

# Phytochemicals in Cancer Chemoprevention: Preclinical and Clinical Studies

Cancer Control  
Volume 31: 1–38  
© The Author(s) 2024  
Article reuse guidelines:  
[sagepub.com/journals-permissions](https://sagepub.com/journals-permissions)  
DOI: 10.1177/10732748241302902  
[journals.sagepub.com/home/ccx](https://journals.sagepub.com/home/ccx)



Nitish Lekhak<sup>1</sup> and Hitesh Kumar Bhattarai, PhD<sup>1</sup>

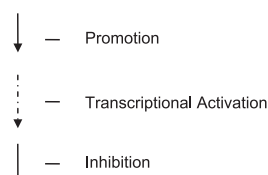
## Abstract

Phytochemicals, chemicals from plants, have garnered huge attention for their potential ability to prevent cancer. In vivo and preclinical models show that they do so often by affecting the hallmarks of cancer. Phytochemicals affect key pathways involved in the survival, genome maintenance, proliferation, senescence, and transendothelial migration of cancer cells. Some phytochemicals, namely antioxidants, can scavenge and quench reactive oxygen species (ROS) to prevent lipid peroxidation and DNA damage. They also trigger apoptosis by stopping the cell cycle at checkpoints to initiate the DNA damage response. Numerous in vitro and in vivo studies suggest that phytochemicals hinder cancer onset and progression by modifying major cell signaling pathways such as JAK/STAT, PI3K/Akt, Wnt, NF-κB, TGF-β, and MAPK. It is a well-known fact that the occurrence of cancer is in itself a very intricate process involving multiple mechanisms concurrently. Cancer prevention using phytochemicals is also an equally complex process that requires investigation and understanding of a myriad of processes going on in the cells and tissues. While many in vitro and preclinical studies have established that phytochemicals may be potential chemopreventive agents of cancer, their role in clinical randomized control trials needs to be established. This paper aims to shed light on the dynamics of chemoprevention using phytochemicals.

## Keywords

cancer, phytochemicals, chemoprevention, antioxidants, signaling pathways, immunotherapy, clinical trials, bioavailability

Received July 18, 2024. Received revised October 11, 2024. Accepted for publication November 11, 2024.



## Introduction

Cancer is a disease caused by abnormal growth and uncontrolled division of cells. It can initiate at any organ and can metastasize to remotely located organs. A diverse array of causative agents aid in causing cancer, ranging from physical agents, namely ultraviolet rays and ionizing radiation, to chemical agents like asbestos, alcohol, and tobacco smoke, and biological agents such as certain bacteria, viruses, and parasites.<sup>1</sup> According to the International Agency for

Research on Cancer (IARC), cancer is among the leading causes of death worldwide in 2022 resulting in 9.7 million fatalities. About 1 in 5 people develop cancer in their lifetime, and approximately 1 in 9 males and 1 in 12 females die from the cancer.<sup>2</sup> In recent times, the mortality rate from cancer has decreased considerably owing to the discoveries in molecular biology, recombinant DNA technology, and immunology.<sup>3,4</sup> Reduction in the number of smokers, maintenance of healthy eating habits, enhancement of living standards and

<sup>1</sup>Department of Biotechnology, Kathmandu University, Dhulikhel, Nepal

### Corresponding Author:

Hitesh Kumar Bhattarai, School of Science, Department of Biotechnology, P. O. Box 6250, Dhulikhel, Nepal.  
Email: [hitesh321@gmail.com](mailto:hitesh321@gmail.com)



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

environmental facilities, and focus of attention towards one's fitness are a few contributors to this heartening trend.

