

REVIEW

Etiology and Pathophysiology

Branched-chain amino acid metabolism: Pathophysiological mechanism and therapeutic intervention in metabolic diseases

Shama Mansoori¹ | Melody Yuen-man Ho¹ | Kelvin Kwun-wang Ng¹ |
Kenneth King-yip Cheng^{1,2} 

¹Department of Health Technology and Informatics, The Hong Kong Polytechnic University, Hong Kong, China

²Hong Kong Polytechnic University Shenzhen Research Institute, Shenzhen, China

Correspondence

Dr. Kenneth K.Y. Cheng, Department of Health Technology and Informatics, The Hong Kong Polytechnic University, Hong Kong, China.

Email: kenneth.ky.cheng@polyu.edu.hk

Funding information

PolyU- Project of Strategic Importance, Grant/Award Number: P0036848; National Natural Science Foundation of China, Grant/Award Numbers: 81970675, 92357305; Research Grant Council Collaborative Research Fund, Grant/Award Number: C5044-23G; General Research Fund (GRF), Hong Kong Research Grant Council (RGC), Grant/Award Number: 15101221; Shenzhen Municipal Science and Technology Innovation Commission, Grant/Award Number: JCYJ20210324130202006; PolyU-Project of RCMI, Grant/Award Number: P0040979

Summary

Branched-chain amino acids (BCAAs), including leucine, isoleucine, and valine, are essential for maintaining physiological functions and metabolic homeostasis. However, chronic elevation of BCAAs causes metabolic diseases such as obesity, type 2 diabetes (T2D), and metabolic-associated fatty liver disease (MAFLD). Adipose tissue, skeletal muscle, and the liver are the three major metabolic tissues not only responsible for controlling glucose, lipid, and energy balance but also for maintaining BCAA homeostasis. Under obese and diabetic conditions, different pathogenic factors like pro-inflammatory cytokines, lipotoxicity, and reduction of adiponectin and peroxisome proliferator-activated receptors γ (PPAR γ) disrupt BCAA metabolism, leading to excessive accumulation of BCAAs and their downstream metabolites in metabolic tissues and circulation. Mechanistically, BCAAs and/or their downstream metabolites, such as branched-chain ketoacids (BCKAs) and 3-hydroxyisobutyrate (3-HIB), impair insulin signaling, inhibit adipogenesis, induce inflammatory responses, and cause lipotoxicity in the metabolic tissues, resulting in multiple metabolic disorders. In this review, we summarize the latest studies on the metabolic regulation of BCAA homeostasis by the three major metabolic tissues—adipose tissue, skeletal muscle, and liver—and how dysregulated BCAA metabolism affects glucose, lipid, and energy balance in these active metabolic tissues. We also summarize therapeutic approaches to restore normal BCAA metabolism as a treatment for metabolic diseases.

KEYWORDS

adipose tissue, BCAAs, diabetes and obesity, liver, metabolic tissues, skeletal muscle

1 | INTRODUCTION

Branched-chain amino acids (BCAAs), leucine, isoleucine, and valine are essential amino acids and constitute approximately 20% of the incorporated amino acids into proteins.^{1,2} BCAAs perform a plethora

of functions including lipid metabolism, gluconeogenesis, and serving as nitrogen donors in different biological processes. Their actions mainly involve several key metabolic pathways such as the mammalian target of rapamycin complex 1 (mTORC1), AMP-activated protein kinase (AMPK), and general control non-derepressible (GCN) 2, as well

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial-NoDerivs](https://creativecommons.org/licenses/by-nc-nd/4.0/) License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2024 The Author(s). *Obesity Reviews* published by John Wiley & Sons Ltd on behalf of World Obesity Federation.

as insulin signaling.³⁻⁷ Despite their essential roles in maintaining physiological functions, chronic elevations of BCAAs and/or their downstream metabolites are associated with a higher risk of cardio-metabolic diseases including type 2 diabetes (T2D), heart attack, stroke, and non-alcoholic fatty liver disease (NAFLD; also now known as metabolic-associated fatty liver diseases [MAFLD]).^{3,8-12} The pathogenic events of these cardiometabolic diseases are intertwined and some of them are overlapped, which include insulin resistance, chronic sterile low-grade inflammation, aberrant metabolic and lipid pathways, etc. Interestingly, BCAAs and their downstream metabolites are known to participate in and exacerbate these pathogenic events in metabolic diseases. For instance, increased levels of BCAAs and/or their immediate catabolic metabolites, such as branched-chain keto acids (BCKAs), or incomplete oxidation of BCAA-derived metabolites (such as 3-hydroxyisobutyrate) can induce insulin resistance, inflammation, and lipotoxicity in the muscle and liver,^{12,13} as well as enhance lipolysis in adipocytes.¹⁴ In contrast, restriction of BCAA consumption improves glucose metabolism, reduces fat mass, and extends lifespan in C57BL/6 J natural aging and obese rodents.¹⁵⁻¹⁷ The beneficial effects of BCAA restriction on glucose metabolism are also observed in humans and rodents with obesity and diabetes.¹⁸⁻²⁰ Therefore, BCAAs and their downstream metabolites not only serve as biomarkers but also as mediators of metabolic diseases and thus represent important therapeutic targets.

In this review, we discuss how BCAAs are metabolized in the major metabolic organs, and how their dysmetabolism contributes to obesity and its associated metabolic diseases. Furthermore, we will summarize the strategies for modifying BCAA metabolism and their therapeutic efficacy against the metabolic diseases that afflict millions of people worldwide.

2 | OVERVIEW OF BCAA METABOLISM

BCAAs represent three out of nine essential amino acids. In the human diet, 20% of total protein intake is derived from BCAAs.^{1,2} Contrary to the oxidation of other amino acids that occurs primarily in the liver, BCAAs uptake, and oxidation take place differentially in numerous tissues because of the low branched-chain amino acid transaminase (BCAT) activity in hepatocytes.²¹ However, due to the presence of BCAT in the non-parenchymal liver cells, some transamination may also happen in the liver.²²

The primary sites for BCAA metabolism include skeletal muscle, adipose tissue, liver, kidney, pancreas, and heart.²³ Once inside the cells, BCAAs can be used for protein synthesis, mitochondrial oxidation, and adding reserve to the amino acid pool.² Additionally, BCAAs can act as key nitrogen donors in the brain for the production of neurotransmitters such as aminobutyric acid (GABA) and glutamate.⁴ BCAAs can also be utilized as the biosynthetic precursors of sterol and glucose.

The BCAA catabolism initiates with the reversible transamination of BCAAs into their respective branched-chain ketoacids (BCKAs) by

BCAT1/2. Global disruption of *Bcat2* but not *Bcat1* leads to excessive accumulation of BCAAs and BCKAs in the mouse model.²⁴ BCKAs are decarboxylated by the multi-subunit enzyme complex alpha-keto acid dehydrogenase (BCKDH) and generate the corresponding acyl-CoA derivatives and nicotinamide adenine dinucleotide (NADH).²⁵ The acyl-CoA derivatives undergo a series of biochemical reactions to produce acetyl-CoA and succinyl-CoA and eventually integrate into the tricarboxylic acid (TCA) cycle (Figure 1).^{23,26} The end products, acetyl-CoA and succinyl-CoA, can serve as substrates for anaplerotic reactions, and acetyl-CoA can also be utilized for lipogenesis in some cell types.

The BCKDH complex is a rate-limiting enzyme and comprises three subunits, including E1, E2, and E3.²⁷ The activity of BCKDH is regulated by post-translational modification, i.e., phosphorylation and dephosphorylation at serine 293 (Ser²⁹³) on the E1 α subunit, positively by the protein phosphatase 1 K, mitochondrial (PPM1K)²⁸ and negatively by the branched-chain ketoacid dehydrogenase kinase (BCKDK).²⁹ Under low BCAA conditions, the BCKDH complex is phosphorylated and inactivated by BCKDK, whereas an increase in the BCAAs/BCKAs ratio allows the activation of BCKDHA by PPM1K-mediated dephosphorylation at Ser²⁹³.²⁹

The pioneer in-vivo ¹³C-isotope study shed light on the quantitative framework of whole-body BCAA disposal in mouse model.²³ Quantitative analysis revealed that skeletal muscle (59%), adipose tissue (19%), and liver (8%) are mainly responsible for BCAA catabolism. On the other hand, the liver, and skeletal muscle both contribute largely to BCAA disposal into proteins.²³ Different conditions can alter the distribution of BCAA oxidation among different tissues. Insulin shifts BCAA oxidation more toward skeletal muscles. Under diabetic conditions, a shift of BCAA oxidation from adipose tissue and liver to skeletal muscle was observed as evidenced by a reduction in the contribution of ¹³C-valine to succinate by ~60% and ~20% in adipose tissues and liver, respectively.²³ This increase of BCAA catabolism in skeletal muscle may be due to attenuated oxidation in adipose tissue and liver, given that muscle has a higher capacity for BCAA oxidation. These findings suggest inter-tissue crosstalk under diabetic conditions, leading to a relocation of BCAA oxidation to tissues with higher efficiency (muscle), to adapt to the metabolic state. A recent BCAA flux study showed that BCAA-derived nitrogen is important in synthesizing non-essential amino acids such as alanine, glutamate, proline, and other metabolites including N-acetyl-Glu, N-acetyl-Asp, and glutathione in BAT via the mitochondrial BCAA carrier (MBC) dependent manner.³⁰ Interestingly, mitochondrial dysfunction-induced blockage of BCAA-derived metabolites generation in BAT led to higher oxidative stress and attenuated insulin signaling in the liver.³⁰ Again, this study indicates the crosstalk between BAT and the liver through BCAA metabolism. Despite these promising data, further exploration of BCAA flux among the metabolic tissues, longitudinal studies on BCAA metabolism in metabolic disease progression, and investigation into spatial and cell-specific BCAA metabolism in key metabolic tissues would deepen our understanding of the interaction/proportional contribution of metabolic tissues in control of BCAA homeostasis.

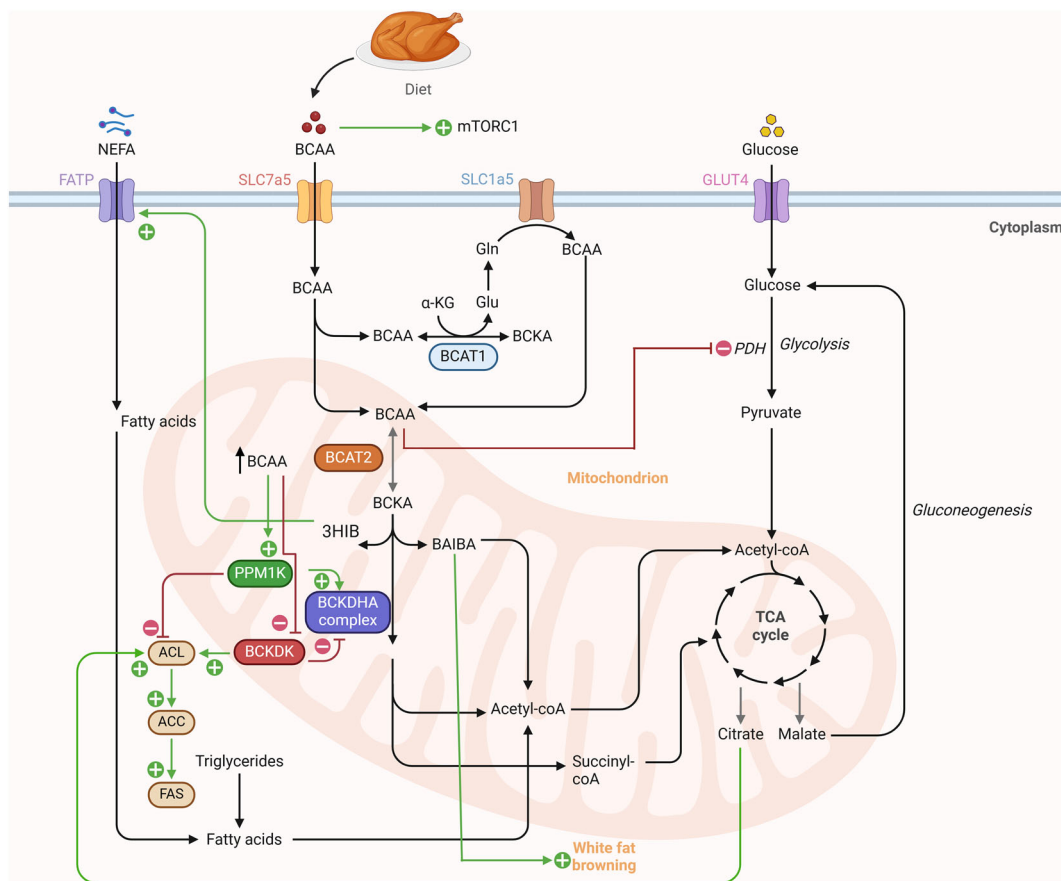


FIGURE 1 Overview of BCAA catabolism and crosstalk with different metabolic pathways. BCAAs are catabolized via multistep reactions. The rate-limiting enzyme BCKDHA is positively and negatively regulated by PPM1K and BCKDK, respectively. BCAA-derived acetyl- and succinyl-CoA can enter the TCA cycle to generate ATP or anaplerosis. BCAAs can affect glucose oxidation by inhibiting the pyruvate dehydrogenase complex. The regulatory enzymes PPM1K and BCKDK also regulate the ATP citrate lyase (involved in lipogenesis) in the liver. Metabolites 3-HIB can alter the fatty acid transport in the endothelial cells of skeletal muscle. Adapted from “Overview of BCAA Catabolic Enzymes”, by BioRender.com (2024). Retrieved from <https://app.biorender.com/biorender-templates>.

3 | ASSOCIATION STUDIES OF BCAAS AND THEIR DOWNSTREAM METABOLITES WITH METABOLIC DISEASES

BCAAs can serve as a predictive risk biomarker for a variety of disorders, including obesity, T2D,^{8,31} and CVD.^{9,10} Accumulating evidence shows a positive association between plasma BCAA levels and obesity/T2D in human subjects from different ethnic groups and various diabetic rodent models, including diet-induced obesity (DIO) rats, *ob/ob* mice, and Zucker rats.^{32–34} The dietary BCAAs intake is also shown to be associated with the risk of developing dyslipidemia in the Chinese population.³⁵ A positive association has been reported between dietary isoleucine and body mass index (BMI) in humans.³⁶ In addition, single nucleotide polymorphisms (SNPs) in different BCAA catabolic genes, including *PPM1K*, *BCAT2*, and *BCKDHA*, are linked with higher serum BCAAs and increased risk of metabolic diseases.^{37–40} On the other hand, the genetic variants linked with insulin resistance are strongly associated with higher circulating BCAA levels in humans.^{41,42} Plasma BCKA levels are also reported to be associated with obesity/T2D and nonalcoholic steatohepatitis (NASH).^{11,32,43} The circulatory level

of 3-HIB (a catabolic intermediate of valine) is positively associated with the incidence of T2D and insulin resistance in human subjects in the EPIC-Norfolk study,^{12,44} and insulin levels and HOMA-IR in the CARBFUNC cohort.⁴⁵ Altogether, human studies show a positive association between BCAAs and their downstream metabolites with obesity/T2D and MAFLD.

4 | THE UNDERLYING CAUSES OF DEFECTIVE BCAA METABOLISM IN OBESITY/T2D

In this section, we will further discuss how aberrant BCAA catabolism in the metabolic tissues contributes to the chronic elevation of BCAAs in metabolic diseases.

The proteins facilitating BCAA catabolism, such as BCKDHA and BCAT2, are reduced in adipose tissue, skeletal muscles, and the liver under obese or diabetic conditions, whereas the negative regulators of BCAA catabolism (such as BCKDK and phospho-BCKDHA at ser 293) are induced, suggesting the blockage of BCAA catabolism.^{46–49}

In addition, the increased level of BCAAs during obesity is partly because of the BCAA catabolic rate drop in adipose tissue and the liver.^{49,50} Further investigation is warranted into monitoring BCAA flux and their metabolic fates (such as to the TCA cycle or protein synthesis) in the metabolic tissues during the development and progression of metabolic diseases.

5 | BCAA METABOLISM IN DIFFERENT TISSUES UNDER PHYSIOLOGICAL AND PATHOPHYSIOLOGICAL CONDITIONS

In the following sections, we will discuss how adipose tissue, skeletal muscle, and liver maintain BCAA homeostasis, and how their aberrant metabolism causes obesity and its related metabolic disorders.

