



## Fractions of Shen-Sui-Tong-Zhi Formula Enhance Osteogenesis Via Activation of $\beta$ -Catenin Signaling in Growth Plate Chondrocytes

Rui Xu<sup>1,2,3</sup>, Qinghe Zeng<sup>1,2</sup>, Chenjie Xia<sup>4</sup>, Jiali Chen<sup>1</sup>, Pinger Wang<sup>1</sup>, Shan Zhao<sup>5</sup>, Wenhua Yuan<sup>1</sup>, Zhaohuan Lou<sup>5</sup>, Houfu Lin<sup>1,2</sup>, Hanting Xia<sup>1,2</sup>, Shuaijie Lv<sup>6</sup>, Taotao Xu<sup>6</sup>, Peijian Tong<sup>6</sup>\*, Mancang Gu<sup>5</sup>\* and Hongting Jin<sup>1</sup>\*

#### **OPEN ACCESS**

Edited by: Jung Chao, China Medical University, Taiwan

#### Reviewed by:

Xin Feng Zheng, Shanghai Jiaotong University, China Rupesh K. Srivastava, All India Institute of Medical Sciences, India Martha Ramírez-Rosas, Universidad Autónoma de Tamaulipas, Mexico

#### \*Correspondence:

Peijian Tong peijiantongzitcm@163.com Mancang Gu gmancang@zcmu.edu.cn Hongting Jin hongtingjin@163.com

#### Specialty section:

This article was submitted to Ethnopharmacology, a section of the journal Frontiers in Pharmacology

Received: 17 May 2021 Accepted: 06 September 2021 Published: 24 September 2021

#### Citation:

Xu R, Zeng Q, Xia C, Chen J, Wang P, Zhao S, Yuan W, Lou Z, Lin H, Xia H, Lv S, Xu T, Tong P, Gu M and Jin H (2021) Fractions of Shen-Sui-Tong-Zhi Formula Enhance Osteogenesis Via Activation of β-Catenin Signaling in Growth Plate Chondrocytes. Front. Pharmacol. 12:711004. doi: 10.3389/fphar.2021.711004 <sup>1</sup>Institute of Orthopedics and Traumatology, The First Affiliated Hospital of Zhejiang Chinese Medical University, Hangzhou, China, <sup>2</sup>The First College of Clinical Medicine, Zhejiang Chinese Medical University, Hangzhou, China, <sup>3</sup>Department of Orthopedic Surgery, Zhejiang Hospital of Integrated Traditional Chinese and Western Medicine, Hangzhou, China, <sup>4</sup>Department of Orthopedic Surgery, Ningbo Medical Center Lihuili Hospital, Ningbo, China, <sup>5</sup>College of Pharmaceutical Science, Zhejiang Chinese Medical University, Hangzhou, China, <sup>6</sup>Department of Orthopedic Surgery, The First Affiliated Hospital of Zhejiang Chinese Medical University, Hangzhou, China

**Background:** Shen-sui-tong-zhi formula (SSTZF) has been used to treat osteoporosis for decades and shows excellent clinical efficacy. This article aims to explore the optimal anti-osteoporotic ingredient and its precise mechanisms in mice models.

**Methods:** In this study, we first screened the optimal anti-osteoporosis fraction of SSTZF extract *in vivo*, and then further explored the mechanism of its effects both *in vivo* and *in vitro*. Ten-week-old female C57BL/6J mice were administrated with each fraction of SSTZF. At 10 weeks after ovariectomy (OVX), femurs were collected for tissue analyses, including histology, micro-CT, biomechanical tests, and immunohistochemistry for ALP, FABP4, and  $\beta$ -catenin. Additionally, we also evaluated the mRNA expression level of ALP and FABP4 and the protein expression level of  $\beta$ -catenin after being treated with SSTZF extract in C<sub>3</sub>H<sub>10</sub>T1/2 cells. Moreover, we investigated the anti-osteoporosis effect of SSTZF extract on mice with  $\beta$ -catenin conditional knockout in growth plate chondrocytes ( $\beta$ -catenin<sup>Gli1ER</sup> mice) through  $\mu$ CT, histology, and immunohistochemistry analyzes.

**Results:** At 10 weeks after treatment, osteoporosis-like phenotype were significantly ameliorated in SSTZF n-butanol extract (SSTZF-NB) group mice, as indicated by increased trabecular bone area and ALP content, and decreased lipid droplet area and FABP4 content. No such improvements were observed after being treated with other extracts, demonstrating that SSTZF-NB is the optimal anti-osteoporosis fraction. Additionally, the elevated  $\beta$ -catenin was revealed in both OVX mice and C<sub>3</sub>H<sub>10</sub>T1/2 cells with SSTZF-NB administered. Furthermore, a significant osteoporosis-like phenotype was observed in  $\beta$ -catenin<sup>Gl/1ER</sup> mice as expected. However, SSTZF-NB failed to rescue the deterioration in  $\beta$ -catenin<sup>Gl/1ER</sup> mice, no significant re-upregulated ALP and downregulated FABP4 were observed after being treated with SSTZF-NB, demonstrating that SSTZF-NB prevents bone loss mainly via  $\beta$ -catenin signaling.

1

**Conclusion:** SSTZF-NB enhances osteogenesis mainly via activation of  $\beta$ -catenin signaling in growth plate chondrocytes. SSTZF-NB is the optimal anti-osteoporosis fraction of SSTZF and it can be considered a salutary alternative therapeutic option for osteoporosis.

Keywords: Shensuitongzhi formula, osteoporosis, osteogenesis, β-catenin signaling, transgenic mice

### **1 INTRODUCTION**

Osteoporosis (OP) is the most common bone disorder around the world. It is characterized by fragile bone fracture, which results from reduced bone mass and deteriorated bone microstructure (Force et al., 2018). Studies have shown that there are 10 million people with osteoporosis in the United States alone, and nearly 34 million people with low bone mass, which means they suffer from an increased risk of osteoporosis (Wu et al., 2019). With the speeding tendency of an aging society, the number of potential OP patients will continuously rise. Currently, the drug for treating OP is still limited due to unexpected side effects, like the impact on the uterus and breast (Cheng et al., 2021). At the moment when Covid-19 is rampant all over the world, it will undoubtedly bring a heavy burden and pressure to the social economy and medical systems (Mattioli et al., 2020). Thus, it is essential to explore and create more alternative therapies for OP treatment (Vandenbroucke et al., 2017). Botanical drugs and their natural extracts attract more and more attention due to the potential anti-osteoporosis effects and the fact that they are relatively safe (Zhang et al., 2016; Zhu et al., 2018).

