatrix}A\sigma_{a1}^{2} & A\sigma_{a12} & 0 & 0\\A\sigma_{a12} & A\sigma_{a2}^{2} & 0 & 0\\0 & 0 & I\sigma_{e1}^{2} & 0\\0 & 0 & 0 & I\sigma_{e2}^{2}\end{bmatrix},$$

where  $\sigma_a 12$  is the additive genetic covariance; and the error covariance was fixed at 0 (e.g., Árnason, 1999; Árnason et al., 2012; Meyer & Thompson, 1984). Variance components were estimated in GIBBS3F90 software.

Second, the following single-trait linear random regression animal model (Model 4) was used (Plaengkaeo et al., 2020):

$$y_{ik} = F_{ik} + \sum_{j=0}^{2} \varphi_j(k) b_j + \sum_{j=0}^{2} \varphi_j(k) a_{ij} + \sum_{j=0}^{2} \varphi_j(k) p e_{ij} + e_{ik},$$

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where  $y_{ik}$  is the record of sow *i*;  $F_{ik}$  is the sum of the macroenvironment effects (farrowing year by parity, 103 levels for Landrace, 102 levels for Large White; farrowing season by parity, 24 levels for both breeds; mating sire breed by parity, 18 levels for both breeds; farm by parity, 42 levels for Landrace, 48 levels for Large White);  $b_j$  is the regression coefficient of the *j*th-order Legendre polynomial at parity *k*,  $\varphi_j(k)$  (*j* = 0, 1, or 2);  $a_{ij}$  is the regression coefficient of  $\varphi_j(k)$  for breeding value of sow *i*;  $pe_{ij}$  is the regression coefficient of  $\varphi_j(k)$  for permanent environmental effect of sow *i*; and  $e_{ik}$  is the error. This model can be described in matrix notation as

$$\mathbf{y} = \mathbf{X}\mathbf{b} + \sum_{j=0}^{2} \mathbf{Z}_{j}\mathbf{a}_{j} + \sum_{j=0}^{2} \mathbf{W}_{j}\mathbf{p}\mathbf{e}_{j} + \mathbf{e}_{j}$$

The mean and variance–covariance of the vectors  $\mathbf{a}_{j}$ ,  $\mathbf{p}\mathbf{e}_{j}$ , and  $\mathbf{e}$  were as follows:

$$E\begin{bmatrix} a_{0} \\ a_{1} \\ a_{2} \\ pe_{0} \\ pe_{1} \\ pe_{2} \\ e \end{bmatrix} = \begin{bmatrix} 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \end{bmatrix} \text{ and }$$

	<b>a</b> 0		$\mathbf{A}\sigma_{a0}^2$	$\mathbf{A}\sigma_{a01}$	$\mathbf{A}\sigma_{a02}$	0	0	0	0
	$a_1$		$A\sigma_{a01}$	$\mathbf{A}\sigma_{a1}^2$	$\mathbf{A}\sigma_{a12}$	0	0	0	0
	<b>a</b> 2		$\mathbf{A}\sigma_{a02}$	$\mathbf{A}\sigma_{a12}$	$A\sigma_{a2}^2$	0	0	0	0
var	pe <sub>0</sub>	=	$ \begin{array}{c} \mathbf{A}\sigma_{a0}^2 \\ \mathbf{A}\sigma_{a01} \\ \mathbf{A}\sigma_{a02} \\ 0 \end{array} $	0	0	$\sigma_{pe0}^2$	$\sigma_{pe01}$	$\sigma_{pe02}$	0
	$\mathbf{pe}_1$		0	0	0	$\sigma_{pe01}$	$\sigma_{pe1}^2$	$\sigma_{pe12}$	0
	$\mathbf{pe}_2$		0	0	0	$\sigma_{pe02}$	$\sigma_{pe12}$	$\sigma_{pe2}^2$	0
	e		0	0	0	0	0	0	$\left \sigma_{e}^{2}\right $

Variance components were estimated in GIBBS3F90 software.

The following two-trait linear-linear animal model (Model 5) was used to estimate the genetic correlation of sow stayability at each parity with NBA, NSB, SVB, LWB, and MWB:

$$\begin{bmatrix} y_1 \\ y_2 \end{bmatrix} = \begin{bmatrix} X_1 & 0 \\ 0 & X_2 \end{bmatrix} \begin{bmatrix} b_1 \\ b_2 \end{bmatrix} + \begin{bmatrix} Z_1 & 0 \\ 0 & Z_2 \end{bmatrix} \begin{bmatrix} a_1 \\ a_2 \end{bmatrix} + \begin{bmatrix} 0 \\ W_2 \end{bmatrix} pe_2 + \begin{bmatrix} e_1 \\ e_2 \end{bmatrix},$$

where **pe** is the vector of permanent environmental effect; **W** is the design matrix relating **y** to **pe**; subscripts 1 corresponds to any sow stayability trait, and subscript 2 corresponds to any of NBA, NSB, SVB, LWB, and MWB. Macro-environmental effects for NBA, NSB, SVB, LWB, and MWB were parity, farrowing year, farrowing season, farm, and mating sire breed (Konta et al., 2019, 2020; Ogawa et al., 2019a, 2019b, 2019c). The mean and variance-covariance of vector  $\mathbf{a_1}$ ,  $\mathbf{a_2}$ ,  $\mathbf{pe_2}$ ,  $\mathbf{e_1}$ , and  $\mathbf{e_2}$  were as follows:

	<b>a</b> 1		[0]		<b>a</b> 1 <sup>-</sup>		$\mathbf{A}\sigma_{a1}^2$	$\mathbf{A}\sigma_{a12}$	0	0	0	
	<b>a</b> 2		0		<b>a</b> 2		$\mathbf{A}\sigma_{a12}$	$\mathbf{A}\sigma_{a2}^2$	0	0	0	
Ε	$\mathbf{pe}_2$	=	0	and var	pe <sub>2</sub>	=	0	0	$\sigma_{pe2}^2$	0	0	,
	<b>e</b> <sub>1</sub>		0	and var	<b>e</b> <sub>1</sub>		0	0	0	$\sigma_{e1}^2$	0	
	<b>e</b> <sub>2</sub>		0		<b>e</b> <sub>2</sub>		0	0	0	0	$I\sigma_{e2}^2$	

where the error covariance was fixed at 0 (e.g., Konta et al., 2019; Ogawa et al., 2020; Wolf et al., 2005). Variance components were estimated in GIBBS3F90 software.