5.1 | BCAA metabolism in the adipose tissue

In addition to lipids, recent studies showed white adipose tissue (WAT)/BAT also plays a crucial role in maintaining multiple nutrient homeostasis, including glucose, retinol, amino acids, irons, etc.⁵¹ WAT acts as a caloric reservoir to store energy in the form of triglycerides

via lipogenesis in the feeding state and releases energy as free fatty acids via lipolysis,⁵² whereas BAT is responsible for adaptive thermogenesis mainly via the action of uncoupling protein 1 (UCP1).

In vitro and ex vivo studies indicated adipose tissue can metabolize a large amount of BCAAs.^{50,53} The uptake of BCAAs into adipocytes is mediated by the transporters Slc1a5, Slc3a2, and Slc7a5.^{54,55} The direct transportation of BCAAs is facilitated by the Slc1a5 transporter. Simultaneously, a heterodimeric plasma membrane complex, formed by Slc3a2 and Slc7a5, allows for the exchange of internal glutamine and asparagine for external BCAAs.^{55,56} The BCAA intermediate metabolites such as isovaleryl-CoA and 2-methylbutyryl-CoA (acyl-CoA) can be utilized to synthesize monomethyl branched-chain fatty acids (mmBCFA). The acyl-CoA precursors are transported from the mitochondria to the cytosol by carnitine acetyltransferase and elongated by the enzyme fatty acid synthase (FASN) in adipose tissue.⁵⁷ Thus, mmBCFA serves as a substrate for de novo lipogenesis in adipose tissue (Figure 2).⁵⁷ In 2016, Green et al, reported that BCAA catabolism is important for adipogenesis, as evidenced by the association between upregulated levels of BCAA catabolic enzymes and peroxisome proliferator-activated receptors (PPAR) γ during early adipogenesis.⁵⁸ BCAAs promote adipocyte differentiation via the regulation of the mTORC1/ribosomal protein S6 kinase (S6K1) and mTORC1/eukaryotic mRNA translation initiation factor 4E-binding

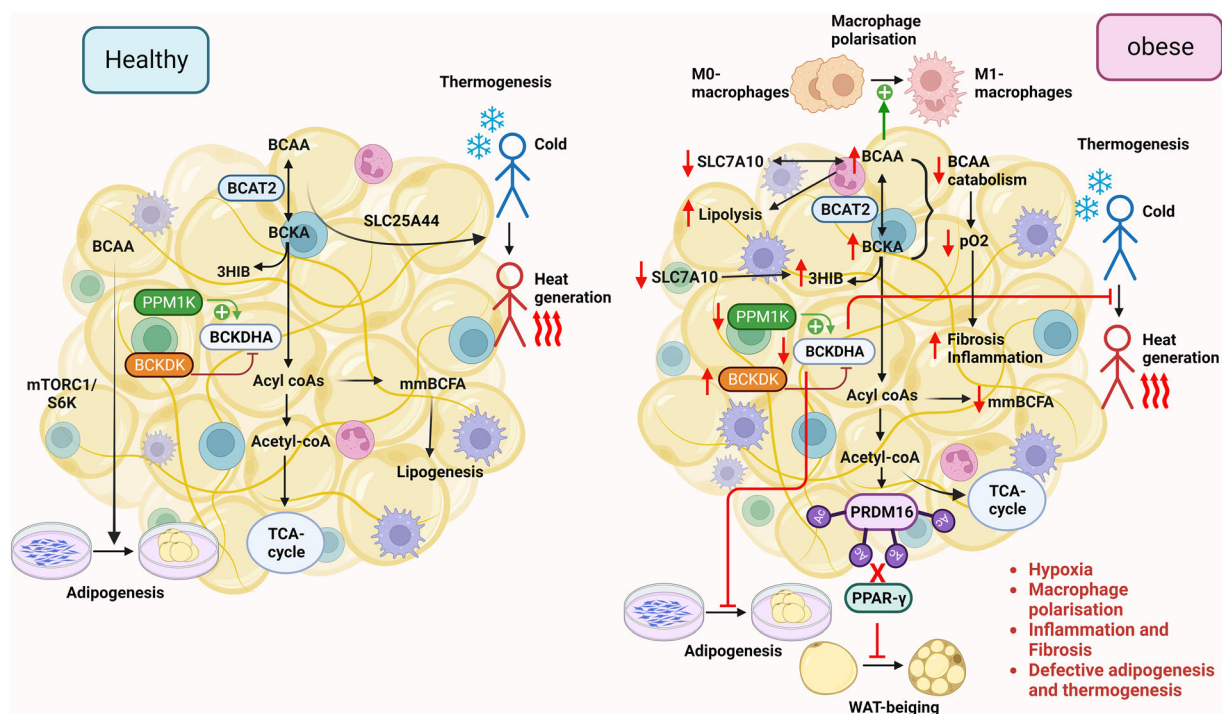


FIGURE 2 BCAA catabolism in adipose tissue under physiological and pathophysiological conditions. Under physiological conditions, BCAAs are a substrate for adipogenesis (by activating mTORC1/S6K1 axis), lipogenesis (via the production of mmBCFA), and thermogenesis. While under pathological conditions, a higher level of BCAAs and their associated metabolites affects different processes in adipose tissue. Elevated BCKA levels suppress the WAT-beiging by inhibiting the binding of PRDM16 to PPAR γ , and promote macrophage polarization. The knockdown of BCKDH impairs adipogenesis and thermogenesis. A reduced rate of BCAA catabolism is also linked with hypoxia and adipose tissue fibrosis. Additionally, the blockage of BCAA transporter SLC7A10 leads to higher production of 3-HIB in adipocytes, and an inverse correlation has been observed in sWAT SLC7A10 expression and BCAA levels. Figure created with BioRender.com.

protein 1 (4E-BP1)/PPAR γ axis.⁵⁹ Further, the mitochondrial protein sirtuin 4 (SIRT4) regulates BCAA catabolism in pre-mature adipocytes by promoting the expression of PPAR γ during early adipogenesis.⁶⁰

BAT is the second largest contributor to BCAA oxidation.²³ Fatty acids and glucose are traditionally regarded as major substrates for thermogenesis in BAT,⁶¹ but a recent study by Yoneshiro et al, unprecedentedly reported the usage of BCAAs as thermogenic substrates.⁶² BCAA clearance from the bloodstream by BAT is stimulated in response to cold exposure and is facilitated by SLC25A44, a newly characterized BCAA transporter in brown adipocytes.⁶² These findings provided a new perspective on the role of BCAA catabolism in driving thermogenesis in BAT.

Several factors are known to regulate BCAA catabolism in adipose tissue, including adiponectin and PPAR γ expression. Adiponectin, the most abundant adipokine, possesses multiple salutary properties, including insulin-sensitizing and anti-inflammatory effects.^{63–65} Adiponectin negatively associates with the risk of obesity/T2D.^{63,66,67} The activity of BCKDH is increased by adiponectin via upregulation of PPM1K expression and AMPK signaling in adipose tissue.⁶⁸ Hypoadiponectinaemia could lead to higher BCAA levels in obesity and T2D, as adiponectin directly regulates the rate-limiting enzyme BCKDH. PPAR γ , which belongs to the family of nuclear hormone receptors, plays an indispensable role in adipose tissue functions.⁶⁹ Pharmacological activation of PPAR γ using thiazolidinediones (an anti-diabetic drug) increases insulin sensitivity and adipogenesis in adipose tissue.⁷⁰ In obese Zucker rats, the administration of PPAR γ agonists upregulated the expression of genes related to BCAA catabolism (22 genes, such as *Bckdhb*, *Dbt*, *Dld*, *Acca2*, *Pcca*, etc.).⁷¹ Moreover, the ability of PPAR ligands to sensitize insulin is strongly associated with changes in the BCAA metabolic pathway in the adipose tissue of obese rats.⁷¹ Indeed, adipocyte-specific deletion of PPAR γ caused higher serum BCAAs and decreased incorporation of BCAAs into triacylglycerol in WAT and BAT in mice, accompanied with glucose intolerance in mouse models. Treatment with rosiglitazone increased BCKDH activity in WAT and BAT and reversed the defects in adipocyte-specific PPAR γ knockout mice.⁷²

5.1.1 | BCAA metabolism in the adipose tissue under pathophysiological conditions

Disruption of BCAA catabolism induces adipose tissue dysfunction. For instance, knockdown of *Bckdha* in pre-adipocytes impairs adipocyte differentiation, as shown by reduced expression of adipogenic genes, including *Pparg*, *Plin4*, *Adipoq*, and *Leptin*.⁵⁸ Under obese conditions, the BCAA flux to mmBCFA is diminished, leading to a reduction of de novo lipogenesis (Figure 2).⁵⁷ On the other hand, the reduced enzymatic activity of BCKDHA (probably due to attenuated protein expression of PPM1K and increased BCKDK expression) was observed in adipose tissues of genetically inherited or dietary-induced obese and diabetic mouse models.⁶⁸ Bariatric surgery significantly reduces serum BCAA levels along with an increase in adipose tissue expression of BCAT2 and BCKDHA proteins in obese individuals.⁴⁸ Consistently,

transplantation of normal epididymal WAT to the mice with *Bcat2* deletion, restores the BCAA levels to a normal level.⁵⁰ The studies support the key role of adipose tissue in systemic BCAA homeostasis.

Under obesity, WAT undergoes hypertrophy and/or hyperplasia to store additional lipids. The deleterious hypertrophic effects are closely associated with exacerbated hypoxia, chronic low-grade inflammation, necrosis, and fibrosis.⁵² As continuous inflammatory activities proceed in hypertrophic adipocytes, WAT loses its plasticity.⁵² Consequently, it fails to remodel in response to nutritional and hormonal signals. Cifarelli et al, reported a negative correlation between adipose tissue oxygenation and plasma BCAA levels in healthy and obese human subjects.⁷³ Interestingly, adipose tissue oxygenation showed a positive association with the BCAA catabolic genes, including *BCKDHB*, *HIBDH*, *HADHA*, *MMUT*, *HMGCL*, *MCC2*, and *ACAT1*, and a negative association with the genes involved in extracellular matrix (ECM) remodeling or fibrosis.⁷³ In obese adipose tissue, hypoxia suppresses BCAA catabolism, reduces BCAA uptake in adipocytes, and channels BCAAs toward lipogenesis.⁵⁷ This association study indicates that obesity-induced hypoxia may attenuate BCAA catabolism, potentially affecting adipose tissue remodeling and lipogenesis (Figure 2). It is currently unknown whether defective BCAA catabolism in adipocytes or other cell types contributes to adipose tissue fibrosis. However, defective BCAA metabolism has been shown to induce cardiac tissue fibrosis by activating the mTOR pathway in myocardial infarction (MI) mouse models.⁷⁴

Adipose tissue is a heterogeneous tissue containing not only adipocytes but also immune cells, stem cells, endothelial cells, etc.⁷⁵ M1 pro-inflammatory and lipid-laden macrophages impair adipose tissue functions, including induction of inflammation, insulin resistance, and promoting lipolysis, whereas M2 anti-inflammatory macrophages exert the opposite effects.^{76,77} Aberrant BCAA metabolism has been reported to control macrophage polarization, therefore potentially involving in adipose tissue-resident macrophage functions (Figure 2). Zhao et al, reported that the rise in BCAA levels promotes the secretion of high mobility group box 1 (HMGB1) nuclear protein, an accelerator of M1 macrophage.^{78,79} In *db/db* diabetic mice, BCAA supplementation led to an increase in the levels of BCKAs, resulting in macrophage hyperactivation. In addition, treatment with BCKAs resulted in increased production of pro-inflammatory cytokines like monocyte chemoattractant protein-1 (MCP-1), tumor necrosis factor- α (TNF- α), and interleukin-1 β (IL-1 β) in bone marrow-derived macrophages (BMDM) through induction of mitochondrial oxidative stress.⁸⁰ Thus, local inflammation is exacerbated in adipose tissue when BCAA catabolism is disrupted. Consistently, TNF- α and IL-6 expressions were suppressed upon boosting BCAA catabolism using 3,6-dichlorobenzothio(2)-carboxylic acid (BT2) treatment in the peritoneal macrophages isolated from *db/db* diabetic mice. Interestingly, TNF- α and IL-1 β have been shown to block leucine catabolism to the lipogenic substrate, but exert no obvious effect on its conversion to proteins in adipocytes.⁸¹ Further, treatment with TNF- α selectively attenuated the expression of BCAA transporter *Slc1a5* in 3T3-L1 adipocytes. Such effects of TNF- α have also been observed in the epididymal fat of HFD mice.⁸¹ These findings

suggest a reciprocal relationship between inflammation and BCAA metabolism in adipose tissue. On one hand, the changes in BCAA metabolism promote M1 macrophage polarization. On the other hand, the inflammatory conditions negatively affect BCAA metabolism. This reciprocal interaction forms a viscous cycle that exacerbates obesity-induced adipose tissue inflammation and metabolic dysfunction.

Defective BCAA catabolism also alters WAT browning and thermogenesis. For instance, genetic deletion of *Bcat2* has been shown to prevent obesity and increase energy expenditure by promoting browning in WAT. BCKAs repress WAT browning by inhibiting the master regulator of beiging, PR domain-containing protein 16 (PRDM16).⁸² At the molecular level, BCKA-derived acetyl-CoA leads to the acetylation of PRDM16 and hence blocks its binding with PPAR γ (Figure 2).⁸³ Moreover, leucine deprivation has been shown to not only reduce inflammation but also promote WAT browning in C57BL/6 mice.⁸⁴ Mechanistically, leucine deprivation activates GCN2 in macrophages. This, in turn, decreases monoamine oxidase A (MAOA) expression, resulting in a higher secretion of norepinephrine from macrophages to adipocytes, hence triggering WAT browning.⁸⁴ In addition, knockout of *Bckdha* weakens fuel oxidation and thermogenesis, resulting in higher body weight gain and glucose intolerance.⁶² Thus, the collective findings indicated a close reciprocal relationship between BCAA metabolism and adipose tissue functions.

5.2 | BCAA metabolism in the skeletal muscle

Skeletal muscle is a major site for glucose uptake, glycogen storage, thermogenesis, and acts as a pool of amino acids. The reserved pool of amino acids can be utilized for protein synthesis or catabolic

processes to support the energy demand.⁸⁵ Skeletal muscle is responsible for about 70% of whole-body glucose uptake via the glucose transporter (GLUT4) in muscle cells through the AKT-TBC1D4 and APPL2-TBC1D1 regulatory axes.⁸⁶⁻⁸⁸

In vivo isotope tracing analysis showed skeletal muscle contributes to approximately 59% of whole-body BCAA catabolism. The highest percentage of BCAA oxidation in skeletal muscle might be due to high transamination activity (~65%).⁸⁹ BCAA catabolism in muscle regulates amino acid homeostasis, as glutamate derived from the BCAT2 reaction can either combine with pyruvate to form alanine or with ammonia to form glutamine (Figure 3).⁹⁰

In healthy mice, acute BCKDK inhibition with BT2 promotes whole-body BCAA oxidation, primarily driven by the upsurge in BCAA oxidation in skeletal muscle.²³ This highlights the significance of skeletal muscle in whole-body BCAA oxidation. BCAAs, particularly leucine, activate the mTORC1 pathway in skeletal muscle.⁹¹ Upon activation, mTORC1 phosphorylates downstream targets such as S6K1 and 4E-BP1, resulting in increased protein synthesis, an increment in both the size and mass of skeletal muscle fiber, and inhibition of skeletal muscle proteolysis (Figure 3).⁹² In addition, supplementation of BCAAs promotes mitochondrial biogenesis and expression of *Sirt1* (the anti-aging molecule) in mice.⁹³

Several conditions regulate BCAA metabolism in skeletal muscle. Endurance exercise enhances BCAA oxidation in skeletal muscle⁹⁴ by activating the BCKDH complex.⁹⁵ Overexpression of PGC-1 α , a master mediator of exercise effects⁹⁶ induces BCAA catabolism by upregulating the expression of *Bcat2* and *Bckdh*.⁹⁷ This induction is mediated through the activation of estrogen-related receptor- α (ERR- α).⁹⁸ Like in adipose tissue, adiponectin also regulates BCAA catabolism in skeletal muscle. Adiponectin knockout mice exhibited higher *Bckdk* expression along with higher BCAA and BCKA levels in the