Traditional Chinese Medicine (TCM) has been widely used for various medical purposes for centuries in East Asia. According to the "kidney dominates bone" theory in TCM, OP is the result of kidney deficiency and decreased marrow. Therefore, the therapeutic strategy should emphasize tonifying kidney and regulating the marrow. Based on the theory mentioned above, the Shen-sui-tong-zhi formula (SSTZF) was devised and has been used for treating bone-related disorders for dozens of years. SSTZF is an experiential effective recipe devised by wellknown doctors of the Zhejiang School and is modified from Yougui Pills, which was first recorded in "jin yue quan shu," a medical classic text written by Jingyue Zhang in the Ming Dynasty. It encompasses Carthamus tinctorius L. (CTL), Rehmannia glutinosa (Gaertn.) DC. (RG), Eucommia ulmoides oliver (EUO), Aconitum carmichaelii debeaux (ACD), Lycium barbarum L. (LBL), Cornus officinalis Siebold & Zucc. (COS), Dioscorea oppositifolia L. (DOL) Glycyrrhiza glabra L. (GGL), and Prunus davidiana (CarriŠre) Franch (PD), Cinnamomum cassia (L.). J.Presl (CCL). In the context of Yougui Pills, the combination of RG, EUO, ACD, LBL, COS, DOL, GGL, and CCL has proven efficacy for bone protection and reveals the potential of activating β-catenin signaling (Yan et al., 2018; Zhang et al., 2019). In addition, several studies have indicated that CTL and its active ingredients could promote BMSCs and differentiate into osteoblasts and show a capacity for bone protection (Kim et al., 2002; Alam et al., 2006; Cui et al., 2019). PD is included to inhibit adipogenesis (Choi et al., 1991a; Choi et al., 1991b; Jung et al., 2017). Combining osteogenic botanical drugs and antiadipogenic botanical drugs with bone-friendly components has offered a promising alternative therapy for bone related disease, especially for OP. Our previous study indicated that SSTZF drug serum promotes osteoblast proliferation and mineralization in the  $\beta$ -catenin related pathway *in vitro*. However, the precise underlying mechanisms and optimal fractions remain unclear, which limits its further exploitation and promotion.

Bone is a living organ in vertebrates. In clinical practice, bone loss and fat accumulation in bone marrow were found in agerelated OP (Pei and Tontonoz, 2004). Even though the mechanism of OP remains unclear, more and more evidence indicates that the unbalance between osteogenesis and adipogenesis plays an important role during the progression (Colaianni et al., 2015; Zou et al., 2020). As is well-known, mesenchymal stem cells (MSCs) are a type of pluripotent stem cell that can differentiate into mesenchymal tissue lineage including osteoblast and adipocyte, the promotion of osteogenesis inhibits adipogenesis and vice versa (Lorthongpanich et al., 2019). During the OP process, the balance between osteogenesis adipogenesis is disturbed and the latter occupies an advantage. The ratio of fat in bone marrow leads to the reduction of osteoblasts, osteoclasts are activated afterward and finally causes the loss of bone mass (Uezumi et al., 2010; Chen et al., 2016; Hu et al., 2018; Liu et al., 2020). There are multitudinous signaling pathways and factors involved in this complex process, there into, the canonical Wnt/β-catenin signaling deserves attention. Research has indicated that β-catenin signaling regulates the differentiation of MSCs as a switch directly (Qiang et al., 2012). To be specific, once the β-catenin was inhibited, the osteogenesis of MSCs was restricted subsequently and the adipogenesis was enhanced on the contrary (de Winter and Nusse, 2021). Therefore, targeting  $\beta$ -catenin could be a potential strategy for OP treatment.

In the current study, we screened the optimal fraction of SSTZF in mice, furthermore, the mechanism of the antiosteoporotic effect of SSTZF extract was elucidated by transgenic mice and cell experiments, *in vivo* and *in vitro* respectively.

#### 2 METHODS AND MATERIALS

#### 2.1 Preparation of SSTZF Extract

All ten botanical drugs in SSTZF (**Table 1**) were provided by the First Affiliated Hospital of Zhejiang Chinese Medical University (Hangzhou China). The process of SSTZF extract preparation includes two parts: a concentrate of SSTZF preparation and secondary extraction with the organic solvent. The specific steps are as follows:

Chinese name	Botanical name	Family	Parts used	Weight (kg)
Hong hua	Carthamus tinctorius L.	Compositae	Flower	1.5
Di huang	Rehmannia glutinosa (Gaertn.) DC.	Plantaginaceae	Root	4.5
Du Zhong	Eucommia ulmoides Oliver.	Eucommiaceae	Bark	3
Fu zi	Aconitum carmichaelii Debeaux	Ranunculaceae	Root	3
Gou qi	Lycium barbarum L.	Solanaceae	Bark	3
Shan zhu yu	Cornus officinalis Siebold & Zucc.	Cornaceae	Fruit	1.5
Shan yao	Dioscorea oppositifolia L.	Dioscoreaceae	Fruit	3
Gan cao	Glycyrrhiza glabra L.	Leguminosae	Root	1.5
Tao ren	Prunus davidiana (CarriŠre) Franch.	Rosaceae	Seed	3
Rou gui	Cinnamomum cassia (L.) J.Presl	Lauraceae	Bark	1.5

#### TABLE 1 | Composition of SSTZ formula.

TABLE 2	Fractions of SSTZE and	<i>i</i> iold
IADLE 2	FIACTIONS OF SSIZE AND	vieia.

