

## Chapter C10

# CORONAVIRUS RECEPTORS

Fumihito Taguchi

*National Institute of Neuroscience, NCNP, 4-1-1 Ogawahigashi, Kodaira, Tokyo 187-8502, Japan*

**Abstract:** The major receptor for murine coronavirus, mouse hepatitis virus (MHV), is identified as a protein, cell-adhesion molecule 1 in the carcinoembryonic antigen family (CEACAM1), which is classified in the immunoglobulin superfamily. There are four CEACAM1 isoforms, with either four or two ectodomains, resulting from an alternative splicing mechanism. CEACAM1 is expressed on the epithelium and in endothelial cells of a variety of tissues and hemopoietic cells, and functions as a homophilic and heterophilic adhesion molecule. It is used as a receptor for some bacteria as well. The N terminal domain participates in mediating homophilic adhesion. This domain is also responsible for binding to the MHV spike (S) protein; the CC' face protruding in this domain interacts with an N terminal region of the S protein composed of 330 amino acids (called S1N330). The binding of CEACAM1 with MHV S protein induces S protein conformational changes and converts fusion-negative S protein to a fusion-positive form. The allelic forms of CEACAM1 found among mouse strains are thought to be an important determinant for mouse susceptibility to MHV.

**Key words:** CEACAM1, cell adhesion molecule, carcinoembryonic antigen, mouse hepatitis virus

## 1. INTRODUCTION

The Coronavirus family includes a number of different viruses that infect a variety of animal species, causing numerous diseases, mainly in organs of the enteric, respiratory and central nervous systems. They are

classified into three distinct groups in terms of serological cross-reactivity and sequence homology. Group I consists of porcine transmissible gastroenteritis virus, human coronavirus (HCoV) 229E, feline infectious peritonitis virus and so on. Group II includes mouse hepatitis virus (MHV), HCoV-OC43, bovine coronavirus and some others. Group III is comprised of avian coronaviruses; infectious bronchitis virus and turkey coronavirus. All of these viruses infect animals in a highly species-specific fashion, although some of them can experimentally infect animals different from their natural hosts. The receptor protein for group I viruses has been revealed to be an aminopeptidase N, while the receptor for MHV in group II is a protein classified in the immunoglobulin (Ig) superfamily. The receptors for other viruses in group II, as well as for viruses in group III, have not yet been identified. In this chapter, I describe the receptor for MHV as well as the interaction of MHV receptor and the virus spike (S) protein. The receptors of other coronaviruses can be found in a review article (1).

## 2. RECEPTORS FOR MHV

### 2.1 Discovery of MHV receptor proteins

MHV infects mice, but few other species. The major target organs are the liver, intestine and central nervous system. This host-species specificity or organ tropism of MHV has been thought to be determined mainly by the cellular receptor for MHV. A series of studies carried out by Kay Holmes and her colleagues, which began with analysis of differential susceptibility to MHV infection among mouse strains, has led to the finding of a major MHV receptor molecule. Boyle et al. found that the plasma membranes isolated from MHV-susceptible BALB/c mouse hepatocytes or enterocytes contained a 110 to 120-kDa protein that binds to MHV particles, but those derived from MHV-resistant SJL mice lacked such a protein (2). This finding suggested that the difference in MHV susceptibility among mouse strains is determined by this protein, presumably the MHV receptor. By using monoclonal antibody (MAb) CC-1 specific to 110-120 kDa protein from BALB/c, they purified a protein of ca. 110 kDa and determined the amino acid sequence in its N terminal region, from which the 110-kDa protein was postulated to be a glycoprotein classified in the carcinoembryonic antigen (CEA) family (3, 4). Finally, they isolated a gene encoding this protein, which was revealed to be cell adhesion molecule 1 in the CEA family of the Ig superfamily [formerly called biliary glycoprotein] (Bgp1) and currently termed CEACAM1] (5). MHV non-permissive BHK cells transfected with this gene were converted to MHV-susceptible cells, indicating that this

molecule is the receptor for MHV. It was also found that MHV-resistant SJL mice express a homologous protein (6, 7).