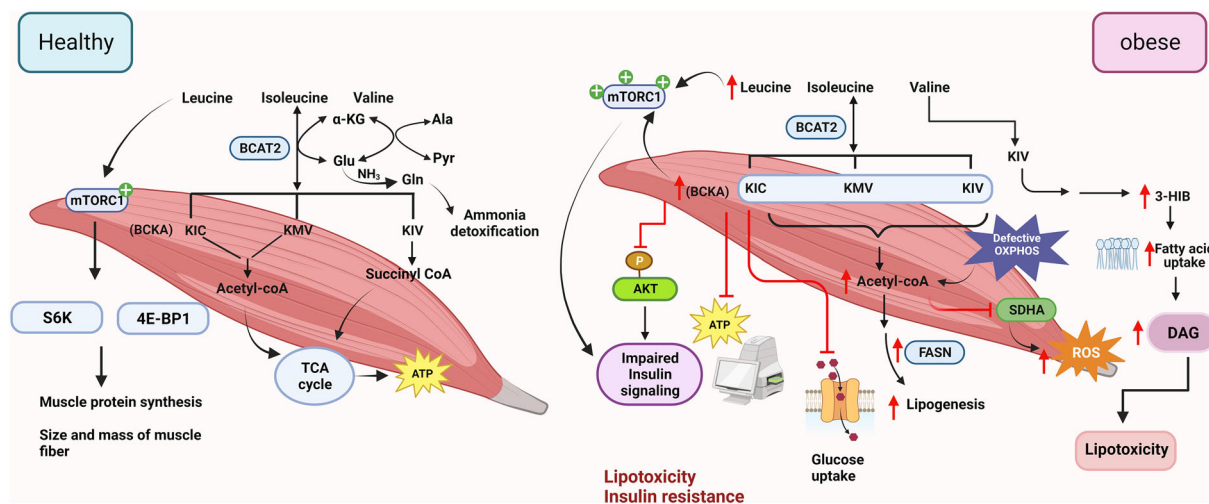


FIGURE 3 Role of BCAA catabolism in skeletal muscle under physiological and pathophysiological conditions. BCAAs induce protein synthesis and increase muscle fiber mass and size by activating the mTORC1 pathway and ATP production. However, chronic elevation of BCAAs/BCKAs leads to hyperactivation of mTORC1, and thus impairment of insulin signaling. BCKAs also suppress insulin-mediated glucose uptake by inhibiting the phosphorylation of AKT. 3-HIB causes lipotoxicity by increasing the transport of fatty acids. Defective OXPHOS can induce more production of acetyl-CoA, which can lead to an increase in de novo lipogenesis and ROS production via blocking mitochondria complex II (succinate dehydrogenase A [SDHA]) in skeletal muscle. Figure created with BioRender.com.

skeletal muscle, indicating impaired BCAA oxidation.⁶⁸ Conversely, treatment with adiponectin rescued HFD-induced elevation of BCAAs in the muscle tissue.⁹⁹

5.2.1 | BCAA metabolism in the skeletal muscle under pathophysiological conditions

BCAAs are required for protein synthesis and growth of skeletal muscle under physiological conditions.¹⁰⁰ However, an increase in BCAA and BCKA levels in skeletal muscles is reported in both early and late stages of T2D, potentially due to the attenuated expression of BCAA catabolic genes.^{32,47,98} Interestingly, hyperglycemia further diminished the catabolic ability of skeletal muscle on BCAA in T2D patients.⁹⁸ In addition, GSEA analysis revealed a positive correlation between insulin sensitivity and the BCAA catabolic pathway in skeletal muscle in human subjects with or without diabetes.¹⁰¹ Defective BCAA catabolism and subsequent elevation of BCAAs lead to persistent activation of the mTORC1/S6K1 pathway thereby blocking insulin signaling in skeletal muscle.^{102,103}

Although the physiological concentration of BCAAs enhances mitochondrial biogenesis,^{93,104} high concentrations (e.g., 20mM) of BCAAs attenuate the expression of mitochondrial biogenesis and oxidation genes such as *Pgc-1 α* , *Cox5a*, and *Atp5*, and glycolytic enzymes (*Ldha*, *Ldhb*, and *Pdh*) in insulin-resistant C2C12 myotubes.¹⁰⁵ In addition, myocyte-specific inhibition of mitochondrial oxidative phosphorylation (OXPHOS) accelerates BCAA catabolism, resulting in increased acetyl-CoA production for lipogenesis. Consequently, the buildup of acetyl-CoA leads to acetylation-mediated inhibition of mitochondrial respiration and subsequent reactive oxygen species (ROS) production and inflammation in the skeletal muscle (Figure 3).¹⁰⁶

The downstream metabolites of BCAAs are also emerging as mediators of insulin resistance. BCKAs hamper insulin signaling in skeletal muscle by impairing insulin-induced phosphorylation of AKT1 at serine residue 473 and insulin receptor substrate 1 (IRS1) at tyrosine residue 612.¹⁰⁷ BCKAs also dampen mitochondrial oxygen consumption and glucose uptake in C2C12 myotubes.¹⁰⁷ The valine metabolic intermediate 3-HIB is increased in the skeletal muscles of *db/db* diabetic mice and diabetic human subjects.¹³ 3-HIB promotes trans-endothelial transport of fatty acids and the build-up of lipotoxic intermediates such as diacylglycerol (DAG), and hence blocks insulin signaling in skeletal muscle.¹³ Ketoisocaproic acid (KIC) generated from leucine, reduces insulin-driven glucose uptake in myotubes in a BCAT2-dependent manner (Figure 3).¹⁰⁸

Diabetic animals also exhibit elevated levels of more distal BCAA downstream metabolites C3 and C5 acylcarnitines in skeletal muscle^{3,109} potentially resulting from a shift in BCAA oxidation from adipose tissue and the liver to the skeletal muscle. In 2009, Newgard et al, demonstrated that supplementation of BCAAs with HFD induces insulin resistance by activating the mTOR and JNK pathways in the skeletal muscle of rats, perhaps due to excessive accumulation of acylcarnitines.³ Unlike in the rat model, supplementation of BCAAs

had no obvious effects on glucose tolerance and insulin sensitivity in mice fed with an HFD or high-fat/high-sucrose diet.¹¹⁰ The discrepancy regarding the effects of BCAAs on rat and mouse models remains elusive.

Histone deacetylase 3 (HDAC3) is an epigenome modifier that regulates circadian rhythms. Muscle-specific deletion of HDAC3 leads to insulin resistance and reduced glucose utilization but increases exercise endurance and resistance in mouse models.¹¹¹ HDAC3 deficiency upregulates BCAA catabolic genes, such as *Bcat2*, *Dbt*, and *Ivd*, and facilitates the utilization of BCAA catabolic machinery for anaplerotic reactions, increased TCA flux, and oxidative metabolism for muscle contraction, but this increase in BCAA may also cause insulin resistance in skeletal muscle.¹¹¹ Taken together, these studies suggest that BCAA metabolism controls both lipid and glucose homeostasis as well as exercise endurance in skeletal muscle.

5.3 | BCAA metabolism in the liver

The liver controls energy metabolism and serves as a metabolic hub for protein, lipid, and glucose homeostasis. This endocrine organ can coordinate with other tissues like adipose tissue and skeletal muscle via the production of hepatokines (liver-derived hormones) in control of whole-body homeostasis.¹¹² The liver can convert carbohydrates into triglycerides via de novo lipogenesis, and it's regulated by carbohydrate responsive-element binding protein- β (ChREBP) and sterol regulatory element-binding proteins (SREBP) family members (SREBP-1c).^{112,113} Owing to the role of the liver in protein metabolism, it participates in various processes including plasma protein formation, deamination of amino acids, amino acid interconversion, and synthesis of urea (Figure 4).¹¹⁴

Unlike other essential amino acids, BCAAs bypass the first-phase metabolism due to the low BCAT2 activity in hepatocytes.²¹ The transamination of BCAAs to their respective BCKAs occurs in extrahepatic tissues, followed by oxidation in the liver, where BCKDH activity is high.² The isotope-tracing experiment showed the liver is the largest contributor to the disposal of BCAAs into protein synthesis.² The balance of BCAAs between the liver and blood is regulated by the hepatic transporter.¹¹⁵ Specifically, the transporter SLC43A1 facilitates BCAA efflux from the liver into the bloodstream.¹¹⁶ Glutamate generated by the BCAT2 reaction is converted into glutamine and transported back to the liver and participates in nitrogen homeostasis. Additionally, the transfer of α -amino nitrogen from glutamate to pyruvate leads to the formation of alanine, which is utilized in hepatic gluconeogenesis. In this way, the shuttle system involving glutamine and alanine enables the transfer of BCAA nitrogen from muscle to the liver for urea synthesis.¹¹⁷

BCKAs derived from extrahepatic tissue undergo complete oxidation in the liver. The liver removes ammonia generated from BCAA catabolism via urea synthesis. BCAA catabolic enzymes BCKDK and PPM1K positively and negatively regulate the activity of ATP citrate lyase (ACLY), thereby controlling hepatic de novo lipogenesis.⁴⁹ BCAAs promote the synthesis of albumin and glycogen by activating

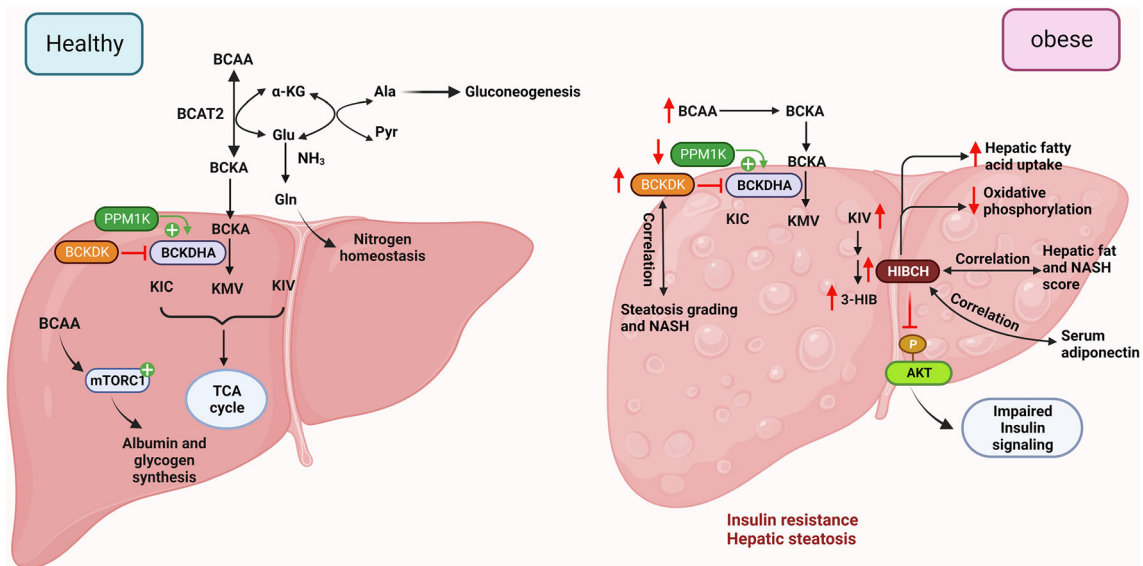


FIGURE 4 BCAA catabolism in the liver under physiological and pathophysiological conditions. Apart from producing ATP, BCAAs generate the substrate for hepatic gluconeogenesis. The liver plays a crucial role in detoxifying the ammonia generated during BCAA catabolism. BCAAs induce the synthesis of liver proteins via mTORC1 activation. On the other hand, elevated levels of BCAAs/BCKAs/3-HIB induce fatty acid uptake, and steatosis, and impair insulin signaling. Figure created with [BioRender.com](#).

the mTOR signaling pathway (Figure 4).¹¹⁸ Thus, BCAA metabolism regulates nitrogen homeostasis, de novo lipogenesis, and protein synthesis in the liver.

5.3.1 | BCAA metabolism in liver tissue under pathophysiological conditions

Elevated levels of plasma BCAAs are associated with different statuses of MAFLD, ranging from steatosis, non-alcoholic steatohepatitis (NASH), and cirrhosis, as well as liver cancer (as shown in Table 1).

During obesity, the hepatic activity of the BCKDH complex is compromised by the hyper-phosphorylation of Ser²⁹³ at the E1 alpha subunit, possibly resulting from the altered expression of *Ppm1k* (decreased) and *Bckdk* (increased).⁶⁸ These changes might cause higher BCAA levels during obesity.

A positive correlation has been observed between plasma BCAAs and liver steatosis stages in subjects with severe obesity and MAFLD in children and adolescents.¹²⁵ In the Prospective Multicenter Imaging Study for Evaluation of Chest Pain (PROMISE), a positive association of BCAAs with hepatic steatosis was also identified.¹²⁶ The metabolomic and transcriptomic data showed higher hepatic BCAA levels and reduced expression of BCKDHA and BCAA transporters in NASH patients with or without fatty liver.¹¹⁵ While hepatic BCAA levels remain unchanged between healthy subjects and those with steatosis (except for valine), a significant increase in BCAAs (% of normal) was observed in NASH, specifically valine (147%), leucine (127%), and isoleucine (137%).¹¹⁵ The association between BCAA levels and hepatic steatosis can be explained by a dose-dependent increase in SREBP1 expression and lipogenic

markers, along with reduced β -oxidation following valine treatment in HepG2 hepatoma cells.¹²⁷ Interestingly, a study in patients with severe obesity found that among BCKAs, α -ketoisovalerate (KIV) and the ratio of BCKA/BCAA showed a positive association with both the steatosis grade and NASH¹¹ indicating the ratio of BCKA/BCAA could serve as a robust biomarker for the liver disease instead of relying on either BCAAs or BCKAs, and defective BCAA catabolism might causatively contribute to the development of MAFLD. Future longitudinal monitoring of BCAAs and BCKAs in the progression of steatosis to NASH and HCC in different populations might indicate the usefulness of BCAA/BCKA for predictive and diagnostic biomarkers for MAFLD.

The hepatic expression of BCAA catabolic genes also showed a strong association with steatosis and the NASH score during obesity. A human study showed hepatic mRNA expression of *BCKDK* and *HIBCH* (the enzyme that produces 3-HIB) is positively associated with steatosis and NASH score, and negatively associated with serum adiponectin levels (Figure 4).^{11,45} Overexpression of *HIBCH* or treatment with 3-HIB leads to increased fatty acid uptake and ROS production in human Huh7 hepatocytes, accompanied by upregulation of genes involved in fatty acid metabolism or lipid storage⁴⁵ siRNA-mediated knockdown of *HIBCH* expression sensitizes insulin action in the hepatocytes, which is similar to the observation of reduced AKT phosphorylation in myocytes and adipocytes treated with 3-HIB.^{12,13} These findings explain the positive association between *HIBCH* expression/3-HIB levels, and hepatic steatosis and NASH in humans. On the other hand, *HIBCH* knockdown also resulted in the upregulation of genes related to TNF- α signaling and hypoxia⁴⁵ yet the underlying causes and their effects on hepatocyte functions remain unknown.

TABLE 1 Studies showing alteration in BCAA or BCKA levels in liver steatosis, MAFLD, NASH, and liver cancer.

