Research Advance

Eating for hunger or pleasure: a Serotonin Model

Zili Yan¹, Yanlin He², Xing Cai³, Gang Shu^{4,*}, and Yong Xu^{1,5,*}

¹ Children's Nutrition Research Center, Department of Pediatrics, Baylor College of Medicine, Houston, TX 77030, USA

² Brain Glycemic and Metabolism Control Department, Pennington Biomedical Research Center, Louisiana State University, Baton Rouge, LA 70808, USA

³ State Key Laboratory of Genetic Resources and Evolution, Kunming Institute of Zoology, Chinese Academy of Sciences, Kunming 650201, China

- ⁴ Guangdong Laboratory of Lingnan Modern Agriculture and Guangdong Province Key Laboratory of Animal Nutritional Regulation, College of Animal Science, South China Agricultural University, Guangzhou 510642, China
- ⁵ Department of Molecular and Cellular Biology, Baylor College of Medicine, Houston, TX 77030, USA

* Correspondence to: Yong Xu, E-mail: yongx@bcm.edu; Gang Shu, E-mail: shugang@scau.edu.cn

Obesity, resulting from an imbalance between energy intake and expenditure, represents a major health crisis to our society, due to its alarmingly high prevalence and comorbidities, including diabetes, cardiovascular diseases, cancer, and COVID-19. Better understanding the neurobiological mechanisms for feeding behavior is essential for developing rational strategies to combat obesity and related comorbidities.

Agouti-related peptide (AgRP) neurons located in the arcuate nucleus of the hypothalamus (ARH) have received perhaps the most attention as a master regulator of feeding behavior. It is well known that AgRP neurons are regulated by multiple hormones that reflect the body's energy storage or nutritional state, e.g., leptin, insulin, ghrelin, and asprosin (Yang and Xu, 2020). AgRP neurons are activated in a caloriedeficient state (Takahashi and Cone, 2005; Yang et al., 2011; Liu et al., 2012), and activations of AgRP neurons can drive eating (Aponte et al., 2011; Krashes et al., 2011). Together, these findings support a physiological feedback pathway that regulates feeding: a calorie-deficient state (e.g. hunger) activates AgRP neurons, which in turn drive eating. However, this 'AgRP model' faces a challenge, as recent in vivo recordings revealed that AgRP neurons decrease



Figure 1 The 'Serotonin Model' illustrates physiological feedback signals to regulate both hunger-driven feeding and non-hunger-driven feeding. (**A**) A schematic illustration of changes in activities of AgRP neurons or 5-HT^{DRN} neurons during feeding. (**B**) A subgroup of 5-HT^{DRN} neurons project to the ARH, inhibiting AgRP neurons via the 5-HT_{1B}R and activating POMC neurons via the 5-HT_{2C}R, to suppress hunger-driven feeding; another subgroup of 5-HT^{DRN} neurons project to and inhibit DA^{VTA} neurons via the 5-HT_{2C}R to suppress non-hungerdriven feeding. The GABA_A receptor and the SK3 potassium channel mediate changes in activities of the ARH-projecting and VTA-projecting 5-HT^{DRN} neurons, respectively, during feeding. 5-HT_{1B}R, 5-HT 1B receptor; 5-HT_{2C}R, 5-HT 2C receptor; DA, dopamine; DRN, dorsal Raphe nucleus; GABA, γ -aminobutyric acid; POMC, proopiomelanocortin; SK3, small conductance calcium-activated potassium channel 3.

their activities dramatically within a few seconds after feeding starts, or even without the food actually being consumed (Betley et al., 2015; Chen et al., 2015; Mandelblat-Cerf et al., 2015). This rapid diminishment of AgRP neuron activity (Figure 1A) raises a question regarding how feeding behavior, which usually lasts for minutes to hours, is sustained. Based on our observations reported in a recent *Molecular Psychiatry* article (He et al., 2021), we propose an alternative 'Serotonin Model', which provides physiological feedback signals for feeding control. The brain serotonin, a neurotransmitter also known as 5-hydroxytryptamine (5-HT), is mainly synthesized by neurons in the midbrain dorsal Raphe nucleus (DRN). We demonstrated that the activation of these 5-HT^{DRN} neurons can inhibit eating (He et al., 2021). Importantly, using the *in vivo* recordings, we found that 5-HT^{DRN} neurons gradually increase their activities during the 2-h feeding period (Figure 1A). In sharp contrast to the rapid and sustained inhibition of AgRP neurons, the activation of 5-HT^{DRN} neurons occurs in a gradual and slow fashion (He et al., 2021). Importantly, the

This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (https://creativecom mons.org/licenses/by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

level of 5-HT^{DRN} neuron activity is correlated to the quantity of food intake (He et al., 2021). Thus, we suggest that 5-HT^{DRN} neurons function as a key component of a negative feedback loop. Low 5-HT^{DRN} neuron activity permits animals to eat; as animals continue eating, 5-HT^{DRN} neurons slowly elevate their activities to eventually terminate the meal.

