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ORIGINAL ARTICLE

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Diagnostic Value of Bile Acids and Fibroblast Growth Factor 21 in Women with Polycystic Ovary Syndrome

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Abstract

Objective: Polycystic ovary syndrome (PCOS) is a heterogeneous disorder characterized by a reduction in fertility and metabolic dysfunction. Unfortunately, due to a lack of clear presentation, it is often a long process of diagnosis. In this study, we investigated bile acids as potential biomarkers.

Materials and Methods: Subjects were recruited and stratified into groups based on body mass index and PCOS status. Biometric data and plasma were acquired to understand bile acid profiles and related markers. *Results:* Taurocholic acid (TCA) and taurodeoxycholic acid were elevated in PCOS subjects with obesity in comparison to controls without PCOS. Fibroblast growth factor 21 (FGF-21), a metabolic regulator implemented in bile acid metabolism, was elevated in PCOS patients and was positively correlated with TCA changes. *Conclusions:* We present evidence suggesting that bile acids may be novel diagnostic targets in obese patients with PCOS while further studies need to delineate the interplay between FGF-21, bile acids, and testosterone in the early detection of PCOS.

Keywords: polycystic ovary syndrome; bile acids; hyocholic acid; obesity; FGF-21; FGF-19

Introduction

Polycystic ovary syndrome (PCOS) is a disorder of metabolic and reproductive function and affects 6%–15% of women of reproductive age. Classic features include chronic anovulation, polycystic ovary morphology on ultrasound, and hyperandrogenism such as hirsutism, acne, or alopecia.¹ Three guidelines that exist are overlapping, yet, both the Androgen Excess Society and the National Institutes of Health criteria require hyperandrogenism as a key diagnostic component.^{2,3} Polycystic ovaries alone can be seen in women

with no other endocrine anomalies; therefore, hyperandrogenism remains a key aspect of this disease. It is known that women with PCOS are at increased risk for development of type 2 diabetes and metabolic syndrome (glucose intolerance, hypertension, dyslipidemia, and central obesity).⁴ It is therefore important to recognize this syndrome in adolescent women, as early intervention may prevent long-term sequelae.

Biomarkers have the potential to aid in earlier diagnosis of this complex disorder. It has been demonstrated that chronic inflammation may be a link

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Table 1. Biometric Data of Recruited Patien

	Control/no PCOS (n = 6)	Obese/no PCOS (n = 6)	Control/ PCOS (n = 7)	Obese/ PCOS (n = 8)
BMI	23.03 ± 1.17	37.1 ± 1.79	23.12 ± 1.31	37.7 ± 1.56
Age	20 ± 0.63	23.17 ± 1.76	21.57 ± 1.4	21.13 ± 0.77
Menarche age	11.8 ± 1.09	13 ± 1	13.8 ± 2.14	12.625 ± 2.97
Testosterone	31.17 ± 3.22	33.5 ± 2.5	41.58 ± 2.87	$48.5 \pm 4.57*$
Insulin	7.98 ± 1.35	$20.53 \pm 4.41 **$	7.24 ± 1.05	21.65 ± 3.92***

A total of 27 patients were included in this study. Insulin data are consistent with metabolic disease and a reduction in insulin sensitivity. (Hyperinsulinemia is a diagnostic marker for type 2 diabetes.)

*p = 0.014, **p = 0.022, ***p = 0.005. BMI, body mass index; PCOS, polycystic ovary syndrome.

between the metabolic disturbances in PCOS and hyperandrogenism.⁵⁻⁷ Many surrogate inflammatory markers have been studied among women with PCOS leading to a large heterogeneity of results.^{8,9} This is further compounded by obesity, as up to 64% of PCOS patients are obese.¹⁰ Obese PCOS subjects have increased androgens compared with normal weight PCOS subjects.¹¹ Obese PCOS subjects also have increased androgens and fasting insulin levels compared with obese controls without PCOS.^{1,6} Despite an increased prevalence of obesity among women with PCOS, it is also known that normal weight PCOS patients also have an increased risk for metabolic disturbance.¹² Hence, identification of biomarkers that are sensitive to metabolic derangements as seen in PCOS is required for early diagnosis.