Healthy eating practices have been linked with lower cancer rates for a long time. While no conclusive evidence suggests any particular food can cure cancer, a nutritious diet encompassing vegetables, fruits, whole grains, and legumes can help lower the risk factors associated with cancer.<sup>5</sup> Different foods have different components that aid in safeguarding against various types of cancer. Some foods, such as tomatoes and grapes, contain chemicals like lycopene and resveratrol, which help fight against prostate, breast, and pancreatic cancers.<sup>6,7</sup> Other foods like whole grains have dietary fibers that protect from colon and digestive tract cancer.<sup>8</sup> Phytochemicals are naturally occurring bioactive compounds in plants that have several health benefits. Cancer cells have 6 distinctive traits defined as hallmarks of cancer. These signature attributes, unique to the cancer cells, are sustained proliferative signaling, evasion of growth suppressors, resistance to apoptosis, replicative immortality, induction of angiogenesis, and metastasis.<sup>9</sup> The phytochemicals in plant-based foods work as chemopreventive agents by mitigating the hallmarks of cancer. These chemicals such as isoflavones, catechins, and lycopene exert inhibitory effects against cancer.<sup>10</sup>

There are generally 2 types of chemopreventive agents of cancer. The blocking agents that suppress the initiation of cancer in healthy individuals reduce DNA damage by blocking endogenous or exogenous carcinogens. They prevent general genomic instability and neoplastic transformation. The mechanism might be decreased uptake of or increased metabolism of procarcinogens; enhanced detoxification of free radicals and electrophiles, and finally, upregulation of DNA repair activity. Other modes of action include the downregulation of inflammatory responses and the production of reactive oxygen and nitrogen species. In some instances, blocking agents act by obstructing epigenetic mechanisms of cancer progression such as methylation and deacetylation. The second type of chemopreventive agent is called a suppressing agent. They act on 1 or more signal transduction pathways that lead to cell proliferation. They also induce increased apoptosis of cancerous lesions and lead to inhibition of angiogenesis.<sup>11</sup>

Based on the stage at which the chemopreventive agent acts, chemopreventive agents can be grouped into 3 categories. The first category of chemoprevention, the primary chemoprevention, involves the prevention of cancer in high-risk and other healthy individuals. These agents prevent cancer biomarkers from appearing. The second category, secondary chemoprevention, includes the prevention of pre-malignant lesions from developing into invasive cancer. These include agents such as non-steroidal anti-inflammatory drugs (NSAIDs) in patients with colorectal cancer. Primary and secondary chemoprevention sometimes fall under the umbrella of primary chemoprevention. Finally, tertiary

chemoprevention involves administering agents that prevent the recurrence of invasive cancer.<sup>11</sup>

