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MK-0677, a Ghrelin Agonist, Alleviates Amyloid Beta-Related Pathology in 5XFAD Mice, an Animal Model of Alzheimer's Disease

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Abstract: Alzheimer's disease (AD) is a progressive neurodegenerative disorder characterized by cognitive deficits, neuroinflammation, and neuronal death. The primary pathogenic cause is believed to be the accumulation of pathogenic amyloid beta (A β) assemblies in the brain. Ghrelin, which is a peptide hormone predominantly secreted from the stomach, is an endogenous ligand for the growth hormone secretagogue-receptor type 1a (GHS-R1a). MK-0677 is a ghrelin agonist that potently stimulates the GHS-R1a ghrelin receptor. Interestingly, previous studies have shown that ghrelin improves cognitive impairments and attenuates neuronal death and neuroinflammation in several neurological disorders. However, it is unknown whether MK-0677 can affect Aβ accumulation or A β -mediated pathology in the brains of patients with AD. Therefore, we examined the effects of MK-0677 administration on AD-related pathology in 5XFAD mice, an Aβ-overexpressing transgenic mouse model of AD. MK-0677 was intraperitoneally administered to three-month-old 5XFAD mice. To visualize AB accumulation, neuroinflammation, and neurodegeneration, thioflavin-S staining and immunostaining with antibodies against A β (4G8), ionized calcium-binding adaptor molecule 1 (Iba-1), glial fibrillary acidic protein (GFAP), neuronal nuclear antigen (NeuN), and synaptophysin were conducted in the neocortex of 5XFAD and wild-type mice, and to evaluate changes of phosphorylated cyclic adenosine monophosphate (cAMP) response element binding protein (pCREB) levels, immunostaining with antibody against pCREB was performed in dentate gyrus of the hippocampus of 5XFAD and wild-type mice. The histological analyses indicated that MK-0677-treated 5XFAD mice showed reduced A β deposition, gliosis, and neuronal and synaptic loss in the deep cortical layers, and inhibited the decrement of pCREB levels in dentate gyrus of the hippocampus compared to vehicle-treated 5XFAD mice. Our results showed that activation of the ghrelin receptor with MK-0677 inhibited the AB burden, neuroinflammation, and neurodegeneration, which suggested that MK-0677 might have potential as a treatment of the early phase of AD.

Keywords: Alzheimer disease; ghrelin; MK-0677; amyloid beta; 5XFAD mice; Ibutamoren

1. Introduction

Alzheimer's disease (AD) is the most preeminent type of dementia. The number of Americans affected by AD is increasing exponentially and is expected to reach 13.8 million by 2050 from 5.4 million in 2016 [1,2]. AD results in various symptoms, such as memory impairments, language disturbances, and psychiatric problems [3]. The more AD progresses, the more daily functioning decreases and neuropsychiatric symptoms increase. According to the amyloid hypothesis, which is the most well-developed of the hypotheses of AD pathogenesis, deposits of the amyloid beta (A β) peptide are considered the major cause of the development of AD [4]. In addition, A β directly causes neurodegeneration, microgliosis, astrocytosis, neurofibrillary tangle deposition, and memory loss [4]. Therefore, targeting A β aggregation and A β -related pathologic changes has been suggested as a potential strategy for preventing AD pathogenesis.

Ghrelin, which is a 28-amino-acid peptide hormone, is mostly released from gastric enteroendocrine cells. The release of ghrelin can be caused by hunger [5]. In addition, des-acyl ghrelin, an inactive form of ghrelin, acts on the growth hormone secretagogue receptor type 1a (GHS-R1a) when it is transformed into acylated ghrelin by ghrelin o-acyltransferase (GOAT) [6]. Ghrelin is expressed in various cells and organs, such as the stomach, testis, ovary, kidney, and small intestine [7–14]. Several studies have suggested that ghrelin-containing neurons are extensively expressed in the central nervous system (CNS) [15–17]. In addition, accumulating evidence for the role of the ghrelin system in cognitive functions has indicated that they are strongly correlated [18,19]. One study reported that the mRNA levels of ghrelin, GOAT, and GHS-R1a were decreased in the temporal lobe of patients with AD [20]. a clinical study has revealed that male patients who were newly diagnosed with AD had a decreased area under the curve for ghrelin levels, while female patients did not show any alterations [21]. Therefore, the cognitive impairments in patients with AD have been suggested to be associated with the altered ghrelin system.

Ghrelin has various effects on physiological functions, such as appetite regulation [22], adiposity [23], glucose metabolism [24], and energy homeostasis in the hypothalamus [25–27]. Moreover, studies have revealed an association between ghrelin and neural function. Previous in vitro studies have demonstrated that ghrelin treatment enhances the synaptic density of dissociated cortical neurons with decreased synapses induced by hypoxia [28], attenuates oxygen/glucose deprivation-induced apoptosis in hypothalamic neurons [29], increases the proliferation of hippocampal neural stem cells via multiple signaling pathways [30], prevents apoptosis signal-regulating kinase 1-mediated apoptosis of rat pheochromocytoma (PC12) cells by increasing heat-shock protein 70 levels [31], and inhibits the increment of inflammatory cytokines induced by fibrillar A β in mice microglial cells [32]. Consistent with the in vitro studies, ghrelin has protective effects on the CNS in vivo. Ghrelin administration ameliorates the neuronal damage of hippocampal neurons in ischemia/reperfusion-injured rats [33] and rats with pilocarpine-induced seizures [34], and inhibits neuronal loss in the substantia nigra pars compacta (SNpc) in a 1-methyl-4-phenyl-1,2,5,6 tetrahydropyridine (MPTP)-induced mouse model of Parkinson's disease [35]. Moreover, ghrelin administration improves spatial learning and memory in animal models. The cognitive improvements induced by ghrelin are mediated by an increased spine synapse density in the CA1 of the hippocampus of rats [36] and the stimulation of adult hippocampal neurogenesis in the subgranular zone of the hippocampus of mice [37]. In accordance with the effects of ghrelin in AD, studies using animal AD models have suggested that ghrelin alleviates $A\beta$ -induced synaptic degeneration, microgliosis, impaired adult hippocampal neurogenesis, and cognitive deficits [38,39]. Taken together, these results surprisingly indicate that ghrelin has beneficial properties against neurotoxicity and pathologic changes that are similar to $A\beta$ -induced pathogenesis.

MK-0677, which is also known as ibutamoren mesylate or L-163,191, is a non-peptide ghrelin agonist with higher efficacy compared to ghrelin when it is bound to GHS-R1a [40–42]. The half-life of MK-0677 is 6 h, which is approximately 12 times longer than ghrelin's 30 min [43,44]. In addition, MK-0677 increases the levels of growth hormone and insulin-like growth factor-1 [45]. Therefore,

the properties of MK-0677 can be used to stimulate the ghrelin receptor, which could be effective for enhancing degenerated cognitive functions. Nevertheless, the effects of MK-0677 on AD pathology in animal models have not yet been reported.

To the finest of our knowledge, only a few studies have investigated a ghrelin agonist (LY444711) or ghrelin antagonist ([D-Lys3]-GHRP-6) in animal models of AD. Moreover, there were considerable discrepancies in the effects of ghrelin agonists on $A\beta$ load in the previous studies. The long-term administration of LY444711 significantly increases cognitive performance and lowers A^β levels in the dentate gyrus in an AD mouse model [46]. However, another study reported that the long-term treatment of LY444711 did not change A β levels in the dentate gyrus, stratum oriens, and olfactory bulb, while cognitive functions were improved in the AD mouse model [47]. Moreover, [D-Lys3] GHRP-6, a ghrelin antagonist, reduced $A\beta$ levels in the hippocampus in rats with monosodium glutamate-induced obesity [48]. In a randomized clinical trial that did not examine A β levels, the administration of MK-0677 did not improve cognitive function in patients with mild to moderate AD [49]. These disparities with the results of the studies using ghrelin agonists or antagonists confuse the development of strategies for treating AD. Therefore, to clarify the effects of MK-0677 on A β and A β -induced pathogenesis, we aimed to test the efficacy of the ghrelin agonist MK-0677 on AD pathology, including AB accumulation, neurodegeneration, and neuroinflammation in mice at the early phase of AD through histological quantification analyses. In addition, we purposed to evaluate the role of MK-0677 in levels of phosphorylation of the cyclic adenosine monophosphate (cAMP) response element binding protein (pCREB), which is responsible for cognitive functioning and neuronal survival [50,51], in dentate gyrus of the hippocampus. Here, we report that the administration of MK-0677 ameliorates Aβ accumulation, neuronal/synaptic loss, microgliosis, and astrogliosis, and inhibits the decrement of the phosphorylation of CREB in the hippocampus without altering significant appetite in three-month-old 5XFAD mice.

