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CD73 represses pro-inflammatory responses in human endothelial cells

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Abstract

Background: CD73 is a 5'-ectonucleotidase that produces extracellular adenosine, which then acts on G protein-coupled purigenic receptors to induce cellular responses. CD73 has been reported to regulate expression of pro-inflammatory molecules in mouse endothelium. Our aim is to determine the function of CD73 in human endothelial cells.

Methods: We used RNAi to deplete CD73 levels in human umbilical cord endothelial cells (HUVECs).

Results: CD73 depletion resulted in a strong reduction in adenosine production, indicating that CD73 is the major source of extracellular adenosine in HUVECs. We find that CD73 depletion induces a similar response to pro-inflammatory stimuli such as the cytokine TNF- α . In CD73-depleted cells, surface levels of the leukocyte adhesion molecules ICAM-1, VCAM-1 and E-selectin increase. This correlates with increased translocation of the transcription factor NF- κ B to the nucleus, which is known to regulate ICAM-1, VCAM-1 and E-selectin expression in response to TNF- α . Adhesion of monocytic cells to endothelial cells is enhanced. In addition, CD73-depleted cells become elongated, have higher levels of stress fibres and increased endothelial permeability, resembling known responses to TNF- α .

Conclusions: These results indicate that CD73 normally suppresses pro-inflammatory responses in human endothelial cells.

Background

CD73 is a 5'-ectonucleotidase that uses extracellular AMP to produce adenosine, and is a GPI-anchored protein that is expressed abundantly on endothelial cells and on a subset of leukocytes [1,2]. CD73^{-/-} mice are viable but have multiple cardiovascular phenotypes [3], including cardioprotection during myocardial ischemia [4], vasoprotection [3,5], increased neointimal plaque formation and increased monocyte adhesion due to upregulation of VCAM-1 on the endothelium [6]. In the cremaster model of ischaemia-reperfusion, leukocyte attachment to the endothelium is significantly increased in CD73^{-/-} mice [3]. Additionally, CD73^{-/-} mice have increased vascular leakage in response to hypoxia [5], lipopolysaccharide (LPS) [7] and cardiac transplantation [8]. Whether these phenotypes are a consequence of reduced adenosine production by endothelial or other cell types is not known, although inhibition of CD73

enzymatic function induces a similar accumulation of neutrophils in lungs following LPS treatment to lack of CD73 [7].

Adenosine generally has anti-inflammatory properties and exerts its effects via G-protein-coupled P1 purinergic receptors [2], although in some cell types purinergic receptors play a pro-inflammatory role [9]. A_{2A} and A_{2B} purinergic receptors activate adenylate cyclase, thereby increasing intracellular cAMP levels, while A₁ and A₃ receptors inhibit cAMP production [10]. In endothelial cells, stimulation of A_{2B} receptors increases endothelial barrier function by decreasing actomyosin contractility and strengthening the intercellular junctions [11,12], and A_{2B}-null mice have increased vascular permeability in response to hypoxia and increased pulmonary leakage after lung injury [13,14]. Adenosine has also been shown to inhibit neutrophil adhesion to the endothelium and transendothelial migration via neutrophil A₂ receptors [15,16], and an inhibitor of CD73-mediated adenosine production was found to enhance migration of lymphocytes across brain microvascular endothelial

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cells [17]. CD73 is therefore proposed to provide an anti-inflammatory signal via adenosine production, leading to increased endothelial barrier function and decreased leukocyte binding.

In addition to increasing endothelial barrier function, adenosine inhibits NF- κ B-mediated upregulation of leukocyte adhesion molecules on endothelial cells including P-selectin, E-selectin and VCAM-1 [18-21]. The regulation of ICAM-1 by adenosine is unclear; while Bouma et al. did not see an adenosine-mediated decrease in ICAM-1 levels [22], others have demonstrated inhibition of ICAM-1 expression in response to adenosine analogues or A_{2A} receptor agonists [18,21].

Although adenosine has multiple effects in protecting human endothelial cells from pro-inflammatory stimuli and CD73 produces adenosine, whether endogenous CD73 contributes to endothelial cell function in the absence of pro-inflammatory stimuli is not clear. In order to investigate how CD73 affects the properties of human endothelial cells, we have used RNAi to reduce CD73 expression. We show that CD73 depletion induces a phenotype similar to that of the pro-inflammatory cytokine TNF- α , including upregulation of leukocyte adhesion molecules, changes to cell shape and the actin cytoskeleton, and increased endothelial permeability.