### 3 | RESULTS AND DISCUSSION

### 3.1 | Descriptive statistics of sow stayability

Mean phenotypic records for sow stayability were similar between breeds (Table 1). STAY12 averaged 0.80, lower than STAY23, STAY34, and STAY45 and higher than STAY56 and STAY67 in both breeds. Means differed among other studies; for example, STAY12 was 0.82 in Landrace and 0.79 in Large White (López-Serrano et al., 2000), 0.85 in Yorkshire (Hong et al., 2019), 0.41 in Landrace (Aasmundstad et al., 2014), 0.63 in Landrace (Abell et al., 2016), and 0.68 in Yorkshire (Le et al., 2016). These differences might be caused in part by the timing of data collection and population management, which both depend on decision making by farmers for culling. In this regard, Tholen et al. (1996) noted different average values of STAY12 among herds.

### 3.2 | Heritability of sow stayability

Heritabilities of sow stayability estimated using Model 1 were low regardless of breed and parity, ranging from 0.065 to 0.119 in Landrace and from 0.061 to 0.157 in Large White, and additive genetic and error variances varied among parity within breed (Table 2). Heritabilities estimated using Model 2 were also low but were higher than those estimated using Model 1, ranging from 0.136 to 0.200 in Landrace and from 0.110 to 0.283 in Large White. SEs of heritability estimates using Model 2 were larger than those obtained using Model 1, and their 95% Cls largely overlapped with each other. Heritabilities estimated using Model 1 converted to a liability scale ranged from 0.118 to 0.275 in Landrace and from 0.122 to 0.355 in Large White, within the range of 95% Cls of the estimates obtained using Model 2 in most cases.

Previous studies have consistently estimated the heritability of purebred sow stayability to be low by using linear model (Aasmundstad et al., 2014; Le et al., 2016; López-Serrano et al., 2000; Poulsen et al., 2020; Serenius et al., 2006; Tholen et al., 1996) and threshold model (Hong et al., 2019; Le et al., 2016). Aasmundstad et al. (2014), López-Serrano et al. (2000), Poulsen et al. (2020), and Tholen et al. (1996) considered information on reproductive performance of sows, such as age at first farrowing and litter size, into model as macro-environment effects. Some studies considered the common litter environmental effects (Aasmundstad et al., 2014; Hong et al., 2019; López-Serrano et al., 2000; Serenius et al., 2006), and Serenius et al. (2006) reported that the heritability was estimated to be slightly lower when using model considering the common litter environmental effects than when using model ignoring it, which were also observed for several body measurement and meat production traits (Ogawa et al., 2021; Ohnishi & Satoh, 2018). Le et al. (2016) estimated the heritability of STAY12 to be higher when using

TABLE 2 Genetic parameters for sow stayability traits estimated by using single-trait linear and threshold animal models (Models 1 and 2)

	Linear model								Threshold model			
	Phenotypi	c variance	:	Heritabilit	Heritability					Heritability		
Trait	Estimate	SE	95%CI	Estimate	SE	95%CI	Converted <sup>a</sup>	Estimate	SE	95%CI		
Landrace												
STAY12	0.139	0.002	0.135 to 0.143	0.105	0.024	0.066 to 0.168	0.217	0.200	0.039	0.117 to 0.274		
STAY23	0.107	0.002	0.103 to 0.110	0.119	0.025	0.077 to 0.170	0.275	0.189	0.034	0.121 to 0.264		
STAY34	0.107	0.002	0.104 to 0.111	0.087	0.021	0.049 to 0.126	0.204	0.141	0.042	0.067 to 0.220		
STAY45	0.124	0.002	0.120 to 0.129	0.107	0.023	0.060 to 0.151	0.237	0.174	0.042	0.105 to 0.272		
STAY56	0.142	0.003	0.136 to 0.148	0.065	0.016	0.041 to 0.104	0.130	0.136	0.040	0.056 to 0.210		
STAY67	0.173	0.004	0.165 to 0.181	0.066	0.025	0.017 to 0.118	0.118	0.154	0.041	0.096 to 0.252		
Large White												
STAY12	0.138	0.002	0.134 to 0.142	0.061	0.015	0.034 to 0.090	0.125	0.200	0.039	0.117 to 0.274		
STAY23	0.112	0.002	0.107 to 0.116	0.157	0.027	0.107 to 0.210	0.355	0.189	0.034	0.121 to 0.264		
STAY34	0.116	0.002	0.112 to 0.121	0.084	0.029	0.043 to 0.145	0.189	0.141	0.042	0.067 to 0.220		
STAY45	0.134	0.003	0.129 to 0.140	0.116	0.028	0.058 to 0.174	0.241	0.174	0.042	0.105 to 0.272		
STAY56	0.144	0.003	0.137 to 0.151	0.092	0.023	0.059 to 0.139	0.176	0.136	0.040	0.056 to 0.210		
STAY67	0.173	0.005	0.164 to 0.182	0.068	0.028	0.027 to 0.128	0.122	0.154	0.041	0.096 to 0.252		

Note: See Table 1 for abbreviations of trait names.

Abbreviations: 95% CI, 95% confidence interval; SE, standard error.