Two other species of glycoprotein, Bgp2 (8) and pregnancy-specific glycoprotein (9), both of which belong to CEA family members, were thereafter found to serve as the MHV receptor in mouse species. However, none of these are as highly efficient as CEACAM1 in terms of receptor functionality or receptor utility by MHV strains. Human CEA glycoprotein works as an MHV receptor as well (10).

## 2.2. Structure and functional regions of major MHV receptor proteins

CEACAM1 is a member of the Ig superfamily and its prototypical 120-kDa glycoprotein consists of four ectodomains (in the order of N, A1, B and A2 from the N terminus), a transmembrane region (TM) and a cytoplasmic tail (Cy) (Fig. 1, 11). The N domain is similar to an Ig-variable domain, and the three other domains resemble a C2 Ig-constant domain. Four different isoforms of CEACAM1 are known to exist, and have been produced by alternative splicing (Fig. 1). Two of the isoforms have 4 ectodomains and the other two have 2 domains, consisting of an N terminal and A2 domains, one of which has either a short or long Cy. The two-domain protein is 48 to 58 kDa in size. CEACAM1 has two allelic forms, CEACAM1<sup>a</sup> and CEACAM1<sup>b</sup> (Fig. 1). The former is expressed in most laboratory mouse strains, while the latter, insofar as is currently known, is expressed only in MHV-resistant SJL mice (12). In wild mice, however, both of those forms are widely distributed (13). The major structural differences between CEACAM1<sup>a</sup> and CEACAM1<sup>b</sup> lie in the N domain, which differs in 29 of its 108 amino acids (6, 7). CEACAM1<sup>a</sup> is 10- to 100-fold higher than CEACAM1<sup>b</sup> in terms of receptor function (14, 15). There is no apparent difference in virus-binding activity as examined by a neutralization test between the 4-ectodomain isoform and the 2-domain CEACAM1<sup>a</sup> (16). However, mice deleting the 4-domain CEACAM1<sup>a</sup> and expressing the 2-domain isoform alone are more resistant to MHV than those expressing both of the 4- and 2-domain isoforms (17). Thus, there could be a difference between them in terms of MHV receptor function in mice. On the contrary, CEACAM1<sup>b</sup> isoform containing 4 domains neutralizes MHV-A59 strain more efficiently than does the isoform containing 2 domains (N and A2 domains), while both of these isoforms showed similar neutralization activity to MHV-JHM strain (16), suggesting a virus-strain specificity in the interaction with CEACAM1.

The N domain is responsible for receptor function (18). Since the CEACAM1 splice variant deleting the A1 and B domains is functional, then it is evident that these domains are not necessary for receptor function. The CEACAM1 isoform containing the N and second A1 domains is also

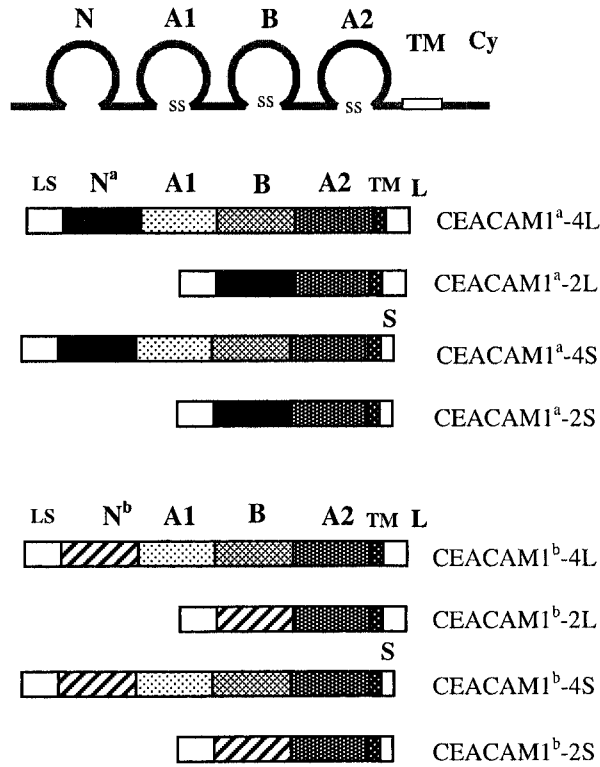


Fig. 1. Schematic structure of a major MHV receptor CECAM1: Four distinct isoforms of CECAM1 exist in each CECAM1<sup>a</sup> and CECAM1<sup>b</sup> allelic forms. There are four or two ectodomains resembling Ig-V or -C domain. Also, long (L) or short (S) cytoplasmic tail (Cy) is known. LS; leader sequence, TM; transmembrane domain.