Methods

Reagents

Human fibronectin, adenosine 5'-monophosphate, TRITC-phalloidin and FITC-dextran (Mr 42 000) were obtained from Sigma-Aldrich; Oligofectamine reagent, AlexaFluor594-labelled goat anti-rabbit and AlexaFluor488-labelled goat anti-mouse antibodies were obtained from Invitrogen; mouse anti-CD73 antibody (4G4) was a gift from Sirpa Jalkanen (Turku, Finland); mouse anti-ICAM-1 antibody (BBIG-I1) was from R&D Systems; mouse anti-VCAM-1 antibody (51-10C9) and mouse anti- β -catenin (AC15) were from BD Pharmingen; mouse anti-E-selectin (CTB202) and rabbit anti-NF- κ B (p65) antibody (C-20) were from Santa Cruz Biotechnology; [³H] adenosine 5'-monophosphate was obtained from GE Healthcare.

Cell Culture

Pooled human umbilical vein endothelial cells (HUVECs) were obtained from Lonza and cultured in flasks pre-coated with 10 μ g/ml human fibronectin in EBM-2 medium with growth factors (Lonza) in an atmosphere of 5% CO₂ and 95% air. The human monocytic cell line THP-1 (ATCC) was cultured in RPMI-1640 medium (Invitrogen) supplemented with 2 mM L-glutamine, 10% heat-inactivated fetal calf serum (FCS), penicillin (100 U/ml) and streptomycin (100 μ g/ml) in an atmosphere of 5% CO₂ and 95% air.

siRNA Transfection

HUVECs were plated on 6-well dishes at 1.5×10^5 cells per well, 24 h prior to transfection. siRNAs (1.25 μ l of 20 μ M stock) were premixed with 4 μ l of Oligofectamine reagent (Invitrogen). The three siRNAs oligonucleotides si1, si2 and si3 targeting human NT5E (CD73) were siGENOME duplexes D-008217-01 (GAACCGUGCUGCUGUAUUGUU), D-008217-02 (GGAAGUCA CUGCCAUGGAAUU) and D-008217-04 (GGACUUU AUUUGCCAUAUAUU) (Dharmacon). The non-targeting control siRNA (siC) was ON-TARGETplus D-001810-01 (UGGUUUACAUGUCGACUAA). Cells were transfected for 4 h at 37°C in 1 ml EBM-2 medium with growth supplements but no antibiotics or FCS. EBM-2 medium (0.5 ml) with growth factors and 6% FCS was then added to each well and cells were incubated overnight. Cells were trypsinized 48 h after transfection and plated on fibronectin-coated 6-well plates (4×10^5 cells per well; flow cytometry or phase-contrast images), 24-well plates (2×10^5 cells per well; thin layer chromatography), coverslips (2×10^5 cells per coverslip; immunofluorescence), black 96-well plates with glass bottom (5×10^4 cells per well; adhesion assay) or Transwells (2×10^5 cells per Transwell; permeability assay). Where indicated, cells were stimulated with 10 ng/ml TNF- α for 15 h. Cells were analyzed 72 h after transfection.

Flow Cytometry

Flow cytometry (FC) was used to detect levels of cell surface receptors in HUVECs. Cells were detached with trypsin/EDTA and washed once with FC flow buffer (0.2% BSA, 0.1% N₃Na in PBS). Cells were then sequentially incubated with 2% BSA in FC buffer (30 min, 4°C), primary antibody (30 min, 4°C) and AlexaFluor488-conjugated goat anti-mouse antibody (20 min, 4°C). To remove the antibodies, cells were washed twice with FC buffer. Samples were measured using a BD FACSCalibur flow cytometer (Becton Dickinson) at 488 nm excitation wavelength and using a 530 nm emission bandpass filter.

CD73 Activity Assay

HUVECs were washed once before adding EGM-2, containing 180 μ M [³H] adenosine 5'-monophosphate (specific activity per well: 37 μ Bq) and 200 μ M unlabelled adenosine 5'-monophosphate (10 min, 37°C). Aliquots of the medium were applied to silica gel 60 ADAMANT™ thin layer chromatography (TLC) plates (Sigma-Aldrich) and were separated using isobutyl alcohol:isoamyl alcohol:2-ethoxyethanol:ammonia:H₂O (ratio 9:6:18:9:15) as a solvent. The TLC plates were developed by exposing to tritium-sensitive film (Kodak BioMax MS film) together with a BioMax TranscreenLE intensifying screen (Kodak). TLC spots were quantified by

densitometry and relative CD73 activity was calculated as ^3H -adenosine/ ^3H -AMP.

