

Antitumour activity of oleanolic acid: A systematic review and meta‑analysis

YING ZENG^{*}, ZHONGLIAN WANG^{*}, JING ZHANG, WEI JIAN and QIAOFEN FU

Department of Radiation Oncology, First Affiliated Hospital of Kunming Medical University, Kunming, Yunnan 650032, P.R. China

Received June 3, 2024; Accepted September 6, 2024

DOI: 10.3892/ol.2024.14715

Abstract. Oleanolic acid (OA), a compound known for its potent antitumour properties, has been the subject of investigations in both cell and animal models. Although OA has good biological activity, its low water solubility and bioavail‑ ability limit its therapeutic use, and therefore translating the potential of OA into the clinical oncology setting remains challenging. The present systematic review and meta‑analysis utilized evidence from animal model studies to gain insights into the antitumour mechanisms of OA to address the gap in understanding, and to provide guidance for future research directions and potential clinical applications. The guidelines outlined in the Preferred Reporting Items for Systematic Reviews and Meta‑Analyses were applied in the present study and a comprehensive search was conducted across the PubMed/MEDLINE, Web of Science, Cochrane Library and Embase databases, with a cut‑off date of June 30, 2023. The primary focus was on randomized controlled trials that used animal models to assess the antitumour effects of OA. The methodological quality appraisal was conducted using the Systematic Review Centre for Laboratory Animal Experimentation risk of bias tool, and tumour volume and weight served as the principal outcome measures. Data were analysed using the RevMan (version 5.3) and Stata SE11 software packages, with an assessment of heterogeneity conducted using the I² statistical test, sensitivity analysis conducted using the leave‑one‑out approach, and evaluation of publication bias performed using Egger's test and funnel plot analysis. The present study demonstrated a significant inhibitory effect of OA intervention on tumour growth and a decrease in tumour weight in animal models. Despite the broad spectrum of

E‑mail: fuqiaofen@163.com

antitumour effects exhibited by OA, further investigations are warranted to optimize the dosage and administration routes of OA to maximize its efficacy in clinical cancer treatment.

Introduction

Cancer has been deemed a notable public health issue by the World Health Organization due to its prominent contributions to global morbidity and mortality (1,2). Moreover, the incidence and mortality rates of cancer are rapidly increasing worldwide. By 2040, the number of cancer cases worldwide is expected to reach 30.2 million, and the number of cancer-related deaths is expected to reach 16.3 million (3). Cancer is a heterogeneous disease and its development involves multiple biological processes and multiple factors, including environmental pollution or immune dysfunction (4,5).

At present, surgery, immunotherapy, and hormone, gene, radiation, laser and targeted therapies are the primary cancer treatment methods (6-8). Despite advances in cancer treatment, drug resistance remains a notable cause of relapse and poor survival in most tumor patients (9). Natural products can be used as important substrates to overcome drug resistance and improve the efficacy of cancer therapy (8). Since the late 1930s, natural products and their derivatives have been recognized as sources of antitumour drugs (10) due to proper‑ ties such as preventing and delaying tumour growth (11,12). Oleanolic acid (OA; 3β‑hydroxyolean‑12‑en‑28‑oic acid), as a representative natural product, has shown marked effects in this field (13).

OA is a pentacyclic triterpene compound that is widely found in the plant kingdom; it exists both as a free acid and as a triterpenoid saponin when joint with sugar chains (14). The richest source of OA is the leaves of the olive plant (Fig. 1A) (15). OA has a diverse range of biological and pharmacological activities, including liver protection (16,17), anti-inflammatory (18,19), antidiabetic (20,21), antiviral (22,23), bidirectional immunity (24) and antitumour effects (14,25‑27). The anticancer effects of OA have been demonstrated at the cellular level and in animal models in numerous types of cancer (Fig. 1B), including osteosarcoma, liver, lung and breast cancer (26,28‑44). The mechanisms underlying the antitumour activity of OA are multifaceted and include the inhibition of cellular proliferation, the promotion of apoptosis, the induction of autophagy, the modulation of cell cycle regulatory proteins, the inhibition of tumour cell

Correspondence to: Professor Qiaofen Fu, Department of Radiation Oncology, First Affiliated Hospital of Kunming Medical University, 295 Xichang Road, Kunming, Yunnan 650032, P.R. China

^{*} Contributed equally

Key words: oleanolic acid, biological products, pentacyclic triterpenes, neoplasms, animal tumour model, meta‑analysis

migration and invasion, and the suppression of angiogenesis (44‑46). Furthermore, OA increases the sensitivity of tumour cells to radiation (47). OA, akin to numerous other triterpenes, exhibits a broad spectrum of pharmacological activities, coupled with low toxicity and favourable tolerance profiles (48). However, high doses or prolonged administration of OA are reported to induce hepatotoxicity (49). According to the Biopharmaceutical Classification System (BCS), OA is categorized as a BCS Class IV drug characterized by exceedingly low aqueous solubility and suboptimal intestinal permeability, which collectively constrain its absorption and bioavailability (50). With further research, the chemical synthesis of OA derivatives and novel dosage forms may markedly improve the water solubility and bioavailability, thus strengthening their antitumour effects and ensuring their biosafety (15,16).

At present, numerous studies have reported the antitumour effects of OA in cell and animal experiments(13,25,27,34,51‑53). However, research on the antitumour effects of OA in clinical applications is currently limited. The present systematic review aimed to evaluate the potential antitumour effects of OA by collecting and analysing data from animal model experiments to provide guidance for further clinical research on the application of OA in cancer.

Materials and methods

Search strategy. The present systematic review and metaanalysis were performed in accordance with the Preferred Reporting Project for Systematic Review and Meta‑Analysis guidelines (54). The PubMed/MEDLINE, Web of Science, Cochrane Library and Embase databases were comprehensively searched up to June 30, 2023, using the following key words: 'Oleanolic acid', 'cancer', 'tumor' and 'tumour'. A total of 12 studies with 190 animals were included in the present study. A specific search strategy was devised for each database based on the Population, Intervention, Condition and Outcome framework and Medical Subject Heading terms (Table SI) (55,56). The present review protocol was not registered.