functional, indicating that the fourth A2 domain is not absolutely critical. Although CECAM1 consisting of N domain alone bound MHV, it did not work as a functional receptor when expressed on CECAM1-negative cells (18). However, the chimeric CECAM1 having the N domain linked to the mouse poliovirus receptor homolog deleting N domain that has a binding specificity to poliovirus served as a functional receptor for MHV (19). As a result of these reports, it is believed that the N domain alone is sufficient for receptor function; however, when expressed on the cell surface, CECAM1 containing the N domain alone was buried among the various molecules expressed on the cell surface because of its shortness and hence it failed to bind to viruses (19). By using a soluble form of

CEACAM1, we have recently found that CEACAM1 with N domain alone converted MHV S protein from a fusion-negative to fusion-positive form (20). Collectively, the N domain is sufficient for the MHV receptor function. Detailed analysis using N domain deletion mutants of CEACAM1 showed that a stretch composed of 19 amino acids (aa34 to 52) in the N domain is particularly important for receptor function (21). The stretch is located in the CC' loop in the N domain composed by  $\beta$ -strands and is supposed to protrude from the N domain (22, 23). The difference in receptor function observed between CEACAM1<sup>a</sup> and CEACAM1<sup>b</sup> results from a 6-amino-acid difference in the above-described 19 amino acids in the N domain (14).

### 2.3. Distribution and biological function of CEACAM1

CEACAM1 is reported to be distributed in various cells in different organs, not only in the target organs of MHV, but also in those in which MHV infection has not been detected. A high level of CEACAM1 expression is reported on epithelium and endothelial cells of a variety of tissues, and in hemopoietic cells, such as monocytes, macrophages, granulocytes and their precursors, the B cells, activated T cells and thymic stromal cells (11, 24). It is also demonstrated on both apical membranes of epithelial cells as well as on sites of cell-cell contact (e.g., between hepatocytes, stratified epithelia, junctional epithelium that forms a transition zone between gingival epithelium and teeth, and between pericytes and endothelial cells of blood vessel walls). Furthermore, during early mouse embryonic development, CEACAM1 is abundantly expressed in endodermal and mesenchymal derivatives (25), but is not detected by immune histochemistry in any type of cells in the brain (26). However, MHV infection in the brain was blocked by anti-CEACAM1 MAb CC-1 (27). Also, a CEACAM1 isoform with 2 domains was detected by RT-PCR in the brain (28). These findings suggest that the CEACAM1 molecule is expressed, albeit in very small amounts, in some cell populations of the brain.

The major biological function of CEACAM1 is cell adhesion. It serves as both a homophilic and heterophilic adhesion molecule (11). Homophilic adhesion, confirmed by *in vitro* studies of rodent and human CEACAM1, is thought to be important in the embryonic organization of the intestinal epithelium and liver hepatocytes, in placental trophoblasts, during muscle and tooth development and vascularization of the central nervous system (24). CEACAM1 also plays an important role in neutrophil activation and adhesion during inflammatory responses (29), lymphoregulation and immunosurveillance (30), angiogenesis (31), and the negative regulation of cell proliferation (11, 24). Heterophilic adhesion of CEACAM1 to other

CEACAM family members has been shown (32). Also, heterophilic adhesion to *Opa* proteins of *Neisseria gonorrhoeae*, *Neisseria meningitidis* and *Haemophilus influenzae* mediates their infections (33, 34), indicating that CEACAM1 is a receptor for those bacteria. This also facilitates bacterial colonization of the gut and bacterial phagocytosis by neutrophils and is involved in the initial tethering of granulocytes to E-selectin on the endothelium prior to their transendothelial migration during inflammatory responses. It was recently shown that homophilic adhesion of CEACAM1 involves N-terminal domain interactions. The GFCC' face of the N domain, which includes the MHV binding site CC' region, is responsible for homophilic interaction (22).