Bile acids are involved in intestinal uptake of dietary lipids and fat-soluble vitamins in addition to acting as signaling molecules for cholesterol and lipid homeostasis.^{13–15} Bile acids also act as signaling molecules to regulate hepatic metabolism. Bile acids are first synthesized in the liver from cholesterol as primary bile acids, cholic acid, and chenodeoxycholic acid. Once synthesized, bile acids can be conjugated (taurinated/ glycinated) and are subsequently stored in the gallbladder and are delivered to the gut after a meal. In the gut, bile acids can undergo bacterial deconjugation and dehydroxylation in the colon, yielding secondary bile acids (lithocholic acid, deoxycholic acid, ursodeoxycholic acid [UDCA]). Ninety-five percent of bile acids in the colon are recycled back to the liver via the enterhepatic circulation (only 5% are excreted into the feces).^{13,15,16}

As several publications have eluded, bile acid levels and bile acid species composition (specifically changes in concentration of secondary and conjugated bile acids such as UDCA and glycoursodeoxycholic acid)

are connected to insulin sensitivity following bariatric surgery.^{14,17,18} Following bariatric surgery, 80%–90% of insulin-insensitive patients experience a reversal in insulin sensitivity.¹⁹⁻²¹ This change is correlated with a change in bile acid pool size and species, suggesting a potential new feature and biomarker potential.¹⁴ As PCOS is associated with metabolic derangements independent of body mass index (BMI), we hypothesize that changes in certain bile acid species levels correlate with PCOS diagnostic criteria and hyperandrogenemia, making them a potential biomarker for diagnosis.

The goal of this study was to look at differences in bile acid species among women with known PCOS and controls, and to account for obesity by stratifying groups by BMI. In addition, we sought to understand the connection of bile acids with other factors associated with metabolic health, such as fibroblast growth factor-21 (FGF-21) and fibroblast growth factor 19 (FGF-19). As the incidence rate of PCOS continues to rise, it is vital to characterize the disease early, and biomarker analysis could potentially expedite this process, decreasing disease progression and improving patient outcomes.

Table 2.	Preva	lence of	a First-	or Second-[Degree Fa	mily
Member	with	Polycysti	ic Ovary	/ Syndrome	or Polycy	/stic
Ovary Sy	ndroi	ne-Relate	d Cond	litions		

	Control (lean and obese) (%)	PCOS (lean and obese) (%)
PCOS	16.7	33.3
Heart disease	25	73.3*
Diabetes	25	80**
Menstrual cycle intervals > 90 days	0	73.3***

PCOS patients reported more family members with heart disease and diabetes. In addition, most PCOS patients reported altered menstrual cycles, while control patients reported normal menstrual cycles. p = 0.021, p = 0.007, p = 0.001.

Methods

This is a cross-sectional study evaluating serum testosterone, insulin, metabolic regulators (FGF-19/FGF-21), and bile acids among young women with and without PCOS, stratified by BMI.

Subjects

This study was conducted among women attending an academic OBGYN practice in Huntington, WV, during 2018/2019 before the COVID-19 global pandemic. Women aged 18–35 years were recruited into one of four subject groups: PCOS with obesity, PCOS without obesity, non-PCOS with obesity, and non-PCOS without obesity. Diagnostic criteria for PCOS included a history of chronic anovulation based on menstrual history, and clinical evidence of hyperandrogenism (acne, hirsutism, or alopecia).

Clinical or serum evidence of hyperandrogenism was included as key diagnostic criteria for study subjects as it is included in the most specific diagnostic guidelines. For the control non-PCOS subjects, inclusion criteria required no clinical evidence of hyperandrogenism (acne, hirsutism, or alopecia) and regular menstrual cycles. All subjects were at least 2 years from menarche. Obesity was defined by a BMI >30 kg/m². This distinction was made to delineate alterations independent and dependent on weight as many of the markers evaluated are subject to change in an obese background.