Fraction	Isolated solvent	Isolated content (g)	Percentage of total crude extract (%)
SSTZF-PE	Petroleum ether	2.8	2.9
SSTZF-EA	Ethyl acetate	10.3	10.8
SSTZF-NB	Normal butanol	15.4	16.2

After soaking in 6 volumes of distilled water for 1 h, CTL, RG, EUO, ACD, LBL, COS, DOL, GGL were mixed (the total dry weight was 21 kg) and in the ratio of 1:3:2:2:2:1:2:1(w/w) for reflux extraction (three times,1.5 h/time). PD and CCL were soaked in 5 volumes 60% EtOH for 1 h and were mixed in 2: 1(w/w) (the total dry weight was 4.5 kg) for reflux extraction (three times, 1.5 h/time). Then the two portions of extracts were completely mixed into 7.5 L solution and then concentrated into the form of concentrated solution (3.4 g crude drug/mL). The procedures mentioned above are the preparation method of SSTZF concentrates. To extract its optimal anti-osteoporosis fraction, organic solvent including petroleum ether, ethyl acetate, and n-butanol solutions were used for further separation and extraction according to molecular polarity. 95 ml of the extract was diluted in 300 ml of distilled water, then, 300 ml of petroleum ether, 300 ml of ethyl acetate, and 300 ml of n-butanol solution were added and mixed in equal proportions respectively. The mixed solution was poured into the separating funnel and allowed to stand at room temperature for 12 h. The precipitation of the emulsion layer was discarded and the supernatant was collected. Each mixed solution was extracted three times in this mode. A rotary evaporator was used to volatilize the organic solvent completely, then the refined extract powders (2.8 g for petroleum ether extract, 10.3 g for ethyl acetate extract, 15.4 g for normal butanol extract) were obtained (Table 2). The powders were redissolved in 300 ml distilled water respectively and stored at -20°C. Each extract fraction was named SSTZF- petroleum ether extract (SSTZF-PE), SSTZF-ethyl acetate extract (SSTZF-EA), and SSTZF-n-butanol extract (SSTZF-NB).

#### **2.2 Experimental Animals**

To construct the OVX mice model, 10 week-old female C57BL/6 mice were purchased from the Experimental Animal Center of

Zhejiang Chinese Medical University (Hangzhou, China). For the sake of constructing growth plate chondrocytes -specific  $\beta$ -catenin conditional knockout mice,  $\beta$ -catenin<sup>flox/flox</sup> mice were crossed with *Gli1-CreER*<sup>T2</sup> transgenic mice to generate *Gli1-CreER*<sup>T2</sup>;  $\beta$ -catenin<sup>flox/flox</sup> mice hereinafter referred to as  $\beta$ -catenin<sup>Gli1ER</sup> mice (**Table 3**). All original mice were purchased from Jackson Lab (Bar Harbor, ME, United States). To avoid gender-dependent differences, only females were selected for further experiments. To induce conditional gene knockout, tamoxifen was injected for three consecutive days (1 mg/10 g body weight, one time a day, intra-peritonelly) in 1-month-old mice. All studies were approved by the Animal Ethics Committee of Zhejiang Chinese Medical University (LZ12H27001).

# 2.3 Experimental Groups and Drug Administration

All C57BL/6 mice were arranged into six groups (n = 6 in each group) randomly: the sham group, OVX group, SSTZF-PE group, SSTZF-EA group, SSTZF-NB group, and SSTZF group. According to the screening results, the transgenic mice were divided into three groups (n = 6 in each group): Cre-negative group,  $\beta$ -catenin<sup>Gli1ER</sup> group, and  $\beta$ -catenin<sup>Gli1ER</sup> + SSTZF-NB group. Both ovaries were removed in all C56BL/6 mice groups except the sham group. Instead, a sham operation that only excised the surrounding fat tissues equally and preserved bilateral ovaries intact was performed. SSTZF-PE, SSTZF-EA, SSTZF-NB were orally administrated to each corresponding group respectively for 10 consecutive weeks (0.2 ml/10 g body weight, once a day) from the day after OVX surgery. After tamoxifen inducement, SSTZF-NB were orally administrated to mice in the  $\beta$ -catenin<sup>Gli1ER</sup> + SSTZF-NB group. Other groups were treated with an identical dosage of PBS.

#### 2.4 µCT Analyses

Samples of the femur from each group were collected and scanned with micro-CT ( $\mu$ CT). The cross-section of the distal femur metaphysis was reconstructed in three dimensions. Bone mineral density (BMD, g/mm<sup>3</sup>), bone volume fraction (BV/TV, %), average trabecular thickness (Tb.Th, mm), the average number of trabecular (1/mm), and average trabecular separation (Tb.Sp, mm) were collected for morphometry quantitative analysis.

Breeding	Desired progeny
<ul> <li>(a) Gli1-CreER × β-catenin<sup>fx/fx</sup></li> <li>(b) Gli1-CreER; β-cateninfx/wt ×β-catenin<sup>fx/fx</sup></li> <li>(c) Gli1-CreER; β-cateninfx/fx ×β-catenin<sup>fx/fx</sup></li> </ul>	(a) Gli1-CreER; β-catenin <sup>fx/wt</sup> (b) Gli1-CreER; β-catenin <sup>fx/fx</sup> (c) Gli1-CreER; β-cateninfx/fx andβ-catenin <sup>fx/fx</sup>

#### 2.5 Biomechanical Testing

To test the modulus of elasticity (MOE) and maximum loading of femurs in six groups of wild type mice, three-point bending test was performed with the use of the Axial-Torsion Fatigue Testing System (Instron, 5569R1412, United States).

# 2.6 Histology, Histomorphometry, and Immunohistochemistry

The preparation for the production of sections was processed as described previously (Xu et al., 2020). Then  $3 \mu m$  sections at the

site of femur metaphysis were cut coronally for Alcian Blue Hematoxylin/Orange G (ABH) staining. The indexes of histomorphometry including the area of lipid droplets and area of trabecular bone were detected with the use of OsteoMetrics software (Decatur, GA). IHC was performed using anti-alkaline phosphatase (ALP, ARIGO, ARG57422 1: 300), anti-fatty acid-binding protein (FABP4, Abcam, ab92501, 1:200), anti- $\beta$ -catenin (HuaBio, ER0805, 1:200). The quantitative analyses for positive staining area were performed by the software of image-pro plus6.0 (Media Cybernetics, Silver Spring, United States).



with Duncan's multiple-range test.