Study population (sample size, study type, and ethnicity)	Key findings	References
Healthy controls and nondiabetic subjects with steatosis or NASH (using untargeted global metabolomic analysis) (n = 60) cross-sectional study, not defined	Higher plasma BCAA levels in NASH	119
Genome-scale metabolic model of hepatocytes from patients with NASH (n = 45), cross-sectional study, not defined	Participation of valine in the appearance of NASH	120
Frozen liver tissue samples from healthy, steatosis, and NASH (with or without fatty liver) subjects (n = 45), cross-sectional study, not defined	Elevation in hepatic valine (147% of normal), leucine (127%), and isoleucine (139%) during the transition from steatosis to NASH	115
Patients with NASH (both male and females) (undergone liver biopsy) (n = 137), cross-sectional study, Japanese population	The plasma BCAA levels in NASH patients decreased with the progression of fibrosis stages	121
MAFLD (with or without obesity) and control subjects (n = 64), cross-sectional study, Italian population	Subjects with MAFLD have elevated plasma levels of isoleucine and valine	122
A cohort study with subjects without MAFLD at baseline (n = 1302), prospective cohort study, Chinese population	At baseline, the total BCAA levels and individual BCAA were higher in subjects with MAFLD as compared to controls	123
Patients with different grades of steatosis with or without NASH (n = 288), cross-sectional study, European population	Positive association of BCKA/BCAA ratio α -ketoisovalerate with the grade of steatosis and NASH.	11
Patients with hepatocellular carcinoma (HCC) (n = 89) cross-sectional study, Egyptians population	Higher plasma BCAAs in patients with HCC	124

Elevated plasma BCAA levels have been observed in patients with HCC and utilized as a biomarker for the progression to HCC development.¹²⁴ Consistent with this finding, the BCAA catabolic pathway was identified as the most significantly downregulated pathway in the HCC cohort.¹²⁸ The dramatic changes in BCAA catabolic genes

are not due to changes in the promoter but somatic copy number variation (CNV) in tumor tissue.¹²⁸ The accumulation of BCAAs, resulting from reduced hepatic BCAA catabolism, leads to the hyperactivation of the mTORC1 pathway in liver tumors. Additionally, sh-RNA-mediated knockdown of *Bckdha* also produces a similar effect on mTORC1 activation, which can be mitigated by treatment with the mTOR inhibitor rapamycin in the AML12 cell line.¹²⁸ Consistent with these findings, BCAA dietary supplementation exacerbates tumor growth in vivo.¹²⁸

Despite the higher levels of BCAA/BCKA observed in liver diseases (Table 1), BCAA supplementation yields several beneficial effects on liver functions, including increased protein synthesis, maintenance of nitrogen balance, and an improved survival rate in cirrhosis patients.¹²⁹ However, different pre-clinical studies have reported conflicting findings regarding the effect of BCAA supplementation.¹³⁰ For instance, in obese/diabetic mouse models, BCAAs supplementation demonstrated beneficial metabolic effects such as reduced body weight and repressed hepatic de novo lipogenesis. However, it can also lead to detrimental effects such as abnormal lipolysis in adipocytes and hepatic inflammation.¹⁴ In the choline-deficient, L-amino acid-defined high-fat diet (CDAHFD)-induced NASH mouse model, BCAA supplementation improved hepatic steatosis via a reduction of *Fasn* expression.¹³¹ In the HFD rat model, the anti-hepatic steatosis effect of BCAA is partly due to changes in gut microbiota. Notably, an increase in *Ruminococcus flavefaciens* (linked to portal acetic acid production) was found.¹³² These effects are negated under cellulose deficiency, as *R. flavefaciens* primarily uses cellulose to produce acetic acid.¹³²

5.4 | BCAA metabolism in heart

The heart is another tissue heavily influenced by BCAA metabolism. Carbohydrates, amino acids, lipids, and ketone bodies can be used as energy substrates to generate ATP in the mitochondria of the heart. The preference of substrate utilization in the heart is context-dependent (such as fatty acid and amino acids contribute to energy production during fasting and lactate during exercise) and pathological condition (glycogen metabolism is essential in the fetal heart).^{133,134} In a healthy heart, the rate of BCAA oxidation within the TCA cycle is relatively low (4%) when compared to other primary sites of BCAA oxidation such as muscle, liver, and adipose tissue.²³ Numerous studies showed a defective BCAA catabolism in heart diseases. Elevated BCAA levels blunted glucose oxidation through inhibition of the PDH complex via O-linked N-acetylglucosamine (O-GlcNAc) modification, thus rendering the heart more susceptible to ischemic injury.¹³⁵ BCKA perfusion in the heart resulted in the inhibition of insulin-induced glucose oxidation with reduced AKT and PDH activity.¹³⁶ This implied the regulatory role of BCAA metabolism in glucose oxidation and insulin signaling in cardiac tissue. Strikingly, defective BCAA catabolism alone can induce contractile dysfunction in the heart without external pathological stressors. The echocardiogram data indicated a decrease in cardiac function in 3-month-old PPM1K KO mice, which

progressively worsened as the mice aged to 18 months.⁹ In addition, global PPM1K deletion also increased susceptibility of heart failure in mice under pathological stress, yet whether the effect is autonomous in cardiomyocytes or systemic elevation of BCAA/BCKA requires further investigation.⁹ Consistently, human patients with dilated cardiomyopathy also showed defects in BCAA catabolic machinery and diminished insulin signaling.^{9,137} A study using ¹³C-labeled KIV flux revealed the fate of BCKA in heart tissue. The findings indicated that under pathological conditions, BCKA is preferentially reaminated back to BCAA, resulting in increased protein synthesis rates and leading to cardiac hypertrophy, rather than being oxidized.¹³⁸ The low expression of BCAA transporter SLC25A44 in the heart contributes to the accumulation of BCAA derived from reamination.¹³⁸ Taken together, intact BCAA catabolic machinery is vital for heart functions.

6 | THERAPEUTIC APPROACHES AGAINST METABOLIC DISEASES BY TARGETING BCAA METABOLISM

Since high circulating BCAA levels and defective BCAA catabolism in the metabolic tissues are causatively linked with diabetes and MAFLD, restoring these defects could offer a feasible and novel approach for these non-communicable and incurable diseases. In this section, we

summarize the approaches, including pharmacological drugs, modulation of gut microbiota, and lifestyle interventions to modulate BCAA metabolism, and discuss their efficacy against metabolic diseases.

6.1 | Use of pharmacological compounds to promote BCAA catabolism

6.1.1 | Pharmacological compounds targeting BCKDK

BT2

BT2, an allosteric inhibitor of BCKDK, enhances the activity of the BCKDH complex, thereby promoting BCAA catabolism.¹³⁹ In *ob/ob* mice, BT2 has been reported to significantly increase BCKDH activity in skeletal muscle, liver, and WAT, leading to a reduction in plasma BCAAs and BCKAs.⁴⁶ Furthermore, BT2 treatment restored BCAA catabolism and improved insulin sensitivity and glucose tolerance in the DIO mouse model.⁴⁶ Notably, BT2 enhances the efficacy of the anti-diabetic drug metformin in *ob/ob* and DIO mice.¹⁴⁰ BT2 treatment also reduced hepatic triglyceride levels in insulin-resistant Zucker fatty rats.⁴⁹ In the MAFLD mouse model fed with a CDAHFD, BT2 treatment alleviated hepatic inflammation and fibrosis (Figure 5).¹⁴¹ Likewise, BT2 treatment attenuated liver fibrosis,

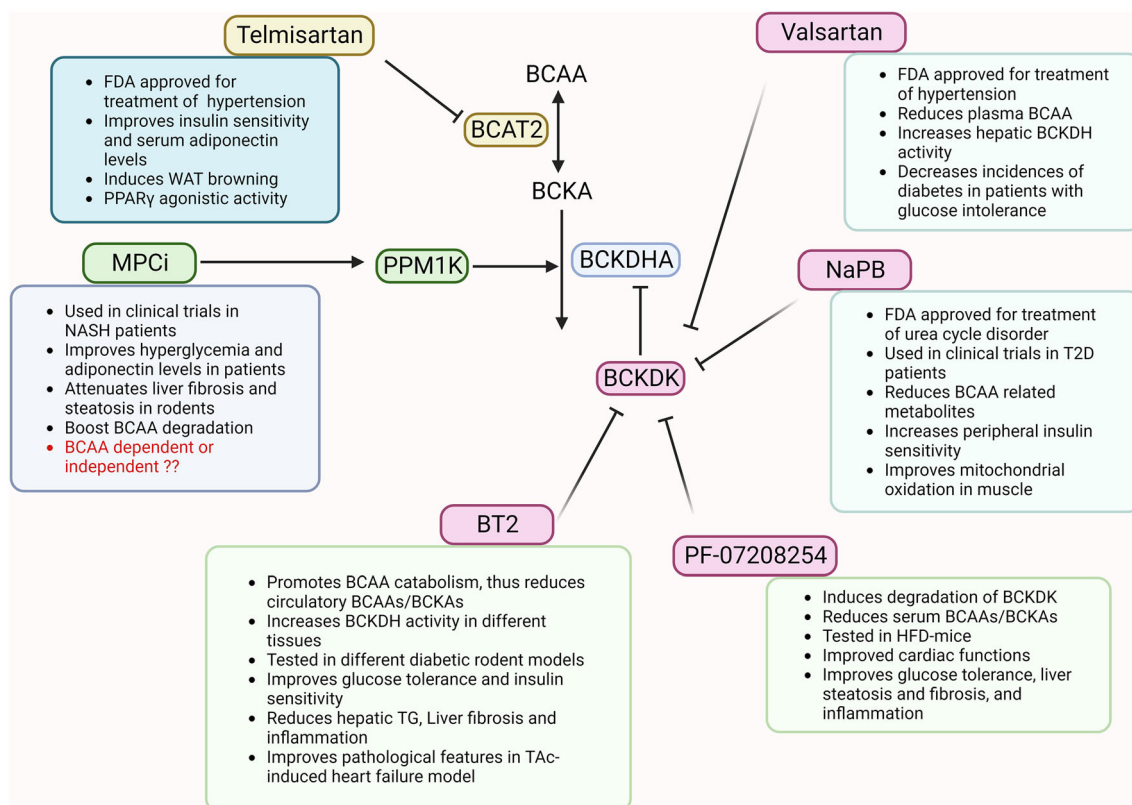


FIGURE 5 Different pharmacological compounds targeting BCAA catabolic machinery. Different pharmacological compounds target BCKCK, PPM1K, and BCAT2 (as shown in the figure) to boost BCAA catabolism and produce beneficial effects against metabolic diseases. Figure created with BioRender.com.

steatosis, and inflammation in low-density lipoprotein receptor knock-out (Ldlr^{-/-}) mice fed with a western diet.¹⁴¹ Thus, these preclinical studies showed that BT2 treatment is effective in alleviating metabolic diseases, but its use in humans has not been tested till date.

Pf-07208254

A new potent BCKDK inhibitor is identified by Flach et al in 2023.¹⁴² PF-07208254 induces degradation of BCKDK by reducing the proximity between BCKDK and the E2 subunit of the BCKDH enzyme complex.¹⁴² Chronic treatment with PF-07208254 showed improvement in metabolic parameters (glucose tolerance, liver steatosis, fibrosis, and inflammation) through reduction in BCAAs, BCKAs, and phospho-BCKDHA levels in HFD-fed mice (Figure 5). PF-07208254 showed similar efficacy as BT2 for the treatment of metabolic diseases, however, it exhibits higher potency (lower half maximal inhibitory concentration [IC₅₀]), due to better protein interaction with non-polar side chains within the BCKDK binding pocket.¹⁴²

Valsartan

It is an angiotensin II receptor blocker used for the treatment of hypertension. It reduces the incidence of diabetes among patients with glucose intolerance and cardiovascular diseases.¹⁴³ Interestingly, valsartan is reported to inhibit BCKDK in a concentration-dependent and ATP-competitive manner.¹⁴⁴ The treatment with valsartan resulted in a reduction in plasma BCAA levels and increased activation of the hepatic BCKDH complex in rats (Figure 5).¹⁴⁴ However, whether the anti-hypertension effects of valsartan are partially mediated by its modulating actions on BCKDK requires further investigation.

Sodium phenylbutyrate (NaPB)

NaPB has been utilized as a pharmacological activator to boost BCAA catabolism in a clinical trial for T2D patients (NetherlandsTrialRegister: NTR7426).¹⁴⁵ NaPB, an FDA-approved drug commonly used for the treatment of urea cycle disorders, acts on the same binding site as BCKDK. Clinical trial data showed that NaPB treatment led to a decrease in BCAA catabolic metabolites, accompanied by improvements in peripheral insulin sensitivity, mitochondrial oxidation in muscle, and whole-body carbohydrate oxidation (Figure 5).¹⁴⁵

6.1.2 | Pharmacological compounds acting on other BCAA enzymes

Mitochondrial pyruvate carrier (MPC) inhibitor

Mitochondrial pyruvate carrier (MPC) is responsible for importing cytosolic pyruvate to the mitochondrial matrix for oxidative phosphorylation. In recent years, MPC has emerged as a promising therapeutic target for the treatment of metabolic diseases. Inhibition or genetic deletion of MPC has demonstrated notable improvements in liver fibrosis, NASH, and BCAA degradation in rodent models.^{146,147} In vitro, MPC inhibition boosts BCAA catabolism by reducing phospho-BCKDHA levels through PPM1K. The effect of MPC inhibitor on

BCKDH activity is mediated via the activation of mTOR and AMPK signaling pathways.¹⁴⁷ Treatment with MPC inhibitor ameliorated hyperglycemia, hepatic steatosis, and HOMA-IR in Zucker diabetic fatty (ZDF) rats (Figure 5). Although MPC inhibitors do not significantly affect BCAA levels, they reduce circulatory 3-HIB levels, which might contribute to the improvement of insulin sensitivity and lipotoxicity in the skeletal muscle and liver.^{13,45,147}

Telmisartan

Telmisartan is an FDA-approved drug for the treatment of hypertension, and it works by blocking angiotensin receptor II. Telmisartan has been shown to improve metabolic parameters including insulin sensitivity, triglyceride levels, adiponectin levels, and the expression of *Ucp1* in obese mice.¹⁴⁸ In addition, telmisartan blocks BCAA catabolism by inhibiting the BCAT2 enzyme and inducing browning of WAT, and protecting against diet-induced obesity.⁸³ The induction of WAT browning has emerged as one of the approaches for the treatment of obesity/diabetes (Figure 5). Due to its pleiotropic beneficial effects, telmisartan may be utilized as a therapeutic against obesity/T2D, pending further evaluation of its efficacy in large clinical trials.

Given their approval by the FDA, re-purposing of Valsartan, Telmisartan, and NaPB for the treatment of T2D holds enormous potential. Nonetheless, large-scale clinical trials are needed to further validate their potential in the treatment of T2D. Testing the newly identified BCKDK inhibitor PF-07208254 on metabolic diseases in an independent laboratory is also urgent.

6.2 | Gut microbiota modulation

The gut microbiota dysbiosis is one of the contributors to high plasma BCAAs in obesity/T2D.¹⁴⁹ The gut microbes *Prevotella copri* and *Bacteroides vulgatus* with BCAA biosynthesis capacity are positively correlated with circulating BCAA levels in human subjects with insulin resistance. Metagenomic analysis showed that the expression of genes related to the synthesis and transport of BCAAs in the gut microbe were increased and decreased, respectively, in subjects with insulin resistance.¹⁴⁹ In line with these observations, the higher abundance of genes involved in BCAA biosynthesis in the gut microbe is also found in HFD- and streptozotocin (STZ)-induced diabetic rats.¹⁵⁰ In a cohort study consisting of morbidly obese women, lower microbial gene richness (MGR) was found in subjects with steatosis, thus pinpointing the role of the gut microbiome in hepatic steatosis. The gut microbiota of obese patients with hepatic steatosis also showed an increased potential for BCAA biosynthesis.¹⁵¹ In addition, phenylacetic acid, a microbial metabolite derived from aromatic amino acid metabolism, enhanced BCAA utilization and lipogenesis in hepatocytes.¹⁵¹ These studies altogether indicate that changes in gut microbiota affect BCAA levels during obesity/T2D. Thus, modulating the gut microbiota can represent as therapeutic approach using phytochemicals (Figure 6).