Exclusion criteria included: (1) metformin use within past 3 months, (2) combination contraceptive use within past 3 months, (3) antibiotic use within 2 weeks of blood draw, (4) lactating, (5) pregnant within past 18 months, (6) known thyroid disease, (7) hyperprolactinemia, (8) liver disease, (9) other androgen disorders such as Cushing's disease, and (10) recent gastrointestinal illness within the past 2 weeks. Detailed clinical surveys were given to delineate medical history, age of menarche, family history specifically of PCOS, and cardiovascular and metabolic diseases such as type 2 diabetes. Once the subjects were recruited, their BMI was calculated using their weight and height. Informed consent was obtained from all study subjects. This study was approved by the Marshall Internal Review Board (1293149).

Blood measurements

Recruited patients were subjected to venipuncture to collect blood for biochemical measurements following an overnight fast. Blood for bile acid analysis was immediately centrifuged for collection of plasma, which was then subsequently frozen at -80° C to maintain integrity. Bile acid plasma was sent to Michigan Metabolomics for analysis. Fasting serum insulin and testosterone levels were measured by Cabell Huntington Hospital Laboratories. FGF-19 and FGF-21 levels were assessed using ELISA (R&D) per the manufacturer's instructions.

Bile acid measurements

Following plasma separation, samples were stored in the -80° C freezer before sending to the University of Michigan Metabolomics Core for analysis. Bile acids were separated and defined using a two-step solvent extraction and liquid chromatography-mass spectrometry separation by reverse-phase liquid chromatography and measurements by electrospray ionization triple quadruple multiple reaction monitoring methods (QQQ MRM).²²

Results

Twenty-seven subjects were recruited and consented for study participation. Subjects were placed into one of four groups based on their medical history with and without PCOS and their BMI (BMI >30 kg/m², obese; BMI <30 kg/m², nonobese). There were 15 subjects who met the clinical criteria for PCOS, and 12 control non-PCOS patients. These are further stratified by BMI in Table 1. There was no difference in age between groups. Subjects with PCOS (nonobese and obese combined) had an average age of menarche of 13.1 (standard deviation [SD] 2.56) compared with mean age of 12.2 (SD 1.13) among controls (p=0.27) (Table 1).

In regard to differentiation between nonobese and obese patients, the average age of menarche was 13.8 and 12.625, respectively (Table 1). Two PCOS subjects noted that they required hormonal therapy to induce menarche. PCOS subjects had a higher prevalence compared with controls of any first- or second-degree relative with PCOS (33.3% vs. 16.7%, p=0.41), heart disease (73.3% vs. 25%, p=0.021), or diabetes (80%) vs. 25%, p = 0.007) (Table 2). PCOS subjects also had a higher reported incidence of menstrual cycle intervals greater than 90 days apart within the past year (73.3% vs. 0%, p=0.001) (Table 2). As predicted with their history, subjects with PCOS had an increase in testosterone levels indicative of hyperandrogenism, independent of obesity (Table 1). Fasting insulin levels were also elevated in PCOS and non-PCOS subjects