Hematoxylin/Orange G staining of the distal femur. (B) The area of lipid droplets. (C) The area of trabecular bone. SSTZF-NB enhances bone strength. (D) The modulus of elasticity. (E) The max-loading. Scale bars: 1000  $\mu$ m A–D: The means not sharing a common letter are significantly different among the groups at p < 0.05 by one-way ANOVA with Duncan's multiple-range test.

## 2.7 Cell Culture

Mesenchymal stem cells line C3H10T1/2 cells (ATCC, Manassas, VA, United States) were cultured in Alpha modified Eagle's medium (Gibco, MD, United States) containing 10% (v/v) fetal bovine serum (FBS) (Sigma, MO, United States) and 1% penicillin and streptomycin (Gibco) at 37°C in 5% CO<sub>2</sub> atmosphere.10%FBS, FBS containing 10 µg/ml SSTZF-NB and FBS containing 50 µg/ml SSTZF-NB were added to corresponding wells separately. With treatment for 72 h, cells were measured for qRT-PCR and western blot analyzes.

## 2.8 Real-Time Quantitative PCR

Total RNA was extracted from  $C_3H_{10}T1/2$  cells with the use of TRIzol reagent (Invitrogen, CA, United States). Subsequently, 2 µg of total RNA was sucked out and used for cDNA synthesis by RevertAid First Strand cDNA Synthesis Kit (Invitrogen, CA, United States) following the manufacturer's instructions. Then, the expression level of *ALP* and *FABP4* was determined by real-

time quantitative PCR [relative to  $\beta$ -actin control with a QuantStudio<sup>TM</sup> 7 Flex Real-Time PCR System (Thermo Scientific, MA, United States)]. Forward and reverse sequence of the target gene are as follows: primer sequences of ALP, forward: 5'-TCCTGACCAAAAACCTCAAAGG-3', reverse: 5'-TCGTTCATGCAGAGCCTGC-3'; sequences of FABP4, forward: 5'-AAATCACCGCAGACGACAGG-3', reverse: 5'-GGCTCATGCCCTTTCATAAAC-3'; sequence of  $\beta$ -actin, forward: 5'-GGAGATTACTGCCCTGGCTCCTA-3', reverse: 5'-GACTCATCGTACTGCTGCTGC-3'.

## 2.9 Western Blot

Total proteins were extracted respectively from  $C_3H_{10}T1/2$  cells, which were cultured in control serum and serum containing SSTZF-NB, by using lysis buffer containing protease and phosphatase inhibitors, then incubated on ice for 30 min and isolated on a 12% SDS-PAGE gel. Samples were transferred on polyvinylidene fluoride membranes and blocked in 5% milk for



1.5 h. The membranes were incubated overnight at 4°C with anti- $\beta$ -actin (1:5,000, Abcam, United Kingdom) and anti- $\beta$ -catenin (1: 1,000 HuaBio, CN). TBS-T was used to wash the membranes, and then membranes were incubated for 1 h with goat anti-rabbit horseradish peroxidase-conjugated secondary antibody (1:5,000, Abcam, MA, United States). The protein bands were visualized with Image Quant LAS 4000 (EG, United States). Finally, ImageJ was used to calculate the densitometry of each band for quantification.

#### 2.10 UPLC/MS Analysis of SSTZF

Sample preparation for the Ultra Performance Liquid Chromatography (UPLC) analysis was as follows: 1 mg of SSTZF-NB was diluted in methanol/water (50/50) solution. Then, 1 ml of the previous liquid was diluted with methanol to 1/10 concentration. The test sample was harvested after being filtered with 0.22 µm membrane. The conditions of UPLC were as follows: the type of ACQUITY UPLC<sup>TM</sup> HSS T3 (100 mm × 2.1 mm, 1.8 µm) chromatographic column was selected. The sample size for injection, rate of flow, and column temperature were 1 µl, 0.3 ml/min, and 30°C respectively. The mobile phase was acetonitrile (A)—0.1% formic acid (B) and the

elution gradients were as below: 00 min ~ 7 min, 15%A-85%B; 7 ~ 15 min, 35%A-65%B; 15-18 min, 60%A-40%B; 18 ~ 23 min, 90%A-10%B; 23 ~ 26 min, 90%A-10%B; 27-29 min, 5% A-95%B.

For Mass Spectrometry (MS),  $MS^E$  continuum mode was selected and the following parameter settings were used: ESI + mode: capillary voltage 3.0 kV; sample cone 40 V; source offset 80 V; source temperature 120°C; desolvation temperature 400°C; cone gas 50 L/h; desolvation gas 800 L/h; nebulizer 6.0Bar. TOF MS and TOF MS/MS were scanned with the mass range of m/z 50–1,500 and 50–1,500, respectively. 100 ng/ml leucine enkephalin solution was used as the calibration standard solution for quality control, and sodium-formate was used to calibrate the instrument. Accurate mass and composition for the precursor and fragment ions were analyzed by using UNIFI software integrated with the instrument. Base peak ion chromatogram of SSTZF-NB is shown in **Supplementary File S1**. The detected compounds are listed in **Supplementary File S2**.

#### 2.11 Statistical Analysis

All data were presented as mean  $\pm$  standard deviation. Student's t-test and one-way ANOVA test followed by the Tukey-Kramer



test were performed using SPSS 24.0 software. \*p < 0.05 was considered statistically significant.

#### **3 RESULTS**

#### 3.1 Screening the Optimal Anti-osteoporotic Fraction of SSTZF Extract

To determine the optimal anti-osteoporotic fraction of SSTZF extract, three kinds of extracts and original SSTZF were orally administrated to the OVX mice respectively. No adverse events happened during the experiment. The 3D reconstruction images revealed that OVX could induce obvious bone loss of the distal femur metaphysis area when compared to sham group mice. SSTZF-PE and the original SSTZF could partially restrain deterioration, however, SSTZF-EA hardly works. Interestingly, SSTZF-NB showed a remarkable inhibitory effect on bone loss when compared to each intervention group (**Figure 1A**). The  $\mu$ CT analysis showed a similar trend. The indexes of BMD, BV/TV, Tb.Th, Tb.N, and Tb.Sp were significantly deteriorated after OVX inducement and significantly ameliorated by SSTZF-NB treatment (**Figures 1B–F**).