<sup>a</sup>Heritability estimates by linear model analyses were converted to a liability scale according to Robertson and Lerner (1949).

threshold model (0.18) than when using linear model (0.08), which was also observed in this study (Table 2). On the other hand, only Abell et al. (2016) estimated the heritability of STAY12 in purebred Landrace sows to be 0.81 using threshold model and denoted that this was not unexpected because selection decisions are made based on the genetic potential of a sow; however they did not estimate the heritability by using linear model. Poulsen et al. (2020) proposed that the difference in estimated heritability among studies could be due to the difference not only in statistical modeling but also in the type and number of farms analyzed (e.g., nucleus vs. commercial). None of the previous studies compared the value of heritability estimated using linear model and converted to a liability scale with that estimated using threshold model.

# 3.3 | Genetic correlation of sow stayability among parities

By Model 3, estimated genetic correlations of sow stayability among parities ranged from -0.138 to 0.685 in Landrace and from -0.490 to 0.779 in Large White (Table 3), with similar additive genetic and error variances to those estimated by single-trait linear animal model analysis. Patterns of estimated genetic correlations seem to differ between breeds and to be less associated with parity than those in the previous study of NBA (Ogawa et al., 2019b). Here, the error covariance was fixed at 0 (e.g., Árnason, 1999; Árnason et al., 2012; Meyer & Thompson, 1984), which might bias estimates of genetic parameters (Árnason et al., 2012). Gates et al. (1999) assessed the performance of multiple- (including binary) trait analysis, estimated the error covariances by computer simulation and "quasi-REML" approach, and discussed the effect of the overall incidence, selection, confounding, and sign of the value of the true error correlation on estimating genetic and error correlations. However, in our study, the record at a lower parity of a sow, which also has a record at a higher parity, is always 1 (e.g., Árnason, 1999; Árnason et al., 2012; Meyer & Thompson, 1984), unlike in Gates et al. (1999). Here, we attempted to estimate the error covariance, but the results were hard to interpret because the estimated additive genetic and error variances were different from those estimated by single-trait analysis, and genetic and error correlations were sometimes both negative.

By Model 4, the estimated genetic correlations ranged from -0.022 to 0.952 in Landrace and tended to decrease as the parities were more distance (Table S1). Those between adjacent parities ranged from 0.733 to 0.952, sometimes lower than the threshold of 0.8 suggested by Robertson (1959). However, the values were inconsistent with those estimated by two-trait model analysis. In Large White, on the other hand, the estimated error variance was very small, and the phenotypic variance was overestimated (Table S1), causing the "blowing up" phenomenon (Hoeschele & Tier, 1995). The possibility of the "extreme category problem" increases in categorical trait analysis when the modeling becomes more complicated (e.g., Hoeschele & Tier, 1995; Luo et al., 2001; Misztal et al., 1989). In this study, the number of records was lower for Large White (Table 1), which might also be a reason why the blowing up phenomenon was observed only for Large White. Macro-environment effects included in the random regression model were set to be similar to those in **TABLE 3** Genetic correlations  $\pm$  standard errors (SEs) and 95% confidence intervals in parentheses of sow stayability traits among parities estimated by using a two-trait linear-linear animal model (Model 3)

Trait	STAY12	STAY23	STAY34	STAY45	STAY56	STAY67
STAY12		$0.756^{**}\pm 0.117$ (0.489 to 0.923)	0.779** ± 0.144 (0.470 to 0.943)	$0.353^*\pm 0.209$ (-0.009 to 0.707)	$0.494^{**}\pm 0.171$ (0.088 to 0.736)	$-0.357^{*}\pm0.232$ (-0.745 to 0.054)
STAY23	$0.225^* \pm 0.162$ (-0.084 to 0.533)		$-0.139 \pm 0.225$ (-0.492 to 0.353)	$0.320^*\pm 0.183$ (-0.070 to 0.649)	$-0.073 \pm 0.170$ (-0.359 to 0.252)	$0.201 \pm 0.238$ (-0.340 to 0.539)
STAY34	$0.105 \pm 0.218$ (-0.343 to 0.473)	$0.625^{**}\pm 0.114$ (0.391 to 0.796)		$0.258^* \pm 0.169$ (-0.124 to 0.561)	$0.336^* \pm 0.171$ (-0.057 to 0.615)	$0.129 \pm 0.328$ (-0.454 to 0.657)
STAY45	$0.278^{**}\pm 0.128$ (0.033 to 0.528)	$0.615^{**}\pm 0.169$ (0.286 to 0.926)	$0.107 \pm 0.175$ (-0.288 to 0.473)		$-0.063 \pm 0.225$ (-0.474 to -0.364)	$-0.490^{**}\pm 0.190$ (-0.746 to 0.017)
STAY56	$-0.138 \pm 0.254$ ( $-0.566$ to 0.454)	$0.394^* \pm 0.198$ (-0.061 to 0.699)	$0.112 \pm 0.310$ (-0.408 to -0.673)	0.654** ± 0.191 (0.287 to 0.905)		$0.021 \pm 0.238$ (-0.509 to 0.415)
STAY67	0.447* ± 0.279 (-0.026 to 0.875)	$0.211 \pm 0.332$ (-0.551 to 0.648)	0.685** ± 0.126 (0.444 to 0.888)	$0.414^{**}\pm 0.126$ (0.171 to 0.642)	$0.351^*\pm 0.249$ (–0.074 to 0.830)	

Note: See Table 1 for abbreviations of trait names. Bottom left, Landrace; top right Large White.

\*The absolute value of the estimate was greater than SE.

\*\*The absolute value of the estimate was greater than 2 SE.

the two-trait animal model, which might make the modeling too complicated. Plaengkaeo et al. (2020) used a simpler model including only the discrete effect of herd-year-season and the linear covariate of age at first farrowing as macro-environment effects. Here, the macro-environment effects in the single-trait model were determined according to previous studies of this population (Konta et al., 2019, 2020; Ogawa et al., 2019a, 2019b, 2019c), which could complicate the analytical model. Using a simpler model might reduce the possibility of blowing up (Hoeschele & Tier, 1995), but the estimated value could be biased owing to confounding (Gates et al., 1999).