THCA	0.20 ± 0.08	0.25 ± 0.14	0.27 ± 0.11	0.41 ± 0.11
T ₀ MCA/ TBMCA	0.90 ± 0.34	0.59 ± 0.27	1.05 ± 0.30	2.06 ± 0.69
TUDCA	0.89 ± 0.21	0.69 ± 0.16	1.06 ± 0.28	1.00 ± 0.21
TCDCA	14.35 ± 3.51	12.27 ± 5.17	10.20 ± 1.50	20.66 ± 3.57
TLCA	0.15 ± 0.03	0.20 ± 0.04	0.27 ± 0.06	0.18 ± 0.05
GCA	0.36 ± 0.10	0.67 ± 0.24	0.35 ± 0.08	0.69 ± 0.11
GUDCA	1.24 ± 0.39	2.54 ± 1.72	1.34 ± 0.45	1.49 ± 0.29
GDCA	2.38 ± 0.90	3.15 ± 1.61	2.79 ± 0.51	2.63 ± 0.53
GCDCA	5.34 ± 1.36	11.00 ± 5.21	4.89 ± 0.54	7.72 ± 0.85
GLCA	0.06 ± 0.02	0.10 ± 0.05	0.13 ± 0.05	0.02 ± 0.01
MCA	0.11 ± 0.03	0.27 ± 0.21	0.11 ± 0.05	0.12 ± 0.03
НСА	0.10 ± 0.02	0.05 ± 0.12	0.21 ± 0.14	0.06 ± 0.02
CA	1.92 ± 0.79	2.25 ± 0.99	0.10 ± 0.41	1.56 ± 0.35
BMCA	0.13 ± 0.06	0.64 ± 0.18	0.34 ± 0.14	0.07 ± 0.03
αMCA	0.11 ± 0.031	0.27 ± 0.20	0.10 ± 0.05	0.10 ± 0.29
HDCA/	1.50 ± 0.56	1.77 ± 0.56	2.67 ± 0.69	2.21 ± 0.94
DCA	21.87 ± 8.75	18.57 ± 3.74	23.63 ± 7.37	20.42 ± 3.30
CDCA	1.85 ± 0.63	2.85 ± 1.74	1.13 ± 0.64	1.09 ± 0.31
	Control/no PCOS	Obese/no PCOS	Control/PCOS	Obese/PCOS

Table 3. Relative Response of Bile Acids Unaltered by Body Mass Index and Polycystic Ovary Syndrome Status

Data are shown as relative rate average±standard error of the mean. aMCA, alpha-muricholic acid; *β*MCA, beta-muricholic acid; *Q*MCA, omega-muricholic acid; CA, chonodeoxycholic acid; DCA, deoxycholic acid; GCA, glycocholic acid; GCDCA, glycocheno-deoxycholic acid; GDCA, glycodeoxycholic acid; GLCA, glycolithocholic acid; GUDCA, glycoursodeoxycholic acid; HCA, hyocholic acid; TCDCA, taurochenodeoxycholic acid; TLCA, taurochenodeoxycholic acid; TUCA, tauromuricholic acid; TUCA, taurochenodeoxycholic acid; TLCA, taurochenodeoxycholic acid; TUCA, taurochenochenodeoxycholic acid; TUCA, taurochenodeoxycholic ac

with obesity, highlighting possible insulin resistance (Table 1).

Fasting bile acid levels in all the four groups of study participants were relatively unchanged (Table 3 and Fig. 1A). A more in-depth analysis of groups of bile acids also proved to be unrelated to PCOS as glycineconjugated bile acids remained unaltered (Table 3 and Fig. 1B). Primary and secondary bile acids were also unaltered in patients with PCOS (Fig. 1C, D). Interestingly taurocholic acid (TCA) levels, a primary bile acid, increased significantly in PCOS patients with obesity compared with obese non-PCOS patients, reflecting a compounding change due to the metabolic state of the patient (Fig. 2A). In addition, taurodeoxycholic acid (TDCA) was only significantly elevated in obese PCOS patients compared with obese non-PCOS patients (Fig. 2B). In total, these data suggest that only some bile acids are altered in PCOS and that obesity with PCOS reflects a different bile acid expression pattern in comparison to PCOS patients who exhibit a normal BMI.

We next evaluated molecular regulators and factors associated with bile acid metabolism. FGF-19 and



FIG. 1. Overall bile acid levels or accumulation of glycine-conjugated bile acids are unchanged in PCOS. Subjects were subjected to a fasting blood draw to elucidate changes in bile acid species and accumulation. **(A)** Primary bile acids were unchanged. **(B)** Glycine-conjugated bile acids are unresponsive to PCOS metabolic disruption. **(C)** Primary bile acid (CDCA and CA) levels were not significantly altered in PCOS. **(D)** Secondary bile acids were unchanged in subjects with/without PCOS. CA, cholic acid; CDCA, chenodeoxycholic acid; PCOS, polycystic ovary syndrome.



taurocholic acid; TCDCA, taurochenodeoxycholic acid; TDCA, taurodeoxycholic acid.