Results of ABH staining and histomorphometry analyses showed that OVX surgery could cause serious trabeculae



deterioration and fat droplet accumulation in the area of the femoral metaphysis, compared to the sham group. However, the lesion could be alleviated with the treatment of SSTZF-NB, but not with other extract fractions. SSTZF could partially improve the deterioration (**Figures 2A–C**).

Outcomes of biomechanical experiments indicated that OVX could lead to a decreased MOE and reduced max-loading when compared with the sham group, and these indexes were improved only in the SSTZF-NB group (**Figures 2D–E**). All data mentioned above testify that SSTZF-NB is the optimal anti-bone-loss fraction of SSTZF extract.

# 3.2 SSTZF-NB Fraction Upregulated the Expression of β-Catenin

To determine the possible molecular mechanism of SSTZF-NB anti-osteoporosis, qRT-PCR was performed to examine the effect of SSTZF-NB on the regulation of ALP and FABP4 mRNA expression in  $C_3H_{10}T1/2$  cells. Surprisingly, we found that ALP was highly expressed while FABP4 was downregulated in  $C_3H_{10}T1/2$  cells with any dose of SSTZF-NB treatment (**Figure 3A**). To provide more evidence of SSTZF-NB



preventing bone loss, western bolt was performed to examine the protein expression level of  $\beta$ -catenin after being treated with SSTZF-NB. As expected,  $\beta$ -catenin was significantly upregulated in the low dose SSTZF-NB treatment group (**Figures 3B,C**). These data indicate SSTZF-NB anti-osteoporosis possibly via enhancing osteogenesis and inhibiting adipogenesis and are related to the activation of  $\beta$ -catenin.

### 3.3 SSTZF-NB Fraction Enhanced Osteogenesis and Inhibited Adipogenesis in OVX Mice

To further determine the molecular mechanism of the SSTZF-NB anti-osteoporotic IHC of ALP (an osteogenic specific matrix protein), FABP4 (a fatty acid-binding protein that specifically labels adipocytes) and  $\beta$ -catenin were used. Compared to the sham group mice, the expression of ALP was significantly downregulated while FABP4 was upregulated in the area of the chondro-osseous junction in the OVX mice. These phenomena were significantly reversed and ameliorated in the SSTZF-NB treatment group (**Figures 4A-D**). Furthermore, the expression level of  $\beta$ -catenin in growth plate chondrocytes was decreased in OVX mice and the reduction was elevated in SSTZF-NB group mice (**Figures 4A,B**).

### 3.4 SSTZF-NB Fraction Failed to Ameliorate Bone Loss and Lipid Drops Accumulation in $\beta$ -catenin<sup>Gli1ER</sup> Mice

To provide conclusive evidence about the role of  $\beta$ -catenin signaling in SSTZF-NB preventing bone loss, growth platespecific  $\beta$ -catenin conditional KO mice ( $\beta$ -catenin<sup>Gli1ER</sup>) were generated. To examine the anti-osteoporotic effect of SSTZF-NB, µCT analyzes were first performed. Significantly bone loss was observed in  $\beta$ -catenin<sup>Gli1ER</sup> group mice as expected through the 3D constructed images when compared with Cre-negative group mice. Contrary to previous results, SSTZF-NB failed to alleviate the abnormality (Figure 5A). The indexes of BMD, BV/TV, Tb.Th, Tb.N, and Tb.Sp were not improved significantly in the  $\beta$ -catenin<sup>Gli1ER</sup> + SSTZF-NB group (Figures 5B–F). Additionally, according to the results of ABH staining, significant bone loss and lipid droplet accumulation at the area of femoral metaphysis were found in  $\beta$ -catenin<sup>Gli1ER</sup> group mice compared to cre-negative mice, especially in the chondro-osseous junction area. These pathological changes were also observed in  $\beta$ -catenin<sup>Gli1ER</sup> + SSTZF-NB group mice according to histologic and histomorphometric analyses (Figures 6A-C). These data indicate that SSTZF-NB could not ameliorate the osteoporosislike changes caused by  $\beta$ -catenin-deficiency.

IHC assay showed that the protein expression of  $\beta$ -catenin decreased significantly in the area of the growth plate chondrocytes in  $\beta$ -catenin<sup>Gli1ER</sup> group mice when contrasted with cre-negative group mice. Furthermore, a decreased protein expression of ALP and an increased protein expression of FABP4, which could lead to osteoporotic-like changes, were also observed in the area of the chondro-osseous junction in  $\beta$ -catenin<sup>Gli1ER</sup> group mice. By contrast, these aberrant osteogeneses (decreased ALP) and adipogenesis (increased FABP4) induced by the inhibition of  $\beta$ -catenin signaling in growth-plate-chondrocytes were not restored by SSTZF-NB. No significant difference in the protein expression of ALP and FABP4 was found in  $\beta$ -catenin<sup>Gli1ER</sup> + SSTZF-NB group mice  $\beta$ -catenin<sup>Gli1ER</sup> compared with group when mice (Figures 7A-D).

## **4 DISCUSSION**

In this study, we confirmed that SSTZF-NB could ameliorate the osteoporotic-like phenotype and analyzed its potential mechanisms in mice models. Both  $\mu$ CT and histological results demonstrated that SSTZF-NB could prevent bone loss and alleviate fat accumulation. The fact that the positive effect of SSTZF-NB on anti-osteoporotic could be blocked when  $\beta$ -catenin was specifically deleted in growth plate chondrocyte which has been reported to have multi-differentiation potential, indicating that the mechanism of SSTZF-NB in preventing bone loss may largely depend on its contribution to the activation of  $\beta$ -catenin signaling in growth plate chondrocyte.