Immunofluorescence and Phase-contrast Microscopy

HUVECs were washed once with PBS and fixed with 4% paraformaldehyde in PBS (20 min, room temperature) and for NF- κ B localisation additionally with 100% ice-cold acetone (5 min, -20°C). After fixation cells were permeabilised with 0.1% Triton X-100 in PBS (5 min, 4°C) and blocked with 2% BSA in PBS (30 min, 22°C). Coverslips were then sequentially incubated with antibodies against NF- κ B (p65) and β -catenin, AlexaFluor488 goat anti-mouse and AlexaFluor594 goat anti-rabbit antibodies and/or with TRITC-phalloidin to visualise F-actin (45 min, 22°C). Coverslips were mounted onto slides using fluorescent mounting medium, and visualised using a LSM 510 laser scanning confocal microscope (Zeiss). Phase-contrast images of siRNA-treated HUVECs in 6-well dishes were generated on a Nikon Eclipse TE2000-E microscope with a Hamamatsu Orca-ER digital camera using Metamorph software.

Cell Adhesion Assay

THP-1 cells were stained with CellTracker Green CMFDA ($1\ \mu\text{M}$, 30 min, 37°C), washed once with PBS and 5×10^6 THP-1 cells were added for 15 min to black 96-well dishes with clear bottom (Corning) containing siRNA-treated HUVECs. The wells were washed twice with PBS and the remaining fluorescence measured in a Fusion α -FP plate reader (Perkin Elmer) at 485 nm excitation wavelength and using a 525/35 nm emission bandpass filter.

Permeability Assay

siRNA-treated HUVECs were cultured to confluency on Transwell filters (Corning; 12 mm diameter, $0.4\ \mu\text{m}$ pore size), cells were washed once with medium and $100\ \mu\text{g}/\text{ml}$ FITC-dextran was applied to the upper chamber. Samples of the medium from the lower chamber were subsequently removed after 80 min and measured in black clear-bottom 96-well plates using a Fusion α -FP plate reader (Perkin Elmer) at 485 nm excitation wavelength and using a 525/35 nm emission bandpass filter.

Statistical Analysis

In order to determine statistical significance, Student's *t*-test with Bonferroni post-test was carried out using GraphPad Prism software <http://www.graphpad.com>.

Results

CD73 is the main source of adenosine production by HUVECs

To investigate the role of CD73 in human endothelial cells, HUVECs were transfected with three different

siRNAs to CD73 (si1, si2 and si3), all of which reduced surface levels of CD73 by at least 70%, whereas a control non-targeting siRNA (siControl; siC) did not affect CD73 levels (Figure 1A). Adenosine is the product of CD73 enzymatic activity. It was constitutively produced by HUVECs, and this was markedly reduced in CD73 knockdown cells (Figure 1B), indicating that CD73 is the major source of extracellular adenosine in these cells.

CD73 regulates adhesion molecule expression in endothelial cells

Pro-inflammatory cytokines up-regulate the expression of the leukocyte adhesion molecules ICAM-1, V-CAM-1 and E-selectin in endothelial cells [19]. To investigate whether CD73 regulates cell surface levels of these adhesion molecules, we tested the effects of CD73 depletion. Unstimulated HUVECs expressed low levels of ICAM-1 on the cell surface, whereas VCAM-1 and E-selectin levels were not above background (data not shown). CD73 depletion induced an increase in ICAM-1, VCAM-1 and E-selectin levels, whereas siControl had no effect (Figure 1C-E). Taken together, these results are consistent with a role of constitutive adenosine production by CD73 in suppressing expression of leukocyte adhesion molecules in endothelial cells.

TNF- α induces ICAM-1, VCAM-1 and E-selectin expression in part through activation of the transcription factor NF- κ B [19]. NF- κ B activity was reported to be increased in endothelial cells derived from CD73 $^{-/-}$ mice, and thus could contribute to upregulation of VCAM-1 levels [6]. To test if NF- κ B activity was increased in HUVECs depleted of CD73, cells were stained with antibodies to NF- κ B. NF- κ B translocates to the nucleus when it is activated [23], and TNF- α , which is well known to stimulate NF- κ B activity, stimulated NF- κ B nuclear translocation in over 60% of HUVECs (Figure 2). CD73 depletion also increased the proportion of cells with nuclear NF- κ B staining (Figure 2). These results suggest that CD73 knockdown induces a pro-inflammatory phenotype in HUVECs, which could be mediated in part by NF- κ B activation.