2. Results

2.1. Ghrelin Agonist Treatment Affected the Food Intake and Body Weight of the Healthy Animals

The ghrelin agonist MK-0677 has acute orexigenic effects in rodents [52]. To establish the optimal doses of MK-0677 that induce biological responses, we injected eight-week-old C57BL/6 mice with 0.1, 1, and 3 mg/kg of MK-0677 cumulatively for 10 days, as described previously [52]. The intraperitoneal administration of 1 and 3 mg/kg of MK-0677 increased food intake compared to that in the vehicle-administered group (n = 5; Figure 1A). In particular, the 3 mg dose significantly increased cumulative food intake during the 10-day administration period. In addition, the body weight of the mice administered MK-0677 at 1 mg/kg (92.6%) and 3 mg/kg (64.1%) was significantly higher than that of the vehicle-treated animals at 10 days (Figure 1B). The administration of MK-0677 to the three-month-old 5XFAD mice (n = 8) showed a tendency for increased cumulative food intake compared to that in the vehicle-treated 5XFAD mice (Figure 1C). However, the changes in body weight after treatment with MK-0677 did not differ significantly (Figure 1D).

2.2. Ghrelin Agonist Treatment Significantly Reduces $A\beta$ Accumulation in the Brains of 5XFAD Mice

To test if activation of the ghrelin receptor affected β -amyloidosis, we performed thioflavin-S staining in the frontal cortex of 5XFAD mice. The quantitative analysis demonstrated that the A β burden in the deep cortical layers was significantly decreased to 23% after the MK-0677 injections in the 5XFAD mice compared to the vehicle-administered 5XFAD mice (Figure 2A). In addition, the 4G8 immunoreactivity in layer V of the frontal cortex showed that MK-0677 administration significantly decreased the 4G8-positive areas to 30% compared with that in the vehicle-injected 5XFAD mice (Figure 2B). These results suggested that activation of the GHS-R1a might have anti-amyloid activity in the brains of patients with AD.



Figure 1. Cumulative food intake and change in body weight after the administration of MK-0677 in C57BL/6 mice and 5XFAD mice. MK-0677 was administered daily to C57BL/6 mice (n = 5) at doses of 0.1, 1, and 3 mg/kg for 10 days and to 5XFAD mice at doses of 5 mg/kg for three weeks. The group injected with MK-0677 exhibited a significant increase in cumulative food intake compared with the control group. Compared with the control group, the significant difference indicators are as follows: 0.1 mg/kg group ([†] p < 0.05), 1 mg/kg group ([‡] p < 0.001), and 3 mg/kg group (* p < 0.05 and ** p < 0.01) (**A**). The body weight changes for 10 days were also significant differences compared to control group) (**B**). MK-0677-treated 5XFAD mice (n = 8) showed tendency to the increase of cumulative food intake induced compared with vehicle-treated 5XFAD mice (n = 8) (**C**). The body weight changes among wild-type mice (n = 8), vehicle- and, MK-0677-treated 5XFAD mice (n = 8) (**C**). The body weight changes among wild-type mice (n = 8), vehicle- and, MK-0677-treated 5XFAD mice (n = 8) (**C**).



Figure 2. MK-0677 treatment significantly reduced A β plaques in the deep cortical layers of 5XFAD mice. The burden of A β was estimated by thoflavin-S staining and immunohistochemical staining for the 4G8 antibody. 5XFAD mice treated with MK-0677 (n = 6) showed a decreased positive area (%) in both thioflavin-S (**A**) and 4G8 (**B**)-stained brains, compared with vehicle-treated 5XFAD mice (n = 7). *** p < 0.001 indicates significant differences between the groups. Scale bar = 50 µm.

2.3. The Ghrelin Agonist Significantly Attenuated Neurodegeneration in the Neocortex of 5XFAD Mice

We examined whether the anti-amyloid actions of MK-0677 affected the neuronal and synaptic loss in the AD brain. To visualize neuronal death and synaptic loss induced by A β , we immunohistochemically stained brain tissues with neuronal nuclear antigen (NeuN) and synaptophysin (SYN) The quantitative analysis showed that the number of NeuN-positive cells and SYN immunoreactivity were significantly decreased in the neocortex of vehicle-treated 5XFAD mice compared with the vehicle-treated wild-type mice (Figure 3). In contrast, the number of NeuN-positive cells per area (Figure 3A) and optical density of SYN (Figure 3B) were significantly higher in the MK-0677-administered 5XFAD mice compared with the vehicle-injected 5XFAD mice. These results demonstrated that neuronal and synaptic loss in the brains of 5XFAD mice was ameliorated by treatment with MK-0677.

2.4. Ghrelin Agonist Significantly Inhibited Neuroinflammation in the Deep Cortical Layers of 5XFAD Mice

To examine the effects of MK-0677 on neuroinflammation induced by A β , we performed immunohistochemical staining for the microglia marker ionized calcium-binding adaptor molecule 1 (Iba-1) and astrocyte marker glial fibrillary acidic protein (GFAP). The quantitative analysis showed that the percentages of the Iba-1- and GFAP-stained areas were markedly increased in the vehicle-administered 5XFAD mice compared with the vehicle-treated wild-type littermate mice. However, the MK-0677-treated 5XFAD mice showed a significant decrease in the Iba-1-positive (Figure 4A) and GFAP-positive (Figure 4B) areas compared with those in the vehicle-treated 5XFAD mice. These results suggested that MK-0677 treatment alleviates the neuroinflammation induced by A β in the AD brain.



Figure 3. MK-0677-treated 5XFAD mice exhibited a significant reduction in neurodegeneration compared with the vehicle group. Immunofluorescent staining was performed to detect the markers of neuronal cells (NeuN) and pre-synaptic terminals (SYN) in layer V of the frontal cortex of wild-type (n = 8) and 5XFAD mice. MK-0677 significantly ameliorated the reduction of the number of NeuN (+) cells (**A**) and optical density of SYN (+) area (**B**) in 5XFAD mice (n = 6), compared with vehicle-treated 5XFAD mice (n = 7). *** p < 0.001 indicates significant differences between the groups. Scale bars are 50 µm in the upper panel and 25 µm in the lower panel.





Figure 4. MK-0677-treated 5XFAD mice exhibited significant inhibition of neuroinflammation compared with vehicle-administered mice. Immunofluorescent staining was performed to detect the markers of microglia (Iba-1) and astrocyte (GFAP) in layer V of the frontal cortex of wild-type (n = 8) and 5XFAD mice. MK-0677 significantly reduced the Iba-1 (+) area (**A**) and GFAP (+) area (**B**) in 5XFAD mice (n = 6), compared with vehicle-treated 5XFAD mice (n = 7). *** p < 0.001 indicates significant differences between the groups. Scale bar = 50 µm.

