



Genome Sequence of *Lelliottia* sp. Strain WAP21, Isolated from Soil in Canola Fields in Victoria, Australia

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ABSTRACT Here, we describe the genome of *Lelliottia* sp. strain WAP21, which was isolated from the soil of canola fields in Australia. The genome has a size of 4.9 Mbp and 4,583 predicted genes, with some potential pathways for metabolism of various carbon sources and metal acquisition.

The genus *Lelliottia* is a relatively newly characterized genus within the *Enterobacteriaceae* family that was reclassified from *Enterobacter* as a novel genus based on multilocus sequence analysis (MLSA) in 2013. *Lelliottia* bacteria are Gram-negative, rod-shaped, facultative anaerobes that have been identified in diverse environments and food (1). *Lelliottia amnigena* was associated with onion bulb decay (2), while *Lelliottia jeotgali* sp. nov. was found in fermented clam (3) and *Lelliottia aquatilis* sp. nov. was identified in a drinking water storage reservoir (4).

Lelliottia sp. strain WAP21 was isolated from a rhizosphere, bulk soil, and canola root sample obtained in Bungalally, Victoria, Australia (coordinates: −36.814969, 142.307717). A suspension was made from the sample with sterile phosphate-buffered saline (PBS), and colonies were obtained by 1,000-fold dilution, spread plating on tryptone-yeast (TY) agar, and aerobic incubation at 22°C for 48 h. Single colonies confirmed with Gram stain were further streak plated on tryptic soy agar (TSA) and incubated aerobically at 30°C for 24 h. Genomic DNA was then extracted with the Monarch genomic DNA purification kit (New England Biolabs). A library was prepared from the extracted DNA with the Nextera Flex library preparation kit and sequenced on a MiSeq platform using the MiSeq reagent kit v2 (300 cycles) (Illumina, San Diego, CA, USA). A total of 1,259,356 paired-end reads with an average length of 150 bp were retained after quality and adapter trimming using Trimmomatic v0.39 (5). Default settings were used for all tools unless otherwise specified. Trimmed reads were *de novo* assembled by SPAdes v3.15.3 with a kmer size of 127 (6).

The genome, with 53 contigs (7), a G+C content of 53.5%, and a total length of 4,940,396 bp (N_{50} , 443,208 bp), was assessed using the QUAST Web interface (<http://cab.cc.spbu.ru/quast>; 8); 4,583 genes were predicted with Prodigal v2.6.3 (9) and annotated with PGAP v5.3 (10).

Classification was performed by GTDB-Tk v1.5.0 against the Genome Taxonomy Database (GTDB) release 202 (11). Strain WAP21 was assigned to the *Lelliottia* genus but failed to be assigned to any existing species, with closest placement average nucleotide identity (ANI) of 93.07% to *L. amnigena* ZB04 (GenBank assembly accession number [GCA_001652505.2](https://genbank.org/assembly/GCA_001652505.2)) and ANIm of 84.94% to *L. amnigena* LMG 2784^T (GenBank assembly accession number [GCA_002553545.1](https://genbank.org/assembly/GCA_002553545.1)) (cutoff value, 95%).

The predicted genes were analyzed with KofamKOALA v2022-03-01 (KEGG release 101.0), and pathway analysis was conducted with an E value setting of 0.01. This analysis showed the presence of diverse carbohydrate metabolism (12). The genome possesses a complete gene module for dissimilatory nitrate reduction (DNRA), suggesting a role for *Lelliottia* sp. strain

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WAP21 in the soil N cycle. One previous study demonstrated that high labile C availability shifts NO₃ consumption from denitrification to DNRA (13, 14).

Data availability. The nucleotide sequence of *Lelliottia* sp. strain WAP21 was submitted to GenBank under accession number [JAIWPG000000000](https://www.ncbi.nlm.nih.gov/nuclseq/JAIWPG000000000) (version [JAIWPG000000000.2](https://www.ncbi.nlm.nih.gov/nuclseq/JAIWPG000000000.2)). The raw reads were deposited in the NCBI Sequence Read Archive (SRA) with accession number [SRR16094769](https://www.ncbi.nlm.nih.gov/sra/SRR16094769).

