

CASE REPORT

Inactivity of imatinib in gastrointestinal stromal tumors (GISTs) harboring a KIT activation-loop domain mutation (exon 17 mutation pN822K)

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¹Division of Chest Medical Oncology, ²Division of Abdominal Surgery, ³Division of Pathology, ⁴Oncology Unit of Thymic cancer, Rare Tumors and Sarcomas, European Institute of Oncology, Milan, Italy **Abstract:** The development of gastrointestinal stromal tumors (GISTs) is largely driven by mutations in the KIT and $PDGFR\alpha$ genes. Imatinib mesylate is an oral small molecular tyrosine kinase inhibitor that mainly targets abl, c-KIT, and $PDGFR\alpha$. Imatinib achieves disease control in approximately 70%–85% of patients with advanced GIST, and the median progression-free survival is 20–24 months. The efficacy of imatinib correlates with tumor kinase mutational status (exon 11 mutations mainly), and some mutations are known to be responsible for primary and secondary imatinib resistance. Beyond these, there are many other mutations that are considered rare and are associated with unknown clinical behavior. In the literature, there are poor and inconsistent data about the inhibitor sensitivity of mutations occurring in the activation-loop domain encoded by exon 17. In this article, we focus on a case of a patient suffering from GIST, harboring an extremely rare KIT activation-loop domain mutation (exon 17 mutation pN822K) treated with imatinib. A review of the literature is also presented.

Keywords: GIST, KIT activation-loop domain mutation, drug resistance, imatinib

Introduction

Gastrointestinal stromal tumors (GISTs) are the most common mesenchymal tumors of the gastrointestinal tract.¹ At diagnosis, majority of the tumors are resectable, but approximately 50% of these cases recur.² The most common sites of primary disease are the stomach (60%) and small intestine (25%).³ Histologically, GISTs have a wide range of morphologies (from spindle cell to epithelioid), but the diagnosis requires an immunopositivity for *KIT* (*CD117*)⁴ or *DOG1*.⁵ Most GISTs, but not all, harbor mutations of *KIT* (approximately 75% of cases),⁶ or *PDGFRα* (approximately 5%–8%).⁷

Imatinib mesylate is an oral small molecular inhibitor of tyrosine kinases; it mainly targets *abl*, *c-KIT*, and *PDGFR*α.⁸ Imatinib achieves disease control in approximately 70%–85% of patients with advanced GIST, and the median progression-free survival is 20–24 months with a median survival of 5 years.^{9,10} The spectrum of activity of imatinib is strongly affected by the driver mutations that the tumor harbors and, in particular, the juxtamembrane domain mutations (encoded by exon 11) are the most sensitive, while the extracellular domain mutations (exon 9) require a higher dose of imatinib to achieve good clinical control.¹⁰ A subgroup analysis from the ACOSOG Z9001 Trial¹¹ (1 year of adjuvant imatinib in a placebo-controlled trial in 645 patients) did not demonstrate that tumor mutation status independently affects relapse-free survival in either the placebo or imatinib arm. In detail, there was a clear benefit of adjuvant imatinib in patients with *KIT* exon 11 deletions (not *KIT* exon 11 insertions or point mutations).

Correspondence: Gianluca Spitaleri European Institute of Oncology, Ripamonti 435, 20141 Milan, Italy Tel +39 025 748 9482 Fax +39 029 437 9235 Email gianluca.spitaleri@ieo.it However, drug resistance caused by secondary *KIT* or *PDGFRA* mutations eventually develops in 90% of cases. ¹² In addition to these mutations, there are many others that are considered rare and are associated with unknown clinical behavior. In this article, we report a case of a patient suffering from GIST harboring a rare *KIT* activation-loop domain mutation (exon 17 mutation pN822K) treated with imatinib.

Case report

In November 2009, a 48-year-old Philippine man without relevant comorbidities was admitted to an Asian hospital to undergo surgical resection of a high-risk ileal 10 cm GIST.

After surgery, he moved to Italy and came to our institute for a second opinion. Tumor tissues for the molecular analysis to sequence *c-KIT* were not available, so he was treated with a 1-year adjuvant treatment with imatinib at a daily dose of 400 mg daily, according to 2009 international guidelines.

On October 2012, a follow-up abdominal computed tomography (CT) scan detected a 45×32 mm unique local relapse, and treatment with 400 mg a day imatinib was administered again. A tumor biopsy was not performed due to the patient's refusal.