2.5. Ghrelin Agonist Significantly Increased Phosphorylation of CREB in Dentate Gyrus of the Hippocampus of 5XFAD Mice

To examine the effects of MK-0677 on the phosphorylation of CREB in dentate gyrus of the hippocampus, we performed immunohistochemical staining for the pCREB. The quantitative analysis showed that pCREB-immunostained cells per length of inner rim, defined as boundaries of hilus and granule layers, were markedly increased in the MK-0677-administered 5XFAD mice compared with the vehicle-treated 5XFAD mice (Figure 5). These result suggested that MK-0677 treatment inhibited the decrement of pCREB in dentate gyrus of the hippocampus in 5XFAD mice.



Figure 5. MK-0677 treatment significantly improved the reduced phosphorylation of CREB in 5XFAD mice. Immunofluorescent staining was performed to detect the phosphorylation form of CREB (pCREB) in dentate gyrus of the hippocampus of wild-type (n = 8) and 5XFAD mice. MK-0677-treated 5XFAD mice (n = 6) showed significantly increased pCREB, compared with vehicle-treated 5XFAD mice (n = 7). * p < 0.05 and *** p < 0.001 indicate significant differences between the groups. Scale bar = 50 µm. CREB = cyclic adenosine monophosphate (cAMP) response element binding protein.

3. Discussion

Because the evidence for a correlation between the ghrelin system and neural function is growing, the roles of ghrelin in neurodegenerative and cognitive impairment diseases have been intensively investigated. Moreover, the effects of ghrelin on the pathological changes resulting from A β -induced pathogenesis might be used as a possible strategy for AD treatment. However, the previous randomized clinical trial reported that MK-0677 had no effects on cognitive function in patients with AD did not assess any pathologic changes, including A β levels [49]. Moreover, the effects of MK-0677 on A β accumulation in studies of ghrelin agonists and antagonists have been conflicting [46–48], which prompted us to examine if a ghrelin agonist reduced A β accumulation and A β -induced pathogenesis in 5XFAD mice. The thioflavin-S staining and immunohistochemical results revealed that MK-0677 reduced A β accumulation, neurodegeneration, and neuroinflammation in the 5XFAD mice, which suggested that MK-0677 ameliorated A β accumulation, as well as A β -induced pathogenesis.

The therapeutic roles of exogenous GHS-R1a agonists in disorders, such as malnutrition, growth hormone deficiency, gastrointestinal hypomotility, and protein-energy wasting, are well known [53–56]. Despite the finding that a ghrelin agonist improved cognitive function in AD patients and animal models, the specific effects on A β accumulation varied [47–49]. We found that the intraperitoneal administration of MK-0677 significantly reduced A β accumulation in the frontal cortex in three-month-old 5XFAD mice. The deposition of A β started in the 5XFAD mice at two months and markedly increased at four months [57]. Moreover, several studies reported that the significant memory impairment was shown in the 5XFAD mice at 6 months of age [58–62].

Considering that A β deposits in the neocortex represent the first of the five phases of A β deposition and the patients with AD in the early phase of AB deposition do not exhibit dementia [63], our data indicated that the administration of a ghrelin agonist in the early stage of AD could alleviate Aβ accumulation before the onset of AD-related symptoms. The results of pCREB immunostaining showed that MK-0677 inhibited the decreased pCREB levels in dentate gyrus of the hippocampus and were corroborated by a previous study reporting the up-regulation of pCREB induced by increased ghrelin levels in the hippocampus [64]. It has been reported that hippocampal pCREB levels were down-regulated in AD patients or animal models of AD [65,66]. Accordingly, the inhibition of decreased pCREB in the hippocampus induced by MK-0677 might indirectly implicate that MK-0677 might conserve cognitive functions against Aβ-induced toxicity. However, these interpretations should be treated with caution because previous studies of ghrelin agonists in AD models and treatment have reported inconsistent findings of the effects of ghrelin agonists on A β accumulation [46,47]. Notably, the two studies had a difference in the diet of the animals: free diet vs. high-glycemic index diet. One possible explanation is that the altered glucose homeostasis induced by the high glycemic index diet influenced A β formation and that the ameliorating effects of ghrelin on glucose homeostasis were not sufficient to reduce the $A\beta$ formation. Furthermore, results from a randomized clinical trial that showed MK-0677 had no cognitive improving effects in AD patients [49] made it hard to interpret our data. This disparity could be explained by the differences of AD stage of subjects between studies. The previous study included AD patients with mild to moderate dementia, whereas ours were mice with a very early stage of AD. Thus, these different results from studies might implicate that MK-0677 may be effective, at least, in an early phase of AD. Another possible limitation could be raised by studies supporting the roles of soluble A β oligomers on neurotoxicity [67]. Further investigations are needed to examine the effect of the ghrelin agonist on A β accumulation in models of altered glucose homeostasis and soluble A β oligomers, as well as behavioral tests for cognitive functions in AD animal models.

We next examined the effects of MK-0677 on neurodegeneration in the neocortex. NeuN and SYN, which are markers of neuronal and synaptic loss, respectively, were examined. The immunohistochemical results suggested that both neuronal/synaptic losses were reduced by the administration of the ghrelin agonist. In accordance with the neurodegenerative pathology in

AD, A β accumulation is known to play key roles in that pathology [68]. The A β accumulation and accompanying neuronal loss were corroborated in a previous study that used the same transgenic mice as our study did [69]. Therefore, these results strongly suggested that treatment with the ghrelin agonist would sufficiently reduce neurodegeneration by inhibiting A β accumulation.

The results of the immunohistochemistry for Iba-1 and GFAP indicated that both microgliosis and astrogliosis were decreased in the deep layers of the neocortices by ghrelin treatment. These results were consistent with previous results that indicated that ghrelin suppressed gliosis in the CNS. The intraperitoneal injections of ghrelin inhibited microgliosis and the expression of inflammatory cytokines and nitric oxide synthase in the brain of an animal model of MPTP-induced Parkinson's disease, and similar effects were found in 1-methyl-4-phenylpyridinium (MPP+)-treated cultured mesencephalic cells [70]. Ghrelin inhibits reactive oxygen species and inflammatory signaling proteins in microglial cells activated by lipopolysaccharide [71]. In addition, ghrelin decreases mRNA levels of tumor necrosis factor- α in primary cultured hypothalamic astrocytes [72]. Furthermore, in studies of the effects of the ghrelin agonist on neuroinflammation in AD model mice, the ghrelin agonist LY444711 subdued microglial activation [46]. However, the effects of the ghrelin agonist on gliosis in AD pathogenesis should be further examined because its effects could differ in AD patients with other conditions, such as an aberrant glucose metabolism [47].

Besides the beneficial effects of ghrelin on A β and A β -induced pathogenesis, ghrelin demonstrates orexigenic properties [14,73]. Ghrelin increases feeding by stimulating neuropeptide Y in the arcuate nucleus, as well as counteracted leptin [26,74]. We demonstrated that the administration of 3 mg/kg of MK-0677 resulted in an 11.6% increase in food intake and 64% weight gain in the C57BL/6 mice compared with the vehicle group. These findings were consistent with those of previous studies which reported that MK-0677 increased food intake and body weight [52,75,76]. Interestingly, MK-0677 did not increase the cumulative food intake or changes in body weight in the 5XFAD mice. Although the mechanisms underlying the lack of effects of ghrelin on food intake and body weight were not clear, it can be assumed that different species and strains may affect the results. Regardless of the specific mechanism, these results indicate that MK-0677 had beneficial effects on A β accumulation and A β -induced pathogenesis without changing appetite. Another possible explanation is suggested by the evidence, proposing that alterations of signaling pathways, such as the adenosine monophosphate (AMP)-activated protein kinase pathway, impair energy homeostasis in AD [77]. Thus, additional studies are needed to identify the effects of ghrelin on signaling proteins involving metabolism.

Hyperphosphorylation of tau and the resulting neurofibrillary tangles (NFTs) is another hallmarker of AD [78]. The A β -induced tau-pathology has been demonstrated by several studies, while its mechanism has not been fully elucidated [79,80]. It has been reported that only pS396 levels were increased in two-month-old 5XFAD mice [81]. However, despite the detection of abnormal tau phosphorylation in the brain, neurofibrillary tangles, the aggregates of hyperphosphorylated tau, are not observed as with other A β -related mutant mice (APP and APP/PS1) [69]. In addition, according to the results from a study using 5XFAD mice at three, six, and nine months of age, endogenous tau did not show age-related differences, while A β -related factors showed significant changes [82].