# 3.4 | Genetic correlations of sow stayability with litter traits at farrowing

Estimated genetic correlations with NBA using Model 5 ranged from 0.159 to 0.595 in Landrace and from -0.014 to 0.556 in Large White, being positive except for the almost negligible correlation of -0.014 with STAY12 in Large White (Table 4). The results imply that selection for NBA would not immediately harm sow stayability. Estimated values tended to be higher when the parity was higher, which seems to be dependent on the sow culling scheme. On the GGP farms, male and female animals with estimated breeding values for NBA and total litter weight at weaning greater than their sires and dams, respectively, are selected as candidates at 20 weeks of age based on their phenotypic performance for 18 body and leg conformation traits only from litters without piglet having hernia or prolapse (Ogawa et al., 2019c). Estimated breeding values of replacement gilts with higher reliabilities can be obtained after recording one or more farrowing performances of their own, and sows in production herd with worse estimated breeding values have more risk of being culled. These facts might affect our results. Hong et al. (2019) estimated the genetic correlation between STAY12 and NBA at first parity to be 0.31 in Yorkshire pigs of a Korean company.

The estimated genetic correlation of sow stavability was negative with NSB and positive with SVB in both breeds, but the absolute values were larger for SVB than for NSB. A possible reason is that SVB includes information not only on NSB but also on NBA. The estimated genetic correlation with MWB was negative. MWB might be higher when NBA is lower or oversized piglets are born. Some of the sows with a higher MWB might be culled because such a farrowing could be related to reproductive disorders including dystocia especially at first parity, although one of the major culling reasons could be lower NBA. The estimated genetic correlation with LWB was negative for STAY12 in Landrace and for STAY12 and STAY23 in Large White but was positive at later parities in both breeds. This difference might be associated with the change in the genetic correlations with NBA and MWB over parity and the fact that total litter weight at weaning, which has a positive genetic correlation with LWB, was considered at selection for sows in this population (Ogawa et al., 2019c).

### 3.5 | General discussion

We estimated genetic parameters for binary traits relating to sow stayability at different parities in purebred Landrace and Large White pigs of a single Japanese pig breeding company, using several different models. We also estimated the variance components by using Models 1, 3, and 5 in AIREMLF90 software (Misztal et al., 2002), and the results were similar (Tables S2, S3, and S4), whereas the REML estimates were diverged when using Model 4 in Large White. Heritability estimates obtained by using Model 1 were low in both breeds and at all parities (Table 2), and the random regression model analysis with Model 4 could not give reliable results (Table S1). These facts indicate the difficulty in binary-trait analysis and the fact that efficiently improving sow stayability by direct selection would be difficult (Aasmundstad et al., 2014; Hong et al., 2019; Le et al., 2016;

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**TABLE 4** Estimated genetic correlations ± standard errors (SEs) and 95% confidence intervals in parentheses of sow stayability traits with litter traits at farrowing by using a two-trait linear animal model (Model 5)

Trait	STAY12	STAY23	STAY34	STAY45	STAY56	STAY67
Landrace						
NBA	$0.159^*\pm 0.082$ (-0.013 to 0.309)	$0.321^{**}\pm 0.067$ (0.188 to 0.458)	0.584** ± 0.057 (0.475 to 0.697)	0.548 <sup>**</sup> ± 0.064 (0.415 to 0.670)	$0.595^{**}\pm 0.086$ (0.420 to 0.741)	$0.422^{**}\pm 0.089$ (0.247 to 0.585)
NSB	$-0.234^{**}\pm 0.083$ (-0.388 to -0.070)	$-0.201^{**}\pm 0.084$ (-0.377 to -0.053)	$-0.060 \pm 0.088$ (-0.218 to 0.124)	$-0.070 \pm 0.080$ (-0.225 to 0.096)	$-0.232^{**}\pm 0.099$ (-0.422 to -0.043)	$-0.484^{**}\pm 0.104$ (-0.659 to -0.263)
SVB	$0.334^{**}\pm 0.081$ (0.158 to 0.475)	$0.246^{**}\pm 0.076$ (0.095 to 0.391)	$0.152^*\pm 0.094$ (-0.040 to 0.335)	0.197** ± 0.088 (0.017 to 0.375)	0.328 <sup>**</sup> ± 0.086 (0.168 to 0.493)	$0.524^{**}\pm 0.105$ (0.330 to 0.708)
LWB	$-0.214^{**}\pm 0.085$ (-0.407 to -0.060)	$0.160^{**}\pm 0.059$ (0.039 to 0.275)	$0.442^{**}\pm 0.065$ (0.325 to 0.574)	$0.475^{**}\pm 0.062$ (0.343 to 0.598)	$0.467^{**}\pm 0.083$ (0.314 to 0.622)	$0.168^*\pm 0.100$ (-0.036 to 0.355)
MWB	$-0.514^{**}\pm 0.079$ (-0.681 to -0.369)	$-0.277^{**}\pm 0.062$ (-0.404 to -0.157)	$-0.236^{**}\pm 0.068$ (-0.357 to -0.100)	$-0.082^{*}\pm0.068$ (-0.214 to 0.056)	$-0.165^*\pm 0.093$ (-0.364 to 0.003)	$-0.395^{**}\pm 0.094$ (-0.563 to -0.232)
Large Whi	te					
NBA	$-0.014 \pm 0.107$ (-0.212 to 0.194)	$0.144^*\pm 0.081$ (-0.012 to 0.288)	$0.479^{**}\pm 0.085$ (-0.302 to 0.656)	$0.363^{**}\pm 0.071$ (0.229 to 0.513)	$0.431^{**}\pm 0.104$ (0.249 to 0.677)	$0.556^{**}\pm 0.098$ (0.381 to 0.754)
NSB	$-0.430^{**}\pm 0.085$ (-0.573 to -0.256)	$-0.212^{**}\pm 0.083$ (-0.360 to -0.039)	$-0.138^*\pm 0.092$ (-0.309 to 0.033)	$-0.270^{**}\pm 0.085$ (-0.443 to -0.104)	$-0.293^{**}\pm 0.092$ (-0.464 to -0.102)	$-0.352^{**}\pm 0.124$ (-0.597 to -0.119)
SVB	$0.474^{**}\pm 0.115$ (0.248 to 0.661)	$0.248^{**}\pm 0.085$ (0.085 to 0.410)	$0.197^{**}\pm 0.096$ (0.005 to 0.376)	0.346** ± 0.088 (0.177 to 0.527)	$0.355^{**}\pm 0.134$ (0.135 to 0.701)	$0.506^{**}\pm 0.126$ (0.260 to 0.743)
LWB	$-0.360^{**}\pm 0.082$ (-0.506 to -0.187)	$-0.052 \pm 0.072$ (-0.186 to 0.088)	$0.229^{**}\pm 0.079$ (0.076 to 0.391)	$0.182^{**}\pm 0.070$ (0.043 to 0.332)	$0.216^{**}\pm 0.105$ (0.030 to 0.450)	$0.235^{**}\pm 0.113$ (0.025 to 0.463)
MWB	$-0.555^{**}\pm 0.078$ (-0.693 to -0.394)	$-0.347^{**}\pm 0.066$ (-0.469 to -0.209)	$-0.453^{**}\pm 0.080$ (-0.602 to -0.298)	$-0.313^{**}\pm 0.076$ (-0.469 to -0.168)	$-0.361^{**}\pm 0.091$ (-0.530 to -0.173)	$-0.446^{**}\pm 0.108$ (-0.655 to -0.235)