Study selection

Inclusion criteria. The inclusion criteria were as follows: i) The study type was a randomized controlled trial using animals; ii) experimental subjects consisted of tumour model mice, without any specific limitations on disease models or modelling methods; iii) the interventions involved the use of OA alone; iv) the outcome indicators included tumour weight and/or tumour volume; v) the studies were published in English; and vi) any type of solid tumour was included in the present study.

Exclusion criteria. The exclusion criteria were as follows: i) The target disease was a non-malignant tumour or cancer; ii) no control group was included; iii) OA was used in combination with other drugs; iv) duplicate publications; v) observational or non‑interventional studies, clinical studies, case reports, reviews, conference papers, systematic reviews, meta-analyses, editorial/letters or patent results; vi) unpublished dissertations; and vii) the full text could not be obtained or the data were incomplete.

Literature screening and data extraction. Based on the inclusion and exclusion criteria, two researchers independently screened the studies using EndNote (version X20; Clarivate plc). Data were from the included studies using Microsoft Excel (Microsoft Corporation) software and a subsequent cross-check was conducted to ensure accuracy. Any disagreements were resolved by consulting a third researcher. The following data were extracted: i) First author, year and country of journal publication; ii) animal species, weight, age, sample size and cancer type; iii) OA dosage, administration method and intervention time; and iv) tumour weight and tumour volume. In studies where the original publication presented the tumour sizes in only a graphical format, the Graph Grabber (version. 2.0.2; Quintessa Ltd.) program was used.

Risk of bias assessment. The Systematic Review Centre for Laboratory Animal Experimentation risk of bias tool (57) was used to evaluate the risk of bias of the included studies. The tool assessed 10 items across different domains, including selection bias, experimental bias, measurement bias, untracking bias and selective reporting bias (57). The analysis was independently performed by YZ and ZW. Any disagreements were resolved by consulting a third investigator. The tool offered three options: 'Yes' for a low bias risk, 'No' for a high bias risk and 'Unclear' when assigning bias was not possible.

Statistical methods. The data (mean ± standard deviation) from multiple intervention groups were merged in accordance with the Cochrane Handbook for Systematic Reviews of Interventions (58,59). The merged data were then analysed using RevMan (version 5.4; The Cochrane Collaboration) and Stata SE11 (StataCorp LP) software. Sensitivity and subgroup analyses were then performed to explore potential sources of heterogeneity and to assess the reliability of the results. A random‑effects model was employed for the meta-analysis. P<0.05 was considered to indicate a statistically significant difference (60). The I² statistical test was used to quantify the degree of heterogeneity among the studies. An I² value of $\geq 50\%$ was considered to indicate significant heterogeneity (58). Publication bias was assessed using Egger's test and funnel plots if ≥10 studies were included for an outcome.

Results

Literature retrieval results. The present search strategy produced a total of 3,654 articles (Fig. 2). Following the preliminary literature screen, 72 articles remained and the comprehensive review ultimately only included the results from 12 studies (27,46,47,51‑53,61‑66), with a total of 190 animals. The full texts of the 12 articles were read by two independent researchers who evaluated them according to the inclusion criteria.

Basic characteristics of the included studies. The present review encompassed 12 studies that addressed seven distinct types of cancer, with contributions from publications across six countries (Fig. 3A and B). Colorectal cancer emerged as the most prevalent form of malignancy within the present dataset, accounting for 34% of all cases. Notably, the journals

A

B Timeline of key studies of OA in cellular and animal models of cancer.

Figure 1. Chemical structure of OA and its antitumour development process. (A) Chemical structure of OA. (B) Timeline of key studies of OA in cellular and animal models of cancer. The figure provides a summary of the first author (Ref.), the molecular mechanisms through which oleanolic acid exerts its antitumor effects, the specific types of cancer investigated, and the cell and animal models utilized in the study. The literature describing only cell lines did not carry out animal studies. Cell lines: MG63, Saos-2 (osteosarcoma); U-87 MG, U-251MG (glioblastoma); HepG2, H22 (hepatoma); Panc-28 (pancreatic cancer); PC-3, DU145, LNCaP (prostate cancer); MCF-7 (breast cancer); T24 (bladder cancer); GBC-SD, NOZ (gallbladder cancer); HCT-15, HT-29, HCT-8, Colo 205 (rectal cancer); MKN‑45, SGC‑7901 (gastric cancer); SW579 (thyroid carcinoma); SKOV3, OVCA420 (ovarian cancer). OA, oleanolic acid; mTOR, mammalian target of rapamycin; MAPK, mitogen-activated protein kinases; ERK, extracellular signal regulated kinases; ROS, reactive oxygen species; NOX2, NADPH oxidase 2; HIF‑1α, hypoxia inducible factor‑1α; YAP, yes‑associated protein; FOXA1, forkhead box A1; PD‑L1, programmed cell death‑ligand 1; EMT, epithelial to mesenchymal transition.

from the United States and the Netherlands each contributed to 25% of the included studies.

The main characteristics of the included studies were summarised (Table I), including the following: i) First author and year; ii) species, strain, sex and age of the animals, of which BALB/c mice (n=10) were the most common, with 7 studies of female mice and 5 studies of male mice; iii) tumour types, which included colorectal (n=4), cervical (n=2), gastric (n=2), breast $(n=1)$ and lung $(n=1)$ cancer, hepatocellular carcinoma $(n=1)$ and melanoma (n=1); iv) dosage of OA (range, $2-150$ mg/kg); v) route of administration, including oral (n=2), intraperitoneal (n=6), intragastric (n=3) or subcutaneous (n=1) injections; vi) duration of drug intervention (range, 10‑30 days); vii) type of control, with normal saline used in 50% of the included studies; and vii) country of journal publication which included the United States (n=3), Netherlands (n=3), England (n=2), Greece (n=2), United Arab Emirates (n=1) and Brazil (n=1).