A CT scan performed after 6 weeks from imatinib onset showed that the lesion increased in size, with no areas of reduced contrast enhancement. Imatinib dosage was then increased to 800 mg a day, but a subsequent CT scan performed after 6 weeks showed no signals of treatment response. The lesion was unique at CT scan and was amenable to radical surgery; on February 27, 2013, the patient underwent surgical disease excision. A large implant of recurrent GIST was visible on the peritoneal surface of the abdominal wall, 8.5 cm in longitudinal diameter. It was apparently increased with respect to the previous CT scan, despite the fact that the patient had not interrupted imatinib administration. An enlarged epiploic appendix of the sigmoid

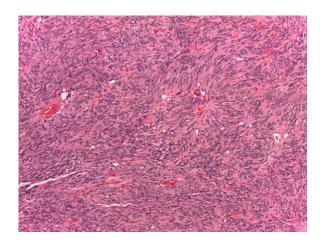


Figure 1 The tumor was composed of a monotonous spindle-cell proliferation. **Notes:** Hematoxylin and eosin stain; original magnification: 10×.

colon was removed for histology, and a peritoneal washing was performed for cytology. The recurrent lesion was eventually radically removed, together with the adherent omentum, taking care not to open the lining capsule surrounding it.

Macroscopically, the tumor was roundish and with a hard consistency; the maximum diameter was 8.5 cm. The cut surface was grayish and dishomogeneous for the presence of hemorrhagic areas. Histologically, the tumor was composed of bland spindle cells (Figures 1 and 2A). There were no areas of tumor necrosis. Many dilated and thrombosed vessels resembling similar findings seen in neurogenic tumors were intermingled within the tumor cells. Immunocytochemical stains revealed strong cytoplasmic expression of CD117, DOG1, and CD34 (Figure 2B and C). No expression was detected for desmin and S100 protein. c-KIT (exons 9, 11, 13, and 17) and PDGFRα (exons 12, 14, and 18) mutational analyses were performed by bidirectional Sanger sequencing, using BigDye Terminator chemistry, on a 3500 Dx Genetic Analyzer. The test results showed a single mutation in exon 17 of the c-KIT gene (pN822K; Figure 3), confirmed in two independent amplifications, while the PDGFR \alpha mutational status

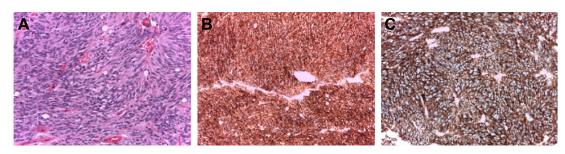


Figure 2 Gastrointestinal stromal tumor cell histology.

Notes: (A) Tumor cells were cytologically bland. No areas of necrosis or atypical mitoses were detected (hematoxylin and eosin stain; original magnification: 10×).

(B) Tumor cells expressed strong and diffuse positivity for CD117 (KIT) (original magnification: 20×). (C) Tumor cells expressed strong and diffuse positivity for DOG-1 (original magnification: 20×).

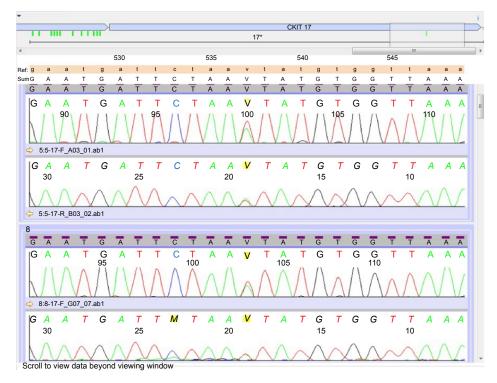


Figure 3 Electropherogram obtained by bidirectional Sanger sequencing of two independent amplifications, both showing a pN822K mutation.

was wild type. Based on the evidence of prior response to imatinib (no tumor shrinkage at instrumental evaluation and no pathological response at the histological report) and the evidence of this rare mutation, treatment with imatinib was not restarted and we decided to begin a clinical—instrumental follow-up every 3–4 months.

At the time of this report, 18 months after surgical resection of the relapsed disease, the patient is still in complete remission.

Discussion

In this article, we have described the case of a patient affected by a GIST harboring an extremely rare *KIT* exon 17 mutation, pN822K, treated with imatinib. The description of this case has significant caveats because we cannot classify this mutation as intrinsic or acquired resistance, since the sample from the first surgery was not available to perform gene sequencing analyses. In our case, the mutation was associated with resistance to imatinib.