### 3. INTERACTION OF CEACAM1 AND MHV S PROTEIN

The MHV S protein comprising a petal-like projection on virion surface is the ligand for the CEACAM1 molecule. The projection is composed of two or three molecules of the S1-S2 heterodimer derived from S protein. S protein is a type I glycoprotein. It is synthesized and cotranslationally glycosylated as a 150-kDa protein, becomes as 180-200 kDa protein after modification of glycans and cleaved by a host-derived proteinase into two subunits, N terminal S1 and C terminal S2 (35). S1 comprises an outer knob-like structure of the spike, and S2 consists of the stem-like part beneath the knob (36). S1 and S2 units are associated by non-covalent linkage, and they can be easily dissociated from each other by denaturing reagents or even during a purification process. Alpha-helices constructed by the heptad repeats in the S2 play an important role for oligomerization of S1-S2 heterodimers, though there is another determinant in the S1 for oligomerization (37). Following its synthesis, the S protein is incorporated into the envelope of viral particles after interaction with viral integral membrane protein in the internal compartments from endoplasmic reticulum to the Golgi apparatus. The S protein is also transported to the plasma membrane.

The N terminal region in the S1 composed of 330 amino acids (S1N330) is responsible for binding to CEACAM1 (38). Among S proteins of a variety of MHV strains, there are three conserved regions in S1N330 (S1N330-I, -II and -III) composed of 10 or more identical amino-acid stretches (39). Site-directed mutagenesis analysis suggested that two of these regions, S1N330-I and -II, located far from one another, are involved in receptor binding (39). Studies using MHV variants containing mutations in S1N330-I confirmed the significance of this region in receptor binding (40). S1N330-III was recently suggested to be responsible for

virus entry into the cell in combination with a region in the S2 (41). Denaturing of S1N330 abolished the receptor-binding activity, indicating that the tertiary structure composed of different regions in the S1N330 or/and the dimerization of S1 which takes place within S1N330 (37) is important.

The interaction of MHV S protein and CEACAM1 leads to the S protein functional conversion from a fusion-negative to a fusion-positive form (42). Recently, it was also shown that this functional activation is accompanied with conformational changes in the S protein; receptor-bound S protein has a fraction resistant to proteinase digestion, while receptor-unbound S protein is susceptible (43). These functional and structural changes of the S protein greatly resemble those of the envelope protein of retroviruses that take place after they bind to their receptors (44, 45), suggesting that MHV enters cells in a fashion similar to that of retroviruses.

#### 4. MHV RECEPTOR AND ITS IMPLICATIONS FOR MOUSE SUSCEPTIBILITY TO MHV

A number of investigators have reported that BALB/c, C57BL and most other mouse strains, later revealed to have a *Ceacam1<sup>a</sup>* (*I<sup>a</sup>*) gene, are susceptible, while SJL mice with a *Ceacam1<sup>b</sup>* (*I<sup>b</sup>*) gene are resistant (46, 47). Genetic analyses indicated that a single dominant gene located on chromosome 7 is responsible for susceptibility to MHV (47). The *Ceacam1* could be a gene determining susceptibility, since 1) the *I<sup>a</sup>* makes mice susceptible and 2) *Ceacam1* is also mapped on chromosome 7 (48). The expression of either *I<sup>a</sup>* or *I<sup>b</sup>* in CEACAM1-negative cells converted them to MHV susceptible, suggesting that allelic differences in receptor proteins were not sufficient to explain the differences in mouse susceptibility to MHV (6, 7). In detailed studies, however, cells transiently expressing *I<sup>a</sup>* were 10 to 100 times more sensitive to MHV than were cells expressing *I<sup>b</sup>* (14, 15), indicating a small, but significant difference between *I<sup>a</sup>* and *I<sup>b</sup>*. This also suggested that the MHV receptor expressed in SJL is still functional. If the receptor allele controls MHV susceptibility, then SJL should be relatively, but not completely, resistant to MHV. SJL mice are, in fact, resistant to MHV when challenged with a low dose of virus, but susceptible when inoculated with a high dose of virus (49, 50). These observations suggest that *Ceacam1* is a gene controlling MHV susceptibility. Of 120 mice of (BALB/c x SJL) F2 and backcrossed mice to SJL, all mice with *I<sup>a</sup>/I<sup>a</sup>* and *I<sup>a</sup>/I<sup>b</sup>* genotypes were susceptible, while all mice with *I<sup>b</sup>/I<sup>b</sup>* genotype were resistant after infection with a low dose of virus (12). This implies the MHV receptor gene and MHV-susceptibility