This study evaluated the antitumor effects of oleanolic acid by tumor weight and tumor volume. But in addition to the information presented in Table I, further *in vitro* and *in vivo* tests

using OA had been performed in the included studies of the present systematic review. A total of 5 studies had 2 intervention groups, in which two doses of OA were tested. Furthermore, 7 studies had only 1 intervention group. The most common *in vitro* tests were cytotoxicity tests $(n=9)$ and flow cytometry $(n=3)$. The most frequent *in vivo* assessments were tumour volume (n=12), tumour weight (n=8) and animal weight (n=8) measurements.

Results of the risk of bias assessment. All studies included in the present review were evaluated using the RoB guideline's assessment of the risk of bias (Table SII). Numerous parameters were described as 'unclear', which indicated that the information reported in the reviewed articles was incomplete or unclear. This lack of clarity was predominantly related to sequence generation, baseline features and covert grouping. Of note, in none of the included studies, the evaluators were blinded regarding the results; however, this deficiency did not affect the accuracy of the results due to the objectivity of the measured results. As a result, these studies were identified as low-risk in the bias risk assessment.

Figure 2. Flow diagram of the present systematic literature search strategy.

Meta‑analysis. Tumour volume and tumour weight were extracted as outcome measures for subsequent analysis to evaluate the antitumour effects of OA in animal experiments. No additional studies were included in this assessment.

Tumour volume. Forest plot analysis demonstrated that 10 studies reported the tumour volumes from 159 animals (Fig. 3C). Statistically significant differences were observed between the OA groups and control groups (mean difference, ‑0.64; 95% CI, ‑0.89 to ‑0.39; P<0.00001). These results

suggest that treatment with OA significantly inhibited tumour growth compared with the control treatment.

Tumour weight. In total, 9 articles assessed differences in tumour weight between the control and OA groups (Fig. 3D). The differences in tumour weight between the OA and control groups were statistically significant (mean difference, ‑0.43; 95% CI, ‑0.58 to ‑0.28; P<0.00001). By the end of the experiments evaluated in the present review, OA‑treated animals had significantly lower tumour weights

 -8

 $\mathbf 0$

 0.5

 $\mathbf{1}$

Precision

 1.5

Figure 3. Analysis of the 12 included studies. Percentage distribution of the studies evaluating the antitumour effects of OA that met the inclusion criteria for this systematic review by (A) type of tumour model and (B) country of journal publication. Forest plots of the effects of OA on (C) tumour volume (cm³) and (D) tumour weight (g). (E) Egger's test of publication bias in the present meta-analysis of tumour volume inhibition by OA treatment. (F) Funnel plot of included studies of tumour volume inhibition by OA treatment. SE, standard error; SMD, standardized mean difference; OA, oleanolic acid; SD, standard deviation; CI, confidence interval; IV, inverse variance; df, degrees of freedom.

 \overline{c}

2.5

 $-i$ ^o

 -5

SMD

 \dot{o}

 $\overline{5}$

6 ZENG *et al*: ANTITUMOUR ACTIVITY OF OLEANOLIC ACID

A, Tumor volume

B, Tumor weight

MD, mean difference; CI, confidence interval.

compared with control animals, regardless of the tumour type.

Analysis of sources of heterogeneity

Sensitivity analysis. Of the included studies, 10 analysed the tumour volume. After excluding one study at a time, the heterogeneity remained unchanged. The I^2 of each pooled analysis remained between 98‑99%, which indicated no significant change in heterogeneity. In the 9 studies that analysed tumour weight, similar results were observed after each study was excluded individually. The I^2 of each pooled analysis remained between 98‑99%, which suggested no significant change in heterogeneity (Table II; Fig. S1).

Subgroup analysis. According to the Cochrane Handbook for Systematic Reviews of Interventions (58), as certain interventions shared a control group, only the total number of participants were divided into subgroup analyses. The original mean and standard deviation were unchanged.

Subgroup analyses were conducted based on the following factors: i) Dose of OA, categorized as low (<50 mg/kg), medium (50 mg/kg) or high (>50 mg/kg); ii) route of administration, including oral gavage, intraperitoneal, intragastric and subcutaneous injections; iii) type of cancer, including hepatocellular carcinoma, melanoma, and colorectal, cervical, gastric, breast and lung cancer; and iv) strains of mice utilized, including Kunming, severe combined immunodeficiency and BALB/c nude or athymic mice.

The subgroup analysis of the effects on the tumour volume demonstrated statistically significant differences among the different treatment regimens, tumour types and mouse strains (Table III). It should be noted that oleanolic acid had no significant effect on tumor volume inhibition in melanoma and gastric cancer (P>0.05). The subgroup analysis of tumour weight indicated that the dose, administration mode, tumour type and mouse strain had significant effects on the outcome (Table IV).

Risk of bias analysis. Due to the limited number of included studies that assessed tumour weight (<10 publications), the evaluation of publication bias was limited to studies that evaluated tumour volume as an outcome. Egger's test was performed to quantify bias in this parameter and a funnel plot

Low, dosages <50 mg/kg; Medium, dosages of 50 mg/kg; High, dosages >50 mg/kg; SMD, standardized mean difference; CI, confidence interval.

was generated to visualize the results (y-intercept, -4.97; 95%) CI, $7.22-2.73$; P=0.001; Fig. 3E and F). Additionally, the funnel plot for tumour volume exhibited asymmetry and skewness, with 3 out of 10 selected studies falling outside the 95% CI. These findings suggested the presence of publication bias in studies that reported the tumour volume.

