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PERSPECTIVES

Capillary Inward-Rectifying K⁺ Crippled in a Mouse Model of Alzheimer's Disease: Phosphatidylinositol 4,5-Bisphosphate to the Rescue!

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A Perspective on: "PIP₂ Improves Cerebral Blood Flow in a Mouse Model of Alzheimer's Disease"

Blood flow to the brain is precisely regulated to match the metabolic activity of neurons.¹ This process, dubbed neurovascular coupling, ensures the appropriate supply of oxygen, glucose, and other substrates necessary for proper brain function.¹ It has become apparent over the past two decades that cerebral blood flow is reduced, and neurovascular coupling is attenuated in a number of brain pathologies, including Alzheimer's disease (AD).^{1–3} However, the time course of dysregulation of cerebral blood flow relative to the onset of cognitive impairment, the underlying mechanisms responsible for the dysregulation of blood flow, and, importantly, if reversal of impaired cerebrovascular function improves cognition in AD remains in question. Mughal et al.4 in this issue of Function provide compelling evidence that reduced membrane phosphatidylinositol 4,5-bisphosphate (PIP₂), an established characteristic of Alzheimer's pathology,⁵ inactivates brain capillary endothelial cell (EC) inward-rectifying K⁺ (K_{IR}2.1) channels, resulting in attenuated neurovascular coupling in the whisker barrel cortex in a murine model of AD in which the mice express five mutant human genes associated with familial AD: three amyloid precursor protein (APP) genes (APPswe, APPflo, and APPlon) and two presenilin 1 (PS1 and PSEN1) genes (PSEN1 M146L and PSEN1 L286V; 5XFAD mouse).⁶ The authors demonstrate that capillary EC K_{IR}2.1 channel function is crippled in this model system and that application of a

PIP₂ analog in patch-clamp experiments completely rescues the channel function. Importantly, they go on to show that K⁺-induced enhancement of red blood cell flux in capillaries, an in vivo test of capillary EC $K_{\mbox{\tiny IR}}2.1$ function, and neurovascular coupling in the somatosensory cortex invoked by whisker stimulation are likewise impaired in the 5XFAD mouse model of AD. Most excitingly, Mughal et al.⁴ demonstrated rescue of capillary EC K_{IR}2.1 function and neurovascular coupling by intravenous (IV) administration of a PIP₂-analog. These data offer hope of dietary or pharmacological restoration of capillary EC membrane PIP₂ levels and restoration of ion channel function impaired by a reduction in PIP₂ in AD. The authors' findings also strongly support this group's contention that capillary EC K_{IR}2.1 channels serve as an important vascular sensor of extracellular [K⁺] released in proportion to neural and glial activity, providing a key signal that couples increases in local neuron electrical activity with increases in capillary blood flow to these active cells.

Several questions remain to be answered. First, what is the time course of capillary EC PIP₂ depletion relative to loss of neurons and, importantly, impaired cognitive function? How early are the capillary K_{IR} 2.1 channels crippled in the progression of AD? Mughal et al.⁴ used 12-month-old 5XFAD mice in their investigations. However, studies in this model have shown impaired cognition and loss of neurons as early as 4–5 months, while changes in cerebral blood flow appear at approximately 7 months.⁷ Is capillary EC K_{IR} 2.1 function also impaired at these time points? Second, does recovery of K_{IR} 2.1 function by addition of exogenous PIP₂ restore or improve cognitive function in

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This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/ licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com this model of AD and at what point does the PIP₂ have to be administered? Third, how selective was the IV administration of the PIP₂ analog to capillary EC? Decreased membrane PIP₂ also has been implicated in reduced synaptic transmission in AD models.^{5,8} Did the IV administration of the PIP₂ analog improve signaling elsewhere in the neurovascular unit (neurons, astrocytes, etc.)? Fourth, while capillary EC K_{IR}2.1 function is impaired by loss of PIP₂.^{4,9} capillary EC TRPV₄ function should be enhanced by loss of PIP₂.⁹ Does, this imply that an increase in capillary EC TRPV₄ activity may contribute, somehow, to impaired neurovascular coupling in AD? Finally, it will be interesting to see if capillary K_{IR}2.1 is also crippled in human AD and whether K_{IR}2.1 function can be restored by PIP₂ supplementation. Obviously, additional research will be required to answer these and other questions that arise from this provocative study. Nonetheless, the paper by Mughal et al.4 provides exciting new information that may help in our fight to combat cerebral pathologies, like AD.

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Conflict of interest statement

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References

- 1. Iadecola C. The neurovascular unit coming of age: a journey through neurovascular coupling in health and disease. *Neuron* 2017;96(1):17–42.
- Kisler K, Nelson AR, Montagne A, Zlokovic BV. Cerebral blood flow regulation and neurovascular dysfunction in Alzheimer disease. Nat Rev Neurosci 2017;18(7):419–434.
- 3. Iadecola C, Gottesman RF. Cerebrovascular alterations in Alzheimer aisease. Circ Res 2018;123(4):406–408.
- 4. Mughal A, Harraz OF, Gonzales AL, Hill-Eubanks D, Nelson MT. PIP2 improves cerebral blood flow in a mouse model of Alzheimer's disease. *Function* 2021;2(2). doi: 10.1093/function/ zqab010.
- 5. Di Paolo G, Kim TW. Linking lipids to Alzheimer's disease: cholesterol and beyond. Nat Rev Neurosci 2011;12(5):284–296.
- 6.Oakley H, Cole SL, Logan S, et al. Intraneuronal beta-amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation. J Neurosci 2006;26(40): 10129–10140.
- 7. Igarashi H, Ueki S, Kitaura H, et al. Longitudinal GluCEST MRI changes and cerebral blood flow in 5xFAD mice. Contrast Media Mol Imaging 2020;2020:8831936. doi: 10.1155/2020/8831936.
- He Y, Wei M, Wu Y, et al. Amyloid beta oligomers suppress excitatory transmitter release via presynaptic depletion of phosphatidylinositol-4,5-bisphosphate. Nature communications 2019; 10(1):1193.
- 9. Harraz OF, Longden TA, Hill-Eubanks D, Nelson MT. PIP2 depletion promotes TRPV4 channel activity in mouse brain capillary endothelial cells. *Elife* 2018;7. doi: 10.7554/eLife.38689.