FGF-21 are part of the fibroblast growth factor family of signaling molecules that are involved in regulating bile acid, glucose, and lipid metabolism.²³⁻²⁵ FGF-21 is a starved state hormone that augments and stages alterations in glucose metabolism to maintain homeostasis. Interestingly, FGF-21 levels are also augmented during metabolic diseases such as obesity and type 2 diabetes have increased concentrations.²³⁻²⁵ It has been previously reported that FGF-19 and FGF-21 levels are augmented in PCOS; however, there are also some reports that FGF-19 and FGF-21 levels are unchanged.²⁶⁻²⁸ In our population, FGF-19 levels were unchanged in the fasted state, in the normal weighted and obese background (Fig. 3A). In both weighted backgrounds, FGF-21 levels were significantly increased by 2.5- to 3-fold (Fig. 3B).

PCOS obese patients with higher levels of FGF-21 also had higher levels of TCA, perhaps suggesting a regulatory role of FGF-21 in bile acid accumulation in PCOS (Fig. 4D). In addition, we also saw a correlation between higher FGF-21 levels with higher testosterone levels, again suggesting a possible interplay in obese patients with PCOS (Fig. 5D). We did not observe any correlation between TCA and FGF-21 or testosterone and FGF-21 in the control subjects (lean or obese) or in PCOS nonobese subjects (Figs. 4A–C and 5A–C). This suggests that obesity in PCOS has a different signaling profile in comparison to PCOS in a lean individual.

Discussion

At the time of sample collection, this was thought to be the first study to elucidate bile acid species and pool in conjunction with other metabolic factors in PCOS patients. Only two bile acids, TCA and TDCA, were altered in PCOS patients compared with controls. Interestingly, TCA has been shown to increase FGF-21 levels in primary rat hepatocytes, perhaps showing a connection between bile acids and FGFs in the PCOS condition.²⁹ Additionally, FGF-21, not FGF-19, was positively correlated with PCOS with and without obesity in our study. Previously, it has been noted that FGF-21 levels are increased in PCOS patients.^{28,30} In Olszanecka-Glinianowicz et al, it is observed that FGF-21 levels are elevated in PCOS patients, regardless of weight, compared with non-PCOS controls.³¹

In addition, FGF-21 levels are elevated in obese PCOS subjects in comparison to lean PCOS subjects, a result we did not see.³¹ This discrepancy in data could be a result of differences in the population or the period at which samples were collected regarding the subjects' menstrual cycle (we did not restrict blood draw to 3–5 days before menstrual cycle as was indicated in Olszanecka-Glinianowicz et al.) In addition, insulin levels were quite different between our data sets (*e.g.*, obese PCOS insulin levels in our study average 21.65 μ IU/mL vs. 12.9 μ IU/mL³¹ in their population), which could also account for differences in FGF-21 as insulin is positive regulator of hepatic FGF-21 levels.²⁹ FGF-19 and FGF-21 are involved in



FIG. 3. FGF-21, not FGF-19, is positively correlated with PCOS. **(A)** FGF-19 levels remain consistent across all the groups. **(B)** In both lean and obese backgrounds, FGF-21 levels are enhanced compared with non-PCOS subjects (*p = 0.0306, **p = 0.038). FGF-19, fibroblast growth factor 19; FGF-21, fibroblast growth factor 21.



FIG. 4. FGF-21 levels correlate with increased levels of TCA in obese PCOS patients. **(A–D)** Using Pearson's correlation analysis, fasting levels of TCA were positively correlated with higher levels of circulating FGF-21 only in patients with PCOS who were also obese **(D)**. (*p = 0.028/r = 0.76).



FIG. 5. FGF-21 levels correlate with increased levels of testosterone. **(A–D)** Using Pearson's correlation analysis, fasting levels of testosterone were positively correlated with higher levels of circulating FGF-21 only in obese PCOS patients **(D)**. (*p = 0.008/r = 0.84).

metabolic balance; they are also implicated in lipid and bile acid metabolism.