Discussion

OA is an important pentacyclic triterpene that is widely found in plants, foods and medicines; it was initially used to treat chronic hepatitis and liver injury (18,67). Žiberna *et al* (68) reported that OA has potential in clinical adjuvant anticancer treatment, supported by its direct anticancer activity, synergistic effect with chemotherapy drugs, inhibition of transporters, enhancement of radiotherapy efficacy, low toxicity and lack of adverse reactions. These attributes suggest the promising potential for OA in clinical cancer therapy.

However, the therapeutic efficacy of oleanolic acid is constrained by its limited water solubility, low bioavailability (69). Structural modification and dosage form optimization could notably increase the water solubility and bioavailability of OA (70). At present, OA derivatives are primarily modified on the C‑3 hydroxyl group, C‑12/C‑13 double bond and C‑28 carboxyl group (71). Novel OA dosage forms consist of nanoparticles, liposomes, solid dispersions and phospholipid complexes (72‑75).

Research has indicated that OA demonstrates antitumour activity across a number of *in vitro* and *in vivo* models, exerting its effects on multiple targets. These targets include, but are not limited to, aldoketo reductase family member 1B10, protein tyrosine phosphatase 1B and cell division cycle 25 phosphatase (76‑78). To the best of our knowledge, no systematic review has evaluated the antitumour effects of OA in animal models. Therefore, the present meta-analysis holds particular significance to address this limitation in the field and lays the groundwork for future clinical explorations of OA.

The present study systematically evaluated the antitumour effects of OA by analysing the tumour volume and weight in experimental mice. Studies have demonstrated that oleanolic acid can significantly reduce tumor weight and inhibit the volumetric growth of most tumors. However, its anti‑tumor efficacy appears suboptimal for certain malignancies, such as gastric cancer and melanoma (P<0.05). This limitation may be attributed to the small sample size, which compromises the reliability of the findings. Future research should aim to increase the sample size and employ robust statistical methodologies to enhance the reliability and clinical applicability of the results. At the same time, further research on the anti-tumor mechanism of oleanolic acid and development of water‑soluble and bioavailable oleanolic acid derivatives will help improve its clinical significance in tumor therapy. OA exerts its antitumour effects through the modulation of various signalling pathways. OA can induce HCT116 colon cancer cell death through the

Table IV. Subgroup analysis of tumor weight.

Low, dosages <50 mg/kg; Medium, dosages of 50 mg/kg; High, dosages >50 mg/kg; SMD, standardized mean difference; CI, confidence interval.

p38/FOXO3a/Sirt6 pathway (27), and induce apoptosis and autophagy in AGS human gastric cancer cells through the PI3K/AKT/mTOR pathway (63). Additionally, OA inhibits the migration and invasion of glioma cells by inactivating the MAPK/ERK signalling pathway (29); it also induces apoptosis in A375SM and A375P melanoma cells through the NF‑κB pathway (65). Furthermore, OA suppresses the proliferation of human bladder cancer cells by targeting the Akt/mTOR/S6K and ERK1/2 signalling pathways (26).

The present meta-analysis revealed substantial heterogeneity, with I² values of 99% for both the tumour volume and weight. A subgroup analysis of the included studies was performed to identify sources of heterogeneity in the data. This indicated that the mode of administration, tumour type and mouse strain were potential sources of heterogeneity. Despite a comprehensive search, the limited number of studies included in the present analysis may have introduced bias. Given the lack of standardization of animal experiments, meta‑analyses of preclinical studies are more heterogeneous compared with clinical studies (57,79). Maganti *et al* (80) conducted a systematic review of preclinical animal studies on the efficacy of CRISPR/Cas9 gene‑edited chimeric antigen receptor T cells against malignant tumours, demonstrating significant heterogeneity ($I^2=96\%$), which may be attributed to the range of cancer types studied. Similarly, a meta‑analysis

by Singh *et al* (81) on the anticancer effects of apigenin on animal models of cancer also demonstrated high heterogeneity. Subgroup analyses demonstrated that the dose of apigenin, tumour model, route of administration and duration of treatment were significant factors that influenced heterogeneity. Furthermore, a sensitivity analysis was performed on the included studies using the leave‑one‑out method in the present review. After excluding one study at a time, the heterogeneity did not change significantly and the I^2 -value of tumour growth inhibition and tumour weight remained between 98‑99%, which indicated the stability of the results. The high heterogeneity in the present study may be attributed to variations in the animal models and experimental conditions used. Since the tumour weight index was included in <10 studies, publication bias was assessed only for the tumour volume, and Egger's test and funnel plot results both indicated publication bias.

The antitumour effect of OA is suggested to be closely related to the immune system. Programmed cell death protein 1/programmed death‑ligand 1 (PD‑L1) blockade therapy is a promising cancer treatment strategy that is considered to have revolutionized the treatment landscape for malignant tumours (82). Lu *et al* (41) reported that OA can restore the effect of T cells on killing gastric cancer cells, achieve DNA hypomethylation and downregulate PD‑L1 by inhibiting the IL-1 β /NF- κ B/TET3 signalling pathway, thus serving

an antitumour role. In addition, Luo *et al* (83) reported that OA nanomicelles showed significant anticancer potential in a tumour‑bearing mouse model and stimulated immune cell infiltration. As the main component of the tumour immune microenvironment, immune‑infiltrating cells can effectively control tumour suppression and immune escape (84). However, through an analysis of the available literature, it was found that the quantification of the antitumour effects of OA in animal experiments has focused mainly on the tumour volume and weight, and has almost never involved an evaluation of immunity. Further studies are needed to clarify the relationship between the antitumour effects of OA and immune regulation.

The present study had several limitations, including the small number of included references and potential publication bias. But it still demonstrates significant strengths and research value. Extensive searches of major medical databases were conducted, and standard search terms were used to retrieve the relevant literature as comprehensively as possible and to reduce bias in selective reporting. An assessment of the risk of bias in the included studies increased the reliability of the results. Through subgroup analyses, the antitumour effects of OA under different conditions were further investigated, which provided guidance for further research and clinical application. In addition, this study also explored the possibility of OA enhancing the anti-tumor effect by modulating the immune system, opening up a new perspective for understanding its mechanism of action. In summary, this study provides solid theoretical support for the anti-tumor potential of oleanolic acid and promotes its transformation from laboratory research to clinical application.