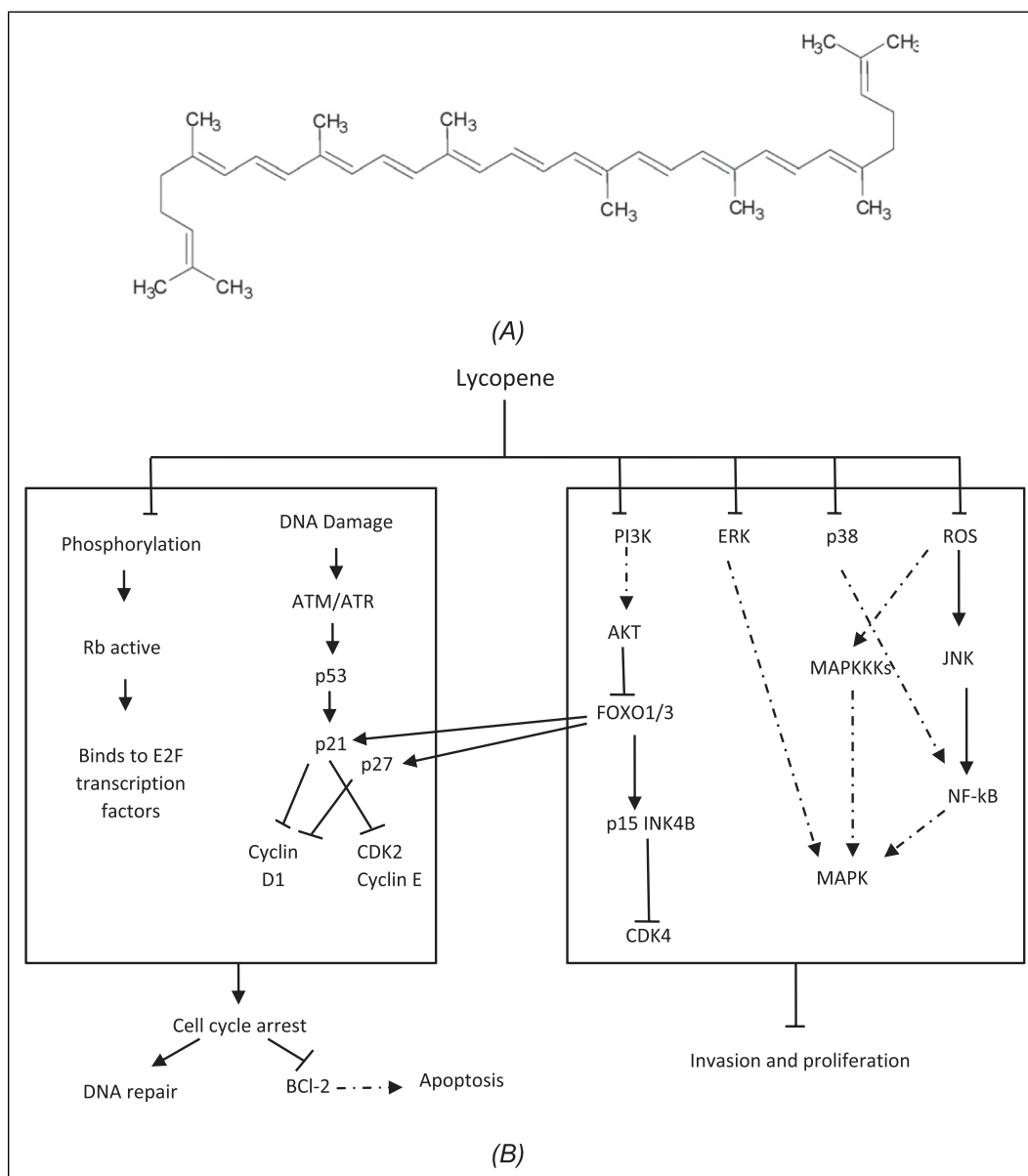
Phytochemicals can act both as suppressing agents or blocking agents in chemoprevention. They can also be primary, secondary, or tertiary chemopreventive agents. In vitro and in vivo studies have identified potential modes of action of certain phytochemicals. Most phytochemicals act on multiple signaling pathways and nodes of cancer development. The role of select phytochemicals—lycopene, resveratrol, sulforaphane, isoflavones, catechin, quercetin, curcumin, luteolin, and apigenin—in chemoprevention has been discussed in this paper.<sup>11</sup>

## Lycopene

Lycopene is a bright-red organic pigment called carotenoid, which gives fruits such as tomato, watermelon, red guava, and papaya their distinctive red color (Figure 1). It is a powerful antioxidant known for health benefits against cardiovascular diseases, hypertension, diabetes, and cancer.<sup>12</sup> When heated, the trans-isomer form of lycopene converts to cis-form and is readily absorbed by the body due to its increased solubility in bile acids.<sup>13</sup> This makes cooked tomatoes an ideal source of lycopene. Lycopene shows a dual impact on oxidative processes, acting as a pro-oxidant at high doses and an antioxidant at low doses. This dual nature of lycopene is vital in studying its potential chemopreventive properties. At low concentrations, it protects against DNA damage from reactive oxygen species, thereby reducing the chance of mutation and cancer.<sup>6</sup> At high doses, it helps destroy diseased cells. Thus, it can act in both primary and tertiary chemoprevention. In in vivo studies, lycopene has been shown to regulate oxidative and inflammatory processes, apoptosis, cell division, angiogenesis, and metastasis.<sup>14</sup>