CD73 depletion induces morphological changes in HUVECs

Since CD73 knockdown induced upregulation of adhesion molecules similar to TNF- α , we tested whether CD73 affected endothelial morphology. We have previously shown that TNF- α induces cell elongation and actin stress fibre formation in HUVECs [24]. CD73 knockdown induced an elongated morphology similar to morphological changes occurring after TNF- α treatment (Figure 3). CD73 depletion also increased stress fibres, although to a lesser extent than $10\ \text{ng}/\text{ml}$ TNF- α (Figure 3). These results

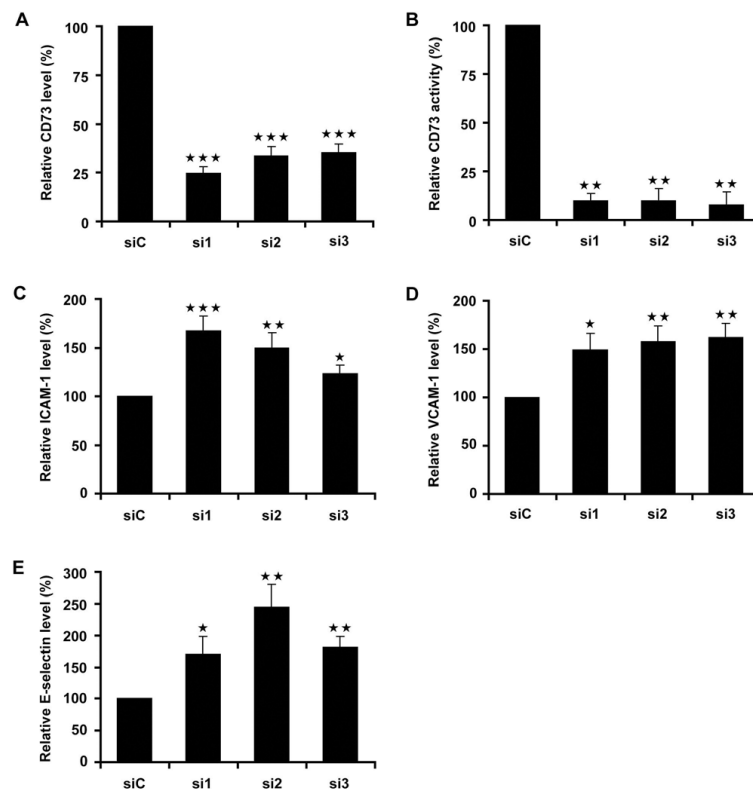


Figure 1 CD73 regulates ICAM-1, VCAM-1 and E-selectin expression. HUVECs were transfected with CD73 siRNAs or control oligonucleotide (siC). **A**, Cell surface expression levels of CD73. **B**, CD73 activity. **C-E**, ICAM-1, VCAM-1 and E-selectin, shown as mean fluorescence of the population. Results were normalised to siC. *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$ determined by Student's t-test and Bonferroni post-test, compared to siC.

further strengthen the hypothesis that CD73 depletion induces a pro-inflammatory phenotype.

CD73 regulates leukocyte adhesion

The increase in adhesion molecule expression in CD73-depleted endothelial cells suggests that leukocyte adhesion could be affected. To study this we incubated THP-1 monocytic leukaemia cells with HUVECs. Adhesion of THP-1 cells to HUVECs was significantly increased by CD73 knockdown (Figure 4A). In contrast, CD73 depletion did not affect THP-1 adhesion to TNF- α -treated HUVECs, reflecting the 4 to 6 fold increase in the levels of ICAM-1, VCAM-1 and E-selectin expression induced by TNF- α alone (data not shown).

Endothelial permeability is increased in CD73-depleted cells

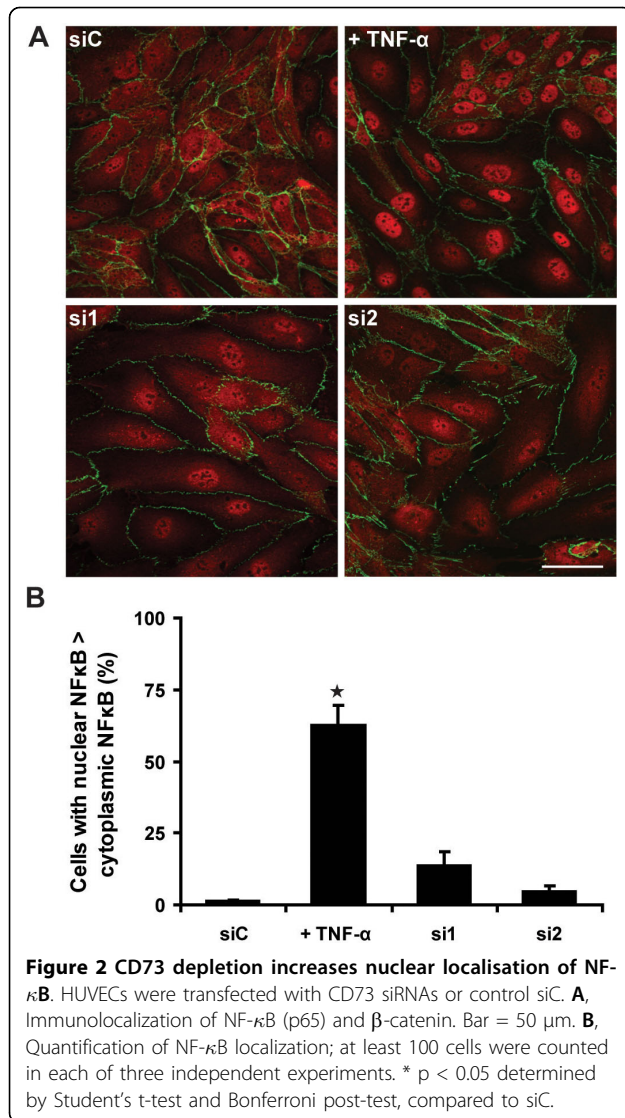
TNF- α is known to increase endothelial permeability in HUVECs [24,25], whereas adenosine, the product of CD73 enzymatic activity, has been shown to reduce permeability [11,12,26]. The decrease in extracellular adenosine production due to CD73 knockdown (Figure 1C) would therefore be predicted to lead to an increase in