Note: See Table 1 for abbreviations of trait names.

\*The absolute value of the estimate was greater than SE.

\*\*The absolute value of the estimate was greater than 2 SE.

López-Serrano et al., 2000; Poulsen et al., 2020; Serenius et al., 2006; Tholen et al., 1996).

As far as we know, this is the first study to estimating genetic correlations of sow stayability with NSB, SVB, LWB, and MWB. Genetic correlations with other traits have been estimated (Hong et al., 2019; Le et al., 2016; López-Serrano et al., 2000; Tholen et al., 1996). Associations with maternal performance and mothering ability, within-litter variation of piglet birth weight and pattern of litter size variation over parity, and resilience indicators should also be investigated (Damgaard et al., 2003; Dobrzański et al., 2020; Gäde et al., 2008; Poulsen et al., 2020).

Factors affecting stayability can be divided into voluntary and involuntary. The latter affects "functional" stayability (e.g., Abe et al., 2020; Oliveira et al., 2020; Valencia-Posadas et al., 2017), which might be a trait to be truly improved. However, it is often difficult to use detailed information about reasons for culling. As a compromise, an indirect evaluation of sow functional stayability could be performed, for example by including the effects of NBA and other indicators possibly relating to voluntary culling in analytical model (Poulsen et al., 2020). In many cases, the final goal of improving sow lifetime productivity is a crossbred population. Previous studies have analyzed the stayability of crossbred sows (Abell et al., 2016; Engblom et al., 2009; Engblom et al., 2016; Poulsen et al., 2020; Serenius et al., 2006). From this perspective, further study will be needed to improve sow lifetime productivity (e.g., Bijma & van Arendonk, 1998; Steyn et al., 2021; Wientjes & Calus, 2017).

### 4 | CONCLUSION

We estimated low heritability of sow stayability in Landrace and Large White populations of a single Japanese pig breeding company, irrespective of model (linear or threshold), breed (Landrace or Large White), and parity (first to seventh). Estimated genetic correlations of sow stayability among parities differed between breeds and models (two-trait linear-linear model and single-trait random regression linear model). Estimated genetic correlations of sow stayability with traits relating to farrowing performance (NBA, NSB, SVB, LWB, & MWB) were considered to reflect of the content of decision making by farmers. In particular, the genetic correlation with NBA was promising in many cases, indicating that improving NBA does not readily inhibit sow stayability.

### ACKNOWLEDGMENTS

This work was supported by a grant from the Ministry of Agriculture, Forestry, and Fisheries of Japan (Development of Breeding Technology for Animal Life Production).

### CONFLICT OF INTEREST

The authors declare that they have no competing interests.