The phytochemical berberine is reported to improve insulin resistance by reducing circulating BCAA levels in HFD-fed mice. This

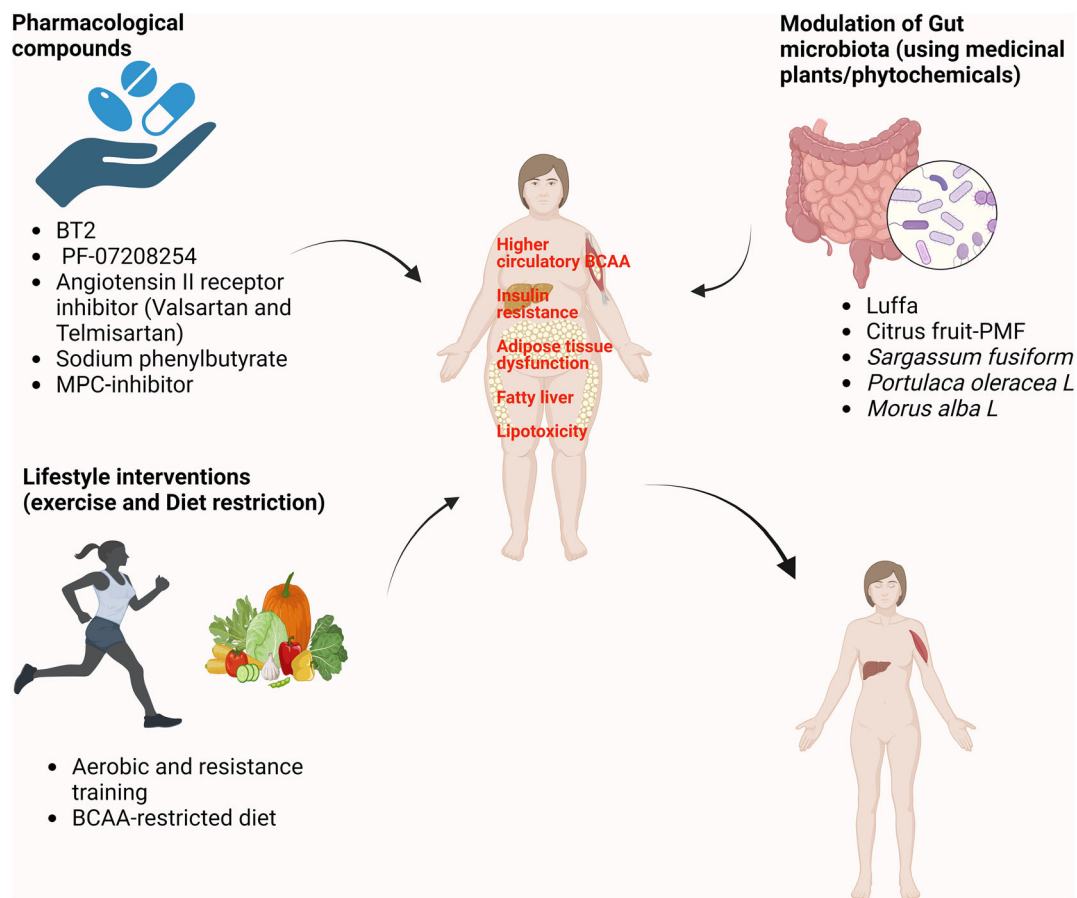


FIGURE 6 Therapeutic interventions for promoting BCAA catabolism or reducing BCAA biosynthesis in metabolic diseases. Figure created with BioRender.com.

effect is mediated through the modulation of the gut microbiota, specifically by decreasing the abundance of gut microbes involved in BCAAs biosynthesis.¹⁵² Additional examples of phytochemicals are discussed below.

The vegetable luffa belongs to the family *Cucurbitaceae* with medicinal value and reduces circulating BCAAs by upregulating BCAA catabolic enzymes in adipose tissue and the liver via gut microbiota modulation in the DIO mouse model.¹⁵³ Luffa treatment suppressed the gut bacteria that are positively associated with serum BCAAs, thereby inhibiting the biosynthesis ability of BCAAs in DIO mice. Furthermore, the beneficial effects of luffa in promoting BCAA catabolism and improving metabolic parameters were abolished when gut microbiota was depleted by the antibiotic treatment.¹⁵³

Citrus peel-derived phytochemical polymethoxyflavones (PMF) extract has been shown to alleviate hepatic steatosis and improve various metabolic parameters, including glucose tolerance, insulin sensitivity, serum cholesterol, and triglyceride level in HFD-fed mice. These effects are mediated through the suppression of the mTOR/P70S6K/SREBP pathway in the liver tissue.^{154,155} Notably, the BCAA biosynthesis pathway in the gut microbiota was identified as the most down-regulated pathway, resulting in lower serum BCAAs in DIO mice treated with PMF-rich extract (PMFE). The protective effects of PMFE were abolished upon antibiotic treatment. Additionally, the

fecal transplantation from PMFE-treated DIO mice to DIO mice resulted in improved metabolic parameters and significantly lower serum BCAA levels,¹⁵⁵ indicating the direct modulation of gut microbiota by PMFE.

Sargassum fusiform (SF-Alg), a type of brown algae, is a traditional Chinese medicine.¹⁵⁶ Administration of SF-Alg results in lower fasting glucose, triglyceride, and total cholesterol levels, as well as increased levels of HDL-cholesterol in diabetic mouse models. Further, histological analysis showed smaller adipocyte size and reduced inflammation in the liver.¹⁵⁷ Mechanistically, the modulation of gut microbiota (increase in *Bacteroidetes* to *Firmicutes* ratio), and reduced colon BCAA levels may be responsible for the beneficial effect of SF-Alg in diabetic mice, yet further investigation is needed.¹⁵⁷

Portulaca oleracea L. (PE) is shown to have an anti-diabetic effect. PE influences gut microbiota composition, specifically the *Firmicutes* to *Bacteroidetes* ratio, inhibits bacterial BCAA biosynthesis, and promotes the expression of BCAA catabolic genes in liver and muscle tissue.¹⁵⁸ *Morus alba L.* (mulberry leaf) extract has been found to possess anti-obesity effects and the ability to modulate the gut microbiota.¹⁵⁹ Administration of *M. alba L.* water extracts (MLE) results in improved metabolic parameters, and reduced inflammation, by altering the abundance of gut bacteria related to BCAA biosynthesis in high-fat, high-sucrose (HFHS) diet-fed mice. Furthermore, MLE

increased the expression of BCAA catabolic genes in the liver, skeletal muscle, and epididymal fat, thus promoting whole-body BCAA catabolism.¹⁶⁰

The aforementioned medicinal herbs or plant-based extracts demonstrate the ability to modify BCAA metabolism by altering the gut microbiota and activating distinct signaling pathways, highlighting their therapeutic potential against metabolic diseases.

6.3 | Lifestyle interventions

Lifestyle interventions, including exercise and diet, are crucial in the management of metabolic diseases (Figure 6).

6.3.1 | Exercise

It is one of the promising approaches in the management of metabolic diseases. Aerobic exercise and resistance training have been shown to have favorable outcomes in T2D management.¹⁶¹ Exercise has long been reported to induce BCAA oxidation. Additionally, endurance exercise can lead to the activation of the BCKDH complex in skeletal muscle and the liver.^{95,162} Strenuous exercise was shown to significantly reduce the serum BCAA levels as compared to the non-exercising control group.¹⁶³ Surprisingly, different individuals respond differently to exercise (response variability) in the context of improvement in glucose tolerance and insulin sensitivity, thus being categorized into two categories: non-responders (ranging from 7% to 69%) and responders.¹⁶⁴ The cause of the response variability to exercise lies in differential modulation in the gut microbiota and its metabolites. For instance, the responders group showed an increase in the biosynthesis of gut-derived short-chain fatty acids (SCFA) and GABA and a decrease in BCAA levels. However, the opposite was observed in the non-responder group.¹⁶⁵ The combination of both exercise and modulation of gut microbiota using phytochemicals might show beneficial outcomes in non-responder groups. Interestingly, the beneficial effect of exercise on insulin sensitivity and inhibition of lipogenesis in sWAT was abolished by BCAA supplementation in obese mice.¹⁶⁶

6.3.2 | Diet restriction

Restriction of BCAAs consumption exerts beneficial effects on metabolic health, including better glycaemic control, body composition, and efficient mitochondrial functions in adipose tissue in rodent models.^{16,19,20} Deprivation of leucine alone for seven days resulted in several metabolic changes in adipose tissue such as higher energy expenditure, suppression of lipogenesis, an increase in fatty acid oxidation in WAT, and a higher *Ucp1* expression in BAT in a mouse model.¹⁶⁷ The effect of leucine on adipose tissue browning can be indirect via neuronal activation through GCN2 and the activating transcription factor 4 (ATF4) switch.¹⁶⁸ It is worth mentioning that BCAA restriction increases the efficacy of metformin in *ob/ob* and DIO mice.¹⁴⁰

In 2019, a randomized controlled trial was performed to assess the effects of a short-term (4-week) BCAAs-reduced (isocaloric) diet on T2D patients. It showed that short-term BCAAs dietary restriction resulted in better postprandial insulin sensitivity, attenuated activation of mTORC1 in adipose tissue, and changes in gut microbiota composition.¹⁸ While the abundance of intestinal *Bacteroidetes* declines and the percentage of *Firmicutes* rises in obesity¹⁶⁹ such changes could be reversed by short-term BCAAs dietary restriction¹⁸ Strikingly, the beneficial effect of the BCAAs restriction is mediated by isoleucine or valine (not by leucine) partially through the activation of the hepatokine fibroblast growth factor 21 (FGF21).^{36,170} The low-protein diet has been shown to upregulate FGF21, which increases energy expenditure and insulin sensitivity by targeting adipose tissue. The beneficial effects of a protein restriction diet can be credited, at least in part, to the reduction of BCAAs intake.^{17,171}

While BCAAs restrictions have shown multiple beneficial effects on metabolic health, it is crucial to acknowledge their vital role in protein synthesis and the maintenance of muscle mass in skeletal muscle.⁹¹ This raises concerns that limiting BCAAs might potentially impact muscle mass, particularly in aging populations, which already suffer from reduced muscle mass and strength, and increased frailty.¹⁷² Moreover, approximately one-third of the amino group in the neurotransmitter glutamate, crucial for brain development and cognitive functions, is derived from BCAAs.¹⁷³ Also, lower plasma valine levels are correlated with accelerated cognitive decline in human subjects.¹⁷⁴ Thus, BCAA deprivation might potentially affect cognitive functions. Furthermore, BCAA restriction has been shown to impair immune function and enhance the susceptibility to pathogens in mice.¹⁷⁵ Of note, obese/diabetic patients display aberrant immunity (i.e. chronic inflammation) and are more susceptible to infection.¹⁷⁶ Considering these potential side effects, future implementation of BCAA restriction in diabetic and obese patients needs to be personalized, with its efficacy and potential side effects closely monitored.

7 | CONTROVERSIAL EFFECTS OF BCAAS AND/OR COMPONENT OF BCAAS

BCAAs restriction exerts multiple salutary effects on aging- and obesity-induced metabolic disorders. Paradoxically, supplementation of leucine also improves metabolic health in obese conditions.¹⁷⁷ For example, leucine supplementation enhances insulin signaling, promotes mitochondrial biogenesis, and reduces lipid accumulation through the activation of PCG1 α via SIRT1-mediated deacetylation in the liver, skeletal muscle, and adipose tissues of male C57BL6/J mice fed with a HFD (45% kcal fat).¹⁷⁸ In addition, leucine supplementation increases sensitivity and secretion of insulin as well as reduces body weight and fat mass in HFD-fed adult mice (60% kcal from fat).¹⁷⁹ Consistently with the notion that leucine might be beneficial, a more recent study revealed that the dietary restriction of valine and isoleucine but not leucine induces metabolic benefits via induction of the hepatokine fibroblast growth factor 21 (FGF21) and its action on

adipose tissue “being” and thermogenesis as well as hepatic insulin sensitivity in C57BL6/J mice fed with a western diet (42% kcal from fat and 0.2% total cholesterol and 34% sucrose by weight).^{16,36} The differential effects of isoleucine, valine, and leucine on metabolic health remain unknown. This may be attributed to their distinct activities on mTOR and their unique downstream catabolic pathways and metabolites.^{180,181} However, some studies have shown beneficial effects of leucine deprivation, such as reduced fat mass through induction of UCP1 in BAT and lipolysis in WAT, as well as modification of gut microbiota composition in the mice.^{167,182}

Recent studies suggest that the therapeutic outcomes of protein restriction, including BCAA restriction, are influenced by factors such as sex, the genetic background of the animals, duration of treatment, age for starting treatment, and ratio of BCAA to tryptophan and threonine in the diet.^{17,183,184} Additionally, different methodologies have been used in studies on leucine supplementation and BCAA restriction. For instance, one study used an isocaloric chow diet with reduced content of BCAA (67% of original concentration), while another study supplemented 1.5% leucine in drinking water under HFD conditions without altering the calorie content or levels of valine and isoleucine.^{36,179} More rigorous experimental designs are needed to compare individual and combination effects of isoleucine, leucine, and valine in deprivation and supplementation in animal studies.

8 | CONCLUSION

As essential nutrients, BCAAs play a vital role in different physiological processes and mediate crosstalk between various metabolic organs. Not only higher plasma BCAAs and downstream metabolites but dysregulated BCAA catabolism in various metabolic tissues is associated with the risk and development of metabolic diseases. However, the individual contribution of defective BCAA catabolism to these tissues in the development of metabolic disease still needs to be explored.

Utilizing the therapeutic approaches that help in boosting the BCAA catabolism or attenuation of BCAA biosynthesis could represent a promising strategy in the future for the management of metabolic diseases. The majority of previous studies focused on the investigation of the first few steps of BCAA catabolism (such as BCAA to BCKA), further investigation of the distal BCAA metabolism and their metabolites, as well as their crosstalk with other metabolic pathways such as the TCA cycle and lipogenesis in different metabolically active cells are attractive topics. A tissue-specific approach for promoting BCAA catabolism in adipose tissues and the liver might not only exert beneficial effects on metabolic diseases but might minimize its potential negative effects on protein synthesis and muscle mass in the skeletal muscle. However, more mechanistic insights are further needed to fully utilize the therapeutic potential of BCAAs.

Important take-to-home messages and open questions are listed below.

Take-to-home messages:

1. Maintenance of the physiological concentration of BCAAs is regulated by the tight coordination between multiple metabolic tissues including adipose tissues, the liver, and the skeletal muscle.
2. The disruption of the BCAA catabolic machinery and altered BCAA metabolic fates in the metabolic tissues lead to elevated circulating BCAA and altered profile of their downstream metabolites, which contribute to the development of metabolic diseases including obesity, diabetes, and the MAFLD.
3. Targeting aberrant BCAA catabolism or BCAA restriction improves metabolic health in rodents and humans.

Open questions:

1. What are the metabolic fates of BCAA in different metabolic tissues under physiological (such as fasting, feeding, and cold environment) and pathological (such as obesity and diabetes) conditions?
2. Apart from the host tissues, aberrant BCAA metabolism in gut microbiota also contributes to systemic BCAA balance and metabolic disorders. Does BCAA metabolism in gut microbiota interact/intertwine with the host's BCAA metabolic pathways? If yes, what are the underlying regulatory mechanisms?
3. Given that BCAA is important for protein synthesis in the skeletal muscle and muscle mass. Is a tissue-specific approach for promoting BCAA catabolism in adipose tissues and the liver sufficient to exert beneficial effects on metabolic diseases? Since this approach avoids the potential deleterious effects on skeletal muscle, particularly in aging population.
4. Both pharmacological and dietary approaches targeting BCAA metabolism show multiple salutary effects on metabolic health in rodents. A recent clinical study showed that BCAA catabolic activator NaBP improves insulin resistance and reduces glucose levels in human diabetic subjects. Further human clinical trials involving a larger sample size, and more diverse populations will provide more evidence about the efficacy, generalization, and safety issues of BCAA-targeting approaches.

AUTHOR CONTRIBUTIONS

Shama Mansoori: Writing- conceptualization, drafting of original manuscript and editing, Kelvin Kwun Wang Ng: Writing, Melody Yuen Man Ho: revision, Kenneth K.Y. Cheng: Writing- review and editing, conceptualization, funding acquisition, and supervision. All authors approved the manuscript for the submission.

ACKNOWLEDGMENTS

We would like to acknowledge the financial support from the National Natural Science Foundation of China (NSFC; grants 81970675 and 92357305 to KKY Cheng), the Research Grant Council for the Collaborative Research Fund (C5044-23G), and the General Research Fund (GRF), Hong Kong Research Grant Council (RGC) (15101221 to KKY Cheng). Additionally, we appreciate the support from the Shenzhen Municipal Science and Technology Innovation Commission (grant JCYJ20210324130202006 to KKY Cheng) and the PolyU-Project of

Strategic Importance (P0036848 to KKY Cheng) and PolyU-Project of RCMI (P0040979 to KKY Cheng).