permeability. In agreement with this, the permeability of HUVEC monolayers was higher following CD73 depletion than in control cells (Figure 4B). The 1.5 to 2-fold increase in permeability following CD73 knockdown was in the same range to that induced by 10 ng/ml TNF- α (2 to 2.5 fold; data not shown and [24])

Discussion

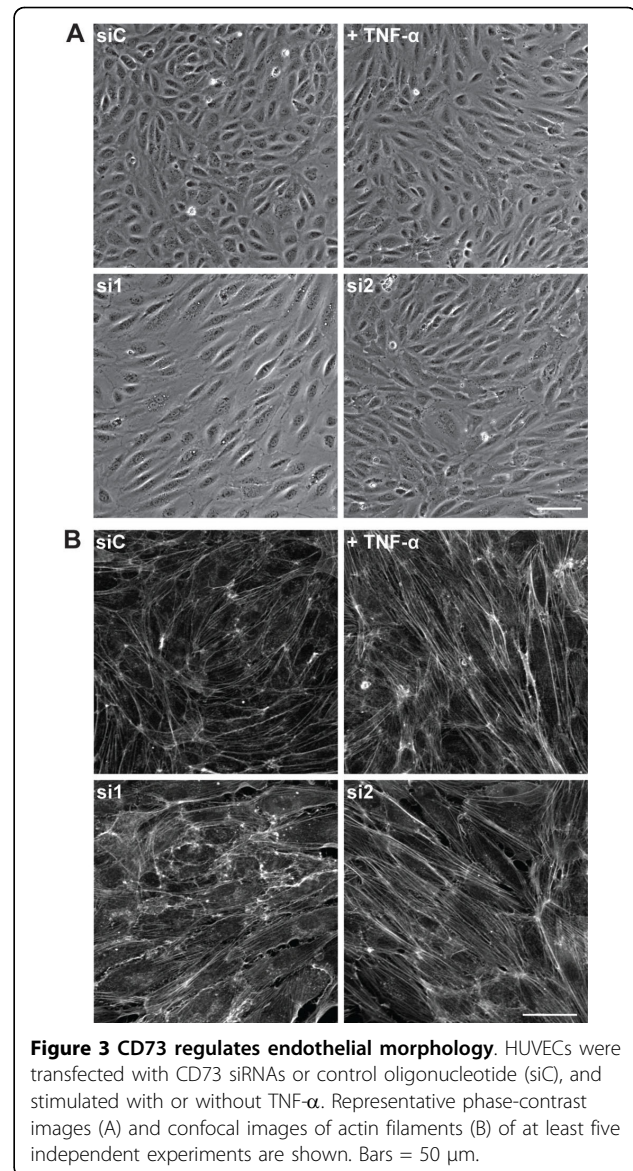
The endothelium of CD73^{-/-} mice has been shown to have increased VCAM-1 levels, but the effect of CD73 depletion on human endothelial cells has not been described. We show here that CD73 normally functions to suppress multiple different aspects of a pro-inflammatory phenotype of endothelial cells, including expression of ICAM-1, VCAM-1 and E-selectin, translocation of the transcription factor NF- κ B to the nucleus, endothelial cell morphology, actin cytoskeletal organisation and permeability. CD73-depleted cells exhibited a similar phenotype to treatment with TNF- α .

Consistent with the lower levels of leukocyte adhesion molecules and leukocyte adhesion we observe in CD73-depleted endothelial cells, leukocyte infiltration in inflammatory situations is reduced in CD73^{-/-} mice [7,27,28].