gene are identical, and if not, they are located within 0.86 cM on chromosome 7. To finally examine whether the receptor gene is identical to the gene that controls MHV susceptibility, gene replacement is a useful strategy. The MHV susceptibility of BALB/c mice in which  $I^a/I^a$  is replaced by  $I^b/I^b$  and SJL mice in which  $I^b/I^b$  is replaced with  $I^a/I^a$  will conclusively establish whether the MHV receptor gene is the gene which controls the MHV susceptibility of mice.

## 5. REFERENCES

- 1) Holmes, K. Coronavirus receptors in "The coronaviridae" ed. by Siddell S. pp55-71 1995 Plenum press.
- 2) Boyle J.F., Weismiller DG., Holmes KV. Genetic resistance to mouse hepatitis virus correlates with absence of virus-binding activity on target tissues. *J. Virol.* 1987;61: 185-89.
- 3) Williams R., Jiang GS, Snyder SW, Frana MF, Holmes KV. Purification of the 110-kilodalton glycoprotein receptor for mouse hepatitis virus (MHV)-A59 from mouse liver and identification of a nonfunctional, homologous protein in MHV-resistant SJL/J mice. *J Virol.* 1990; 64: 3817-23
- 4) Williams RK, Jiang GS, Holmes KV. Receptor for mouse hepatitis virus is a member of the carcinoembryonic antigen family of glycoproteins. *Proc Natl Acad Sci U S A.*1991; 88: 5533-36.
- 5) Dveksler GS, Pensiero MN, Cardellichio CB, Williams RK, Jiang GS, Holmes KV. et al. Cloning of the mouse hepatitis virus (MHV) receptor: expression in human and hamster cell lines confers susceptibility to MHV. *J Virol.* 1991; 65: 6881-91.
- 6) Dveksler GS, Dieffenbach CW, Cardellichio CB, McCuaig K, Pensiero MN, Jiang GS. et al. Several members of the mouse carcinoembryonic antigen-related glycoprotein family are functional receptors for the coronavirus mouse hepatitis virus-A59. *J Virol.* 1993; 67: 1-8.
- 7) Yokomori K, Lai MM. The receptor for mouse hepatitis virus in the resistant mouse strain SJL is functional: implications for the requirement of a second factor for viral infection. *J Virol.* 1992;66: 6931-38.
- 8) Nedellec P, Dveksler GS, Daniels E, Turbide C, Chow B, Basile AA, et al. Bgp2, a new member of the carcinoembryonic antigen-related gene family, encodes an alternative receptor for mouse hepatitis viruses. *J Virol.*1994; 68: 4525-37.
- 9) Chen DS, Asanaka M, Yokomori K, Wang F, Hwang SB, Li HP, et al.. A pregnancy-specific glycoprotein is expressed in the brain and serves as a receptor for mouse hepatitis virus. *Proc Natl Acad Sci U S A.* 1995; 92: 12095-99.
- 10) Chen DS, Asanaka M, Chen FS, Shively JE, Lai MM. Human carcinoembryonic antigen and biliary glycoprotein can serve as mouse hepatitis virus receptors. *J Virol.* 1997; 71: 1688-91.