Lycopene helps contain cancer cells by the arrest/apoptosis of tumor cells and inhibition of invasion. It reduces phosphorylation in Rb, allowing the active Rb to bind with E2F transcription factors, which helps arrest the cell cycle at the G1/S transition. When DNA damage is detected, ATM/ATR kinases are activated, and p53 is phosphorylated, triggering p21 and ultimately leading to cell cycle arrest by blocking cyclin D1, CDC2, and cyclin E. The arrested cell can have 2 fates; it can either undergo a DNA repair process or inactivate Bcl-2 proteins which leads to a series of steps in apoptotic cascade causing apoptosis. Lycopene also obstructs the PI3K/AKT, enabling FOXO3 stimulation, which promotes the expression of p15, p21, and p27 to trigger cell cycle arrest and apoptosis. Lycopene confines the invasive cancer cells by inhibiting the MAPK and NF- $\kappa$ B pathways. By limiting the ERK, p38, and ROS, it thwarts JNK and MAPK kinase kinases (MAPKKKs) to halt MAPK and NF- $\kappa$ B pathways (Figure 1B).<sup>15,16</sup>

Lycopene has been long studied for its chemopreventive and anticancer properties, especially against prostate cancer. It accumulates at a much higher concentration in prostate tissues



**Figure 1.** (A) Lycopene, (B) Modulation of cell signaling pathways by lycopene.

than in other tissues.<sup>17</sup> This tendency provides a reasonable explanation as to why the anticancer activities of lycopene are higher for prostate cancer than for other cancers. According to a study, lycopene subdues the phosphorylation effects on the Retinoblastoma protein (Rb), activates the tumor suppressor protein, p53, and inhibits the expression of cyclin D1 in G0/G1 phases, thereby arresting the cell cycle.<sup>18</sup> Additionally, the research suggests that lycopene inhibits the growth of non-neoplastic PrEC (Prostate epithelial cells) in vitro. Various papers indicate that this action of lycopene against prostate cancer might also be due to the alteration and inhibition of multiple pathways, including the PI3K/Akt pathway,<sup>17</sup> NF-kB signaling pathway,<sup>14</sup> AKT/mTOR pathway, MAPKs pathway, and JAK/STATs pathway.<sup>19</sup> A study by Yang et al

demonstrated that lycopene supplementation suppresses prostate tumor cell growth by decreasing the VEGF levels in plasma. The results of this study revealed that these effects are probably linked to the reduction of proliferation and the interference of insulin-like growth factor 1 signaling. Furthermore, inhibiting VEGF by lycopene could mean that antitumor activities of lycopene involve anti-angiogenesis.<sup>20</sup>

While a majority of the research indicates that lycopene mainly works wonders in prostate cancer, some papers mention that it has beneficial effects against other forms of cancer, such as breast cancer, by acting synergistically with other anticancer drugs like quinacrine that work by increasing the levels of ATP-binding cassette (APC), DAB2, GSK-3 $\beta$  and Axin, decreasing  $\beta$ -Catenin, p-GSK3 $\beta$  (ser 9) and

CK1 and inhibiting Wnt-TCF signaling.<sup>21</sup> Other sources suggest that it has positive results in cancers associated with inflammation such as *Helicobacter pylori*-induced gastric cancer, colitis-associated colorectal cancer, and pancreatic cancer related to chronic pancreatitis,<sup>22</sup> owing to its anti-inflammatory properties. However, these studies do not provide concrete evidence to support the claim that lycopene has the primary role in preventing inflammation-associated cancers, as none quantify its anti-tumor role.

### Immunomodulatory Effects of Lycopene

It has been demonstrated that lycopene modifies immune response through various mechanisms. According to preliminary data on immunomodulation, lycopene modifies dendritic cell responsiveness by downregulating the expression of common surface dendritic cell markers such as CD80, CD86, and MHC II molecules.<sup>23</sup> Regulatory T cells are crucial for the destruction of malignant cells. Increased T cell counts secrete inhibitory cytokines such as IL-10 and TGF- $\beta$ , which suppress effector cell function. In vitro studies have demonstrated a significant decrease in IL-10 and TGF- $\beta$  levels upon increasing the lycopene concentration from 0.5  $\mu\text{mol l}^{-1}$  to 5  $\mu\text{mol l}^{-1}$ .<sup>24</sup> Additional alterations in the concentrations of TNF- $\alpha$ , IL-1, IL-6, IL-8, and TIL-2, which are important T cell stimulants, were also noted.<sup>23,24</sup> Moreover, lycopene may improve T cell activation and differentiation by increasing the ratios of CD4+/CD8+, IFN $\gamma$ + /CD8+, perforin+/CD8+, and granzyme B+/CD8+ while having little effect on DNMT1 and DNMT3b.<sup>23</sup> Another study observed that lycopene may enhance the generation of serum immunoglobulin G and spleen B lymphocytes, improving cells' overall immunological function.<sup>16</sup>