FGF-21 has been explored as an antidiabetic hepatokine for the treatment of type 2 diabetes with somewhat promising results.³² FGF-21 has been shown to have tissue-specific effects, but ultimately pharmacological overexpression of FGF-21 recapitulates metabolic starvation, thus promoting fuel utilization and decrease in adiposity.²³ Indeed, pharmacological overexpression beyond physiological levels could be a promising research goal to explore for the treatment of PCOS. Finally, during the course of our data collection and analysis period, other groups looked at bile acids, FGF-19 and FGF-21 levels in PCOS patients with varying recruitment criteria and conditions.^{28,33,34} FGF-19 levels have been published as decreased or unchanged in patients with PCOS.²⁷ As our recruitment population was from an underserved rural area with high levels of diabetes and obesity, it is possible that distribution patterns of bile acids are affected heterogeneously depending on environmental queues.

Strengths of this study include stratification of PCOS subjects and controls by BMI, the inclusion of younger women, and the inclusion of both hormonal studies and potential biomarkers. Weaknesses include small sample size; however, this did allow for control of other variables such as the influence of medications, which could augment molecular pathways. We also examined bile acids in coordination with other metabolic changes. Therefore, showing correlation of bile acids with known diagnostic markers of PCOS further strengthened our study. PCOS is a heterogeneous disorder that often takes years to diagnose.

The goal of this study was to better define other signaling markers that could in fact be used to more efficiently diagnose PCOS. According to our data, there were not significant changes in bile acid total pool with only changes in two individual species, yet FGF-21 levels were consistently elevated among PCOS patients independent of obesity in coordination with TCA levels. This could be further explored as a dual marker for diagnosis. Future research will explore the use of these specific bile acid species and FGF-21 as early diagnostic markers of PCOS in adolescent women.

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Disclaimer

The content is solely the responsibility of the authors and does not necessarily represent the official views of the National Institutes of Health.

Author Disclosure Statement

No competing financial interests exist.

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References

- Legro RS, Arslanian SA, Ehrmann DA, et al. Diagnosis and treatment of polycystic ovary syndrome: An Endocrine Society clinical practice guideline. J Clin Endocrinol Metab 2013;98(12):4565–4592; doi: 10.1210/jc.2013-2350
- 2. Azziz R, Carmina E, Dewailly D, et al. The Androgen Excess and PCOS Society criteria for the polycystic ovary syndrome: The complete task force report. Fertil Steril 2009;91(2):456–488.
- Johnson T, Kaplan L, Ouyang P, et al. National Institutes of Health Evidence-Based Methodology Workshop on Polycystic Ovary Syndrome (PCOS). National Institutes of Health; 2012:1; pp. 1–14.
- O'Brien RF, Emans SJ. Polycystic ovary syndrome in adolescents. J Pediatr Adolesc Gynecol 2008;21(3):119–128; doi: 10.1016/j.jpag.2007.07.007
- Duleba AJ, Dokras A. Is PCOS an inflammatory process? Fertil Steril 2012; 97(1):7–12; doi:10.1016/j.fertnstert.2011.11.023
- Gonzalez F. Inflammation in polycystic ovary syndrome: Underpinning of insulin resistance and ovarian dysfunction. Steroids 2012;77(4):300–305; doi: 10.1016/j.steroids.2011.12.003
- Deligeoroglou E, Vrachnis N, Athanasopoulos N, et al. Mediators of chronic inflammation in polycystic ovarian syndrome. Gynecol Endocrinol 2012;28(12):974–978; doi: 10.3109/09513590.2012.683082
- Escobar-Morreale HF, Luque-Ramirez M, Gonzalez F. Circulating inflammatory markers in polycystic ovary syndrome: A systematic review and metaanalysis. Fertil Steril 2011;95(3):1048–1058 e1041–e1042.
- 9. Toulis KA, Goulis DG, Farmakiotis D, et al. Adiponectin levels in women with polycystic ovary syndrome: A systematic review and a meta-analysis. Hum Reprod Update 2009;15(3):297–307; doi: 10.1093/humupd/dmp006
- Bulletins—Gynecology ACoP. ACOG Practice Bulletin No. 108: Polycystic ovary syndrome. Obstet Gynecol 2009;114(4):936–949; doi: 10.1097/AOG .0b013e3181bd12cb
- 11. McCartney CR, Prendergast KA, Chhabra S, et al. The association of obesity and hyperandrogenemia during the pubertal transition in girls: Obesity as a potential factor in the genesis of postpubertal hyperandrogenism. J Clin Endocrinol Metab 2006;91(5):1714–1722; doi: 10.1210/jc.2005-1852
- Yildirim B, Sabir N, Kaleli B. Relation of intra-abdominal fat distribution to metabolic disorders in nonobese patients with polycystic ovary syndrome. Fertil Steril 2003;79(6):1358–1364; doi: 10.1016/s0015-0282(03)00265-6
- Noel OF, Still CD, Argyropoulos G, et al. Bile acids, FXR, and metabolic effects of bariatric surgery. J Obes 2016;2016(3):4390254; doi: 10.13039/ 100000002