The development of GISTs is largely driven by mutations in the KIT and $PDGFR\alpha$ genes. KIT and $PDGFR\alpha$ secondary mutations, or nonsensitive primary mutations, represent the principal cause of resistance to imatinib; other mechanisms can involve the activation of different pathways (eg, k-ras and BRAF). $^{6,13-17}$

Regarding the epidemiology, mutations affecting the kinase domain in untreated primary resected GISTs are very rare (each <1%), mainly occurring at codon 642 in exon 13 and at codons 816, 820, and 823 in exon 17. Table 1 depicts the frequency (0.4%–4.5%) of c-KIT exon 17 mutations, as reported by several retrospective series of imatinib-naïve GIST patients. $^{19-27}$

Very little data are available regarding imatinib sensitivity in GISTs harboring kinase-domain mutations. In a smaller study by Bozzi et al,28 it was found that two out 18 imatinib-treated patients had secondary exon 17 KIT mutations (N822K, D816H) associated with exon 11 KIT mutations (dup W577-R586, Ex11:V559A). In a Korean study of 290 patients, ²⁹ two patients were enrolled with primary exon 17 mutations (A794T and G812D), and both attained a partial response. However, the paper did not report the long-term outcomes of these patients. Conversely, the development of subsequent resistance to imatinib is more frequent, as was demonstrated by a meta-analysis of ten trials (>1,000 patients).³⁰ It was shown that the prevalence of second-site KIT or PDGFRA mutations was 61.3%; specifically, the prevalence of exon 17 mutation was the most common, at 54.5%.

In a Phase II study of imatinib conducted with 147 patients with GIST,³¹ molecular analyses were performed

Table I Frequency of KIT exon 17 mutations in untreated patients with GIST from retrospective studies

References	Patient (N)	Exon 17 mutations (N)	Type(s) of mutation	Primitive site
Subramanian et al ¹⁹	26	I (3.8%)	N822K	Gastric
Wasag et al ²⁰	28	I (3.5%)	N822K	Duodenum
Rossi et al ²¹	167	I (0.6%)	Not reported	Not reported
Arne et al ²²	204	I (0.5%)	Not reported	Not reported
Kern et al ²³	95	2 (2.1%)	N822K; G826E	Gastric
Calabuig-Fariñas et al ²⁴	154	3 (1.9%)	D816V; N822K; V823D	Not reported
Wozniak et al ²⁵	427	2 (0.4%)	N822K; N822H	Not reported
Kang et al ²⁶	22	I (4.5%)	Not reported	Not reported
Joensuu et al ²⁷	1,505	10 (0.6%)	Not reported	6/10 nongastric

Abbreviations: GIST, gastrointestinal stromal tumor; N, number.

using samples from ten patients with primary resistance and 33 patients with secondary resistance. Uncommon kinase mutations were most common in GISTs with secondary resistance when compared to those with primary resistance tumors (67% vs 10%, respectively; P=0.002).

Recently, a retrospective analysis of 93 patients treated with neoadjuvant imatinib³² showed that exons 9, 13, and 17 mutations were rare, and that they seem to confer a major risk of recurrence.

Two preclinical studies have attempted to predict the association between exon 17 mutations and inhibitor sensitivity, yielding contradictory results.^{33,34} The first study with a xenograft model of GIST harboring an exon 17 D816H mutation has shown a response to intermittent or continuous imatinib treatment;³³ the second study explored the response of an exon 17 (A818T) knock-in mouse to imatinib, showing that it was refractory.³⁴

In the literature, there are a few clinical reports about GISTs harboring exon 17 mutations showing a worse prognosis when treated with imatinib or sunitinib.^{35–38} In 2006, Loughrey et al³⁵ described a case of a female patient affected by small-intestine GIST who progressed after 1 year of treatment of imatinib, and the mutational analysis performed over a peritoneal disease sample revealed a *KIT* exon 11, 21 amino acid deletion (identical to that found in the primary tumor) in association with an exon 17 point mutation resulting in an Asn822Lys substitution in the kinase domain. In 2009, a retrospective analysis showed that four out of ten

patients refractory to imatinib harbored a mutation in exon 17 (T2467G).³⁶ Subsequently, in another retrospective series, a different mutation in exon 17 (Y823D) was found in nine out 18 refractory patients.³⁷

In 2010, a case was described of a patient with advanced gastric GIST harboring an exon 11 *KIT* mutation who, after a 3-year imatinib treatment, developed two new liver lesions harboring two different *KIT* exon 14 (c.2096C[T]) and *KIT* exon 17 (c.2554T[G]) mutations with dissociated clinical behavior (both failed to benefit from increasing imatinib dose, while the exon 14 lesion benefited from a sunitinib switch).³⁸