### Lycopene Human Studies

Sui et al conducted an umbrella meta-analysis review to analyze and evaluate the association between carotenoids and cancer outcomes.<sup>25</sup> (Table 1) A total of 4 meta-analyses were reported on the association between lycopene intake and total cancer, among which 2 were statistically significant, suggesting a moderate impact of lycopene on total cancer risk. Ten meta-analyses reported on prostate cancer identified 5 statistically significant studies highlighting the role of lycopene in prostate cancer prevention. Lycopene intake also showed positive results in esophageal/larynx/oral cavity cancer. The association between the risk of some other forms of cancer and lycopene intake was also statistically significant, but more clinical trials need to be conducted to verify the results.<sup>25</sup>

### Resveratrol

Resveratrol is a bioactive compound belonging to the class of polyphenols called stilbenes (Figure 2). Usually occurring in

the trans-isomeric form in plants, the trans-resveratrol converts into a more bioactive form of dihydro-resveratrol when consumed orally.<sup>51</sup> The compound found predominantly in the skin of red grapes is produced as a protective agent by the grapes in response to environmental stressors, infection, or injury.<sup>52</sup> This phytoalexin antioxidant, produced primarily by red grapes to combat the damage done by UV radiation, shows promising results in the fight against skin cancer usually triggered by ultraviolet radiation and oxidative damage.<sup>52</sup> Several in vitro studies have shown that resveratrol has cytotoxic effects against skin, colon, breast, stomach, reproductive organs, liver, and thyroid cancer cells.<sup>53</sup> Resveratrol has been observed to reverse drug resistance in various cancer cells by making them responsive to anticancer drugs.<sup>54</sup>