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REFERENCES

- Brady C, Cleenwerck I, Venter S, Coutinho T, De Vos P. 2013. Taxonomic evaluation of the genus *Enterobacter* based on multilocus sequence analysis (MLSA): proposal to reclassify *E. nimipressuralis* and *E. amnigenus* into *Lelliottia* gen. nov. as *Lelliottia nimipressuralis* comb. nov. and *Lelliottia amnigena* comb. nov., respectively, *E. gergoviae* and *E. pyrinus* into *Pluralibacter* gen. nov. as *Pluralibacter gergoviae* comb. nov. and *Pluralibacter pyrinus* comb. nov., respectively, *E. cowanii*, *E. radincintans*, *E. oryzae* and *E. arachidis* into *Kosakonia* gen. nov. as *Kosakonia cowanii* comb. nov., *Kosakonia radincintans* comb. nov., *Kosakonia oryzae* comb. nov. and *Kosakonia arachidis* comb. nov., respectively, and *E. turicensis*, *E. helveticus* and *E. pulveris* into *Cronobacter* as *Cronobacter zurichensis* nom. nov., *Cronobacter helveticus* comb. nov. and *Cronobacter pulveris* comb. nov., respectively, and emended description of the genera *Enterobacter* and *Cronobacter*. *Syst Appl Microbiol* 36:309–319. <https://doi.org/10.1016/j.syapm.2013.03.005>.
- Liu S, Tang Y, Wang D, Lin N, Zhou J. 2016. Identification and characterization of a new *Enterobacter* onion bulb decay caused by *Lelliottia amnigena* in China. *App Microbiol Open Access* 2:114. <https://doi.org/10.4172/2471-9315.1000114>.
- Yuk K-J, Kim Y-T, Huh C-S, Lee J-H. 2018. *Lelliottia jeotgali* sp. nov., isolated from a traditional Korean fermented clam. *Int J Syst Evol Microbiol* 68:1725–1731. <https://doi.org/10.1099/ijsem.0.002737>.
- Kämpfer P, Glaeser SP, Packroff G, Behringer K, Exner M, Chakraborty T, Schmuthausen RM, Doijad S. 2018. *Lelliottia aquatilis* sp. nov., isolated from drinking water. *Int J Syst Evol Microbiol* 68:2454–2461. <https://doi.org/10.1099/ijsem.0.002854>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- Bosi E, Donati B, Galardini M, Brunetti S, Sagot M-F, Lió P, Crescenzi P, Fani R, Fondi M. 2015. MeDuSa: a multi-draft based scaffolder. *Bioinformatics* 31:2443–2451. <https://doi.org/10.1093/bioinformatics/btv171>.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Hyatt D, Chen G-L, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <https://doi.org/10.1186/1471-2105-11-119>.
- Li W, O'Neill KR, Haft DH, DiCuccio M, Chetvermin V, Badretdin A, Coulouris G, Chitsaz F, Derbyshire MK, Durkin AS, Gonzales NR, Gwadz M, Lanczycki CJ, Song JS, Thanki N, Wang J, Yamashita RA, Yang M, Zheng C, Marchler-Bauer A, Thibaud-Nissen F. 2021. RefSeq: expanding the Prokaryotic Genome Annotation Pipeline reach with protein family model curation. *Nucleic Acids Res* 49:D1020–D1028. <https://doi.org/10.1093/nar/gkaa1105>.
- Chaumeil P-A, Mussig AJ, Hugenholtz P, Parks DH. 2019. GTDB-Tk: a toolkit to classify genomes with the Genome Taxonomy Database. *Bioinformatics* 36:1925–1927. <https://doi.org/10.1093/bioinformatics/btz848>.
- Aramaki T, Blanc-Mathieu R, Endo H, Ohkubo K, Kanehisa M, Goto S, Ogata H. 2020. KofamKOALA: KEGG Ortholog assignment based on profile HMM and adaptive score threshold. *Bioinformatics* 36:2251–2252. <https://doi.org/10.1093/bioinformatics/btz859>.
- Friedl J, De Rosa D, Rowlings D, Grace P, Müller C, Scheer C. 2018. Dissimilatory nitrate reduction to ammonium (DNRA), not denitrification dominates nitrate reduction in subtropical pasture soils upon rewetting. *Soil Biol Biochem* 125:340–349. <https://doi.org/10.1016/j.soilbio.2018.07.024>.
- Li X, Qian W, Hou L, Liu M, Chen Z, Tong C. 2020. Soil organic carbon controls dissimilatory nitrate reduction to ammonium along a freshwater-oligohaline gradient of Min River Estuary, southeast China. *Mar Pollut Bull* 160:111696. <https://doi.org/10.1016/j.marpolbul.2020.111696>.