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

ORCID

Kenneth King-yip Cheng  <https://orcid.org/0000-0002-7274-0839>

REFERENCES

- Layman DK. The role of leucine in weight loss diets and glucose homeostasis. *J Nutr.* 2003;133(1):261s-267s. doi:10.1093/jn/133.1.261S
- Biswas D, Duffley L, Pulinilkunnil T. Role of branched-chain amino acid-catabolizing enzymes in intertissue signaling, metabolic remodeling, and energy homeostasis. *FASEB j.* 2019;33(8):8711-8731. doi:10.1096/fj.201802842RR
- Newgard CB, An J, Bain JR, et al. A branched-chain amino acid-related metabolic signature that differentiates obese and lean humans and contributes to insulin resistance. *Cell Metab.* 2009;9(4):311-326. doi:10.1016/j.cmet.2009.02.002
- Hutson SM, Sweatt AJ, Lanoue KF. Branched-chain [corrected] amino acid metabolism: implications for establishing safe intakes. *J Nutr.* 2005;135(6):1557s-1564s. doi:10.1093/jn/135.6.1557S
- Anthony JC, Yoshizawa F, Anthony TG, Vary TC, Jefferson LS, Kimball SR. Leucine stimulates translation initiation in skeletal muscle of postabsorptive rats via a rapamycin-sensitive pathway. *J Nutr.* 2000;130(10):2413-2419. doi:10.1093/jn/130.10.2413
- Saha AK, Xu XJ, Lawson E, et al. Downregulation of AMPK accompanies leucine- and glucose-induced increases in protein synthesis and insulin resistance in rat skeletal muscle. *Diabetes.* 2010;59(10):2426-2434. doi:10.2337/db09-1870
- Xiao F, Huang Z, Li H, et al. Leucine deprivation increases hepatic insulin sensitivity via GCN2/mTOR/S6K1 and AMPK pathways. *Diabetes.* 2011;60(3):746-756. doi:10.2337/db10-1246
- Wang TJ, Larson MG, Vasani RS, et al. Metabolite profiles and the risk of developing diabetes. *Nat Med.* 2011;17(4):448-453. doi:10.1038/nm.2307
- Sun H, Olson KC, Gao C, et al. Catabolic defect of branched-chain amino acids promotes heart failure. *Circulation.* 2016;133(21):2038-2049. doi:10.1161/CIRCULATIONAHA.115.020226
- Tobias DK, Lawler PR, Harada PH, et al. Circulating branched-chain amino acids and incident cardiovascular disease in a prospective cohort of US women. *Circ Genom Precis Med.* 2018;11(4):e002157. doi:10.1161/CIRCGEN.118.002157
- Grenier-Larouche T, Coulter Kwee L, Deleaye Y, et al. Altered branched-chain α -keto acid metabolism is a feature of NAFLD in individuals with severe obesity. *JCI Insight.* 2022;7(15):7. doi:10.1172/jci.insight.159204
- Nilsen MS, Jersin R, Ulvik A, et al. 3-Hydroxyisobutyrate, a strong marker of insulin resistance in type 2 diabetes and obesity that modulates White and Brown adipocyte metabolism. *Diabetes.* 2020;69(9):1903-1916. doi:10.2337/db19-1174
- Jang C, Oh SF, Wada S, et al. A branched-chain amino acid metabolite drives vascular fatty acid transport and causes insulin resistance. *Nat Med.* 2016;22(4):421-426. doi:10.1038/nm.4057
- Zhang F, Zhao S, Yan W, et al. Branched chain amino acids cause liver injury in obese/diabetic mice by promoting adipocyte lipolysis and inhibiting hepatic autophagy. *EBioMedicine.* 2016;13:157-167. doi:10.1016/j.ebiom.2016.10.013
- Xiao F, Yu J, Guo Y, et al. Effects of individual branched-chain amino acids deprivation on insulin sensitivity and glucose metabolism in mice. *Metabolism.* 2014;63(6):841-850. doi:10.1016/j.metabol.2014.03.006
- Cummings NE, Williams EM, Kasza I, et al. Restoration of metabolic health by decreased consumption of branched-chain amino acids. *J Physiol.* 2018;596(4):623-645. doi:10.1113/JP275075
- Richardson NE, Konon EN, Schuster HS, et al. Lifelong restriction of dietary branched-chain amino acids has sex-specific benefits for frailty and lifespan in mice. *Nat Aging.* 2021;1(1):73-86. doi:10.1038/s43587-020-00006-2
- Karusheva Y, Koessler T, Strassburger K, et al. Short-term dietary reduction of branched-chain amino acids reduces meal-induced insulin secretion and modifies microbiome composition in type 2 diabetes: a randomized controlled crossover trial. *Am J Clin Nutr.* 2019;110(5):1098-1107. doi:10.1093/ajcn/nqz191
- White PJ, Lapworth AL, An J, et al. Branched-chain amino acid restriction in Zucker-fatty rats improves muscle insulin sensitivity by enhancing efficiency of fatty acid oxidation and acyl-glycine export. *Mol Metab.* 2016;5(7):538-551. doi:10.1016/j.molmet.2016.04.006
- Fontana L, Cummings NE, Arriola Apelo SI, et al. Decreased consumption of branched-chain amino acids improves metabolic health. *Cell Rep.* 2016;16(2):520-530. doi:10.1016/j.celrep.2016.05.092
- Hutson SM, Wallin R, Hall TR. Identification of mitochondrial branched chain aminotransferase and its isoforms in rat tissues. *J Biol Chem.* 1992;267(22):15681-15686. doi:10.1016/S0021-9258(19)49589-6
- Ding C, Li Y, Guo F, et al. A cell-type-resolved liver proteome. *Mol Cell Proteomics.* 2016;15(10):3190-3202. doi:10.1074/mcp.M116.060145
- Neinast MD, Jang C, Hui S, et al. Quantitative analysis of the whole-body metabolic fate of branched-chain amino acids. *Cell Metab.* 2019;29(2):417-29.e4. doi:10.1016/j.cmet.2018.10.013
- Patrick M, Gu Z, Zhang G, et al. Metabolite formation regulates branched-chain amino acid oxidation and homeostasis. *Nat Metab.* 2022;4(12):1775-1791. doi:10.1038/s42255-022-00689-4
- Brosnan JT, Brosnan ME. Branched-chain amino acids: enzyme and substrate regulation. *J Nutr.* 2006;136(1):207s-211s. doi:10.1093/jn/136.1.207S
- Vanweert F, Schrauwen P, Phielix E. Role of branched-chain amino acid metabolism in the pathogenesis of obesity and type 2 diabetes-related metabolic disturbances BCAA metabolism in type 2 diabetes. *Nutr Diabetes.* 2022;12(1):35. doi:10.1038/s41387-022-00213-3
- AE A, Chuang JL, Wynn RM, Turley S, Chuang DT, Hol WG. Crystal structure of human branched-chain alpha-ketoacid dehydrogenase and the molecular basis of multi-enzyme complex deficiency in maple syrup urine disease. *Structure.* 2000;8(3):277-291. doi:10.1016/S0969-2126(00)00105-2
- Lu G, Sun H, She P, et al. Protein phosphatase 2Cm is a critical regulator of branched-chain amino acid catabolism in mice and cultured cells. *J Clin Invest.* 2009;119(6):1678-1687. doi:10.1172/JCI38151
- Shimomura Y, Nanaumi N, Suzuki M, Popov KM, Harris RA. Purification and partial characterization of branched-chain alpha-ketoacid dehydrogenase kinase from rat liver and rat heart. *Arch Biochem Biophys.* 1990;283(2):293-299. doi:10.1016/0003-9861(90)90645-F
- Verkerke ARP, Wang D, Yoshida N, et al. BCAA-nitrogen flux in brown fat controls metabolic health independent of thermogenesis. *Cell.* 2024; 187: 2359-74, 2359-2374, doi:10.1016/j.cell.2024.03.030.
- Buerger T, Steinfeldt J, Ruyoga G, et al. Metabolomic profiles predict individual multidisease outcomes. *Nat Med.* 2022;28(11):2309-2320. doi:10.1038/s41591-022-01980-3
- She P, Olson KC, Kadota Y, et al. Leucine and protein metabolism in obese Zucker rats. *PLoS ONE.* 2013;8(3):e59443. doi:10.1371/journal.pone.0059443
- Ramzan I, Ardavani A, Vanweert F, Mellett A, Atherton PJ, Idris I. The association between circulating branched chain amino acids and

- the temporal risk of developing type 2 diabetes mellitus: a Systematic Review & Meta-Analysis. *Nutrients*. 2022;14(20):4411. doi:10.3390/nu14204411
34. Lynch CJ, Adams SH. Branched-chain amino acids in metabolic signalling and insulin resistance. *Nat Rev Endocrinol*. 2014;10(12):723-736. doi:10.1038/nrendo.2014.171
 35. Giesbertz P, Padberg I, Rein D, et al. Metabolite profiling in plasma and tissues of Ob/Ob and db/db mice identifies novel markers of obesity and type 2 diabetes. *Diabetologia*. 2015;58(9):2133-2143. doi:10.1007/s00125-015-3656-y
 36. Yu D, Richardson NE, Green CL, et al. The adverse metabolic effects of branched-chain amino acids are mediated by isoleucine and valine. *Cell Metab*. 2021;33(5):905-22.e6. doi:10.1016/j.cmet.2021.03.025
 37. Xu M, Qi Q, Liang J, et al. Genetic determinant for amino acid metabolites and changes in body weight and insulin resistance in response to weight-loss diets: the preventing overweight using novel dietary strategies (POUNDS LOST) trial. *Circulation*. 2013;127(12):1283-1289. doi:10.1161/CIRCULATIONAHA.112.000586
 38. Vargas-Morales JM, Guizar-Heredia R, Méndez-García AL, et al. Association of BCAT2 and BCKDH polymorphisms with clinical, anthropometric and biochemical parameters in young adults. *Nutr Metab Cardiovasc Dis*. 2021;31(11):3210-3218. doi:10.1016/j.numecd.2021.07.011
 39. Hu W, Liu Z, Yu W, et al. Effects of PPM1K rs1440581 and rs7678928 on serum branched-chain amino acid levels and risk of cardiovascular disease. *Ann Med*. 2021;53(1):1316-1326. doi:10.1080/07853890.2021.1965204
 40. Lotta LA, Scott RA, Sharp SJ, et al. Genetic predisposition to an impaired metabolism of the branched-chain amino acids and risk of type 2 diabetes: a Mendelian randomisation analysis. *PLoS Med*. 2016;13(11):e1002179. doi:10.1371/journal.pmed.1002179
 41. Wang Q, Holmes MV, Davey Smith G, Ala-Korpela M. Genetic support for a causal role of insulin resistance on circulating branched-chain amino acids and inflammation. *Diabetes Care*. 2017;40(12):1779-1786. doi:10.2337/dc17-1642
 42. Mahendran Y, Jonsson A, Have CT, et al. Genetic evidence of a causal effect of insulin resistance on branched-chain amino acid levels. *Diabetologia*. 2017;60(5):873-878. doi:10.1007/s00125-017-4222-6
 43. Menni C, Fauman E, Erte I, et al. Biomarkers for type 2 diabetes and impaired fasting glucose using a nontargeted metabolomics approach. *Diabetes*. 2013;62(12):4270-4276. doi:10.2337/db13-0570
 44. Mardinoglu A, Gogg S, Lotta LA, et al. Elevated plasma levels of 3-Hydroxyisobutyric acid are associated with incident type 2 diabetes. *EBioMedicine*. 2018;27:151-155. doi:10.1016/j.ebiom.2017.12.008
 45. Bjune MS, Lawrence-Archer L, Laupsa-Borge J, et al. Metabolic role of the hepatic valine/3-hydroxyisobutyrate (3-HIB) pathway in fatty liver disease. *EBioMedicine*. 2023;91:104569. doi:10.1016/j.ebiom.2023.104569
 46. Zhou M, Shao J, Wu CY, et al. Targeting BCAA catabolism to treat obesity-associated insulin resistance. *Diabetes*. 2019;68(9):1730-1746. doi:10.2337/db18-0927
 47. Hernández-Alvarez MI, Díaz-Ramos A, Berdasco M, et al. Early-onset and classical forms of type 2 diabetes show impaired expression of genes involved in muscle branched-chain amino acids metabolism. *Sci Rep*. 2017;7(1):13850. doi:10.1038/s41598-017-14120-6
 48. She P, Van Horn C, Reid T, Hutson SM, Cooney RN, Lynch CJ. Obesity-related elevations in plasma leucine are associated with alterations in enzymes involved in branched-chain amino acid metabolism. *Am J Physiol Endocrinol Metab*. 2007;293(6):E1552-E1563. doi:10.1152/ajpendo.00134.2007
 49. White PJ, McGarrah RW, Grimsrud PA, et al. The BCKDH kinase and phosphatase integrate BCAA and lipid metabolism via regulation of ATP-citrate Lyase. *Cell Metab*. 2018;27(6):1281-93.e7. doi:10.1016/j.cmet.2018.04.015
 50. Herman MA, She P, Peroni OD, Lynch CJ, Kahn BB. Adipose tissue branched chain amino acid (BCAA) metabolism modulates circulating BCAA levels. *J Biol Chem*. 2010;285(15):11348-11356. doi:10.1074/jbc.M109.075184
 51. Luo L, Liu M. Adipose tissue in control of metabolism. *J Endocrinol*. 2016;231:R77-R99.
 52. Choe SS, Huh JY, Hwang IJ, Kim JI, Kim JB. Adipose tissue remodeling: its role in energy metabolism and metabolic disorders. *Front Endocrinol (Lausanne)*. 2016;7:30. doi:10.3389/fendo.2016.00030
 53. Tischler ME, Goldberg AL. Leucine degradation and release of glutamine and alanine by adipose tissue. *J Biol Chem*. 1980;255(17):8074-8081. doi:10.1016/S0021-9258(19)70609-7
 54. Scalise M, Pochini L, Console L, Lusso MA, Indiveri C. The human SLC1A5 (ASCT2) amino acid transporter: from function to structure and role in cell biology. *Front Cell Dev Biol*. 2018;6:96. doi:10.3389/fcell.2018.00096
 55. Utsunomiya-Tate N, Endou H, Kanai Y. Cloning and functional characterization of a system ASC-like Na⁺-dependent neutral amino acid transporter*. *J Biol Chem*. 1996;271(25):14883-14890. doi:10.1074/jbc.271.25.14883
 56. Segawa H, Fukasawa Y, Miyamoto K, Takeda E, Endou H, Kanai Y. Identification and functional characterization of a Na⁺-independent neutral amino acid transporter with broad substrate selectivity. *J Biol Chem*. 1999;274(28):19745-19751. doi:10.1074/jbc.274.28.19745
 57. Wallace M, Green CR, Roberts LS, et al. Enzyme promiscuity drives branched-chain fatty acid synthesis in adipose tissues. *Nat Chem Biol*. 2018;14(11):1021-1031. doi:10.1038/s41589-018-0132-2
 58. Green CR, Wallace M, Divakaruni AS, et al. Branched-chain amino acid catabolism fuels adipocyte differentiation and lipogenesis. *Nat Chem Biol*. 2016;12(1):15-21. doi:10.1038/nchembio.1961
 59. Melnik BC. Leucine signaling in the pathogenesis of type 2 diabetes and obesity. *World J Diabetes*. 2012;3(3):38-53. doi:10.4239/wjd.v3.i3.38
 60. Zaganjor E, Yoon H, Spinelli JB, et al. SIRT4 is an early regulator of branched-chain amino acid catabolism that promotes adipogenesis. *Cell Rep*. 2021;36(2):109345. doi:10.1016/j.celrep.2021.109345
 61. Sun H, Wang Y. A new branch connecting thermogenesis and diabetes. *Nat Metab*. 2019;1(9):845-846. doi:10.1038/s42255-019-0112-1
 62. Yoneshiro T, Wang Q, Tajima K, et al. BCAA catabolism in brown fat controls energy homeostasis through SLC25A44. *Nature*. 2019;572(7771):614-619. doi:10.1038/s41586-019-1503-x
 63. Turer AT, Scherer PE. Adiponectin: mechanistic insights and clinical implications. *Diabetologia*. 2012;55(9):2319-2326. doi:10.1007/s00125-012-2598-x
 64. Yamauchi T, Kamon J, Minokoshi Y, et al. Adiponectin stimulates glucose utilization and fatty-acid oxidation by activating AMP-activated protein kinase. *Nat Med*. 2002;8(11):1288-1295. doi:10.1038/nm788
 65. Cheng KK, Lam KS, Wang B, Xu A. Signaling mechanisms underlying the insulin-sensitizing effects of adiponectin. *Best Pract Res Clin Endocrinol Metab*. 2014;28(1):3-13. doi:10.1016/j.beem.2013.06.006
 66. Li S, Shin HJ, Ding EL, van Dam RM. Adiponectin levels and risk of type 2 diabetes: a systematic review and meta-analysis. *Jama*. 2009;302(2):179-188. doi:10.1001/jama.2009.976
 67. Weyer C, Funahashi T, Tanaka S, et al. Hypoadiponectinemia in obesity and type 2 diabetes: close association with insulin resistance and hyperinsulinemia. *J Clin Endocrinol Metab*. 2001;86(5):1930-1935. doi:10.1210/jcem.86.5.7463