Regarding the focus of our report, the exon 17 N822K mutation is extremely rare and, so far, it is associated with unpredictable clinical behavior. Table 2 summarizes the description of all case reports about GIST harboring KIT exon 17 mutation N822K. In 2009, Nishida et al³⁹ reported that four patients with GIST harboring secondary/tertiary exon 17 mutations (D816H, W823D, D822K, plus N822K) did not benefit from sunitinib treatment. In 2011, Hanson et al⁴⁰ described a complex case of a female patient affected from a small-intestine rhabdoid GIST harboring KIT exon 11 579–580 insertion. At the excision of the mass, peritoneal metastases were noted, and she was subsequently treated with 1 year of imatinib. Then, 3 years later, there was an abdominal recurrence harboring an exon 17 N822K mutation, together with an exon 11 mutation. After the excision of the mass, she was treated with 2 years of sunitinib in the absence of a

Table 2 Summary of case reports focusing on KIT exon 17 pN822K mutation

References	Patient (N)	Primitive site(s)	Type(s) of mutation	Refractory to
Nishida et al ³⁹	4	Three small intestine; stomach	D816H; W823D; D822K; N822K	Imatinib; sunitinib
Hanson et al40	1	Small intestine	N822K	Imatinib
Gao et al ⁴¹	4	Not reported	N822K	Imatinib; sunitinib
Singeltary et al42	1	Rectum	N822K	Imatinib; sunitinib
Our case	1	Small intestine	N822K	Imatinib

Abbreviation: N, number.

macroscopic evidence of the disease. Later, a third recurrence was excised (sunitinib was given in absence of the macroscopic disease so it cannot be speculated if the disease was responsive to sunitinib) and this did not harbor an exon 17 mutation. The authors speculated that there was a preexisting clone harboring an exon 17 mutation that was selected following imatinib treatment. In 2013, a retrospective analysis of 38 patients who developed imatinib resistance were documented 12 cases with exon 17 mutations (including also N822K mutations), and all the patients with these secondary mutations failed to benefit from switching treatment to sunitinib.⁴¹

Interestingly, a second case of a 54-year-old white male with a rectal GIST harboring an N822K mutation at c-KIT exon 17 was described.⁴² The patient did not respond to imatinib (neither when administered at 400 mg daily nor at 800 mg daily), nor to sunitinib. After two lines of treatment, a third line was attempted with sorafenib, which resulted in a short-lasting partial response (progression with new lesions at 6-month treatment); so far, it was decided that imatinib be added to sorafenib, attaining 2-year disease stabilization.

This case report documented imatinib resistance in a patient with a rectal GIST harboring a *c-KIT* N8222K mutation, which was proven both by radiological disease progression and with the absence of pathological features of imatinib response.

Conclusion

In conclusion, this report suggests that patients with GIST harboring this rare mutation can be resistant to imatinib. Nowadays, sunitinib remains the standard of care for imatinib-refractory GISTs, regardless the status of their secondary KIT mutation. However, genotype analysis showed that patients with a secondary KIT mutation involving the activation-loop domain have poor progression-free survival and overall survival. 43,44 Novel tyrosine kinase inhibitors (nilotinib, regorafenib, sorafenib) have been tested in preclinical and clinical settings. 45-48 Two Phase III randomized trials (nilotinib, regorafenib)^{45,46} and two Phase II trials (sorafenib)^{47,48} have shown a modest benefit of these drugs in patients with imatinib/sunitinib-refractory GIST. Subgroup analyses of secondary mutations of KIT were not performed to check if these drugs could be active in the presence of activation-loop domain mutations. A rebiopsy should be encouraged to better define the mechanisms of acquired resistance, and an effort should be made to retrospectively and prospectively analyze the outcomes of patients with GIST harboring rare mutations treated in clinical trials.

Preclinical studies tested the sensitivity of imatinibresistant GIST cells to PKC412 and found that *KIT-V654A* and *PDGFRA-D842V* mutants were sensitive to *PKC412*. ⁴⁹ Indeed, preclinical studies could help to select drugs active for loop domain mutations: a molecular modeling analysis showed that the fragment deletion of exon 11 and the point mutation on exon 17 would lead to a shift of *KIT* conformational equilibrium toward an active form, for which nilotinib and sorafenib bind with higher affinity than imatinib and sunitinib; thus far, they could be tested in these settings. ⁵⁰

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Author contributions

All authors contributed toward data analysis, drafting and revising the paper and agree to be accountable for all aspects of the work.

Disclosure

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