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### REFERENCES

- Aasmundstad, T., Olsen, D., Sehested, E., & Vangen, O. (2014). The genetic relationships between conformation assessment of gilts and sow production and longevity. *Livestock Science*, 167, 33–40. https://doi. org/10.1016/j.livsci.2014.05.004
- Abe, H., Hagiya, K., Yamaguchi, S., Nakagawa, S., Gotoh, Y., Baba, T., & Kawahara, T. (2020). Genetic parameters and trends of cow livability in Holsteins in Hokkaido, Japan. *Nihon Chikusan Gakkaiho*, *91*, 93–102. https://doi.org/10.2508/chikusan.91.93 (in Japanese)
- Abell, C. E., Fernando, R. L., Serenius, T. V., Rothchild, M. F., Gray, K. A., & Stalder, K. J. (2016). Genetic relationship between purebred and crossbred sow longevity. *Journal of Animal Science and Biotechnology*, 7, 51. https://doi.org/10.1186/s40104-016-0112-x
- Albertsdóttir, E., Árnason, T., Eriksson, S., Sigurdsson, Á., & Fikse, W. F. (2012). Effects of integrated genetic evaluations for Icelandic horses on predictive ability, accuracy and selection bias. *Journal of Animal Breeding and Genetics*, 129, 41–49. https://doi.org/10.1111/j.1439-0388.2011.00940.x
- Árnason, T. (1999). Genetic evaluation of Swedish standard-bred trotters for racing performance and racing status. *Journal of Animal Breeding and Genetics*, 116, 387–398. https://doi.org/10.1046/j.1439-0388. 1999.00202.x
- Árnason, T., Albertsdóttir, E., Fikse, W. F., Eriksson, S., & Sigurdsson, Á. (2012). Estimation of genetic parameters and response to selection for a continuous trait subject to culling before testing. *Journal of Animal Breeding and Genetics*, 129, 50–59. https://doi.org/10.1111/j. 1439-0388.2011.00941.x
- Bijma, P., & Jensen, J. (1996). Genetic analysis of herd life and stayability in Danish dairy cattle. Proceedings International workshop on genetic improvement of functional traits in cattle, Gembloux, Belgium. Uppsala, Sweden: Interbull.
- Bijma, P., & van Arendonk, J. A. M. (1998). Maximizing genetic gain for the sire line of a crossbreeding scheme utilizing both purebred and crossbred information. *Animal Science*, 66, 529–542. https://doi.org/ 10.1017/S135772980000970X
- Bugislaus, A. -E., Roehe, R., Willms, F., & Kalm, E. (2005). Multivariate genetic analysis to account for preselection and disqualified races in the genetic evaluation of racing performances in German trotters. *Acta Agriculturae Scandinavica, Section a – Animal Science, 55*, 49–56. https://doi.org/10.1080/09064700500239545
- Damgaard, L. H., Rydhmer, L., Løvendahl, P., & Grandinson, K. (2003). Genetic parameters for within-litter variation in piglet birth weight and change in within litter variation during suckling. *Journal of Animal Science*, 81, 604–610. https://doi.org/10.2527/2003.813604x
- Dempster, E. R., & Lerner, I. M. (1950). Heritability of threshold characters. *Genetics*, 35, 212–236.
- Dobrzański, J., Mulder, H. A., Knol, E. F., Szwaczkowski, T., & Sell-Kubiak, E. (2020). Estimation of litter size variability phenotypes in large white sows. *Journal of Animal Breeding and Genetics*, 137, 559–570. https://doi.org/10.1111/jbg.12465
- Engblom, L., Calderón Díaz, J. A., Nikkilä, M., Gray, K., Harms, P., Fix, J., Tsuruta, S., Mabry, J., & Stalder, K. (2016). Genetic analysis of sow longevity and sow lifetime reproductive traits using censored data. *Journal of Animal Breeding and Genetics*, 133, 138–144. https://doi. org/10.1111/jbg.12177

- Engblom, L., Kubdeheim, N., Schneider, M. P., Dalin, A.-M., & Andersson, K. (2009). Genetics of crossbred sow longevity. *Animal*, *3*, 783–790. https://doi.org/10.1017/S175173110900411X
- Fennewald, D. J., Weaber, R. L., & Lamberson, W. R. (2018). Genotype by environment interaction for stayability of Red Angus in the United States. *Journal of Animal Science*, 93, 422–429. https://doi.org/10. 1093/jas/skx080
- Gäde, S., Bennewitz, J., Kirchner, K., Looft, H., Knap, P. W., Thaller, G., & Kalm, E. (2008). Genetic parameters for maternal behaviour traits in sows. *Livestock Science*, 114, 31–41. https://doi.org/10.1016/j.livsci. 2007.04.006
- Gates, P., Johansson, K., & Danell, B. (1999). "Quasi-REML" correlation estimates between production and health traits in the presence of selection and confounding: A simulation study. *Journal of Animal Science*, 77, 558–568. https://doi.org/10.2527/1999.773558x
- Gianola, D. (1982). Theory and analysis of threshold characters. *Journal of Animal Science*, *54*, 1079–1096. https://doi.org/10.2527/jas1982. 5451079x
- Hoeschele, I., & Tier, B. (1995). Estimation of variance components of threshold characters by marginal posterior modes and means via Gibbs sampling. *Genetics Selection Evolution*, 27, 519–540. https://doi.org/10.1186/1297-9686-27-6-519
- Hong, J. K., Kim, Y. M., Cho, K. H., Cho, E. S., Lee, D. H., & Choi, T. J. (2019). Genetic association between sow longevity and social genetic effects on growth in pigs. *Asian-Australasian Journal of Animal Sciences*, 32, 1077–1083. https://doi.org/10.5713/ajas.18.0789
- Jamrozik, J., Fatehi, J., & Schaeffer, L. R. (2008). Comparison of models for genetic evaluation of survival traits in dairy cattle: A simulation study. *Journal of Animal Breeding and Genetics*, 125, 75–83. https://doi.org/10.1111/j.1439-0388.2007.00712.x
- Jamrozik, J., McGrath, S., Kemp, R. A., & Miller, S. P. (2013). Estimates of genetic parameters for stayability to consecutive calvings of Canadian Simmentals by random regression models. *Journal of Animal Science*, 91, 3634–3643. https://doi.org/10.2527/jas.2012-6126
- Kang, J. H., Lee, E. A., Hong, K. C., & Kim, J. M. (2018). Regulatory gene network from a genome-wide association study for sow lifetime productivity traits. *Animal Genetics*, 49, 254–258. https://doi.org/10. 1111/age.12640
- Konta, A., Ogawa, S., Kimata, M., Ishii, K., Uemoto, Y., & Satoh, M. (2019). A study on the potential for improving number born alive using teat number in pig female breeds. *Nihon Chikusan Gakkaiho*, 90, 207–212. https://doi.org/10.2508/chikusan.90.207 (in Japanese)
- Konta, A., Ogawa, S., Kimata, M., Ishii, K., Uemoto, Y., & Satoh, M. (2020). Comparison of two models to estimate genetic parameters for number of born alive in pigs. *Animal Science Journal*, *91*, e13417. https://doi.org/10.1111/asj.13417
- Langlois, B., & Vrijenhoek, T. (2004). Qualification status and estimation of breeding values in French trotters. *Livestock Production Science*, 89, 187–194. https://doi.org/10.1016/j.livprodsci.2004.02.001
- Le, T. H., Madsen, P., Lundeheim, N., Nilsson, K., & Norberg, E. (2016). Genetic association between leg conformation in young pigs and sow longevity. *Journal of Animal Breeding and Genetics*, 133, 283–290. https://doi.org/10.1111/jbg.12193
- Lee, M. A., Cullen, N. G., Newman, S. A. N., Dodds, K. G., McEwan, J. C., & Shackell, G. H. (2015). Genetic analysis and genomic selection of stayability and productive life in New Zealand ewes. *Journal of Ani*mal Science, 93, 3268–3277. https://doi.org/10.2527/jas.2014-8259
- López-Serrano, M., Reinsch, N., Looft, H., & Kalm, E. (2000). Genetic correlations of growth, backfat thickness and exterior with stayability in large white and landrace sows. *Livestock Production Science*, 64, 121–131. https://doi.org/10.1016/S0301-6226(99)00169-4
- Luo, M. F., Boettcher, P. J., Schaeffer, L. R., & Dekkers, J. C. M. (2001). Bayesian inference for categorical traits with an application to variance component estimation. *Journal of Dairy Science*, *84*, 694–704. https://doi.org/10.3168/jds.S0022-0302(01)74524-9