- 11) Beauchemin N, Draber P, Dveksler G, Gold P, Gray-Owen S, Grunert F. et al. Redefined nomenclature for members of the carcinoembryonic antigen family. *Exp Cell Res.* 1999; 252: 243-49.
- 12) Ohtsuka N, Taguchi F. Mouse susceptibility to mouse hepatitis virus infection is linked to viral receptor genotype. *J Virol.* 1997; 71: 8860-63.
- 13) Ohtsuka N, Tsuchiya K, Honda E, Taguchi F. A study on mouse hepatitis virus receptor genotype in the wild mouse. *Adv. Exp. Med. Biol.* 2001; 494: 237-40
- 14) Rao PV, Kumari S, Gallagher TM. Identification of a contiguous 6-residue determinant in the MHV receptor that controls the level of virion binding to cells. *Virology.* 1997; 229: 336-48.
- 15) Ohtsuka N, Yamada YK, Taguchi F. Difference in virus-binding activity of two distinct receptor proteins for mouse hepatitis virus. *J Gen Virol.* 1996; 77: 1683-92.
- 16) Zelus BD, Wessner DR, Williams RK, Pensiero MN, Phibbs FT, de Souza M, et al. Purified, soluble recombinant mouse hepatitis virus receptor, Bgp1(b), and Bgp2 murine coronavirus receptors differ in mouse hepatitis virus binding and neutralizing activities. *J Virol.* 1998; 72: 7237-44.
- 17) Blau D., Turbide C, Tremblay M, Olson M, Letourneau S, Michaliszyn E, et al. Targeted disruption of the Ceacam1 (MHVR) gene leads to reduced susceptibility of mice to mouse hepatitis virus infection. *J Virol.* 2001; 75: 8173-86.
- 18) Dveksler GS, Pensiero MN, Dieffenbach CW, Cardellichio CB, Basile AA, Elia P.E, et al. Mouse hepatitis virus strain A59 and blocking antireceptor monoclonal antibody bind to the N-terminal domain of cellular receptor. *Proc Natl Acad Sci U S A.* 1993; 90: 1716-20.
- 19) Dveksler GS, Basile AA, Cardellichio CB, Holmes KV. Mouse hepatitis virus receptor activities of an MHVR/MPH chimera and MHVR mutants lacking N-linked glycosylation of the N-terminal domain. *J. Virol.* 1995; 69: 543-46
- 20) Miura H, Taguchi F. (unpublished observation)
- 21) Wessner DR, Shick PC, Lu JH, Cardellichio CB, Gagneten SE, Beauchemin N, et al. Mutational analysis of the virus and monoclonal antibody binding sites in MHVR, the cellular receptor of the murine coronavirus mouse hepatitis virus strain A59. *J Virol.* 1998; 72: 1941-48.
- 22) Watt SM, Teixeira AM, Zou GQ, Doyonnas R, Zhang Y, Grunert F, et al. Homophilic adhesion of human CEACAM1 involves N-terminal domain interactions: structural analysis of the binding site. *Blood.* 2002; 98: 1469-79
- 23) Tan K, Zelus BD, Meijers R, Liu J, Berfelson J, Duke N. et al. Crystal structure of murine sCEACAM1a [1,4]: a coronavirus receptor in the CEA family. *EMBO J.* 2002; 21: 2076-86