68. Lian K, Du C, Liu Y, et al. Impaired adiponectin signaling contributes to disturbed catabolism of branched-chain amino acids in diabetic mice. *Diabetes*. 2015;64(1):49-59. doi:10.2337/db14-0312
69. Kilroy GE, Zhang X, Floyd ZE. PPAR-gamma AF-2 domain functions as a component of a ubiquitin-dependent degradation signal. *Obesity (Silver Spring)*. 2009;17(4):665-673. doi:10.1038/oby.2008.616
70. Anghel SI, Wahli W. Fat poetry: a kingdom for PPAR gamma. *Cell Res*. 2007;17(6):486-511. doi:10.1038/cr.2007.48
71. Hsiao G, Chapman J, Ofrecio JM, et al. Multi-tissue, selective PPAR γ modulation of insulin sensitivity and metabolic pathways in obese rats. *Am J Physiol Endocrinol Metab*. 2011;300(1):E164-E174. doi:10.1152/ajpendo.00219.2010
72. Blanchard PG, Moreira RJ, Castro É, et al. PPAR γ is a major regulator of branched-chain amino acid blood levels and catabolism in white and brown adipose tissues. *Metabolism*. 2018;89:27-38. doi:10.1016/j.metabol.2018.09.007
73. Cifarelli V, Beeman SC, Smith GI, et al. Decreased adipose tissue oxygenation associates with insulin resistance in individuals with obesity. *J Clin Invest*. 2020;130(12):6688-6699. doi:10.1172/JCI141828
74. Wang W, Zhang F, Xia Y, et al. Defective branched chain amino acid catabolism contributes to cardiac dysfunction and remodeling following myocardial infarction. *Am J Physiol Heart Circ Physiol*. 2016;311(5):H1160-H1169. doi:10.1152/ajpheart.00114.2016
75. Tchkonina T, Morbeck DE, Von Zglinicki T, et al. Fat tissue, aging, and cellular senescence. *Aging Cell*. 2010;9(5):667-684. doi:10.1111/j.1474-9726.2010.00608.x
76. Wu KK, Cheung SW, Cheng KK. NLRP3 Inflammasome activation in adipose tissues and its implications on metabolic diseases. *Int J Mol Sci*. 2020;21(11):4184. doi:10.3390/ijms21114184
77. Cho S, Ying F, Sweeney G. Sterile inflammation and the NLRP3 inflammasome in cardiometabolic disease. *Biom J*. 2023;46(5):100624. doi:10.1016/j.bj.2023.100624
78. Zhao S, Zhou L, Wang Q, et al. Elevated branched-chain amino acid promotes atherosclerosis progression by enhancing mitochondrial-to-nuclear H(2)O(2)-disulfide HMGB1 in macrophages. *Redox Biol*. 2023;62:102696. doi:10.1016/j.redox.2023.102696
79. Son M, Porat A, He M, et al. C1q and HMGB1 reciprocally regulate human macrophage polarization. *Blood*. 2016;128(18):2218-2228. doi:10.1182/blood-2016-05-719757
80. Liu S, Li L, Lou P, et al. Elevated branched-chain α -keto acids exacerbate macrophage oxidative stress and chronic inflammatory damage in type 2 diabetes mellitus. *Free Radic Biol Med*. 2021;175:141-154. doi:10.1016/j.freeradbiomed.2021.08.240
81. Burrill JS, Long EK, Reilly B, et al. Inflammation and ER stress regulate branched-chain amino acid uptake and metabolism in adipocytes. *Mol Endocrinol*. 2015;29(3):411-420. doi:10.1210/me.2014-1275
82. Seale P, Conroe HM, Estall J, et al. Prdm16 determines the thermogenic program of subcutaneous white adipose tissue in mice. *J Clin Invest*. 2011;121(1):96-105. doi:10.1172/JCI44271
83. Ma QX, Zhu WY, Lu XC, et al. BCAA-BCKA axis regulates WAT browning through acetylation of PRDM16. *Nat Metab*. 2022;4(1):106-122. doi:10.1038/s42255-021-00520-6
84. Wang F, Xiao F, Du L, et al. Activation of GCN2 in macrophages promotes white adipose tissue browning and lipolysis under leucine deprivation. *FASEB j*. 2021;35(6):e21652. doi:10.1096/fj.202100061RR
85. Wolfe RR. The underappreciated role of muscle in health and disease. *Am J Clin Nutr*. 2006;84(3):475-482. doi:10.1093/ajcn/84.3.475
86. Stump CS, Henriksen EJ, Wei Y, Sowers JR. The metabolic syndrome: role of skeletal muscle metabolism. *Ann Med*. 2006;38(6):389-402. doi:10.1080/07853890600888413
87. Sakamoto K, Holman GD. Emerging role for AS160/TBC1D4 and TBC1D1 in the regulation of GLUT4 traffic. *Am J Physiol Endocrinol Metab*. 2008;295(1):E29-E37. doi:10.1152/ajpendo.90331.2008
88. Cheng KK, Zhu W, Chen B, et al. The adaptor protein APPL2 inhibits insulin-stimulated glucose uptake by interacting with TBC1D1 in skeletal muscle. *Diabetes*. 2014;63(11):3748-3758. doi:10.2337/db14-0337
89. Suryawan A, Hawes JW, Harris RA, Shimomura Y, Jenkins AE, Hutson SM. A molecular model of human branched-chain amino acid metabolism. *Am J Clin Nutr*. 1998;68(1):72-81. doi:10.1093/ajcn/68.1.72
90. Holeček M. Branched-chain amino acids in health and disease: metabolism, alterations in blood plasma, and as supplements. *Nutr Metab (Lond)*. 2018;15(1):33. doi:10.1186/s12986-018-0271-1
91. Gran P, Cameron-Smith D. The actions of exogenous leucine on mTOR signalling and amino acid transporters in human myotubes. *BMC Physiol*. 2011;11(1):10. doi:10.1186/1472-6793-11-10
92. Bodine SC, Stitt TN, Gonzalez M, et al. Akt/mTOR pathway is a crucial regulator of skeletal muscle hypertrophy and can prevent muscle atrophy in vivo. *Nat Cell Biol*. 2001;3(11):1014-1019. doi:10.1038/ncb1101-1014
93. D'Antona G, Ragni M, Cardile A, et al. Branched-chain amino acid supplementation promotes survival and supports cardiac and skeletal muscle mitochondrial biogenesis in middle-aged mice. *Cell Metab*. 2010;12(4):362-372. doi:10.1016/j.cmet.2010.08.016
94. Shimomura Y, Murakami T, Nakai N, Nagasaki M, Harris RA. Exercise promotes BCAA catabolism: effects of BCAA supplementation on skeletal muscle during exercise. *J Nutr*. 2004;134(6):1583s-1587s. doi:10.1093/jn/134.6.1583S
95. Shimomura Y, Fujii H, Suzuki M, Murakami T, Fujitsuka N, Nakai N. Branched-chain alpha-keto acid dehydrogenase complex in rat skeletal muscle: regulation of the activity and gene expression by nutrition and physical exercise. *J Nutr*. 1995;125(6 Suppl):1762s-1765s. doi:10.1093/jn/125.suppl_6.1762S
96. Puigserver P, Spiegelman BM. Peroxisome proliferator-activated receptor-gamma coactivator 1 alpha (PGC-1 alpha): transcriptional coactivator and metabolic regulator. *Endocr Rev*. 2003;24(1):78-90. doi:10.1210/er.2002-0012
97. Hatazawa Y, Tadaishi M, Nagaie Y, et al. PGC-1alpha-mediated branched-chain amino acid metabolism in the skeletal muscle. *PLoS ONE*. 2014;9(3):e91006. doi:10.1371/journal.pone.0091006
98. Sjögren RJO, Rizo-Roca D, Chibalin AV, et al. Branched-chain amino acid metabolism is regulated by ERR α in primary human myotubes and is further impaired by glucose loading in type 2 diabetes. *Diabetologia*. 2021;64(9):2077-2091. doi:10.1007/s00125-021-05481-9
99. Liu Y, Turdi S, Park T, et al. Adiponectin corrects high-fat diet-induced disturbances in muscle metabolomic profile and whole-body glucose homeostasis. *Diabetes*. 2013;62(3):743-752. doi:10.2337/db12-0687
100. Yoshizawa F. Regulation of protein synthesis by branched-chain amino acids in vivo. *Biochem Biophys Res Commun*. 2004;313(2):417-422. doi:10.1016/j.bbrc.2003.07.013
101. Lerin C, Goldfine AB, Boes T, et al. Defects in muscle branched-chain amino acid oxidation contribute to impaired lipid metabolism. *Mol Metab*. 2016;5(10):926-936. doi:10.1016/j.molmet.2016.08.001
102. Yoon MS. The emerging role of branched-chain amino acids in insulin resistance and metabolism. *Nutrients*. 2016;8(7):405. doi:10.3390/nu8070405
103. Crossland H, Smith K, Idris I, Phillips BE, Atherton PJ, Wilkinson DJ. Exploring mechanistic links between extracellular branched-chain amino acids and muscle insulin resistance: an in vitro approach. *Am J Physiol Cell Physiol*. 2020;319(6):C1151-C1157. doi:10.1152/ajpcell.00377.2020
104. Vaughan RA, Garcia-Smith R, Gannon NP, Bisoffi M, Trujillo KA, Conn CA. Leucine treatment enhances oxidative capacity through

- complete carbohydrate oxidation and increased mitochondrial density in skeletal muscle cells. *Amino Acids*. 2013;45(4):901-911. doi:[10.1007/s00726-013-1538-5](https://doi.org/10.1007/s00726-013-1538-5)
105. Rivera ME, Rivera CN, Vaughan RA. Excess branched-chain amino acids alter myotube metabolism and substrate preference which is worsened by concurrent insulin resistance. *Endocrine*. 2022;76(1):18-28. doi:[10.1007/s12020-021-02939-z](https://doi.org/10.1007/s12020-021-02939-z)
 106. Sánchez-González C, Nuevo-Tapióles C, Herrero Martín JC, et al. Dysfunctional oxidative phosphorylation shunts branched-chain amino acid catabolism onto lipogenesis in skeletal muscle. *EMBO j*. 2020;39(14):e103812. doi:[10.15252/embj.2019103812](https://doi.org/10.15252/embj.2019103812)
 107. Biswas D, Dao KT, Mercer A, et al. Branched-chain ketoacid overload inhibits insulin action in the muscle. *J Biol Chem*. 2020;295(46):15597-15621. doi:[10.1074/jbc.RA120.013121](https://doi.org/10.1074/jbc.RA120.013121)
 108. Moghei M, Tavajohi-Fini P, Beatty B, Adegoke OA. Ketoisocaproic acid, a metabolite of leucine, suppresses insulin-stimulated glucose transport in skeletal muscle cells in a BCAT2-dependent manner. *Am J Physiol Cell Physiol*. 2016;311(3):C518-C527. doi:[10.1152/ajpcell.00062.2016](https://doi.org/10.1152/ajpcell.00062.2016)
 109. Newgard CB. Interplay between lipids and branched-chain amino acids in development of insulin resistance. *Cell Metab*. 2012;15(5):606-614. doi:[10.1016/j.cmet.2012.01.024](https://doi.org/10.1016/j.cmet.2012.01.024)
 110. Lee J, Vijayakumar A, White PJ, et al. BCAA supplementation in mice with diet-induced obesity alters the metabolome without impairing glucose homeostasis. *Endocrinology*. 2021;162(7). doi:[10.1210/endo.2021-0062](https://doi.org/10.1210/endo.2021-0062)
 111. Hong S, Zhou W, Fang B, et al. Dissociation of muscle insulin sensitivity from exercise endurance in mice by HDAC3 depletion. *Nat Med*. 2017;23(2):223-234. doi:[10.1038/nm.4245](https://doi.org/10.1038/nm.4245)
 112. Rui L. Energy metabolism in the liver. *Compr Physiol*. 2014;4(1):177-197. doi:[10.1002/cphy.c130024](https://doi.org/10.1002/cphy.c130024)
 113. Horton JD, Goldstein JL, Brown MS. SREBPs: activators of the complete program of cholesterol and fatty acid synthesis in the liver. *J Clin Invest*. 2002;109(9):1125-1131. doi:[10.1172/JCI0215593](https://doi.org/10.1172/JCI0215593)
 114. Charlton MR. Protein metabolism and liver disease. *Baillieres Clin Endocrinol Metab*. 1996;10(4):617-635. doi:[10.1016/S0950-351X\(96\)80771-3](https://doi.org/10.1016/S0950-351X(96)80771-3)
 115. Lake AD, Novak P, Shipkova P, et al. Branched chain amino acid metabolism profiles in progressive human nonalcoholic fatty liver disease. *Amino Acids*. 2015;47(3):603-615. doi:[10.1007/s00726-014-1894-9](https://doi.org/10.1007/s00726-014-1894-9)
 116. Boday S, Fotiadis D, Stoeger C, Kanai Y, Palacín M. The small SLC43 family: facilitator system I amino acid transporters and the orphan ECG1. *Mol Aspects Med*. 2013;34(2-3):638-645. doi:[10.1016/j.mam.2012.12.006](https://doi.org/10.1016/j.mam.2012.12.006)
 117. Mallet LE, Exton JH, Park CR. Control of gluconeogenesis from amino acids in the perfused rat liver. *J Biol Chem*. 1969;244(20):5713-5723. doi:[10.1016/S0021-9258\(18\)63618-X](https://doi.org/10.1016/S0021-9258(18)63618-X)
 118. Ijichi C, Matsumura T, Tsuji T, Eto Y. Branched-chain amino acids promote albumin synthesis in rat primary hepatocytes through the mTOR signal transduction system. *Biochem Biophys Res Commun*. 2003;303(1):59-64. doi:[10.1016/S0006-291X\(03\)00295-X](https://doi.org/10.1016/S0006-291X(03)00295-X)
 119. Kalhan SC, Guo L, Edmison J, et al. Plasma metabolomic profile in nonalcoholic fatty liver disease. *Metabolism*. 2011;60(3):404-413. doi:[10.1016/j.metabol.2010.03.006](https://doi.org/10.1016/j.metabol.2010.03.006)
 120. Mardinoglu A, Agren R, Kampf C, Asplund A, Uhlen M, Nielsen J. Genome-scale metabolic modelling of hepatocytes reveals serine deficiency in patients with non-alcoholic fatty liver disease. *Nat Commun*. 2014;5(1):3083. doi:[10.1038/ncomms4083](https://doi.org/10.1038/ncomms4083)
 121. Kawanaka M, Nishino K, Oka T, et al. Tyrosine levels are associated with insulin resistance in patients with nonalcoholic fatty liver disease. *Hepat Med*. 2015;7:29-35. doi:[10.2147/HMER.S79100](https://doi.org/10.2147/HMER.S79100)
 122. Gaggini M, Carli F, Rosso C, et al. Altered amino acid concentrations in NAFLD: impact of obesity and insulin resistance. *Hepatology*. 2018;67(1):145-158. doi:[10.1002/hep.29465](https://doi.org/10.1002/hep.29465)
 123. Guo F, Chen R, Kong L, et al. Effects of serum branched-chain amino acids on nonalcoholic fatty liver disease and subsequent cardiovascular disease. *Hepatol Int*. 2022;16(6):1424-1434. doi:[10.1007/s12072-022-10387-8](https://doi.org/10.1007/s12072-022-10387-8)
 124. Nezami Ranjbar MR, Luo Y, Di Poto C, et al. GC-MS based plasma metabolomics for identification of candidate biomarkers for hepatocellular carcinoma in Egyptian cohort. *PLoS ONE*. 2015;10(6):e0127299. doi:[10.1371/journal.pone.0127299](https://doi.org/10.1371/journal.pone.0127299)
 125. Lischka J, Schanzer A, Hojreh A, et al. A branched-chain amino acid-based metabolic score can predict liver fat in children and adolescents with severe obesity. *Pediatr Obes*. 2021;16(4):e12739. doi:[10.1111/ijpo.12739](https://doi.org/10.1111/ijpo.12739)
 126. Zhao E, Giamberardino SN, Pagidipati NJ, et al. Branched-chain amino acids in computed tomography-defined adipose depots and coronary artery disease: a PROMISE trial biomarker substudy. *J Am Heart Assoc*. 2023;12(11):e028410. doi:[10.1161/JAHA.122.028410](https://doi.org/10.1161/JAHA.122.028410)
 127. Ni Y, Qian L, Siliceo SL, et al. Resistant starch decreases intrahepatic triglycerides in patients with NAFLD via gut microbiome alterations. *Cell Metab*. 2023;35(9):1530-47.e8. doi:[10.1016/j.cmet.2023.08.002](https://doi.org/10.1016/j.cmet.2023.08.002)
 128. Ericksen RE, Lim SL, McDonnell E, et al. Loss of BCAA catabolism during carcinogenesis enhances mTORC1 activity and promotes tumor development and progression. *Cell Metab* 2019; 29: 1151-65. doi:[10.1016/j.cmet.2018.12.020](https://doi.org/10.1016/j.cmet.2018.12.020)
 129. Holecsek M. Branched-chain amino acid supplementation in treatment of liver cirrhosis: updated views on how to attenuate their harmful effects on cataplerosis and ammonia formation. *Nutrition*. 2017;41:80-85. doi:[10.1016/j.nut.2017.04.003](https://doi.org/10.1016/j.nut.2017.04.003)
 130. Hamill MJ, Afeyan R, Chakravarthy MV, Tramontin T. Endogenous metabolic modulators: emerging therapeutic potential of amino acids. *iScience*. 2020;23(10):101628. doi:[10.1016/j.isci.2020.101628](https://doi.org/10.1016/j.isci.2020.101628)
 131. Honda T, Ishigami M, Luo F, et al. Branched-chain amino acids alleviate hepatic steatosis and liver injury in choline-deficient high-fat diet induced NASH mice. *Metabolism*. 2017;69:177-187. doi:[10.1016/j.metabol.2016.12.013](https://doi.org/10.1016/j.metabol.2016.12.013)
 132. Iwao M, Gotoh K, Arakawa M, et al. Supplementation of branched-chain amino acids decreases fat accumulation in the liver through intestinal microbiota-mediated production of acetic acid. *Sci Rep*. 2020;10(1):18768. doi:[10.1038/s41598-020-75542-3](https://doi.org/10.1038/s41598-020-75542-3)
 133. Kolwicz SC Jr, Purohit S, Tian R. Cardiac metabolism and its interactions with contraction, growth, and survival of cardiomyocytes. *Circ Res*. 2013;113(5):603-616. doi:[10.1161/CIRCRESAHA.113.302095](https://doi.org/10.1161/CIRCRESAHA.113.302095)
 134. Wang LY, Chen C. Energy metabolism homeostasis in cardiovascular diseases. *J Geriatr Cardiol*. 2021;18(12):1044-1057. doi:[10.11909/j.issn.1671-5411.2021.12.006](https://doi.org/10.11909/j.issn.1671-5411.2021.12.006)
 135. Li T, Zhang Z, Kolwicz SC Jr, et al. Defective branched-chain amino acid catabolism disrupts glucose metabolism and sensitizes the heart to ischemia-reperfusion injury. *Cell Metab*. 2017;25(2):374-385. doi:[10.1016/j.cmet.2016.11.005](https://doi.org/10.1016/j.cmet.2016.11.005)
 136. Uddin GM, Karwi QG, Pherwani S, et al. Deletion of BCATm increases insulin-stimulated glucose oxidation in the heart. *Metabolism*. 2021;124:154871. doi:[10.1016/j.metabol.2021.154871](https://doi.org/10.1016/j.metabol.2021.154871)
 137. Uddin GM, Zhang L, Shah S, et al. Impaired branched chain amino acid oxidation contributes to cardiac insulin resistance in heart failure. *Cardiovasc Diabetol*. 2019;18(1):86. doi:[10.1186/s12933-019-0892-3](https://doi.org/10.1186/s12933-019-0892-3)
 138. Walejko JM, Christopher BA, Crown SB, et al. Branched-chain alpha-ketoacids are preferentially reaminated and activate protein synthesis in the heart. *Nat Commun*. 2021;12(1):1680. doi:[10.1038/s41467-021-21962-2](https://doi.org/10.1038/s41467-021-21962-2)
 139. Tso SC, Gui WJ, Wu CY, et al. Benzothioephene carboxylate derivatives as novel allosteric inhibitors of branched-chain α -ketoacid dehydrogenase kinase. *J Biol Chem*. 2014;289(30):20583-20593. doi:[10.1074/jbc.M114.569251](https://doi.org/10.1074/jbc.M114.569251)