- 14. Albaugh VL, Banan B, Ajouz H, et al. Bile acids and bariatric surgery. Mol Aspects Med 2017;56(Suppl. 2):75–89; doi: 10.1016/j.mam.2017.04.001
- Browning MG, Pessoa BM, Khoraki J, et al. Changes in bile acid metabolism, transport, and signaling as central drivers for metabolic improvements after bariatric surgery. Curr Obes Rep 2019;8(2):175–184; doi: 10.1007/s13679-019-00334-4
- Penney NC, Kinross J, Newton RC, et al. The role of bile acids in reducing the metabolic complications of obesity after bariatric surgery: A systematic review. Int J Obes (Lond) 2015;39(11):1565–1574; doi: 10.1038/ ijo.2015.115
- Nemati R, Lu J, Dokpuang D, et al. Increased bile acids and FGF19 after sleeve gastrectomy and Roux-en-Y Gastric Bypass correlate with improvement in type 2 diabetes in a randomized trial. Obes Surg 2018;28(9): 2672–2686; doi: 10.13039/100012746
- Flynn CR, Albaugh VL, Abumrad NN. Metabolic effects of bile acids: Potential role in bariatric surgery. Cell Mol Gastroenterol Hepatol 2019; 8(2):235–246; doi: 10.13039/100000062
- Xu G, Song M. Recent advances in the mechanisms underlying the beneficial effects of bariatric and metabolic surgery. Surg Obes Relat Dis 2021;17(1):231–238; doi: 10.13039/100000057
- Buchwald H, Avidor Y, Braunwald E, et al. Bariatric surgery: A systematic review and meta-analysis. JAMA 2004;292(14):1724–1737; doi: 10.1001/ jama.292.14.1724
- Ashrafian H, le Roux CW, Darzi A, et al. Effects of bariatric surgery on cardiovascular function. Circulation 2008;118(20):2091–2102; doi: 10.1161/circulationaha.107.721027
- 22. Griffiths WJ, Sjovall J. Bile acids: Analysis in biological fluids and tissues. J Lipid Res 2010;51(1):23-41; doi: 10.1194/jlr.r001941-jlr200
- Dolegowska K, Marchelek-Mysliwiec M, Nowosiad-Magda M, et al. FGF19 subfamily members: FGF19 and FGF21. J Physiol Biochem 2019;75(2): 229–240; doi: 10.13039/501100008781
- Degirolamo C, Sabba C, Moschetta A. Therapeutic potential of the endocrine fibroblast growth factors FGF19, FGF21 and FGF23. Nat Rev Drug Discov 2016;15(1):51–69; doi: 10.1038/nrd.2015.9
- Lan T, Morgan DA, Rahmouni K, et al. FGF19, FGF21, and an FGFR1/beta-Klotho-Activating Antibody Act on the nervous system to regulate body weight and glycemia. Cell Metab 2017;26(5):709–718.e3; doi: 10.1016/ j.cmet.2017.09.005
- 26. Wang D, Zhu W, Li J, et al. Serum concentrations of fibroblast growth factors 19 and 21 in women with gestational diabetes mellitus: Association with insulin resistance, adiponectin, and polycystic ovary syndrome history. PLoS One 2013;8(11):e81190; doi: 10.1371/journal.pone.0081190
- 27. Ramanjaneya M, Bensila M, Bettahi I, et al. Dynamic changes in circulating endocrine FGF19 subfamily and fetuin-A in response to intralipid and insulin infusions in healthy and PCOS women. Front Endocrinol (Lausanne) 2020;11:568500; doi: 10.3389/fendo.2020.568500
- Cheng F, Ng NYH, Tam CHT, et al. Association between FGF19, FGF21 and lipocalin-2, and diabetes progression in PCOS. Endocr Connect 2021; 10(10):1243–1252; doi: 10.1530/ec-21-0082
- Cyphert HA, Ge X, Kohan AB, et al. Activation of the farnesoid X receptor induces hepatic expression and secretion of fibroblast growth factor 21. J Biol Chem 2012;287(30):25123–25138; doi: 10.1074/jbc.m112.375907
- Gorar S, Culha C, Uc ZA, et al. Serum fibroblast growth factor 21 levels in polycystic ovary syndrome. Gynecol Endocrinol 2010;26(11):819–826; doi: 10.3109/09513590.2010.487587
- Olszanecka-Glinianowicz M, Madej P, Wdowczyk M, et al. Circulating FGF21 levels are related to nutritional status and metabolic but not hormonal disturbances in polycystic ovary syndrome. Eur J Endocrinol 2015;172(2):173–179; doi: 10.1530/eje-14-0539
- Jimenez V, Jambrina C, Casana E, et al. FGF21 gene therapy as treatment for obesity and insulin resistance. EMBO Mol Med 2018;10(8):e8791; doi: 10.15252/emmm.201708791
- Zhang B, Shen S, Gu T, et al. Increased circulating conjugated primary bile acids are associated with hyperandrogenism in women with polycystic ovary syndrome. J Steroid Biochem Mol Biol 2019;189(Suppl. 1):171–175; doi: 10.13039/501100012166
- 34. Yang X, Wu R, Qi D, et al. Profile of bile acid metabolomics in the follicular fluid of PCOS patients. Metabolites 2021;11(12):845; doi: 10.3390/ metabo11120845