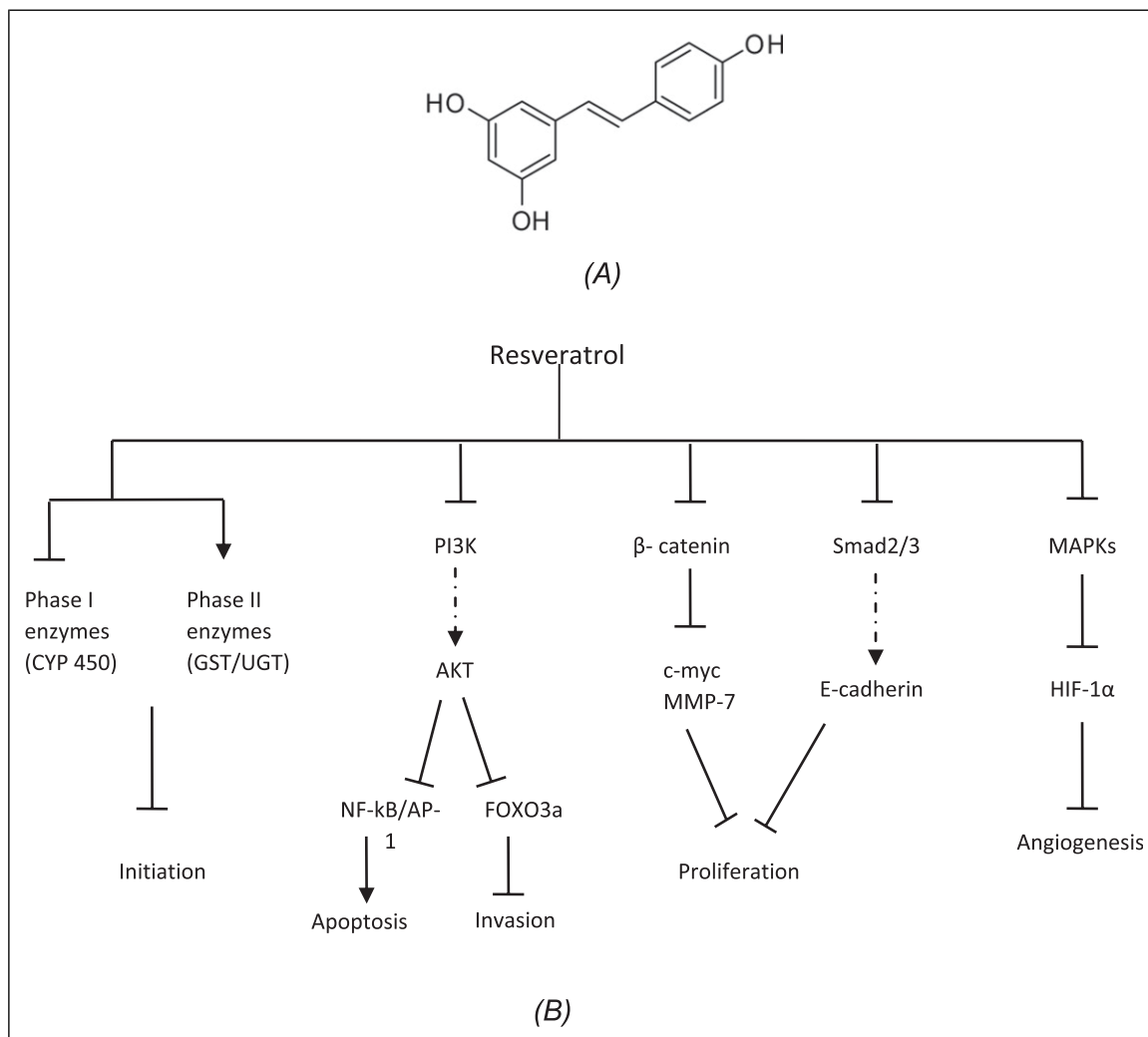
Resveratrol blocks Phase I enzymes (CYP450) to reduce the activation of carcinogens and induces Phase II enzymes (GST, UGT) to enhance the detoxification of carcinogens, thereby thwarting the cancer initiation process. It inhibits the PI3K/Akt pathway by inhibiting PI3K and, consequently inhibiting NF- $\kappa$ B/AP-1 and FOXO3a, leading to apoptosis and inhibition of invasion, respectively. Moreover, resveratrol halts the proliferation of cancer cells by disrupting the Wnt/ $\beta$ -catenin and TGF $\beta$ -Smad pathways. It checks proliferation by suppressing  $\beta$ -catenin, which restricts the transcription of c-myc and MMP-7, proteins necessary for proliferation. Similarly, the obstruction of Smad2/3 upregulates E-cadherin to stop the proliferation. Resveratrol also inhibits MAPKs, which hinders HIF-1 $\alpha$  to help prevent angiogenesis (Figure 2B).<sup>53,55,56</sup>

Resveratrol safeguards against cancer by scavenging and quenching the DNA-damaging reactive species, and preventing tumor initiation, progression, and metastasis. Resveratrol impedes the accumulation of ROS after exposure to oxidative agents like tobacco smoke condensate (TAR) and hydrogen peroxide.<sup>57</sup> It shows a protective action against lipid peroxidation and DNA damage by scavenging and quenching the ROS such as hydroxyl, superoxide, metal/enzymatic induced radicals, and radicals of cellular origin.<sup>58</sup> Attia further demonstrated the radical scavenging activity of resveratrol in their study on the effects of resveratrol on oxidative damage in genomic DNA and apoptosis induced by cisplatin. They observed that the cisplatin-induced genotoxicity and apoptosis were lessened in mice's resveratrol-treated somatic and germinal cells.<sup>59</sup> Induction of apoptosis mediated by the alteration of cyclin and cyclin-dependent kinase to halt the cell cycle is another mechanism by which resveratrol protects against cancer.<sup>60</sup> Resveratrol downregulates the cyclin D1/Cdk4 complex, resulting in the arrest of the cell cycle at the G0/G1 phase. Meanwhile, it enhances the expression of cyclin E and cyclin A, which stops the cell cycle at the G2/M and S phases.<sup>61</sup> A similar study by Kim et al on lung carcinoma A549 cells demonstrated the effects of resveratrol in S phase arrest of the cell cycle by reducing Rb