Endothelial CD73 is important for these responses [28], although lymphocyte CD73 also contributes to reducing cardiac graft rejection [8]. In lymphocytes it has been suggested that CD73 has non-enzymatic functions in modulating the clustering of the integrin LFA-1 or in inhibiting apoptosis, but so far no such role of CD73 has been described in endothelial cells [1,29]. However, an A_{2B} adenosine receptor agonist rescues the defect in lymphocyte recruitment to lymph nodes in $CD73^{-/-}$ mice [28], indicating that in this case the phenotype is probably due to decreased levels of adenosine.

It is likely that the signalling pathway whereby CD73 and adenosine suppress leukocyte adhesion molecule expression differs from that regulating morphology and endothelial permeability. The regulation of endothelial permeability and stress fibre levels by adenosine is attributed to an increase in cAMP, which in turn



induces both inhibition of RhoA, and hence decreases actomyosin contractility and stress fibre formation, and activation of Rap1, thereby strengthening adherens junction integrity [30]. Although the mechanistic basis for adenosine-mediated inhibition of leukocyte adhesion molecule expression is less clear, it is possible that it also involves cAMP production, since increased cAMP inhibits TNF- α - and thrombin-induced transcription of NF- κ B-regulated genes, including ICAM-1 and VCAM-1 [31,32], an effect that could be mediated through cAMP-induced repression of p38 MAPK activity [31].

It is not clear whether the pro-inflammatory phenotypic changes we observe in response to CD73 depletion represent the constitutive activity of an intrinsic signalling pathway in endothelial cells that is suppressed by

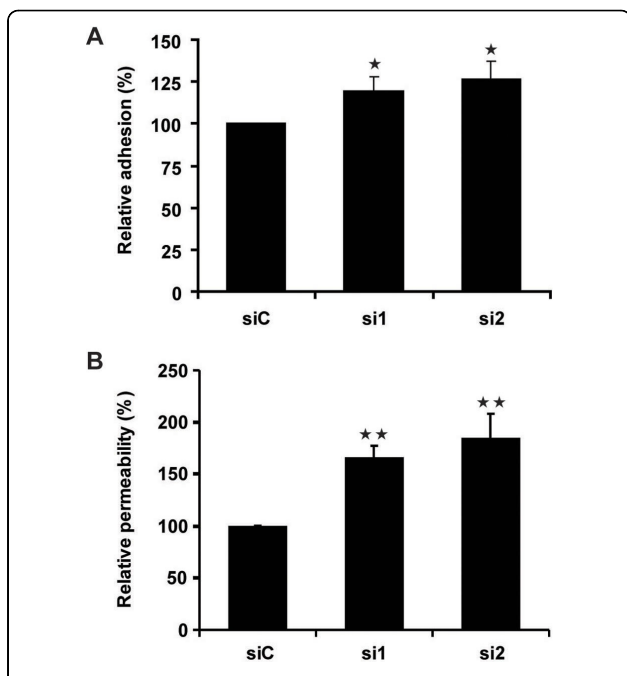


Figure 4 CD73 depletion increases monocyte adhesion to endothelial cells and endothelial permeability. HUVECs were transfected with CD73 siRNAs or control siC. A, Adhesion of THP-1 cells to HUVECs was measured after 15 min. B, Monolayer permeability was determined on Transwell filters. Results were normalised to the respective control (siC). **p < 0.01, *p < 0.05, determined by Student's t-test and Bonferroni post-test, as compared to siC.

CD73 and adenosine or are mediated by an external stimulus. It is possible that HUVECs themselves produce some TNF- α or other pro-inflammatory cytokines, although TNF- α production by endothelial cells is normally only induced by inflammatory stimuli such as LPS or interleukin 1 β [33,34]. In the future it would be interesting to determine whether the anti-inflammatory effects of CD73 are mediated by alterations in the constitutive activity of GTPases such as RhoA or Rap1. It will also be important to investigate whether the effects of reduced CD73 expression we report with human endothelial cells in vitro correlate with in vivo observations on human endothelium.

Conclusions

CD73 depletion in HUVECs induces a pro-inflammatory phenotype similar to low levels of TNF- α , including increased expression of leukocyte adhesion molecules and changes in endothelial morphology. Since we found that HUVECs normally produce extracellular adenosine and that this is predominantly due to CD73, it is likely that reduced levels of adenosine are responsible for the phenotypes we observe upon CD73 knockdown.