The present review highlighted the antitumour potential of OA in animal models. Future studies are necessary to expand the sample size, unify the experimental design and further explore how OA serves an antitumour role by regulating the immune system. Future research could include determining the optimal dose and route of administration of OA, and developing derivatives to improve their solubility and bioavailability, while evaluating their long-term safety and toxicity. In addition, combination therapy, the standardization of preclinical models, molecular mechanism analyses, personalized medicine strategies, drug delivery system innovation and clinical trial design are key directions for promoting the clinical application of OA. Ultimately, the discovery and validation of biomarkers that predict OA responses will improve the development of precision medicine and provide novel strategies for cancer treatment.

Acknowledgements

Not applicable.

Funding

This present study was supported by the National Natural Science Foundation of China (grant no. 82160529), the Applied Basic Science Research Foundation of Yunnan Province (grant no. 202201AT070294), the Yunnan Revitalization Talent Support Program (grant no. RLQB20220014) and the 535 Talent Project of the First Affiliated Hospital of Kunming Medical University (grant no. 2023535D17).

Availability of data and materials

The data generated in the present study may be requested from the corresponding author.

Authors' contributions

YZ and ZW were responsible for screening the data and writing the draft paper. JZ and WJ completed the data extraction and analysis of the results. QF was responsible for a thorough review of the manuscript prior to submission. QF and YZ jointly planned the overall structural design of the paper. YZ and ZW confirm the authenticity of all the raw data. All authors read and approved the final version of the manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

- 1. Russo A, Cardile V, Graziano ACE, Avola R, Montenegro I, Cuellar M, Villena J and Madrid A: Antigrowth activity and induction of apoptosis in human melanoma cells by Drymis winteri forst extract and its active components. Chem Biol Interact 305: 79‑85, 2019.
- 2. Yuan R, Hou Y, Sun W, Yu J, Liu X, Niu Y, Lu JJ and Chen X: Natural products to prevent drug resistance in cancer chemo-
therapy: A review. Ann N Y Acad Sci 1401: 19-27, 2017.
- 3. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A and Bray F: Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 71: 209‑249, 2021.
- 4. Hung KC, Huang TC, Cheng CH, Cheng YW, Lin DY, Fan JJ and Lee KH: The expression profile and prognostic significance of metallothionein genes in colorectal cancer. Int J Mol Sci 20: 3849, 2019.
- 5. Gao L, Yang T, Xue Z and Chan CKD: Hot spots and trends in the relationship between cancer and obesity: A systematic review and knowledge graph analysis. Life (Basel) 13: 337, 2023.
- 6. Baskar R, Lee KA, Yeo R and Yeoh KW: Cancer and radiation therapy: Current advances and future directions. Int J Med Sci 9: 193‑199, 2012.
- 7. Wang X, Zhang H and Chen X: Drug resistance and combating drug resistance in cancer. Cancer Drug Resist 2: 141‑160, 2019.
- 8. Ramos A, Sadeghi S and Tabatabaeian H: Battling chemoresis‑ tance in cancer: Root causes and strategies to uproot them. Int J Mol Sci 22: 9451, 2021.
- 9. Vasseur S and Guillaumond F: Lipids in cancer: A global view of the contribution of lipid pathways to metastatic formation and treatment resistance. Oncogenesis 11: 46, 2022.
- 10. Newman DJ and Cragg GM: Natural products as sources of new drugs from 1981 to 2014. J Nat Prod 79: 629‑661, 2016.
- 11. Bishayee A and Sethi G: Bioactive natural products in cancer prevention and therapy: Progress and promise. Semin Cancer Biol 40‑41: 1‑3, 2016.
- 12. Akter R, Afrose A, Rahman MR, Chowdhury R, Nirzhor SSR, Khan RI and Kabir MT: A comprehensive analysis into the therapeutic application of natural products as SIRT6 modulators in alzheimer's disease, aging, cancer, inflammation, and diabetes. Int J Mol Sci 22: 4180, 2021.