Concerning the signaling molecules involved in the abnormal phosphorylation of tau such as ghrelin treatment glycogen synthase kinase 3 beta (GSK-3), c-Jun N-terminal kinase (JNK), and cyclin-dependent kinases 5 (CDK5) [83], ghrelin treatment inactivated GSK-3β in spinal cord motoneurons [84]. In addition, treatment of ghrelin decreased the phosphorylation of c-Jun N-terminal kinase (JNK) in stimulated macrophages [85,86], hepatocytes [87], and hippocampal neurons [88]. Furthermore, although the relationship between ghrelin and CDK5 is not known, treatment of ghrelin increased the protein expression of CDK2 and Cyclin a in hippocampal neural stem cells [89]. Therefore, although tauopathy such as NFTs is inevident in 5XFAD, further studies are needed to determine the effect of ghrelin agonist MK-0677 on the phosphorylation of tau using other AD model mice.

An agonist needs its receptor for its action. GHS-R1a and GHSR-1 mRNA are expressed in various regions of the brain [90,91]. In patients with AD, even though GHS-R1a expression was decreased,

a certain level of GHS-R1a was expressed [20]. Thus, the administration of exogenous ghrelin agonist decreased A β accumulation and improved A β -induced pathogenesis by stimulating GHS-R1a in the brain.

In the present study, the ghrelin agonist MK-0677 showed high potential for decreasing the burden of amyloid plaques. To solidify this result, studies on the precise molecular mechanisms of the A β -lowering effects of MK-0677 are essential. In other words, additional studies are needed to investigate the effects of MK-0677 both on amyloid precursor protein processing through the modulation of α -, β -, and γ -secretase, and the expression or activity of A β -degrading enzymes, such as neprilysin and insulin-degrading enzymes.

In conclusion, we demonstrated for the first time that the ghrelin agonist MK-0677 suppressed $A\beta$ pathology and $A\beta$ -mediated pathology in vivo. Our results suggest that MK-0677, at least, may be a promising therapeutic agent for the early phase of AD and that activation of the ghrelin receptor can be a therapeutic target for the treatment of AD.

4. Materials and Methods

4.1. Animals and Drug Treatment

Three-month-old male transgenic mice with five familial AD mutations (5XFAD) and B6JSLF1 mice were purchased from The Jackson Laboratory (Bar Harbor, ME, USA). The C57BL/6 mice were obtained from Korean Animal Technology (Koatech, Pyeongtaek-si, Gyeonggi-do, Korea). The 5XFAD mice had mutations in the amyloid precursor protein (*APP*) gene (Swe^{K670N,M671L}, Lon^{V717I}, and Flo^{I716V}) and presenilin 1 (*PSEN1*) gene (M146L and L286V) regulated by the Thy1 promoter [69]. The number of mice per group was five for the C57BL/6 mice and six to eight for the 5XFAD and wild-type littermates. All mice were given access to food ad libitum and maintained under a 12-h light/dark cycle. The experimental procedures were designed in accordance with the National Institutes of Health guide for the care and use of laboratory animals [92] and performed under supervision of the Institutional Animal Care and Use Guidelines of Konyang University (project code:P-15-21-A-01; 30 November 2015).

Inasmuch as 5XFAD mice exhibit detectable A β deposits at two months of age [57,69], MK-0677 (Bio-Techne Corporation, Minneapolis, MN, USA) or a vehicle was administered at the age of three months. MK-0677 was dissolved in saline and administered by intraperitoneal injections at a dose of 5 mg/kg daily for three weeks. The vehicle groups were injected with saline in parallel. The animals were sacrificed one day after the last administration. Food intake and body weight were measured before administration during the period of treatment.

4.2. Brain Tissue Preparation

The animals were anesthetized and transcardially perfused with 0.05 M phosphate-buffered saline (PBS) and then fixed with ice-cold 4% paraformaldehyde in 0.1 M phosphate buffer (PB). The brain tissue was removed, postfixed in 0.1 M PB containing 4% paraformaldehyde (Sigma-Aldrich Corporation, St. Louis, MO, USA) for 20 h at 4 °C, and then immersed in 30% sucrose in 0.05 M PBS for three days at 4 °C for cryoprotection. The samples were sliced into 30-µm serial coronal sections on a cryostat (Leica Biosystems Nussloch GmbH, Nussloch, Germany). The serial coronal sections were stored in storing solution (25% ethylene glycol and 25% glycerol in 0.05 M PB) at 4 °C until use for the histological analysis.

4.3. Thioflavin-S Staining

Six brain sections at 210–240- μ m intervals were extracted from each mouse from the region between –2.6 mm and –4.3 mm to the bregma with reference to Paxinos and Franklin's the Mouse Brain in Stereotaxic Coordinates [93]. To label A β , the stored brain sections were washed three times for five minutes in PBS. The sections were incubated in filtered thioflavin-S solution (1%; Sigma-Aldrich Corporation, St. Louis, MO, USA) for 15 minutes and washed in 80% and 70% ethanol for one minute each. After washing three times for five minutes each in PBS, the sections were mounted on ProbeOnTM Plus Microscope Slides (Thermo Fisher Scientific Inc., Waltham, MA, USA) and coverslipped with the FluoroshieldTM with DAPI (Sigma-Aldrich Corporation, St. Louis, MO, USA).

4.4. Immunofluorescence Labeling

Six brain sections at 210–240-µm intervals were extracted from each mouse from the region between -2.6 mm and -4.3 mm to the bregma with reference to Paxinos and Franklin's the Mouse Brain in Stereotaxic Coordinates [93]. To visualize the immunoreactivity of A β , NeuN, Iba-1, SYN, GFAP, and pCREB, free-floating sections were incubated overnight at 4 °C with the mouse anti-4G8 antibody (1:2000; BioLegend, San Diego, CA, USA), mouse anti-NeuN antibody (1:100; Merck KGaA, Darmstadt, Germany), goat anti-Iba1 antibody (1:500; Abcam plc, Cambridge, UK), mouse anti-SYN antibody (1:500; Sigma-Aldrich Corporation, St. Louis, MO, USA), rat anti-GFAP (1:200; Thermo Fisher Scientific Inc., Waltham, MA, USA), or mouse anti-pCREB antibody (1:1000; MERCK, Kenilworth, NJ, USA). After washing three times for five minutes in PBS, the sections were incubated with the goat Alexa 488-conjugated anti-mouse IgG (1:200; Thermo Fisher Scientific Inc., Waltham, MA, USA) or donkey Alexa 594-conjugated anti-rabbit IgG (1:200; Thermo Fisher Scientific Inc., Waltham, MA, USA) for 1 h at room temperature. The tissue sections were mounted on ProbeOnTM Plus Microscope Slides (Thermo Fisher Scientific Inc., Waltham, MA, USA) and coverslipped with FluoroshieldTM with DAPI (Sigma-Aldrich Corporation, St. Louis, MO, USA).

4.5. Image Acquisition and Quantification

The labeled tissues were imaged and analyzed with a Zeiss LSM 700 Meta confocal microscope (Carl Zeiss AG, Oberkochen, Germany). We examined amyloid plaques, neurodegeneration and neuroinflammation in the deep cortical layers of the frontal cortex, and pCREB in dentate gyrus of the hippocampus. To quantify the fraction of the stained areas and immunoreactivity, we analyzed the images using ImageJ software (National Institutes of Health, Bethesda, MD, USA), as described in our previous study [94].

4.6. Statistical Analysis

The Graph Pad Prism 5 software (GraphPad Software, La Jolla, CA, USA) was used for data representation and statistical analysis. All results are shown as means \pm standard error of the mean. The differences between the treatments were analyzed by one-way analysis of variance, which was followed by Fisher's least significant differences post hoc test or an independent *t*-test. *p* values less than 0.05 were considered statistically significant.