TCM is a popular and effective therapeutic method for treating osteoporosis. Shen-sui-tong-zhi formula is a classical anti-osteoporosis TCM that has a favorable curative effect in



images of  $\beta$ -catenin (B), ALP (C) and FABP4 (D). \*p < 0.001.

China. Although there have been many studies for the identification of the ten botanical drugs contained in SSTZ formula (Qu et al., 2016; Liang et al., 2019; Zheng et al., 2019;

Wang et al., 2020), many phytochemicals exist in formula at the same time. Therefore, choosing the appropriate extraction method and combining the active ingredients is important for enhancing the curative effect and avoiding excessive intake of unnecessary chemical compounds (Wang et al., 2008). In this study, according to molecular polarity, petroleum ether, ethyl acetate, and n-butanol were selected as the extraction media for different fractions of the SSTZ formula. µCT analysis, biomechanical tests, and histomorphometry analysis consistently indicated that SSTZF-NB is the optimal anti-osteoporosis fraction. Then, UPLC/MS was used to identify its material basis. Unsurprisingly, a considerable number of the detected chemical components were reported to have the potential of promoting osteogenesis and anti-osteoporosis, like liquiritigenin, formononetin, etc. (Uchino et al., 2015; Mansoori et al., 2016; Gautam et al., 2017; Carnovali et al., 2020). In our study, the combination of these active components presented a satisfactory effect of preventing bone loss and restraining fat accumulation.

It is well known that  $\beta$ -catenin plays a key role in regulating osteogenic differentiation and adipogenic differentiation from MSCs. Moreover, the abnormality of osteogenesis and adipogenesis is a crucial part of the pathological progress of osteoporosis (Wu et al., 2018; Cao et al., 2019). Our research is consistent with previous studies, in which excessive fat accumulation and serious bone loss were observed in the area of chondro-osseous junction, additionally combined with a decreased protein expression of ALP and an increased expression of FABP4. Meanwhile, the expression of  $\beta$ -catenin was also downregulated in growth plate chondrocytes. Recent lineage tracing research indicates that Gli1-expressed growth plate chondrocytes have the properties of progenitor cells, which can differentiate into osteoblasts for osteogenesis, not only chondrogenesis (Shi et al., 2017; Haraguchi et al., 2018). Therefore, to further confirm whether SSTZF-NB could cause antiosteoporosis when  $\beta$ -catenin signaling was inhibited in growth plate chondrocytes,  $\beta$ -catenin<sup>Gli1ER</sup> mice were generated and utilized. However, the positive effect of SSTZF-NB on antiosteoporosis was almost blocked, neither osteogenesis nor adipogenesis, two of the most important aspects of osteoporosis, were ameliorated following SSTZF-NB administration in  $\beta$ -catenin<sup>Gli1ER</sup> mice. These results demonstrated that SSTZF-NB enhances osteogenesis and then acts as an anti-osteoporotic function mainly via activation of β-catenin signaling.

#### **5 CONCLUSION**

The data of this study illuminated the potentiation and molecular mechanism of SSTZF-NB in treating osteoporosis. Based on the aforementioned findings, we conclude that SSTZF-NB enhances osteogenesis mainly via activation of  $\beta$ -catenin signaling in

#### REFERENCES

Alam, M. R., Kim, S. M., Lee, J. I., Chon, S. K., Choi, S. J., Choi, I. H., et al. (2006). Effects of Safflower Seed Oil in Osteoporosis Induced-Ovariectomized Rats. *Am. J. Chin. Med.* 34 (4), 601–612. doi:10.1142/S0192415X06004132 growth plate chondrocytes. Our findings provide a salutary alternative preventive option for osteoporosis.

## DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/**Supplementary Material**, further inquiries can be directed to the corresponding authors.

#### ETHICS STATEMENT

The animal study was reviewed and approved by the Animal Ethics Committee of Zhejiang Chinese Medical University.

#### AUTHOR CONTRIBUTIONS

Study design: RX and HJ. Data collection: RX, QZ, CX, JC, PW, SZ, TX, and WY. Data analysis and interpretation: RX, HL, HX, SL, and ZL. Drafting the article or revising it critically for important intellectual content: RX. Final approval of the article: PT, MG, and HJ.

### FUNDING

This research has been partially supported by the Natural Science Foundation of China (Grant nos. 81904221, 81973869, 81904219, 81904223, and 81873324), the State Administration of Traditional Chinese Medicine of Zhejiang Province (Grant nos. 2018ZZ011, 2019ZQ018, 2020ZA035, and 2021ZZ014), the Health Commission of Zhejiang Province (Grant no. 2019RC225), the Young and Middle-Aged People Research Innovation Fund of Zhejiang University of Traditional Chinese Medicine (Grant no. KC201932), the Excellent Youth Development Plan of Zhejiang University of Traditional Chinese Medicine (Grant no. Q2019Y01), and the Young Elite Scientists Sponsorship Program by CACM (Grant nos. CACM-2018-QNRC2-C03).

#### SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2021.711004/ full#supplementary-material

Cao, H. J., Li, C. R., Wang, L. Y., Ziadlou, R., Grad, S., Zhang, Y., et al. (2019). Effect and Mechanism of Psoralidin on Promoting Osteogenesis and Inhibiting Adipogenesis. *Phytomedicine* 61, 152860. doi:10.1016/j.phymed.2019.152860

Carnovali, M., Banfi, G., and Mariotti, M. (2020). Liquiritigenin Reduces Osteoclast Activity in Zebrafish Model of Glucocorticoid-Induced Osteoporosis. J. Pharmacol. Sci. 143 (4), 300–306. doi:10.1016/j.jphs.2020.06.001