- 24) Obrink B. CEA adhesion molecules: multifunctional proteins with signal-regulatory properties. *Curr Opin Cell Biol.* 1997; 9: 616-26.
- 25) Daniels ., Letourneau S, Turbide C, Kuprina N., Rudinskaya T., Yazova AC, et al. Biliary glycoprotein 1 expression during embryogenesis: correlation with events of epithelial differentiation, mesenchymal-epithelial interactions, absorption, and myogenesis. *Dev Dyn.* 1996; 206: 272-90.
- 26) Godfraind C, Langreth SG, Cardellichio CB, Knobler R, Coutelier JP, Dubois-Dalcq M, et al. Tissue and cellular distribution of an adhesion molecule in the carcinoembryonic antigen family that serves as a receptor for mouse hepatitis virus. *Lab Invest.* 1995; 73: 615-27
- 27) Smith A, Cardellichio CB, Winograd DF, de Souza MS, Barthold SW, Holmes KV. Monoclonal antibody to the receptor for murine coronavirus MHV-A59 inhibits viral replication in vivo. *J Infect. Dis.* 1991;163: 879-82
- 28) Yokomori K, Lai MM. Mouse hepatitis virus utilizes two carcinoembryonic antigens as alternative receptors. *J Virol.* 1992; 66: 6194-99.
- 29) Skubitz KM, Campbell KD, Skubitz APN, CD66a, CD66b, CD66c and CD66d each independently stimulate neutrophils. *J. Leukoc. Biol.* 1996; 60: 106-17
- 30) Morale V, Christ A, Watt SM, Kim HS Johnson KW, Utku N. et al. Regulation of human intestinal intraepithelial lymphocyte cytolytic function by biliary glycoprotein (CD66a). *J. Immunol.* 1999; 163: 1363-70
- 31) Ergun S, Kilik N, Ziegeler G, Hansen A, Nollau P, Gotze J, et al. CEA-related cell adhesion molecule 1: a potent angiogenic factor and a major effector of vascular endothelial growth factor. *Mol Cell.* 2000; 5: 311-20.
- 32) Stocks SC, Kerr MA, Haslett C, Dransfield I. CD66-dependent neutrophil activation: a possible mechanism for vascular selectin-mediated regulation of neutrophil adhesion. *J. Leukoc. Biol* 1995; 58: 40-8
- 33) Virji M, Watt S., Barker ., Makepeace K, Doyonnas R. The N-domain of the human CD66a adhesion molecule is a target for Opa proteins of *Neisseria meningitidis* and *Neisseria gonorrhoeae*. *Mol Microbiol.* 1996; 22: 929-39.
- 34) Virji M, Evans D, Griffith J, Hill D., Serino L, Hadfield A. et al. Carcinoembryonic antigens are targeted by diverse strains of typable and non-typable *Haemophilus influenzae*. *Mol. Microbiol.* 2000;36: 784-95
- 35) Sturman LS, Ricard CS, Holmes KV. Proteolytic cleavage of the E2 glycoprotein of murine coronavirus: Activation of cell fusing activity of virions by trypsin and separation of two different 90K cleavage fragments. *J. Virol.* 1985; 56: 904-11
- 36) De Groot RJ, W. Luytjes MC, Horzinek BAM, .Van der Zeijst S, SpaanWJM, Lenstra JA. Evidence for a coiled-coil structure in the spike of coronaviruses. *J. Mol. Biol.* 1987;196: 963-66
- 37) Lewicki D, Gallagher T Quaternary structure of coronavirus spikes in complex with CEACAM cellular receptors. *J. Boil. Chem.* 2002; 277:19727-34

- 38) Kubo H, Yamada YK, Taguchi F. Localization of neutralizing epitopes and the receptor-binding site within the amino-terminal 330 amino acids of the murine coronavirus spike protein. *J Virol.* 1994; 68: 5403-10.
- 39) Suzuki H, Taguchi F. Analysis of the receptor-binding site of murine coronavirus spike protein. *J Virol.* 1996; 70: 2632-36.
- 40) Saeki K, Ohtsuka N, Taguchi F. Identification of spike protein residues of murine coronavirus responsible for receptor-binding activity by use of soluble receptor-resistant mutants. *J Virol.* 1987; 71: 9024-31.
- 41) Matsuyama S, Taguchi F. Communication between S1N330 and a region in S2 of murine coronavirus spike protein is important for virus entry into cells expressing CEACAM1b receptor. *Virology* 2002; 295: 160-71
- 42) Taguchi F, Matsuyama S. Soluble receptor potentiates receptor-independent infection by murine coronavirus. *J. Virol.* 2002; 76: 950-58
- 43) Matsuyama S, Taguchi F. Receptor-induced conformational changes of murine coronavirus spike protein. *J. Virol.* 2002;76: in press.
- 44) Damico R, Bates P. Soluble receptor -induced retraviral infection of receptor-deficient cells. *J. Virol.* 2000; 74: 6469-75.
- 45) Chen DC, Kim PS. HIV entry and its inhibition. *Cell*, 1998;29: 681-84
- 46) Stohlman SA, Frelinger JA. Resistance to fatal central nervous system disease by mouse hepatitis virus, strain JHM. 1. Genetic analysis. *Immunogenet.* 1978; 6: 277-81
- 47) Smith MS, Click RE, Plagemann GW. Control of mouse hepatitis virus replication in macrophages by a recessive gene on chromosome 7. *J. Immunol.* 1984; 134: 428-32
- 48) Mouse genome database
- 49) Knobler RL, Haspel MV, Oldstone MAB. Mouse hepatitis virus type 4 (JHM strain)-induced fatal central nervous system disease. 1. Genetic control and the murine neuron as the susceptible site of disease. *J. Exp. Med.* 1981; 153: 832-43
- 50) Taguchi F. (unpublished observation)