- Martinez, G. E., Koch, R. M., Cundiff, L. V., Gregory, K. E., Kachman, S. D., & Van Vleck, L. D. (2005). Genetic parameters for stayability, stayability at calving, and stayability at weaning to specified ages for Hereford cows. *Journal of Animal Science*, 83, 2033–2042. https://doi.org/10.2527/2005.8392033x
- McIntyre, S. B., Newman, S. A. N., & Young, E. A. (2012). Genetic and phenotypic parameters for stayability in a New Zealand research flock. *Proceedings of the New Zealand Society of Animal Production*, 72, 152–155.
- Meyer, K., & Thompson, R. (1984). Bias in variance and covariance component estimators due to selection on a correlated trait. *Zeitschrift für Tierzüchtung und Züchtungsbiologie*, 101, 33–50. https://doi.org/10.1111/j.1439-0388.1984.tb00020.x
- Misztal, I., Gianola, D., & Foulley, J. L. (1989). Computing aspects of a nonlinear method of sire evaluation for categorical data. *Journal of Dairy Science*, 72, 1557–1568. https://doi.org/10.3168/jds.S0022-0302(89)79267-5
- Misztal, I., Tsuruta, S., Strabel, T., Auvray, B., Druet, T., & Lee, D. H. (2002). BLUPF90 and related programs. Proceedings of the Seventh World Congress of Genetics Applied to Livestock Production. France: Montpellier.
- Ogawa, S., Konta, A., Kimata, M., Ishii, K., Uemoto, Y., & Satoh, M. (2019a). Estimation of genetic parameters for farrowing traits in purebred landrace and large white pigs. *Animal Science Journal*, *90*, 23–28. https://doi.org/10.1111/asj.13120
- Ogawa, S., Konta, A., Kimata, M., Ishii, K., Uemoto, Y., & Satoh, M. (2019b). Genetic parameter estimation for number born alive at different parities in landrace and large white pigs. *Animal Science Journal*, *90*, 1111–1119. https://doi.org/10.1111/asj.13252
- Ogawa, S., Konta, A., Kimata, M., Ishii, K., Uemoto, Y., & Satoh, M. (2019c). Genetic relationship of litter traits between farrowing and weaning in landrace and large white pigs. *Animal Science Journal*, 90, 1510–1516. https://doi.org/10.1111/asj.13304
- Ogawa, S., Ohnishi, C., Ishii, K., Uemoto, Y., & Satoh, M. (2020). Genetic relationship between litter size traits at birth and body measurement and production traits in purebred Duroc pigs. *Animal Science Journal*, 91, e13497. https://doi.org/10.1111/asj.13497
- Ogawa, S., Yazaki, N., Ohnishi, C., Ishii, K., Uemoto, Y., & Satoh, M. (2021). Maternal effect on body measurement and meat production traits in purebred Duroc pigs. *Journal of Animal Breeding and Genetics*, 138, 237–245. https://doi.org/10.1111/jbg.12505
- Ohnishi, C., & Satoh, M. (2018). Estimation of genetic parameters for performance and body measurement traits in Duroc pigs selected for average daily gain, loin muscle area, and backfat thickness. *Livestock Science*, 214, 161–166. https://doi.org/10.1016/j.livsci.2018.05.022
- Oliveira, H. R., Brito, L. F., Miller, S. P., & Schenkel, F. S. (2020). Using random regression models to genetically evaluate functional longevity traits in North American Angus cattle. *Animals*, 10, 2410. https://doi.org/10.3390/ani10122410
- Paixão, G., Martins, Â., Esteves, A., Payan-Carreira, R., & Carolino, N. (2019). Genetic parameters for reproductive, longevity and lifetime production traits in Bísaro pigs. *Livestock Science*, 225, 129–134. https://doi.org/10.1016/j.livsci.2019.05.010
- Plaengkaeo, S., Duangjinda, M., & Stalder, K. J. (2020). Identifying early indicator traits for sow longevity using a linear-threshold model in Thai large white and landrace females. *Animal Bioscience*, 34, 20–25. https://doi.org/10.5713/ajas.19.0855
- Poulsen, B. G., Nielsen, B., Ostersen, T., & Christensen, O. F. (2020). Genetic associations between stayability and longevity in commercial crossbred sows, and stayability in multiplier sows. *Journal of Animal Science*, 98, 1–8. https://doi.org/10.1093/jas/skaa183
- Putz, A. M., Tiezzi, F., Maltecca, C., Gray, K. A., & Knauer, M. T. (2015). Variance component estimates for alternative litter size traits in swine. *Journal of Animal Science*, 93, 5153–5163. https://doi.org/10. 2527/jas.2015-9416