5. Conclusions

The results of thioflavin-S staining and immunohistochemical staining results showed that the administration of MK-0677, a ghrelin agonist, attenuated A β deposition and A β -mediated pathologies in the neocortex of three-month-old 5XFAD mice. Moreover, MK-0677 inhibited the decreased pCREB levels in dentate gyrus of the hippocampus. Taken together, we suggest that MK-0677 might be have potential for the treatment of early phase AD.

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References

- 1. Alzheimer's Association. 2016 Alzheimer's disease facts and figures. Alzheimers Dement. 2016, 12, 459–509.
- 2. Alzheimer's Association. 2017 Alzheimer's disease facts and figures. *Alzheimers Dement.* 2017, 13, 325–373.
- McKhann, G.M.; Knopman, D.S.; Chertkow, H.; Hyman, B.T.; Jack, C.R., Jr.; Kawas, C.H.; Klunk, W.E.; Koroshetz, W.J.; Manly, J.J.; Mayeux, R.; et al. The diagnosis of dementia due to Alzheimer's disease: Recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimers Dement.* 2011, 7, 263–269. [CrossRef] [PubMed]
- 4. Selkoe, D.J.; Hardy, J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Mol. Med.* **2016**, *8*, 595–608. [CrossRef] [PubMed]
- Ariyasu, H.; Takaya, K.; Tagami, T.; Ogawa, Y.; Hosoda, K.; Akamizu, T.; Suda, M.; Koh, T.; Natsui, K.; Toyooka, S.; et al. Stomach is a major source of circulating ghrelin, and feeding state determines plasma ghrelin-like immunoreactivity levels in humans. *J. Clin. Endocrinol. Metab.* 2001, *86*, 4753–4758. [CrossRef] [PubMed]
- 6. Delhanty, P.J.; Neggers, S.J.; van der Lely, A.J. Mechanisms in endocrinology: Ghrelin: The differences between acyl- and des-acyl ghrelin. *Eur. J. Endocrinol.* **2012**, *167*, 601–608. [CrossRef] [PubMed]
- 7. Tanaka, M.; Hayashida, Y.; Nakao, N.; Nakai, N.; Nakashima, K. Testis-specific and developmentally induced expression of a ghrelin gene-derived transcript that encodes a novel polypeptide in the mouse. *Biochim. Biophys. Acta* **2001**, *1522*, 62–65. [CrossRef]
- Caminos, J.E.; Tena-Sempere, M.; Gaytan, F.; Sanchez-Criado, J.E.; Barreiro, M.L.; Nogueiras, R.; Casanueva, F.F.; Aguilar, E.; Dieguez, C. Expression of ghrelin in the cyclic and pregnant rat ovary. *Endocrinology* 2003, 144, 1594–1602. [CrossRef] [PubMed]
- Mori, K.; Yoshimoto, A.; Takaya, K.; Hosoda, K.; Ariyasu, H.; Yahata, K.; Mukoyama, M.; Sugawara, A.; Hosoda, H.; Kojima, M.; et al. Kidney produces a novel acylated peptide, ghrelin. *FEBS Lett.* 2000, 486, 213–216. [CrossRef]
- Korbonits, M.; Bustin, S.A.; Kojima, M.; Jordan, S.; Adams, E.F.; Lowe, D.G.; Kangawa, K.; Grossman, A.B. The expression of the growth hormone secretagogue receptor ligand ghrelin in normal and abnormal human pituitary and other neuroendocrine tumors. *J. Clin. Endocrinol. Metab.* 2001, *86*, 881–887. [CrossRef] [PubMed]
- Date, Y.; Kojima, M.; Hosoda, H.; Sawaguchi, A.; Mondal, M.S.; Suganuma, T.; Matsukura, S.; Kangawa, K.; Nakazato, M. Ghrelin, a novel growth hormone-releasing acylated peptide, is synthesized in a distinct endocrine cell type in the gastrointestinal tracts of rats and humans. *Endocrinology* 2000, 141, 4255–4261. [CrossRef] [PubMed]
- Hattori, N.; Saito, T.; Yagyu, T.; Jiang, B.H.; Kitagawa, K.; Inagaki, C. GH, GH receptor, GH secretagogue receptor, and ghrelin expression in human T cells, B cells, and neutrophils. *J. Clin. Endocrinol. Metab.* 2001, 86, 4284–4291. [CrossRef] [PubMed]
- 13. Volante, M.; Allia, E.; Gugliotta, P.; Funaro, A.; Broglio, F.; Deghenghi, R.; Muccioli, G.; Ghigo, E.; Papotti, M. Expression of ghrelin and of the GH secretagogue receptor by pancreatic islet cells and related endocrine tumors. *J. Clin. Endocrinol. Metab.* **2002**, *87*, 1300–1308. [CrossRef] [PubMed]
- 14. Kojima, M.; Hosoda, H.; Date, Y.; Nakazato, M.; Matsuo, H.; Kangawa, K. Ghrelin is a growth-hormone-releasing acylated peptide from stomach. *Nature* **1999**, 402, 656–660. [CrossRef] [PubMed]
- 15. Lu, S.; Guan, J.L.; Wang, Q.P.; Uehara, K.; Yamada, S.; Goto, N.; Date, Y.; Nakazato, M.; Kojima, M.; Kangawa, K.; et al. Immunocytochemical observation of ghrelin-containing neurons in the rat arcuate nucleus. *Neurosci. Lett.* **2002**, *321*, 157–160. [CrossRef]
- Cowley, M.A.; Smith, R.G.; Diano, S.; Tschop, M.; Pronchuk, N.; Grove, K.L.; Strasburger, C.J.; Bidlingmaier, M.; Esterman, M.; Heiman, M.L.; et al. The distribution and mechanism of action of ghrelin in the CNS demonstrates a novel hypothalamic circuit regulating energy homeostasis. *Neuron* 2003, *37*, 649–661. [CrossRef]
- 17. Hou, Z.; Miao, Y.; Gao, L.; Pan, H.; Zhu, S. Ghrelin-containing neuron in cerebral cortex and hypothalamus linked with the DVC of brainstem in rat. *Regul. Pept.* **2006**, *134*, 126–131. [CrossRef] [PubMed]
- Gahete, M.D.; Cordoba-Chacon, J.; Kineman, R.D.; Luque, R.M.; Castano, J.P. Role of ghrelin system in neuroprotection and cognitive functions: Implications in Alzheimer's disease. *Peptides* 2011, 32, 2225–2228. [CrossRef] [PubMed]