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Authors' contributions

JKGG and AJR designed the study. JGG carried out all experimental work and prepared the figures. JKGG and AJR wrote the manuscript. Both authors have read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Jalkanen S, Salmi M: VAP-1 and CD73, endothelial cell surface enzymes in leukocyte extravasation. *Arterioscler Thromb Vasc Biol* 2008, **28**:18-26.
2. Yegutkin GG: Nucleotide- and nucleoside-converting ectoenzymes: Important modulators of purinergic signalling cascade. *Biochim Biophys Acta* 2008, **1783**:673-694.
3. Koszalka P, Ozuyaman B, Huo Y, Zerneck A, Flogel U, Braun N, Buchheiser A, Decking UK, Smith ML, Sevigny J, Gear A, Weber AA, Molojavji A, Ding Z, Weber C, Ley K, Zimmermann H, Godecke A, Schrader J: Targeted disruption of cd73/ecto-5'-nucleotidase alters thromboregulation and augments vascular inflammatory response. *Circ Res* 2004, **95**:814-821.
4. Eckle T, Krahn T, Grenz A, Kohler D, Mittelbronn M, Ledent C, Jacobson MA, Osswald H, Thompson LF, Unertl K, Eltzschig HK: Cardioprotection by ecto-5'-nucleotidase (CD73) and A2B adenosine receptors. *Circulation* 2007, **115**:1581-1590.
5. Thompson LF, Eltzschig HK, Ibla JC, Wiele Van De CJ, Resta R, Morote-Garcia JC, Colgan SP: Crucial role for ecto-5'-nucleotidase (CD73) in vascular leakage during hypoxia. *J Exp Med* 2004, **200**:1395-1405.
6. Zerneck A, Bidzhekov K, Ozuyaman B, Fraemohs L, Liehn EA, Luscher-Firzlaff JM, Luscher B, Schrader J, Weber C: CD73/ecto-5'-nucleotidase protects against vascular inflammation and neointima formation. *Circulation* 2006, **113**:2120-2127.
7. Reutershan J, Vollmer I, Stark S, Wagner R, Ngamsri KC, Eltzschig HK: Adenosine and inflammation: CD39 and CD73 are critical mediators in LPS-induced PMN trafficking into the lungs. *FASEB J* 2009, **23**:473-82.
8. Hasegawa T, Bouis D, Liao H, Visovatti SH, Pinsky DJ: Ecto-5' nucleotidase (CD73)-mediated adenosine generation and signaling in murine cardiac allograft vasculopathy. *Circ Res* 2008, **103**:1410-1421.
9. Ham J, Rees DA: The adenosine a2b receptor: its role in inflammation. *Endocr Metab Immune Disord Drug Targets* 2008, **8**:244-254.
10. Shryock JC, Belardinelli L: Adenosine and adenosine receptors in the cardiovascular system: biochemistry, physiology, and pharmacology. *Am J Cardiol* 1997, **79**:2-10.
11. Comerford KM, Lawrence DW, Synnestevedt K, Levi BP, Colgan SP: Role of vasodilator-stimulated phosphoprotein in PKA-induced changes in endothelial junctional permeability. *FASEB J* 2002, **16**:583-585.
12. Srinivas SP, Satpathy M, Gallagher P, Lariviere E, Van Driessche W: Adenosine induces dephosphorylation of myosin II regulatory light chain in cultured bovine corneal endothelial cells. *Exp Eye Res* 2004, **79**:543-551.
13. Eckle T, Faigle M, Grenz A, Laucher S, Thompson LF, Eltzschig HK: A2B adenosine receptor dampens hypoxia-induced vascular leak. *Blood* 2008, **111**:2024-2035.
14. Eckle T, Grenz A, Laucher S, Eltzschig HK: A2B adenosine receptor signaling attenuates acute lung injury by enhancing alveolar fluid clearance in mice. *J Clin Invest* 2008, **118**:3301-3315.
15. Cronstein BN: Adenosine, an endogenous anti-inflammatory agent. *J Appl Physiol* 1994, **76**:5-13.
16. Wakai A, Wang JH, Winter DC, Street JT, O'Sullivan RG, Redmond HP: Adenosine inhibits neutrophil vascular endothelial growth factor release and transendothelial migration via A2B receptor activation. *Shock* 2001, **15**:297-301.