- Robertson, A. (1959). The sampling variance of the genetic correlation coefficient. *Biometrics*, 15, 469-485. https://doi.org/10.2307/ 2527750
- Robertson, A., & Lerner, I. M. (1949). The heritability of all-or-none traits: Viability of poultry. *Genetics*, 34, 395–411. https://doi.org/10.1093/ genetics/34.4.395
- Rohrer, G. A., Cross, A. J., Lents, C. A., Miles, J. R., Nonneman, D. J., & Rempel, L. A. (2017). Genetic improvement of sow lifetime productivity. *Journal of Animal Science*, 95, 11–12. https://doi.org/10.2527/ asasmw.2017.026
- Sasaki, Y., & Koketsu, Y. (2008). Sows having high lifetime efficiency and high longevity associated with herd productivity in commercial herds. *Livestock Science*, 118, 140–146. https://doi.org/10.1016/j.livsci. 2007.12.029
- Scholtens, M. R., Lopez-Villalobos, N., Garrick, D. J., & Blair, H. T. (2018). Heritability of longevity in New Zealand dairy goats. Proceedings of the New Zealand Society of Animal Production, 78, 11–15.
- Serenius, T., & Stalder, K. J. (2004). Genetics of length of productive life and lifetime prolificacy in the Finnish landrace and large white pig populations. *Journal of Animal Science*, 82, 3111–3117. https://doi. org/10.2527/2004.82113111x
- Serenius, T., & Stalder, K. J. (2006). Selection for sow longevity. Journal of Animal Science, 84, E166–E171. https://doi.org/10.2527/2006. 8413\_supplE166x
- Serenius, T., Stalder, K. J., & Puonti, M. (2006). Impact of dominance effects on sow longevity. *Journal of Animal Breeding and Genetics*, 123, 355–361. https://doi.org/10.1111/j.1439-0388.2006. 00614.x
- Silva, J. A. V. II, Eler, J. P., Ferraz, J. B. S., Golden, B. L., & Oliveira, H. N. (2003). Heritability estimate for stayability in nelore cows. *Livestock Production Science*, 70, 97–101. https://doi.org/10.1016/S0301-6226(02)00149-5
- Sobczyńska, M., Blicharski, T., & Tyra, M. (2013). Relationships between longevity, lifetime productivity, carcass traits and conformation in polish maternal pig breeds. *Journal of Animal Breeding and Genetics*, 130, 361–371. https://doi.org/10.1111/jbg.12024
- Sölkner, J., & Ducrocq, V. (1999). The survival kit: A tool for analysis of survival data. Proceedings international workshop on EU concerted action Genetic Improvement of Functional Traits in Cattle. Jouy-en-Josas, France: Interbull.
- Stalder, K. J., Lacy, C., Cross, T. L., & Conatser, G. E. (2003). Financial impact of average parity of culled females in a breed-to-wean swine operation using replacement gilt net present value analysis. *Journal* of Swine Health and Production, 11, 69–74.
- Steyn, Y., Lourenco, D. A., Chen, C. Y., Valente, B. D., Holl, J., Herring, W. O., & Misztal, I. (2021). Optimal definition of contemporary groups for crossbred pigs in a joint purebred and crossbred genetic evaluation. *Journal of Animal Science*, 99, 1–8. https://doi. org/10.1093/jas/skaa396
- Tholen, E., Bunter, K. L., Hermesch, S., & Graser, H. -U. (1996). The genetic foundation of fitness and reproduction traits in Australian pig populations 2. Relationships between weaning to conception interval, farrowing interval, stayability, and other common reproduction and production traits. Australian Journal of Agricultural Research, 47, 1275–1290. https://doi.org/10.1071/AR9961275
- Tsuruta, S., & Misztal, I. (2006). THRGIBBSF90 for estimation of variance components with threshold and linear models. Proc. 8th World Congress Gen. Appl. Livest. Prod., Belo Horizonte, Brazil. CD-ROM communication 27-31.
- Valencia-Posadas, M., Torrero-Garza, Y., José Torres-Vázquez, J. A., Ángel-Sahagún, C. A., Gutiérrez-Chávez, A. J., Shepard, L., & Montaldo, H. H. (2017). Genetic parameters for functional stayability to 24 and 36 months of age and first lactation milk yield in dairy goats. *Small Ruminant Research*, 149, 209–213. https://doi.org/10. 1016/j.smallrumres.2017.02.010

#### 10 of 10 WILEY\_ Animal Science

VanRaden, P. M., Wright, J. R., Tooker, M. E., & Norman, H. D. (2016). Value of selecting for cow and calf livability. *Interbull Bulletin*, 50, 30–33.

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Journal

- Veerkamp, R. F., Brotherstone, S., Engel, B., & Meuwissen, T. H. E. (2001). Analysis of censored survival data using random regression models. *Animal Science*, 72, 1–10. https://doi.org/10.1017/ S1357729800055491
- Visscher, P., Thompson, R., Yazdi, H., Hill, W. G., & Brotherstone, S. (1999). Proceedings of the genetic improvement of functional traits in cattle longevity. Jouy-en-Jossas, France: Interbull.
- Wientjes, Y. C. J., & Calus, M. P. L. (2017). BOARD INVITED REVIEW: The purebred-crossbred correlation in pigs: A review of theory, estimates, and implications. *Journal of Animal Science*, 95, 3467–3478. https://doi.org/10.2527/jas.2017.1669
- Wolf, J., Žáková, E., & Groeneveld, E. (2005). Genetic parameters for a joint genetic evaluation of production and reproduction traits in pigs. *Czech Journal of Animal Science*, 50, 96–103. https://doi.org/10. 17221/4001-CJAS

### SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of this article.

How to cite this article: Ogawa, S., Kimata, M., Ishii, K., Uemoto, Y., & Satoh, M. (2021). Genetic analysis for sow stayability at different parities in purebred Landrace and Large White pigs. *Animal Science Journal*, *92*(1), e13599. <u>https://doi.</u> org/10.1111/asj.13599