- 19. Kim, C.; Kim, S.; Park, S. Neurogenic Effects of Ghrelin on the Hippocampus. *Int. J. Mol. Sci.* 2017, *18*. [CrossRef] [PubMed]
- 20. Gahete, M.D.; Rubio, A.; Cordoba-Chacon, J.; Gracia-Navarro, F.; Kineman, R.D.; Avila, J.; Luque, R.M.; Castano, J.P. Expression of the ghrelin and neurotensin systems is altered in the temporal lobe of Alzheimer's disease patients. *J. Alzheimers Dis.* **2010**, *22*, 819–828. [CrossRef] [PubMed]
- 21. Theodoropoulou, A.; Metallinos, I.C.; Psyrogiannis, A.; Vagenakis, G.A.; Kyriazopoulou, V. Ghrelin and leptin secretion in patients with moderate Alzheimer's disease. *J. Nutr. Health Aging* **2012**, *16*, 472–477. [CrossRef] [PubMed]
- 22. Pradhan, G.; Samson, S.L.; Sun, Y. Ghrelin: Much more than a hunger hormone. *Curr. Opin. Clin. Nutr. Metab. Care* 2013, *16*, 619–624. [CrossRef] [PubMed]
- 23. Tschop, M.; Smiley, D.L.; Heiman, M.L. Ghrelin induces adiposity in rodents. *Nature* **2000**, 407, 908–913. [CrossRef] [PubMed]
- 24. Heppner, K.M.; Piechowski, C.L.; Muller, A.; Ottaway, N.; Sisley, S.; Smiley, D.L.; Habegger, K.M.; Pfluger, P.T.; Dimarchi, R.; Biebermann, H.; et al. Both acyl and des-acyl ghrelin regulate adiposity and glucose metabolism via central nervous system ghrelin receptors. *Diabetes* **2014**, *63*, 122–131. [CrossRef] [PubMed]
- 25. Gil-Campos, M.; Aguilera, C.M.; Canete, R.; Gil, A. Ghrelin: a hormone regulating food intake and energy homeostasis. *Br. J. Nutr.* **2006**, *96*, 201–226. [CrossRef] [PubMed]
- 26. Nakazato, M.; Murakami, N.; Date, Y.; Kojima, M.; Matsuo, H.; Kangawa, K.; Matsukura, S. a role for ghrelin in the central regulation of feeding. *Nature* **2001**, *409*, 194–198. [CrossRef] [PubMed]
- 27. Ukkola, O.; Poykko, S. Ghrelin, growth and obesity. Ann. Med. 2002, 34, 102–108. [CrossRef] [PubMed]
- 28. Stoyanova, I.I.; Hofmeijer, J.; van Putten, M.; le Feber, J. Acyl Ghrelin Improves Synapse Recovery in an In Vitro Model of Postanoxic Encephalopathy. *Mol. Neurobiol.* **2016**, *53*, 6136–6143. [CrossRef] [PubMed]
- Chung, H.; Kim, E.; Lee, D.H.; Seo, S.; Ju, S.; Lee, D.; Kim, H.; Park, S. Ghrelin inhibits apoptosis in hypothalamic neuronal cells during oxygen-glucose deprivation. *Endocrinology* 2007, 148, 148–159. [CrossRef] [PubMed]
- 30. Chung, H.; Li, E.; Kim, Y.; Kim, S.; Park, S. Multiple signaling pathways mediate ghrelin-induced proliferation of hippocampal neural stem cells. *J. Endocrinol.* **2013**, *218*, 49–59. [CrossRef] [PubMed]
- Yang, M.; Hu, S.; Wu, B.; Miao, Y.; Pan, H.; Zhu, S. Ghrelin inhibits apoptosis signal-regulating kinase 1 activity via upregulating heat-shock protein 70. *Biochem. Biophys. Res. Commun.* 2007, 359, 373–378. [CrossRef] [PubMed]
- Bulgarelli, I.; Tamiazzo, L.; Bresciani, E.; Rapetti, D.; Caporali, S.; Lattuada, D.; Locatelli, V.; Torsello, A. Desacyl-ghrelin and synthetic GH-secretagogues modulate the production of inflammatory cytokines in mouse microglia cells stimulated by beta-amyloid fibrils. *J. Neurosci. Res.* 2009, *87*, 2718–2727. [CrossRef] [PubMed]
- 33. Liu, Y.; Wang, P.S.; Xie, D.; Liu, K.; Chen, L. Ghrelin reduces injury of hippocampal neurons in a rat model of cerebral ischemia/reperfusion. *Chin. J. Physiol.* **2006**, *49*, 244–250. [PubMed]
- 34. Xu, J.; Wang, S.; Lin, Y.; Cao, L.; Wang, R.; Chi, Z. Ghrelin protects against cell death of hippocampal neurons in pilocarpine-induced seizures in rats. *Neurosci. Lett.* **2009**, *453*, 58–61. [CrossRef] [PubMed]
- 35. Andrews, Z.B.; Erion, D.; Beiler, R.; Liu, Z.W.; Abizaid, A.; Zigman, J.; Elsworth, J.D.; Savitt, J.M.; DiMarchi, R.; Tschoep, M.; et al. Ghrelin promotes and protects nigrostriatal dopamine function via a UCP2-dependent mitochondrial mechanism. *J. Neurosci.* **2009**, *29*, 14057–14065. [CrossRef] [PubMed]
- 36. Diano, S.; Farr, S.A.; Benoit, S.C.; McNay, E.C.; da Silva, I.; Horvath, B.; Gaskin, F.S.; Nonaka, N.; Jaeger, L.B.; Banks, W.A.; et al. Ghrelin controls hippocampal spine synapse density and memory performance. *Nat. Neurosci.* **2006**, *9*, 381–388. [CrossRef] [PubMed]
- Li, E.; Chung, H.; Kim, Y.; Kim, D.H.; Ryu, J.H.; Sato, T.; Kojima, M.; Park, S. Ghrelin directly stimulates adult hippocampal neurogenesis: Implications for learning and memory. *Endocr. J.* 2013, 60, 781–789. [CrossRef] [PubMed]
- Moon, M.; Choi, J.G.; Nam, D.W.; Hong, H.S.; Choi, Y.J.; Oh, M.S.; Mook-Jung, I. Ghrelin ameliorates cognitive dysfunction and neurodegeneration in intrahippocampal amyloid-beta1-42 oligomer-injected mice. *J. Alzheimers Dis.* 2011, 23, 147–159. [CrossRef] [PubMed]
- 39. Moon, M.; Cha, M.Y.; Mook-Jung, I. Impaired hippocampal neurogenesis and its enhancement with ghrelin in 5XFAD mice. *J. Alzheimers Dis.* **2014**, *41*, 233–241. [CrossRef] [PubMed]