- Chen, Q., Shou, P., Zheng, C., Jiang, M., Cao, G., Yang, Q., et al. (2016). Fate Decision of Mesenchymal Stem Cells: Adipocytes or Osteoblasts? *Cell Death Differ* 23 (7), 1128–1139. doi:10.1038/cdd.2015.168
- Cheng, X., Zhao, K., Zha, X., Du, X., Li, Y., Chen, S., et al. (2021). Opportunistic Screening Using Low-Dose CT and the Prevalence of Osteoporosis in China: A Nationwide, Multicenter Study. *J. Bone Miner Res.* 36 (3), 427–435. doi:10.1002/ jbmr.4187
- Choi, J. S., Suh, S. S., Young, H. S., and Park, H. J. (1991a). Hypolipemic and Hypoglycemic Activities of Prunus Davidiana in High Fat-Fed Rats. Arch. Pharm. Res. 14 (1), 44–47. doi:10.1007/BF02857813
- Choi, J. S., Yokozawa, T., and Oura, H. (1991b). Antihyperlipidemic Effect of Flavonoids from Prunus Davidiana. J. Nat. Prod. 54 (1), 218–224. doi:10.1021/ np50073a022
- Colaianni, G., Sun, L., Zaidi, M., and Zallone, A. (2015). The "love Hormone" Oxytocin Regulates the Loss and Gain of the Fat-Bone Relationship. *Front. Endocrinol. (Lausanne)* 6, 79. doi:10.3389/fendo.2015.00079
- Cui, D., Zhao, D., and Huang, S. (2019). Beneficial Contribution of a Safflower (Carthamus tinctorius L.) Polysaccharide on Steroid-Induced Avascular Necrosis of the Femoral Head in Rats. *Int. J. Biol. Macromol* 123, 581–586. doi:10.1016/j.ijbiomac.2018.11.046
- de Winter, T. J. J., and Nusse, R. (2021). Running against the Wnt: How Wnt/ $\beta$ -Catenin Suppresses Adipogenesis. Front Cell Dev Biol 9, 627429. doi:10.3389/fcell.2021.627429
- Force, U. S. P. S. T., Curry, S. J., Krist, A. H., Owens, D. K., Barry, M. J., Caughey, A. B., et al. (2018). Screening for Osteoporosis to Prevent Fractures: US Preventive Services Task Force Recommendation Statement. *JAMA* 319 (24), 2521–2531. doi:10.1001/jama.2018.7498
- Gautam, J., Khedgikar, V., Kushwaha, P., Choudhary, D., Nagar, G. K., Dev, K., et al. (2017). Formononetin, an Isoflavone, Activates AMP-Activated Protein Kinase/β-Catenin Signalling to Inhibit Adipogenesis and Rescues C57BL/6 Mice from High-Fat Diet-Induced Obesity and Bone Loss. *Br. J. Nutr.* 117 (5), 645–661. doi:10.1017/S0007114517000149
- Haraguchi, R., Kitazawa, R., Imai, Y., and Kitazawa, S. (2018). Growth Plate-Derived Hedgehog-Signal-Responsive Cells Provide Skeletal Tissue Components in Growing Bone. *Histochem. Cell Biol* 149 (4), 365–373. doi:10.1007/s00418-018-1641-5
- Hu, L., Yin, C., Zhao, F., Ali, A., Ma, J., and Qian, A. (2018). Mesenchymal Stem Cells: Cell Fate Decision to Osteoblast or Adipocyte and Application in Osteoporosis Treatment. *Int. J. Mol. Sci.* 19 (2), 360. doi:10.3390/ijms19020360
- Jung, H. A., Ali, M. Y., Bhakta, H. K., Min, B. S., and Choi, J. S. (2017). Prunin Is a Highly Potent Flavonoid from Prunus Davidiana Stems that Inhibits Protein Tyrosine Phosphatase 1B and Stimulates Glucose Uptake in Insulin-Resistant HepG2 Cells. Arch. Pharm. Res. 40 (1), 37–48. doi:10.1007/s12272-016-0852-3
- Kim, H. J., Bae, Y. C., Park, R. W., Choi, S. W., Cho, S. H., Choi, Y. S., et al. (2002). Bone-protecting Effect of Safflower Seeds in Ovariectomized Rats. *Calcif Tissue Int.* 71 (1), 88–94. doi:10.1007/s00223-001-1080-4
- Liang, Y., Li, Y., Sun, A., and Liu, X. (2019). Chemical Compound Identification and Antibacterial Activity Evaluation of Cinnamon Extracts Obtained by Subcritical N-Butane and Ethanol Extraction. *Food Sci. Nutr.* 7 (6), 2186–2193. doi:10.1002/fsn3.1065
- Liu, Z. Z., Hong, C. G., Hu, W. B., Chen, M. L., Duan, R., Li, H. M., et al. (2020). Autophagy Receptor OPTN (Optineurin) Regulates Mesenchymal Stem Cell Fate and Bone-Fat Balance during Aging by Clearing FABP3. *Autophagy*, 1–17. doi:10.1080/15548627.2020.1839286
- Lorthongpanich, C., Thumanu, K., Tangkiettrakul, K., Jiamvoraphong, N., Laowtammathron, C., Damkham, N., et al. (2019). YAP as a Key Regulator of Adipo-Osteogenic Differentiation in Human MSCs. *Stem Cell Res Ther* 10 (1), 402. doi:10.1186/s13287-019-1494-4
- Mansoori, M. N., Tyagi, A. M., Shukla, P., Srivastava, K., Dev, K., Chillara, R., et al. (2016). Methoxyisoflavones Formononetin and Isoformononetin Inhibit the Differentiation of Th17 Cells and B-Cell Lymphopoesis to Promote Osteogenesis in Estrogen-Deficient Bone Loss Conditions. *Menopause* 23 (5), 565–576. doi:10.1097/GME.000000000000646
- Mattioli, A. V., Sciomer, S., Cocchi, C., Maffei, S., and Gallina, S. (2020). Quarantine during COVID-19 Outbreak: Changes in Diet and Physical Activity Increase the Risk of Cardiovascular Disease. *Nutr. Metab. Cardiovasc. Dis.* 30 (9), 1409–1417. doi:10.1016/j.numecd.2020.05.020