- 13. Wei J, Liu H, Liu M, Wu N, Zhao J, Xiao L, Han L, Chu E and rouracil in pancreatic cancer cells. Oncol Rep 28: 1339-1345, 2012.
- 14. Sultana N and Ata A: Oleanolic acid and related derivatives as medicinally important compounds. J Enzyme Inhib Med Chem 23: 739‑756, 2008.
- 15. Baer‑Dubowska W, Narożna M and Krajka‑Kuźniak V: Anti-cancer potential of synthetic oleanolic acid derivatives and their conjugates with NSAIDs. Molecules 26: 4957, 2021.
- 16. Liu J: Oleanolic acid and ursolic acid: Research perspectives. Ethnopharmacol 100: 92-94, 2005
- Prasad S, Kalra N and Shukla Y: Hepatoprotective effects of lupeol and mango pulp extract of carcinogen induced alteration in Swiss albino mice. Mol Nutr Food Res 51: 352‑359, 2007.
- 18. Singh GB, Singh S, Bani S, Gupta BD and Banerjee SK: Anti-inflammatory activity of oleanolic acid in rats and mice. J Pharm Pharmacol 44: 456‑458, 1992.
- 19. Pádua TA, de Abreu BSSC, Costa TEMM, Nakamura MJ, Valente LMM, Henriques MDG, Siani AC and Rosas EC: Anti-inflammatory effects of methyl ursolate obtained from a chemically derived crude extract of apple peels: Potential use in rheumatoid arthritis. Arch Pharm Res 37: 1487‑1495, 2014.
- 20. Alqahtani A, Hamid K, Kam A, Wong KH, Abdelhak Z, Razmovski‑Naumovski V, Chan K, Li KM, Groundwater PW and Li GQ: The pentacyclic triterpenoids in herbal medicines and their pharmacological activities in diabetes and diabetic complications. Curr Med Chem 20: 908‑931, 2013.
- 21. Teodoro T, Zhang L, Alexander T, Yue J, Vranic M and Volchuk A: Oleanolic acid enhances insulin secretion in pancre‑ atic beta‑cells. FEBS Lett 582: 1375‑1380, 2008.
- 22. Yu F, Wang Q, Zhang Z, Peng Y, Qiu Y, Shi Y, Zheng Y, Xiao S, Wang H, Huang X, et al: Development of oleanane-type triterpenes as a new class of HCV entry inhibitors. J Med Chem 56: 4300‑4319, 2013.
- 23. Cichewicz RH and Kouzi SA: Chemistry, biological activity, and chemotherapeutic potential of betulinic acid for the prevention and treatment of cancer and HIV infection. Med Res Rev 24: 90‑114, 2004.
- 24. Martín R, Carvalho‑Tavares J, Hernández M, Arnés M, Ruiz‑Gutiérrez V and Nieto ML: Beneficial actions of oleanolic acid in an experimental model of multiple sclerosis: A potential therapeutic role. Biochem Pharmacol 79: 198-208, 2010.
25. Zhao X, Liu M and Li D: Oleanolic acid suppresses the prolifera-
- tion of lung carcinoma cells by miR-122/Cyclin G1/MEF2D axis. Mol Cell Biochem 400: 1‑7, 2015.
- 26. Mu DW, Guo HQ, Zhou GB, Li JY and Su B: Oleanolic acid suppresses the proliferation of human bladder cancer by Akt/mTOR/S6K and ERK1/2 signaling. Int J Clin Exp Pathol 8: 13864‑13870, 2015.
- 27. Potočnjak I, Šimić L, Vukelić I, Batičić L and Domitrović R: Oleanolic acid induces HCT116 colon cancer cell death through the p38/FOXO3a/Sirt6 pathway. Chem Biol Interact 363: 110010, 2022.
- 28. Zhou R, Zhang Z, Zhao L, Jia C, Xu S, Mai Q, Lu M, Huang M, Wang L, Wang X, *et al*: Inhibition of mTOR signaling by oleanolic acid contributes to its anti-tumor activity in osteosarcoma cells. J Orthop Res 29: 846‑852, 2011.
- 29. Guo G, Yao W, Zhang Q and Bo Y: Oleanolic acid suppresses migration and invasion of malignant glioma cells by inactivating MAPK/ERK signaling pathway. PLoS One 8: e72079, 2013.
- 30. Wang X, Bai H, Zhang X, Liu J, Cao P, Liao N, Zhang W, Wang Z and Hai C: Inhibitory effect of oleanolic acid on hepatocellular carcinoma via ERK-p53-mediated cell cycle arrest and mitochondrial-dependent apoptosis. Carcinogenesis 34: 1323‑1330, 2013.
- 31. Wei J, Liu M, Liu H, Wang H, Wang F, Zhang Y, Han L and Lin X: Oleanolic acid arrests cell cycle and induces apoptosis via ROS‑mediated mitochondrial depolarization and lysosomal membrane permeabilization in human pancreatic cancer cells. J Appl Toxicol 33: 756‑765, 2013.
- 32. Liu J, Zheng L, Wu N, Ma L, Zhong J, Liu G, Ma L, Zhong J, Liu G and Lin X: Oleanolic acid induces metabolic adaptation in cancer cells by activating the AMP-activated protein kinase pathway. J Agric Food Chem 62: 5528‑5537, 2014.
- 33. Liu J, Wu N, Ma LN, Zhong JT, Liu G, Zheng LH and Lin XK: p38 MAPK signaling mediates mitochondrial apoptosis in cancer cells induced by oleanolic acid. Asian Pac J Cancer Prev 15: 4519‑4525, 2014.
- 34. Li HF, Wang XA, Xiang SS, Hu YP, Jiang L, Shu YJ, Li ML, Wu XS, Zhang F, Ye YY, *et al*: Oleanolic acid induces mitochon‑ drial‑dependent apoptosis and G0/G1 phase arrest in gallbladder cancer cells. Drug Des Devel Ther 9: 3017‑3030, 2015.
- 35. Li X, Song Y, Zhang P, Zhu H, Chen L, Xiao Y and Xing Y: Oleanolic acid inhibits cell survival and proliferation of prostate cancer cells in vitro and in vivo through the PI3K/Akt pathway. Tumour Biol 37: 7599‑7613, 2016.
- 36. Xu Y, Shu B, Tian Y, Wang G, Wang Y, Wang J and Dong Y: Oleanolic acid induces osteosarcoma cell apoptosis by inhibition of Notch signaling. Mol Carcinog 57: 896-902, 2018.
- 37. GuoY, HanB, LuoK, RenZ, CaiL and SunL: NOX2‑ROS‑HIF‑1α signaling is critical for the inhibitory effect of oleanolic acid on rectal cancer cell proliferation. Biomed Pharmacother 85: 733‑739, 2017.