- 40. Bailey, A.R.; Smith, R.G.; Leng, G. The nonpeptide growth hormone secretagogue, MK-0677, activates hypothalamic arcuate nucleus neurons in vivo. *J. Neuroendocrinol.* **1998**, *10*, 111–118. [CrossRef] [PubMed]
- 41. Patchett, A.A.; Nargund, R.P.; Tata, J.R.; Chen, M.H.; Barakat, K.J.; Johnston, D.B.; Cheng, K.; Chan, W.W.; Butler, B.; Hickey, G.; et al. Design and biological activities of L-163,191 (MK-0677): a potent, orally active growth hormone secretagogue. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 7001–7005. [CrossRef] [PubMed]
- 42. Bennett, K.A.; Langmead, C.J.; Wise, A.; Milligan, G. Growth hormone secretagogues and growth hormone releasing peptides act as orthosteric super-agonists but not allosteric regulators for activation of the G protein Galpha(o1) by the Ghrelin receptor. *Mol. Pharmacol.* **2009**, *76*(4), 802–811. [CrossRef] [PubMed]
- 43. Howick, K.; Griffin, B.T.; Cryan, J.F.; Schellekens, H. From Belly to Brain: Targeting the Ghrelin Receptor in Appetite and Food Intake Regulation. *Int. J. Mol. Sci.* **2017**, *18*. [CrossRef] [PubMed]
- 44. Tolle, V.; Bassant, M.H.; Zizzari, P.; Poindessous-Jazat, F.; Tomasetto, C.; Epelbaum, J.; Bluet-Pajot, M.T. Ultradian rhythmicity of ghrelin secretion in relation with GH, feeding behavior, and sleep-wake patterns in rats. *Endocrinology* **2002**, *143*, 1353–1361. [CrossRef] [PubMed]
- Chapman, I.M.; Bach, M.A.; Van Cauter, E.; Farmer, M.; Krupa, D.; Taylor, A.M.; Schilling, L.M.; Cole, K.Y.; Skiles, E.H.; Pezzoli, S.S.; et al. Stimulation of the growth hormone (GH)-insulin-like growth factor I axis by daily oral administration of a GH secretogogue (MK-677) in healthy elderly subjects. *J. Clin. Endocrinol. Metab.* **1996**, *81*, 4249–4257. [CrossRef] [PubMed]
- 46. Dhurandhar, E.J.; Allison, D.B.; van Groen, T.; Kadish, I. Hunger in the absence of caloric restriction improves cognition and attenuates Alzheimer's disease pathology in a mouse model. *PLoS ONE* **2013**, *8*, e60437. [CrossRef] [PubMed]
- Kunath, N.; van Groen, T.; Allison, D.B.; Kumar, A.; Dozier-Sharpe, M.; Kadish, I. Ghrelin agonist does not foster insulin resistance but improves cognition in an Alzheimer's disease mouse model. *Sci. Rep.* 2015, *5*, 11452. [CrossRef] [PubMed]
- 48. Madhavadas, S.; Kutty, B.M.; Subramanian, S. Amyloid beta lowering and cognition enhancing effects of ghrelin receptor analog [D-Lys (3)] GHRP-6 in rat model of obesity. *Indian J. Biochem. Biophys.* **2014**, *51*, 257–262. [PubMed]
- Sevigny, J.J.; Ryan, J.M.; van Dyck, C.H.; Peng, Y.; Lines, C.R.; Nessly, M.L.; Group, M.K.P.S. Growth hormone secretagogue MK-677: No clinical effect on AD progression in a randomized trial. *Neurology* 2008, 71, 1702–1708. [CrossRef] [PubMed]
- 50. Bartolotti, N.; Segura, L.; Lazarov, O. Diminished CRE-Induced Plasticity is Linked to Memory Deficits in Familial Alzheimer's Disease Mice. *J. Alzheimers Dis.* **2016**, *50*, 477–489. [CrossRef] [PubMed]
- 51. Ortega-Martinez, S. a new perspective on the role of the CREB family of transcription factors in memory consolidation via adult hippocampal neurogenesis. *Front. Mol. Neurosci.* **2015**, *8*, 46. [CrossRef] [PubMed]
- Patel, K.; Joharapurkar, A.; Dhanesha, N.; Patel, V.; Kshirsagar, S.; Raval, P.; Raval, S.; Jain, M.R. Thyroid hormone modulates food intake and glycemia via ghrelin secretion in Zucker fatty rats. *Drug Res. (Stuttg.)* 2014, 64, 523–529. [CrossRef] [PubMed]
- 53. Su, J.; Geng, J.; Bao, J.; Tang, Y.; Liu, M.; Yu, H.; Han, Y.; Huang, W.; Zhou, S. Two ghrelin receptor agonists for adults with malnutrition: a systematic review and meta-analysis. *Nutr. J.* **2016**, *15*, 97. [CrossRef] [PubMed]
- 54. Garcia, J.M.; Swerdloff, R.; Wang, C.; Kyle, M.; Kipnes, M.; Biller, B.M.; Cook, D.; Yuen, K.C.; Bonert, V.; Dobs, A.; et al. Macimorelin (AEZS-130)-stimulated growth hormone (GH) test: Validation of a novel oral stimulation test for the diagnosis of adult GH deficiency. *J. Clin. Endocrinol. Metab.* 2013, *98*, 2422–2429. [CrossRef] [PubMed]
- Charoenthongtrakul, S.; Giuliana, D.; Longo, K.A.; Govek, E.K.; Nolan, A.; Gagne, S.; Morgan, K.; Hixon, J.; Flynn, N.; Murphy, B.J.; et al. Enhanced gastrointestinal motility with orally active ghrelin receptor agonists. *J. Pharmacol. Exp. Therap.* 2009, 329, 1178–1186. [CrossRef] [PubMed]
- Campbell, G.A.; Patrie, J.T.; Gaylinn, B.D.; Thorner, M.O.; Bolton, W.K. Oral ghrelin receptor agonist MK-0677 increases serum insulin-like growth factor 1 in hemodialysis patients: a randomized blinded study. *Nephrol. Dialysis Transpl.: Off. Publ. Eur. Dialysis Transpl. Assoc.—Eur. Renal Assoc.* 2018, 33, 523–530. [CrossRef] [PubMed]
- 57. Girard, S.D.; Baranger, K.; Gauthier, C.; Jacquet, M.; Bernard, A.; Escoffier, G.; Marchetti, E.; Khrestchatisky, M.; Rivera, S.; Roman, F.S. Evidence for early cognitive impairment related to frontal cortex in the 5XFAD mouse model of Alzheimer's disease. *J. Alzheimers Dis.* 2013, *33*, 781–796. [CrossRef] [PubMed]

- 58. Kimura, R.; Ohno, M. Impairments in remote memory stabilization precede hippocampal synaptic and cognitive failures in 5XFAD Alzheimer mouse model. *Neurobiol. Dis.* **2009**, *33*, 229–235. [CrossRef] [PubMed]
- 59. Ohno, M. Failures to reconsolidate memory in a mouse model of Alzheimer's disease. *Neurobiol. Learn. Memory* **2009**, *92*, 455–459. [CrossRef] [PubMed]
- Kaczorowski, C.C.; Sametsky, E.; Shah, S.; Vassar, R.; Disterhoft, J.F. Mechanisms underlying basal and learning-related intrinsic excitability in a mouse model of Alzheimer's disease. *Neurobiol. Aging* 2011, 32, 1452–1465. [CrossRef] [PubMed]
- 61. Devi, L.; Ohno, M. Genetic reductions of beta-site amyloid precursor protein-cleaving enzyme 1 and amyloid-beta ameliorate impairment of conditioned taste aversion memory in 5XFAD Alzheimer's disease model mice. *Eur. J. Neurosci.* **2010**, *31*, 110–118. [CrossRef] [PubMed]
- Girard, S.D.; Jacquet, M.; Baranger, K.; Migliorati, M.; Escoffier, G.; Bernard, A.; Khrestchatisky, M.; Feron, F.; Rivera, S.; Roman, F.S.; et al. Onset of hippocampus-dependent memory impairments in 5XFAD transgenic mouse model of Alzheimer's disease. *Hippocampus* 2014, 24, 762–772. [CrossRef] [PubMed]
- Thal, D.R.; Rub, U.; Orantes, M.; Braak, H. Phases of a β-deposition in the human brain and its relevance for the development of AD. *Neurology* 2002, *58*, 1791–1800. [CrossRef] [PubMed]
- 64. Isokawa, M. The role of ghrelin in the hippocampal neuron plasticity. *Alzheimers Dement.: J. Alzheimers Assoc.* **2009**, *5*, P172. [CrossRef]
- 65. Yamamoto-Sasaki, M.; Ozawa, H.; Saito, T.; Rosler, M.; Riederer, P. Impaired phosphorylation of cyclic AMP response element binding protein in the hippocampus of dementia of the Alzheimer type. *Brain Res.* **1999**, *824*, 300–303. [CrossRef]
- 66. Pugazhenthi, S.; Wang, M.; Pham, S.; Sze, C.I.; Eckman, C.B. Downregulation of CREB expression in Alzheimer's brain and in Aβ-treated rat hippocampal neurons. *Mol. Neurodegen.* 2011, *6*, 60. [CrossRef] [PubMed]
- Ferreira, S.T.; Lourenco, M.V.; Oliveira, M.M.; De Felice, F.G. Soluble amyloid-beta oligomers as synaptotoxins leading to cognitive impairment in Alzheimer's disease. *Front. Cell. Neurosci.* 2015, *9*, 191. [CrossRef] [PubMed]
- Wirths, O.; Bayer, T.A. Intraneuronal Aβ accumulation and neurodegeneration: Lessons from transgenic models. *Life Sci.* 2012, *91*, 1148–1152. [CrossRef] [PubMed]
- 69. Oakley, H.; Cole, S.L.; Logan, S.; Maus, E.; Shao, P.; Craft, J.; Guillozet-Bongaarts, A.; Ohno, M.; Disterhoft, J.; Van Eldik, L.; et al. Intraneuronal β-amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: Potential factors in amyloid plaque formation. *J. Neurosci.* 2006, 26, 10129–10140. [CrossRef] [PubMed]
- 70. Moon, M.; Kim, H.G.; Hwang, L.; Seo, J.H.; Kim, S.; Hwang, S.; Kim, S.; Lee, D.; Chung, H.; Oh, M.S.; et al. Neuroprotective effect of ghrelin in the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine mouse model of Parkinson's disease by blocking microglial activation. *Neurotoxicity Res.* 2009, 15, 332–347. [CrossRef] [PubMed]
- 71. Lee, J.Y.; Yune, T.Y. Ghrelin inhibits oligodendrocyte cell death by attenuating microglial activation. *Endocrinol. Metab. (Seoul Korea)* **2014**, *29*, 371–378. [CrossRef] [PubMed]
- 72. Garcia-Caceres, C.; Fuente-Martin, E.; Diaz, F.; Granado, M.; Argente-Arizon, P.; Frago, L.M.; Freire-Regatillo, A.; Barrios, V.; Argente, J.; Chowen, J.A. The opposing effects of ghrelin on hypothalamic and systemic inflammatory processes are modulated by its acylation status and food intake in male rats. *Endocrinology* **2014**, 155, 2868–2880. [CrossRef] [PubMed]
- 73. Inui, A. Ghrelin: An orexigenic and somatotrophic signal from the stomach. *Nat. Rev. Neurosci.* 2001, 2, 551–560. [CrossRef] [PubMed]
- 74. Chen, H.Y.; Trumbauer, M.E.; Chen, A.S.; Weingarth, D.T.; Adams, J.R.; Frazier, E.G.; Shen, Z.; Marsh, D.J.; Feighner, S.D.; Guan, X.M.; et al. Orexigenic action of peripheral ghrelin is mediated by neuropeptide Y and agouti-related protein. *Endocrinology* **2004**, *145*, 2607–2612. [CrossRef] [PubMed]
- Murphy, M.G.; Plunkett, L.M.; Gertz, B.J.; He, W.; Wittreich, J.; Polvino, W.M.; Clemmons, D.R. MK-677, an orally active growth hormone secretagogue, reverses diet-induced catabolism. *J. Clin. Endocrinol. Metab.* 1998, *83*, 320–325. [CrossRef] [PubMed]
- 76. Nass, R.; Pezzoli, S.S.; Oliveri, M.C.; Patrie, J.T.; Harrell, F.E., Jr.; Clasey, J.L.; Heymsfield, S.B.; Bach, M.A.; Vance, M.L.; Thorner, M.O. Effects of an oral ghrelin mimetic on body composition and clinical outcomes in healthy older adults: a randomized trial. *Ann. Internal Med.* 2008, 149, 601–611. [CrossRef]