17. Niemela J, Ifergan I, Yegutkin GG, Jalkanen S, Prat A, Airas L: **IFN-beta regulates CD73 and adenosine expression at the blood-brain barrier.** *Eur J Immunol* 2008, **38**:2718-2726.
18. McPherson JA, Barringhaus KG, Bishop GG, Sanders JM, Rieger JM, Hesselbacher SE, Gimple LW, Powers ER, Macdonald T, Sullivan G, Linden J, Sarembock IJ: **Adenosine A(2A) receptor stimulation reduces inflammation and neointimal growth in a murine carotid ligation model.** *Arterioscler Thromb Vasc Biol* 2001, **21**:791-796.
19. De Martin R, Hoeth M, Hofer-Warbinek R, Schmid JA: **The transcription factor NF- κ B and the regulation of vascular cell function.** *Arterioscler Thromb Vasc Biol* 2000, **20**:E83-88.
20. Minguet S, Huber M, Rosenkranz L, Schamel WW, Reth M, Brummer T: **Adenosine and cAMP are potent inhibitors of the NF- κ B pathway downstream of immunoreceptors.** *Eur J Immunol* 2005, **35**:31-41.
21. Walker G, Langheinrich AC, Dennhauser E, Bohle RM, Dreyer T, Kreuzer J, Tillmanns H, Braun-Dullaeus RC, Haberbosch W: **3-deazaadenosine prevents adhesion molecule expression and atherosclerotic lesion formation in the aortas of C57BL/6J mice.** *Arterioscler Thromb Vasc Biol* 1999, **19**:2673-2679.
22. Bouma MG, Wildenberg van den FA, Buurman WA: **Adenosine inhibits cytokine release and expression of adhesion molecules by activated human endothelial cells.** *Am J Physiol* 1996, **270**:C522-529.
23. Karin M, Greten FR: **NF- κ B: linking inflammation and immunity to cancer development and progression.** *Nat Rev Immunol* 2005, **5**:749-759.
24. McKenzie JA, Ridley AJ: **Roles of Rho/ROCK and MLCK in TNF- α -induced changes in endothelial morphology and permeability.** *J Cell Physiol* 2007, **213**:221-228.
25. Wojciak-Stothard B, Entwistle A, Garg R, Ridley AJ: **Regulation of TNF- α -induced reorganization of the actin cytoskeleton and cell-cell junctions by Rho, Rac, and Cdc42 in human endothelial cells.** *J Cell Physiol* 1998, **176**:150-165.
26. Lennon PF, Taylor CT, Stahl GL, Colgan SP: **Neutrophil-derived 5'-adenosine monophosphate promotes endothelial barrier function via CD73-mediated conversion to adenosine and endothelial A2B receptor activation.** *J Exp Med* 1998, **188**:1433-1443.
27. Mills JH, Thompson LF, Mueller C, Waickman AT, Jalkanen S, Niemela J, Airas L, Bynoe MS: **CD73 is required for efficient entry of lymphocytes into the central nervous system during experimental autoimmune encephalomyelitis.** *Proc Natl Acad Sci USA* 2008, **105**:9325-9330.
28. Takedachi M, Qu D, Ebisuno Y, Oohara H, Joachims ML, McGee ST, Maeda E, McEver RP, Tanaka T, Miyasaka M, Murakami S, Krahn T, Blackburn MR, Thompson LF: **CD73-generated adenosine restricts lymphocyte migration into draining lymph nodes.** *J Immunol* 2008, **180**:6288-6296.
29. Mikhailov A, Sokolovskaya A, Yegutkin GG, Amdahl H, West A, Yagita H, Lahesmaa R, Thompson LF, Jalkanen S, Blokhin D, Eriksson JE: **CD73 participates in cellular multiresistance program and protects against TRAIL-induced apoptosis.** *J Immunol* 2008, **181**:464-475.
30. Vandenbroucke E, Mehta D, Minshall R, Malik AB: **Regulation of endothelial junctional permeability.** *Ann N Y Acad Sci* 2008, **1123**:134-145.
31. Rahman A, Anwar KN, Minhajuddin M, Bijli KM, Javaid K, True AL, Malik AB: **cAMP targeting of p38 MAP kinase inhibits thrombin-induced NF- κ B activation and ICAM-1 expression in endothelial cells.** *Am J Physiol Lung Cell Mol Physiol* 2004, **287**:L1017-1024.
32. Ollivier V, Parry GC, Cobb RR, de Prost D, Mackman N: **Elevated cyclic AMP inhibits NF- κ B-mediated transcription in human monocytic cells and endothelial cells.** *J Biol Chem* 1996, **271**:20828-20835.
33. Nilsen EM, Johansen FE, Jahnsen FL, Lundin KE, Scholz T, Brandtzaeg P, Haraldsen G: **Cytokine profiles of cultured microvascular endothelial cells from the human intestine.** *Gut* 1998, **42**:635-642.
34. Imaizumi T, Itaya H, Fujita K, Kudoh D, Kudoh S, Mori K, Fujimoto K, Matsumiya T, Yoshida H, Satoh K: **Expression of tumor necrosis factor- α in cultured human endothelial cells stimulated with lipopolysaccharide or interleukin-1 α .** *Arterioscler Thromb Vasc Biol* 2000, **20**:410-415.

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