140. Zhao X, Zhang X, Pei J, Liu Y, Niu W, Sun H. Targeting BCAA metabolism to potentiate metformin's therapeutic efficacy in the treatment of diabetes in mice. *Diabetologia*. 2023;66(11):2139-2153. doi:10.1007/s00125-023-05985-6
141. Bollinger E, Peloquin M, Libera J, et al. BDK inhibition acts as a catabolic switch to mimic fasting and improve metabolism in mice. *Mol Metab*. 2022;66:101611. doi:10.1016/j.molmet.2022.101611
142. Roth Flach RJ, Bollinger E, Reyes AR, et al. Small molecule branched-chain ketoacid dehydrogenase kinase (BDK) inhibitors with opposing effects on BDK protein levels. *Nat Commun*. 2023;14(1):4812. doi:10.1038/s41467-023-40536-y
143. McMurray JJ, Holman RR, Haffner SM, et al. Effect of valsartan on the incidence of diabetes and cardiovascular events. *N Engl J Med*. 2010;362(16):1477-1490. doi:10.1056/NEJMoa1001121
144. Kitaura Y, Shindo D, Ogawa T, Sato A, Shimomura Y. Antihypertensive drug valsartan as a novel BDK inhibitor. *Pharmacol Res*. 2021;167:105518. doi:10.1016/j.phrs.2021.105518
145. Vanweert F, Neinast M, Tapia EE, et al. A randomized placebo-controlled clinical trial for pharmacological activation of BCAA catabolism in patients with type 2 diabetes. *Nat Commun*. 2022;13(1):3508. doi:10.1038/s41467-022-31249-9
146. McCommis KS, Hodges WT, Brunt EM, et al. Targeting the mitochondrial pyruvate carrier attenuates fibrosis in a mouse model of nonalcoholic steatohepatitis. *Hepatology*. 2017;65(5):1543-1556. doi:10.1002/hep.29025
147. Ferguson D, Eichler SJ, Yiew NKH, et al. Mitochondrial pyruvate carrier inhibition initiates metabolic crosstalk to stimulate branched chain amino acid catabolism. *Mol Metab*. 2023;70:101694. doi:10.1016/j.molmet.2023.101694
148. Araki K, Masaki T, Katsuragi I, Tanaka K, Kakuma T, Yoshimatsu H. Telmisartan prevents obesity and increases the expression of uncoupling protein 1 in diet-induced obese mice. *Hypertension*. 2006;48(1):51-57. doi:10.1161/01.HYP.0000225402.69580.1d
149. Pedersen HK, Gudmundsdottir V, Nielsen HB, et al. Human gut microbes impact host serum metabolome and insulin sensitivity. *Nature*. 2016;535(7612):376-381. doi:10.1038/nature18646
150. Chen H, Nie Q, Hu J, Huang X, Yin J, Nie S. Multiomics approach to explore the amelioration mechanisms of Glucomannans on the metabolic disorder of type 2 diabetic rats. *J Agric Food Chem*. 2021;69(8):2632-2645. doi:10.1021/acs.jafc.0c07871
151. Hoyles L, Fernández-Real JM, Federici M, et al. Molecular phenomics and metagenomics of hepatic steatosis in non-diabetic obese women. *Nat Med*. 2018;24(7):1070-1080. doi:10.1038/s41591-018-0061-3
152. Yue SJ, Liu J, Wang AT, et al. Berberine alleviates insulin resistance by reducing peripheral branched-chain amino acids. *Am J Physiol Endocrinol Metab*. 2019;316(1):E73-e85. doi:10.1152/ajpendo.00256.2018
153. Zhang L, Yue Y, Shi M, et al. Dietary *Luffa cylindrica* (L.) Roem promotes branched-chain amino acid catabolism in the circulation system via gut microbiota in diet-induced obese mice. *Food Chem*. 2020;320:126648. doi:10.1016/j.foodchem.2020.126648
154. Guo J, Tao H, Cao Y, Ho CT, Jin S, Huang Q. Prevention of obesity and type 2 diabetes with aged citrus Peel (Chenpi) extract. *J Agric Food Chem*. 2016;64(10):2053-2061. doi:10.1021/acs.jafc.5b06157
155. Zeng SL, Li SZ, Xiao PT, et al. Citrus polymethoxyflavones attenuate metabolic syndrome by regulating gut microbiome and amino acid metabolism. *Sci Adv*. 2020;6(1):eaax6208. doi:10.1126/sciadv.aax6208
156. Hu P, Li Z, Chen M, et al. Structural elucidation and protective role of a polysaccharide from *Sargassum fusiforme* on ameliorating learning and memory deficiencies in mice. *Carbohydr Polym*. 2016;139:150-158. doi:10.1016/j.carbpol.2015.12.019
157. Liu J, Wu S, Cheng Y, et al. *Sargassum fusiforme* alginate relieves hyperglycemia and modulates intestinal microbiota and metabolites in type 2 diabetic mice. *Nutrients*. 2021;13(8):2887. doi:10.3390/nu13082887
158. Bao M, Hou K, Xin C, et al. *Portulaca oleracea* L. extract alleviated type 2 diabetes via modulating the gut microbiota and serum branched-chain amino acid metabolism. *Mol Nutr Food Res*. 2022;66:e2101030.
159. Li Q, Liu F, Liu J, Liao S, Zou Y. Mulberry leaf polyphenols and fiber induce synergistic Antiobesity and display a modulation effect on gut microbiota and metabolites. *Nutrients*. 2019;11(5):1017. doi:10.3390/nu11051017
160. Zheng XX, Li DX, Li YT, et al. Mulberry leaf water extract alleviates type 2 diabetes in mice via modulating gut microbiota-host co-metabolism of branched-chain amino acid. *Phytother Res*. 2023;37(8):3195-3210. doi:10.1002/ptr.7822
161. Grøntved A, Rimm EB, Willett WC, Andersen LB, Hu FB. A prospective study of weight training and risk of type 2 diabetes mellitus in men. *Arch Intern Med*. 2012;172(17):1306-1312. doi:10.1001/archinternmed.2012.3138
162. Kobayashi R, Shimomura Y, Murakami T, et al. Hepatic branched-chain alpha-keto acid dehydrogenase complex in female rats: activation by exercise and starvation. *J Nutr Sci Vitaminol (Tokyo)*. 1999;45(3):303-309. doi:10.3177/jnsv.45.303
163. Xiao W, Chen P, Liu X, Zhao L. The impaired function of macrophages induced by strenuous exercise could not be ameliorated by BCAA supplementation. *Nutrients*. 2015;7(10):8645-8656. doi:10.3390/nu7105425
164. Böhm A, Weigert C, Staiger H, Häring HU. Exercise and diabetes: relevance and causes for response variability. *Endocrine*. 2016;51(3):390-401. doi:10.1007/s12020-015-0792-6
165. Liu Y, Wang Y, Ni Y, et al. Gut microbiome fermentation determines the efficacy of exercise for diabetes prevention. *Cell Metab*. 2020;31(1):77-91.e5. doi:10.1016/j.cmet.2019.11.001
166. Zhang H, Xiang L, Huo M, et al. Branched-chain amino acid supplementation impairs insulin sensitivity and promotes lipogenesis during exercise in diet-induced obese mice. *Obesity (Silver Spring)*. 2022;30(6):1205-1218. doi:10.1002/oby.23394
167. Cheng Y, Meng Q, Wang C, et al. Leucine deprivation decreases fat mass by stimulation of lipolysis in white adipose tissue and upregulation of uncoupling protein 1 (UCP1) in brown adipose tissue. *Diabetes*. 2010;59(1):17-25. doi:10.2337/db09-0929
168. Yuan F, Jiang H, Yin H, et al. Activation of GCN2/ATF4 signals in amygdalar PKC- δ neurons promotes WAT browning under leucine deprivation. *Nat Commun*. 2020;11(1):2847. doi:10.1038/s41467-020-16662-2
169. Ley RE, Turnbaugh PJ, Klein S, Gordon JI. Microbial ecology: human gut microbes associated with obesity. *Nature*. 2006;444(7122):1022-1023. doi:10.1038/4441022a
170. Fisher FM, Maratos-Flier E. Understanding the physiology of FGF21. *Annu Rev Physiol*. 2016;78(1):223-241. doi:10.1146/annurev-physiol-021115-105339
171. Solon-Biet SM, McMahon AC, Ballard JW, et al. The ratio of macronutrients, not caloric intake, dictates cardiometabolic health, aging, and longevity in ad libitum-fed mice. *Cell Metab*. 2014;19(3):418-430. doi:10.1016/j.cmet.2014.02.009
172. Wilkinson DJ, Piasecki M, Atherton PJ. The age-related loss of skeletal muscle mass and function: measurement and physiology of muscle fibre atrophy and muscle fibre loss in humans. *Ageing Res Rev*. 2018;47:123-132. doi:10.1016/j.arr.2018.07.005
173. Yudkoff M. Brain metabolism of branched-chain amino acids. *Glia*. 1997;21(1):92-98. doi:10.1002/(SICI)1098-1136(199709)21:13.0.CO;2-W
174. Polis B, Samson AO. Role of the metabolism of branched-chain amino acids in the development of Alzheimer's disease and other metabolic disorders. *Neural Regen Res*. 2020;15(8):1460-1470. doi:10.4103/1673-5374.274328

175. Calder PC. Branched-chain amino acids and immunity. *J Nutr.* 2006; 136(1):288S-293S. doi:[10.1093/jn/136.1.288S](https://doi.org/10.1093/jn/136.1.288S)
176. Kim EJ, Ha KH, Kim DJ, Choi YH. Diabetes and the risk of infection: a National Cohort Study. *Diabetes Metab J.* 2019;43(6):804-814. doi:[10.4093/dmj.2019.0071](https://doi.org/10.4093/dmj.2019.0071)
177. Yao K, Duan Y, Li F, et al. Leucine in obesity: therapeutic prospects. *Trends Pharmacol Sci.* 2016;37(8):714-727. doi:[10.1016/j.tips.2016.05.004](https://doi.org/10.1016/j.tips.2016.05.004)
178. Li H, Xu M, Lee J, He C, Xie Z. Leucine supplementation increases SIRT1 expression and prevents mitochondrial dysfunction and metabolic disorders in high-fat diet-induced obese mice. *Am J Physiol Endocrinol Metab.* 2012;303(10):E1234-E1244. doi:[10.1152/ajpendo.00198.2012](https://doi.org/10.1152/ajpendo.00198.2012)
179. Binder E, Bermudez-Silva FJ, Andre C, et al. Leucine supplementation protects from insulin resistance by regulating adiposity levels. *PLoS ONE.* 2013;8(9):e74705. doi:[10.1371/journal.pone.0074705](https://doi.org/10.1371/journal.pone.0074705)
180. Wolfson RL, Chantranupong L, Saxton RA, et al. Sestrin2 is a leucine sensor for the mTORC1 pathway. *Science.* 2016;351(6268):43-48. doi:[10.1126/science.aab2674](https://doi.org/10.1126/science.aab2674)
181. Sheen JH, Zoncu R, Kim D, Sabatini DM. Defective regulation of autophagy upon leucine deprivation reveals a targetable liability of human melanoma cells in vitro and in vivo. *Cancer Cell.* 2011; 19(5):613-628. doi:[10.1016/j.ccr.2011.03.012](https://doi.org/10.1016/j.ccr.2011.03.012)
182. Niu Y, Hu X, Song Y, et al. Blautia Coccoides is a newly identified bacterium increased by leucine deprivation and has a novel function in improving metabolic disorders. *Adv Sci (Weinh).* 2024;11(18): e2309255. doi:[10.1002/advs.202309255](https://doi.org/10.1002/advs.202309255)
183. Green CL, Pak HH, Richardson NE, et al. Sex and genetic background define the metabolic, physiologic, and molecular response to protein restriction. *Cell Metab.* 2022;34(2):209-26.e5. doi:[10.1016/j.cmet.2021.12.018](https://doi.org/10.1016/j.cmet.2021.12.018)
184. Solon-Biet SM, Cogger VC, Pulpitel T, et al. Branched chain amino acids impact health and lifespan indirectly via amino acid balance and appetite control. *Nat Metab.* 2019;1(5):532-545. doi:[10.1038/s42255-019-0059-2](https://doi.org/10.1038/s42255-019-0059-2)

How to cite this article: Mansoori S, Ho MY, Ng KK, Cheng KK. Branched-chain amino acid metabolism: Pathophysiological mechanism and therapeutic intervention in metabolic diseases. *Obesity Reviews.* 2025;26(2):e13856. doi:[10.1111/obr.13856](https://doi.org/10.1111/obr.13856)