- 77. Caberlotto, L.; Lauria, M.; Nguyen, T.P.; Scotti, M. The central role of AMP-kinase and energy homeostasis impairment in Alzheimer's disease: a multifactor network analysis. *PLoS ONE* 2013, *8*, e78919. [CrossRef] [PubMed]
- Bloom, G.S. Amyloid-beta and tau: The trigger and bullet in Alzheimer disease pathogenesis. *JAMA Neurol.* 2014, 71, 505–508. [CrossRef] [PubMed]
- 79. Hardy, J.; Selkoe, D.J. The amyloid hypothesis of Alzheimer's disease: Progress and problems on the road to therapeutics. *Science* **2002**, *297*, 353–356. [CrossRef] [PubMed]
- 80. Stancu, I.C.; Vasconcelos, B.; Terwel, D.; Dewachter, I. Models of β-amyloid induced Tau-pathology: The long and "folded" road to understand the mechanism. *Mol. Neurodegener.* **2014**, *9*, 51. [CrossRef] [PubMed]
- Kanno, T.; Tsuchiya, A.; Nishizaki, T. Hyperphosphorylation of Tau at Ser396 occurs in the much earlier stage than appearance of learning and memory disorders in 5XFAD mice. *Behav. Brain Res.* 2014, 274, 302–306. [CrossRef] [PubMed]
- Maarouf, C.L.; Kokjohn, T.A.; Whiteside, C.M.; Macias, M.P.; Kalback, W.M.; Sabbagh, M.N.; Beach, T.G.; Vassar, R.; Roher, A.E. Molecular Differences and Similarities Between Alzheimer's Disease and the 5XFAD Transgenic Mouse Model of Amyloidosis. *Biochem. Insights* 2013, *6*, 1–10. [CrossRef] [PubMed]
- Yoshida, H.; Hastie, C.J.; McLauchlan, H.; Cohen, P.; Goedert, M. Phosphorylation of microtubule-associated protein tau by isoforms of c-Jun N-terminal kinase (JNK). *J. Neurochem.* 2004, 90(2), 352–358. [CrossRef] [PubMed]
- 84. Lim, E.; Lee, S.; Li, E.; Kim, Y.; Park, S. Ghrelin protects spinal cord motoneurons against chronic glutamate-induced excitotoxicity via ERK1/2 and phosphatidylinositol-3-kinase/Akt/glycogen synthase kinase-3beta pathways. *Exp. Neurol.* **2011**, *230*, 114–122. [CrossRef] [PubMed]
- Li, B.; Zeng, M.; He, W.; Huang, X.; Luo, L.; Zhang, H.; Deng, D.Y. Ghrelin protects alveolar macrophages against lipopolysaccharide-induced apoptosis through growth hormone secretagogue receptor 1a-dependent c-Jun N-terminal kinase and Wnt/beta-catenin signaling and suppresses lung inflammation. *Endocrinology* 2015, 156, 203–217. [CrossRef] [PubMed]
- 86. Liu, C.; Huang, J.; Li, H.; Yang, Z.; Zeng, Y.; Liu, J.; Hao, Y.; Li, R. Ghrelin accelerates wound healing through GHS-R1a-mediated MAPK-NF-kappaB/GR signaling pathways in combined radiation and burn injury in rats. *Sci. Rep.* **2016**, *6*, 27499. [CrossRef] [PubMed]
- Mao, Y.; Wang, J.; Yu, F.; Li, Z.; Li, H.; Guo, C.; Fan, X. Ghrelin protects against palmitic acid or lipopolysaccharide-induced hepatocyte apoptosis through inhibition of MAPKs/iNOS and restoration of Akt/eNOS pathways. *Biomed. Pharmacother.* 2016, *84*, 305–313. [CrossRef] [PubMed]
- Zhang, R.; Yang, G.; Wang, Q.; Guo, F.; Wang, H. Acylated ghrelin protects hippocampal neurons in pilocarpine-induced seizures of immature rats by inhibiting cell apoptosis. *Mol. Biol. Rep.* 2013, 40, 51–58. [CrossRef] [PubMed]
- 89. Chung, H.; Park, S. Ghrelin regulates cell cycle-related gene expression in cultured hippocampal neural stem cells. *J. Endocrinol.* **2016**, 230, 239–250. [CrossRef] [PubMed]
- 90. Zigman, J.M.; Jones, J.E.; Lee, C.E.; Saper, C.B.; Elmquist, J.K. Expression of ghrelin receptor mRNA in the rat and the mouse brain. *J. Comp. Neurol.* **2006**, *494*, 528–548. [CrossRef] [PubMed]
- Li, E.; Kim, Y.; Kim, S.; Sato, T.; Kojima, M.; Park, S. Ghrelin stimulates proliferation, migration and differentiation of neural progenitors from the subventricular zone in the adult mice. *Exp. Neurol.* 2014, 252, 75–84. [CrossRef] [PubMed]
- 92. Council, N.R. *Guide for the Care and Use of Laboratory Animals;* National Academies Press: Washington, DC, USA, 2010.
- 93. Paxinos, G. *Paxinos and Franklin's the Mouse Brain in Stereotaxic Coordinates*; Elsevier/Academic Press: Boston, MA, USA, 2013.
- 94. Jeon, S.G.; Kim, Y.J.; Kim, K.A.; Mook-Jung, I.; Moon, M. Visualization of Altered Hippocampal Connectivity in an Animal Model of Alzheimer's Disease. *Mol. Neurobiol.* **2018**. [CrossRef] [PubMed]



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