- 38. Li Y, Xu Q, Yang W, Wu T and Lu X: Oleanolic acid reduces aerobic glycolysis‑associated proliferation by inhibiting yes-associated protein in gastric cancer cells. Gene 712: 143956, 2019.
- 39. Duan L, Yang Z, Jiang X, Zhang J and Guo X: Oleanolic acid inhibits cell proliferation migration and invasion and induces SW579 thyroid cancer cell line apoptosis by targeting fork‑ head transcription factor A. Anticancer Drugs 30: 812‑820, 2019.
- 40. Kim GJ, Jo HJ, Lee KJ, Choi JW and An JH: Oleanolic acid induces p53‑dependent apoptosis via the ERK/JNK/AKT pathway in cancer cell lines in prostatic cancer xenografts in mice. Oncotarget 9: 26370‑26386, 2018.
- 41. Lu X, Li Y, Yang W, Tao M, Dai Y, Xu J and Xu Q: Inhibition of NF‑κB is required for oleanolic acid to downregulate PD‑L1 by promoting DNA demethylation in gastric cancer cells. J Biochem Mol Toxicol 35: e22621, 2021.
- 42. Zeng Z, Yu J, Jiang Z and Zhao N: Oleanolic acid (OA) targeting UNC5B inhibits proliferation and EMT of ovarian cancer cell and increases chemotherapy sensitivity of niraparib. J Oncol 2022: 5887671, 2022.
- 43. Ren J, Yan J, Raza F, Zafar H, Wan H, Chen X, Cui Q, Li H and Wang X: A synergistic combination of oleanolic acid and apatinib to enhance antitumor effect on liver cancer cells and protect against hepatic injury. Recent Pat Anticancer Drug Discov 19: 199‑208, 2024.
- 44. Hosny S, Sahyon H, Youssef M and Negm A: Oleanolic acid tion of mitochondrial-mediated apoptosis and autophagy. Nutr Cancer 73: 968‑982, 2021.
- 45. Tang ZY, Li Y, Tang YT, Ma XD and Tang ZY: Anticancer activity of oleanolic acid and its derivatives: Recent advances in evidence, target profiling and mechanisms of action. Biomed Pharmacother 145: 112397, 2022.
- 46. Liang Z, Pan R, Meng X, Su J, Guo Y, Wei G, Zhang Z and He K: Transcriptome study of oleanolic acid in the inhibition of breast tumor growth based on high-throughput sequencing. Aging (Albany NY) 13: 22883‑22897, 2021.
- 47. Wang H, Zhong W, Zhao J, Zhang H, Zhang Q, Liang Y, Chen S, Liu H, Zong S, Tian Y, *et al*: Oleanolic acid inhibits epithelial‑mesenchymal transition of hepatocellular carcinoma by promoting iNOS dimerization. Mol Cancer Ther 18: 62‑74, 2019.
- 48. Bednarczyk‑Cwynar B, Wachowiak N, Szulc M, Kamińska E, Bogacz A, Bartkowiak‑Wieczorek J, Zaprutko L and Mikolajczak PL: Strong and long‑lasting antinociceptive and anti-inflammatory conjugate of naturally occurring oleanolic acid and aspirin. Front Pharmacol 7: 202, 2016.
- 49. Feng H, Wu YQ, Xu YS, Wang KX, Qin XM and Lu YF: LC-MS-based metabolomic study of oleanolic acid-induced hepatotoxicity in mice. Front Pharmacol 11: 747, 2020.
50. Liu Y, Luo X, Xu X, Gao N and Liu X: Preparation, charac-
- terization and in vivo pharmacokinetic study of PVP-modified oleanolic acid liposomes. Int J Pharm 517: 1‑7, 2017.
- 51. Li L, Wei L, Shen A, Chu J, Lin J and Peng J: Oleanolic acid modulates multiple intracellular targets to inhibit colorectal cancer growth. Int J Oncol 47: 2247-2254, 2015.
- 52. Li L, Lin J, Sun G, Wei L, Shen A, Zhang M and Peng J: Oleanolic acid inhibits colorectal cancer angiogenesis *in vivo* and *in vitro* via suppression of STAT3 and Hedgehog pathways. Mol Med Rep 13: 5276‑5282, 2016.
- 53. Jiang X, Shi M, Sui M, Yuan Y, Zhang S, Xia Q and Zhao K: Oleanolic acid inhibits cervical cancer Hela cell proliferation through modulation of the ACSL4 ferroptosis signaling pathway. Biochem Biophys Res Commun 545: 81‑88, 2021.
- 54. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, Shamseer L, Tetzlaff JM, Akl EA, Brennan SE, et al: The PRISMA 2020 statement: An updated guideline for reporting systematic reviews. BMJ 372: n71, 2021.
- 55. Page MJ, Moher D, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, Shamseer L, Tetzlaff JM, Akl EA, Brennan SE, et al: PRISMA 2020 explanation and elaboration: Updated guidance and exemplars for reporting systematic reviews. BMJ 372: n160, 2021.
- 56. Richardson WS, Wilson MC, Nishikawa J and Hayward RS: The well-built clinical question: A key to evidence-based decisions. ACP J Club 123: A12‑A13, 1995.
- 57. Hooijmans CR, Rovers MM, de Vries RBM, Leenaars M, Ritskes‑Hoitinga M and Langendam MW: SYRCLE's risk of bias tool for animal studies. BMC Med Res Methodol 14: 43, 2014.
- 58. Higgins JPT, Thomas J, Chandler J, Cumpston M, Li T, Page MJ and Welch VA (eds): Cochrane handbook for systematic reviews of interventions version 6.2 [updated February 2021]. Cochrane, 2021. Available from: https://training.cochrane.org/handbook.
- 59. Cao LM, Sun ZX, Makale EC, Du GK, Long WF and Huang HR: Antitumor activity of fucoidan: A systematic review and meta‑analysis. Transl Cancer Res 10: 5390‑5405, 2021.
- 60. Vesterinen HM, Sena ES, Egan KJ, Hirst TC, Churolov L, Currie GL, Antonic A, Howells DW and Macleod MR: Meta‑analysis of data from animal studies: a practical guide. J Neurosci Methods 221: 92‑102, 2014.