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Abbreviations Used

 α MCA = alpha-muricholic acid β MCA = beta-muricholic acid ω MCA = omega-muricholic acid BMI = body mass index $\mathsf{CA}=\mathsf{cholic}\;\mathsf{acid}$ $\mathsf{CDCA} = \mathsf{chenodeoxycholic} \; \mathsf{acid}$ $\mathsf{DCA} = \mathsf{deoxycholic} \ \mathsf{acid}$ FGF-19 = fibroblast growth factor 19

- FGF-21 = fibroblast growth factor 21GCA = glycocholic acid GCDCA = glycochenodeoxycholic acid GDCA = glycodeoxycholic acid GLCA = glycolithocholic acid GUDCA = glycoursodeoxycholic acid HCA = hyocholic acid PCOS = polycystic ovary syndrome
 - SD = standard deviationTCA = taurocholic acid
- $\mathsf{TCDCA} = \mathsf{taurochenodeoxycholic} \mathsf{acid}$
- TDCA = taurodeoxycholic acid THCA = 3α , 7α , 12α -trihydroxycholestanoic acid
- $\mathsf{TLCA} = \mathsf{taurolithocholic} \; \mathsf{acid}$
- TMCA = tauromuricholic acid
- $\mathsf{TUDCA} = \mathsf{tauroursodeoxycholic} \ \mathsf{acid}$
- UDCA = ursodeoxycholic acid

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