- Pei, L., and Tontonoz, P. (2004). Fat's Loss Is Bone's Gain. J. Clin. Invest. 113 (6), 805–806. doi:10.1172/JCI21311
- Qiang, Y. W., Heuck, C. J., Shaughnessy, J. D., Jr., Barlogie, B., and Epstein, J. (2012). Proteasome Inhibitors and Bone Disease. *Semin. Hematol.* 49 (3), 243–248. doi:10.1053/j.seminhematol.2012.04.011
- Qu, C., Wang, L. Y., Jin, W. T., Tang, Y. P., Jin, Y., Shi, X. Q., et al. (2016). Comparative Analysis of the Effects of Hydroxysafflor Yellow A and Anhydrosafflor Yellow B in Safflower Series of Herb Pairs Using Prep-HPLC and a Selective Knock-Out Approach. *Molecules* 21 (11), 1480. doi:10.3390/molecules21111480
- Shi, Y., He, G., Lee, W. C., McKenzie, J. A., Silva, M. J., and Long, F. (2017). Gli1 Identifies Osteogenic Progenitors for Bone Formation and Fracture Repair. *Nat. Commun.* 8 (1), 2043. doi:10.1038/s41467-017-02171-2
- Uchino, K., Okamoto, K., Sakai, E., Yoneshima, E., Iwatake, M., Fukuma, Y., et al. (2015). Dual Effects of Liquiritigenin on the Proliferation of Bone Cells: Promotion of Osteoblast Differentiation and Inhibition of Osteoclast Differentiation. *Phytother Res.* 29 (11), 1714–1721. doi:10.1002/ ptr.5416
- Uezumi, A., Fukada, S., Yamamoto, N., Takeda, S., and Tsuchida, K. (2010). Mesenchymal Progenitors Distinct from Satellite Cells Contribute to Ectopic Fat Cell Formation in Skeletal Muscle. *Nat. Cell Biol* 12 (2), 143–152. doi:10.1038/ncb2014
- Vandenbroucke, A., Luyten, F. P., Flamaing, J., and Gielen, E. (2017). Pharmacological Treatment of Osteoporosis in the Oldest Old. *Clin. Interv. Aging* 12, 1065–1077. doi:10.2147/CIA.S131023
- Wang, X. L., Wang, N. L., Zhang, Y., Gao, H., Pang, W. Y., Wong, M. S., et al. (2008). Effects of Eleven Flavonoids from the Osteoprotective Fraction of Drynaria Fortunei (KUNZE) J. SM. On Osteoblastic Proliferation Using an Osteoblast-like Cell Line. Chem. Pharm. Bull. (Tokyo) 56 (1), 46–51. doi:10.1248/cpb.56.46
- Wang, Z., Peng, S., Peng, M., She, Z., Yang, Q., and Huang, T. (2020). Adsorption and Desorption Characteristics of Polyphenols from Eucommia Ulmoides Oliv. Leaves with Macroporous Resin and its Inhibitory Effect on α-amylase and α-glucosidase. Ann. Transl Med. 8 (16), 1004. doi:10.21037/atm-20-5468
- Wu, Q., Xu, Y., and Lin, G. (2019). Trends and Disparities in Self-Reported and Measured Osteoporosis Among US Adults, 2007-2014. J. Clin. Med. 8 (12), 2052. doi:10.3390/jcm8122052
- Wu, Y., Xie, L., Wang, M., Xiong, Q., Guo, Y., Liang, Y., et al. (2018). Mettl3mediated m6A RNA Methylation Regulates the Fate of Bone Marrow Mesenchymal Stem Cells and Osteoporosis. *Nat. Commun.* 9 (1), 4772. doi:10.1038/s41467-018-06898-4
- Xu, R., Luo, C., Ge, Q., Ying, J., Zhang, P., Xia, C., et al. (2020). Radix Rehmanniae Praeparata Promotes Bone Fracture Healing through Activation of TGF-β Signaling in Mesenchymal Progenitors. *Biomed. Pharmacother.* 130, 110581. doi:10.1016/j.biopha.2020.110581
- Yan, C. L., Li, S. H., An, F. Y., Liu, Y. Q., Xia, P. F., Ma, Z. M., et al. (2018). The Therapeutic Effects of Yougui Pill on Knee Osteoarthritis and the Expression of Wnt Signal Pathway Related Factors in Rats. *Zhongguo Ying Yong Sheng Li Xue Za Zhi* 34 (2), 177–181. doi:10.12047/ j.cjap.5603.2018.043
- Zhang, N. D., Han, T., Huang, B. K., Rahman, K., Jiang, Y. P., Xu, H. T., et al. (2016). Traditional Chinese Medicine Formulas for the Treatment of Osteoporosis: Implication for Antiosteoporotic Drug Discovery. J. Ethnopharmacol 189, 61–80. doi:10.1016/j.jep.2016.05.025
- Zhang, P., Xu, H., Wang, P., Dong, R., Xia, C., Shi, Z., et al. (2019). Yougui Pills Exert Osteoprotective Effects on Rabbit Steroid-Related Osteonecrosis of the Femoral Head by Activating  $\beta$ -catenin. *Biomed. Pharmacother.* 120, 109520. doi:10.1016/j.biopha.2019.109520
- Zheng, Z. G., Xu, Y. H., Liu, F., Zhao, T. T., Wang, R. X., Huang, P. Y., et al. (2019). Screening Bioactive Components of Glycyrrhiza Uralensis Fisch. With Isolated Perfused Lung Extraction and HPLC-ESI-MSn Analysis. J. Pharm. Biomed. Anal. 169, 127-132. doi:10.1016/ j.jpba.2019.03.007
- Zhu, B., Zhang, Q. L., Hua, J. W., Cheng, W. L., and Qin, L. P. (2018). The Traditional Uses, Phytochemistry, and Pharmacology of Atractylodes Macrocephala Koidz.: A Review. J. Ethnopharmacol 226, 143–167. doi:10.1016/j.jep.2018.08.023

Zou, W., Rohatgi, N., Brestoff, J. R., Li, Y., Barve, R. A., Tycksen, E., et al. (2020). Ablation of Fat Cells in Adult Mice Induces Massive Bone Gain. *Cell Metab* 32 (5), 801–e6. doi:10.1016/j.cmet.2020.09.011

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Xu, Zeng, Xia, Chen, Wang, Zhao, Yuan, Lou, Lin, Xia, Lv, Xu, Tong, Gu and Jin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.