Feeding can be driven by hunger (a state of nutritional deficit) to ensure survival. Feeding can also be triggered by the hedonic properties of certain foods in the absence of nutritional deficit. Dysregulations of hunger-driven feeding and hedonic feeding both contribute to the development of obesity (Kenny, 2011; Alonso-Alonso et al., 2015). It has been suggested that neurocircuits controlling these two types of feeding behaviors are not completely dissociable (Rossi and Stuber, 2018). Consistent with this notion, we found that 5-HTDRN neurons can regulate a hunger-driven feeding and a non-hunger-driven feeding in animals (He et al., 2021). However, our study further illustrated two largely segregated subgroups of 5-HT^{DRN} neurons: one subgroup send projections to the ARH and specifically inhibit feeding behavior driven by hunger, while the other subgroup of 5-HTDRN neurons project to the ventral tegmental area and reduce the intake of a high palatable diet in the nonhungry state (Figure 1B). Interestingly, these two subgroups of 5-HTDRN neurons both display slow activation during the course of hunger-driven feeding and nonhunger-driven feeding, respectively; however, they use distinct ion channels to achieve these changes (Figure 1B).

In summary, our findings support a 'Serotonin Model' that provides physiological feedback signals to regulate both hunger-driven feeding and non-hungerdriven feeding. We further identified distinct 5-HT^{DRN}-originated neurocircuits and disparate ion channels that can regulate these two types of feeding behaviors. These results provide a necessary framework for the development of a precision medication approach to treat obesity resulting from overeating for hunger or for pleasure.

[We acknowledge Dr Wei Wang (Zhongkai University of Agriculture and Engineering) for the illustration in Figure 1B. The investigators were supported by grants from the National Institutes of Health (NIH; P01DK113954, R01DK115761, R01DK117281, R01DK125480, and R01DK120858 to Y.X.; P20GM135002 to Y.H.), USDA/CRIS (51000-064-01S to Y.X.), and the American Diabetes Association (1-17-PDF-138 to Y.H.).]

References

- Alonso-Alonso, M., Woods, S.C., Pelchat, M., et al. (2015). Food reward system: current perspectives and future research needs. Nutr. Rev. 73, 296–307.
- Aponte, Y., Atasoy, D., and Sternson, S.M. (2011). AGRP neurons are sufficient to orchestrate feeding behavior rapidly and without training. Nat. Neurosci. 14, 351–355.

- Betley, J.N., Xu, S., Cao, Z.F., et al. (2015). Neurons for hunger and thirst transmit a negative-valence teaching signal. Nature *521*, 180–185.
- Chen, Y., Lin, Y.C., Kuo, T.W., et al. (2015). Sensory detection of food rapidly modulates arcuate feeding circuits. Cell *160*, 829–841.
- He, Y., Cai, X., Liu, H., et al. (2021). 5-HT recruits distinct neurocircuits to inhibit hunger-driven and non-hunger-driven feeding. Mol. Psychiatry, doi: 10.1038/s41380-021-01220-z.
- Kenny, P.J. (2011). Reward mechanisms in obesity: new insights and future directions. Neuron 69, 664–679.
- Krashes, M.J., Koda, S., Ye, C., et al. (2011). Rapid, reversible activation of AgRP neurons drives feeding behavior in mice. J. Clin. Invest. 121, 1424–1428.
- Liu, T., Kong, D., Shah, B.P., et al. (2012). Fasting activation of AgRP neurons requires NMDA receptors and involves spinogenesis and increased excitatory tone. Neuron *73*, 511–522.
- Mandelblat-Cerf, Y., Ramesh, R.N., Burgess, C.R., et al. (2015). Arcuate hypothalamic AgRP and putative POMC neurons show opposite changes in spiking across multiple timescales. eLife 4, e07122.
- Rossi, M.A., and Stuber, G.D. (2018). Overlapping brain circuits for homeostatic and hedonic feeding. Cell Metab. *27*, 42–56.
- Takahashi, K.A., and Cone, R.D. (2005). Fasting induces a large, leptin-dependent increase in the intrinsic action potential frequency of orexigenic arcuate nucleus neuropeptide Y/Agoutirelated protein neurons. Endocrinology *146*, 1043–1047.
- Yang, Y., Atasoy, D., Su, H.H., et al. (2011). Hunger states switch a flip–flop memory circuit via a synaptic AMPK-dependent positive feedback loop. Cell 146, 992–1003.
- Yang, Y., and Xu, Y. (2020). The central melanocortin system and human obesity. J. Mol. Cell Biol. 12, 785–797.