- 61. Tang S, Gao D, Zhao T, Zhou J and Zhao X: An evaluation of the anti‑tumor efficacy of oleanolic acid‑loaded PEGylated liposomes. Nanotechnology 24: 235102, 2013.
- 62. Niu G, Sun L, Pei Y and Wang D: Oleanolic acid inhibits colorectal cancer angiogenesis by blocking the VEGFR2 signaling pathway. Anticancer Agents Med Chem 18: 583‑590, 2018.
- 63. Lee JH, Yoo ES, Han SH, Jung GH, Han EJ, Jung SH, Kim BS, Cho SD, Nam JS, Choi C, *et al*: Oleanolic acid induces apoptosis and autophagy via the PI3K/AKT/mTOR pathway in AGS human gastric cancer cells. J Funct Foods 87: 104854, 2021.
- 64. Gao YS, Yuan Y, Song G and Lin SQ: Inhibitory effect of ursolic acid and oleanolic acid from Eriobotrya fragrans on A549 cell viability in vivo. Genet Mol Res 15, 2016.
- 65. Woo JS, Yoo ES, Kim SH, Lee JH, Han SH, Jung SH, Jung GH and Jung JY: Anticancer effects of oleanolic acid on human melanoma cells. Chem Biol Interact 347: 109619, 2021.
- 66. Nie H, Wang Y, Qin Y and Gong XG: Oleanolic acid induces autophagic death in human gastric cancer cells in vitro and in vivo. Cell Biol Int 40: 770‑778, 2016.
- 67. Liu J, Liu Y, Madhu C and Klaassen CD: Protective effects of oleanolic acid on acetaminophen‑induced hepatotoxicity in mice. J Pharmacol Exp Ther 266: 1607‑1613, 1993.
- 68. Žiberna L, Šamec D, Mocan A, Nabavi SF, Bishayee A, Farooqi AA, Sureda A and Nabavi SM: Oleanolic acid alters multiple cell signaling pathways: implication in cancer prevention and therapy. Int J Mol Sci 18: 643, 2017.
- 69. Zhong YY, Chen HS, Wu PP, Zhang BJ, Yang Y, Zhu QY, Zhang CG and Zhao SQ: Synthesis and biological evaluation of novel oleanolic acid analogues as potential $α$ -glucosidase inhibitors. Eur J Med Chem 164: 706‑716, 2019.
- 70. Yang R, Huang X, Dou J, Zhai G and Su L: Self‑microemulsifying drug delivery system for improved oral bioavailability of oleanolic acid: Design and evaluation. Int J Nanomedicine 8: 2917‑2926, 2013.
- 71. Tang C, Chen Y, Bai S and Yang G: Advances in the study of structural modification and biological activities of oleanolic acid. Chin J Org Chem 33: 46‑65, 2013.
- 72. Jiang Q, Yang X, Du P, Zhang H and Zhang T: Dual strategies to improve oral bioavailability of oleanolic acid: Enhancing water-solubility, permeability and inhibiting cytochrome P450 isozymes. Eur J Pharm Biopharm 99: 65‑72, 2016.
- 73. Gao N, Guo M, Fu Q and He Z: Application of hot melt extrusion to enhance the dissolution and oral bioavailability of oleanolic acid. Asian J Pharm Sci 12: 66‑72, 2017.
- 74. Xia X, Liu H, Lv H, Zhang J, Zhou J and Zhao Z: Preparation, characterization, and in vitro/vivo studies of oleanolic acid‑loaded lactoferrin nanoparticles. Drug Des Devel Ther 11: 1417‑1427, 2017.
- 75. Wei CT, Wang YW, Wu YC, Lin LW, Chen CC, Chen CY and Kuo SM: Reparative efficacy of liposome-encapsulated oleanolic acid against liver inflammation induced by fine ambient particulate matter and alcohol in mice. Pharmaceutics 14: 1108, 2022.
- 76. Lu Y, Zheng W, Lin S, Guo F, Zhu Y, Wei Y, Liu X, Jin S, Jin L and Li Y: Identification of an oleanane‑type triterpene hedragonic acid as a novel farnesoid X receptor ligand with liver protective effects and anti-inflammatory activity. Mol Pharmacol 93: 63-72, 2018.
- 77. Li Y, Yu Y, Jin K, Gao L, Luo T, Sheng L, Shao X and Li J: Synthesis and biological evaluation of novel thiadiazole amides as potent Cdc25B and PTP1B inhibitors. Bioorg Med Chem Lett 24: 4125-4128, 2014.
- 78. Feng MT, Wang T, Liu AH, Li J, Yao LG, Wang B, Guo YW and Mao SC: $\overline{PTP1B}$ inhibitory and cytotoxic \overline{C} -24 epimers of Δ28‑24‑hydroxy stigmastane‑type steroids from the brown alga Dictyopteris undulata Holmes. Phytochemistry 146: 25‑35, 2018.
- 79. de Oliveira TV, Stein R, de Andrade DF and Beck RCR: Preclinical studies of the antitumor effect of curcumin-loaded polymeric nanocapsules: A systematic review and meta‑analysis. Phytother Res 36: 3202‑3214, 2022.
- 80. Maganti HB, Kirkham AM, Bailey AJM, Shorr R, Kekre N, Pineault N and Allan DS: Use of CRISPR/Cas9 gene editing to improve chimeric antigen-receptor T cell therapy: A systematic review and meta-analysis of preclinical studies. Cytotherapy 24: 405‑412, 2022.
- 81. Singh D, Gupta M, Sarwat M and Siddique HR: Apigenin in cancer prevention and therapy: A systematic review and meta-analysis of animal models. Crit Rev Oncol Hematol 176: 103751, 2022.
- 82. Sun JY, Zhang D, Wu S, Xu M, Zhou X, Lu XJ and JiJ: Resistance to PD-1/PD-L1 blockade cancer immunotherapy: Mechanisms, predictive factors, and future perspectives. Biomark Res 8: 35, 2020.
- 83. Luo QW, Yao L, Li L, Yang Z, Zhao MM, Zheng YZ, Zhuo FF, Liu TT, Zhang XW, Liu D, *et al*: Inherent capability of self‑assembling nanostructures in specific proteasome activation for cancer cell pyroptosis. Small 19: e2205531, 2023.
- 84. Zhou Z, Song Q, Yang Y, Wang L and Wu Z: Comprehensive landscape of RRM2 with immune infiltration in pan-cancer. Cancers (Basel) 14: 2938, 2022.

Copyright © 2024 Zeng et al. This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International (CC BY-NC-ND 4.0) License.