**Table 1.** Lycopene Clinical Studies.

Author and year	Type of Cancer	N	Type of Studies	Type of Metric	Summary Effect Size (95% CI)	Model	I <sup>2</sup>	Egger's P Value	Statistically Significant
Yin et al, 2022 <sup>26</sup>	Digestive system tumors	5	RCT	OR	0.93 (0.81, 1.08)	Random	0	NR	No
Aune et al, 2018 <sup>27</sup>	Total cancer	3	Cohort	RR	0.81 (0.54, 1.21)	Random	0.655	0.13	No
Psaltopoulou et al, 2018 <sup>28</sup>	Non-Hodgkin's lymphoma	3	CC	RR	1.00 (0.86, 1.16)	Random	0	NR	No
He et al, 2018 <sup>29</sup>	Breast cancer mortality	3	CC, cohort	RR	0.74 (0.53, 1.03)	Random	0	NR	No
Catano et al, 2018 <sup>30</sup>	Prostate cancer	24	CC, cohort	RR	0.90 (0.85, 0.95)	Random	0.04	NR	Yes
Chen et al, 2017 <sup>31</sup>	Non- Hodgkin's lymphoma	7	CC, cohort	RR	0.99 (0.88, 1.12)	Random	0	>0.05	No
Panic et al, 2017 <sup>32</sup>	Colorectal cancer	4	CC	OR	0.92 (0.46, 1.83)	Random	0.947	NR	No
Panic et al, 2017 <sup>32</sup>	Colon cancer	2	CC	OR	0.95 (0.79, 1.15)	Random	0	NR	No
Panic et al, 2017 <sup>32</sup>	Rectal cancer	2	CC	OR	0.82 (0.57, 1.16)	Random	0	NR	No
Panic et al, 2017 <sup>32</sup>	Colorectal cancer	3	Cohort	OR	0.94 (0.71, 1.24)	Random	0.622	NR	No
Rowles et al, 2017 <sup>33</sup>	Prostate cancer	21	CC, cohort	RR	0.88 (0.79, 0.99)	Random	0.567	0.13	Yes
Rowles et al, 2017 <sup>33</sup>	Prostate cancer	17	CC, cohort	RR	0.88 (0.79, 0.98)	Random	0.262	0.064	Yes
Chen et al, 2016 <sup>34</sup>	Pancreatic cancer	6	CC, cohort	OR	0.85 (0.73, 1.00)	Random	0	NR	No
Zhou et al, 2016 <sup>35</sup>	Gastric cancer	5	CC	OR	0.94 (0.73, 1.21)	Random	0.696	NR	No
Zhou et al, 2016 <sup>35</sup>	Gastric cancer	4	Cohort	OR	0.80 (0.60, 1.07)	Random	0	NR	No
Abar et al, 2016 <sup>36</sup>	Lung cancer	6	CC, cohort	RR	0.68 (0.54, 0.87)	Random	0	0	Yes
Wang et al, 2016 <sup>37</sup>	Colorectal cancer	15	CC, cohort	RR	0.94 (0.80, 1.10)	Random	0.805	0.864	No
Huang et al, 2016 <sup>38</sup>	Pancreatic cancer	8	CC, cohort	OR	0.84 (0.73, 0.97)	Random	0	0.857	Yes
Leoncini et al, 2015 <sup>39</sup>	Head and neck cancer	1	CC	OR	0.60 (0.32, 1.11)	Random	NR	NR	No
Leoncini et al, 2015 <sup>39</sup>	Oral cavity and pharynx	4	CC	OR	0.74 (0.56, 0.98)	Random	0.145	NR	Yes
Leoncini et al, 2015 <sup>39</sup>	Larynx	4	CC	OR	0.50 (0.28, 0.89)	Random	0.659	NR	Yes
Wang et al, 2015 <sup>40</sup>	Prostate cancer	13	CC, cohort	RR	0.88 (0.76, 1.02)	Random	0.2361	NR	No
Chen et al, 2015 <sup>41</sup>	Prostate cancer	13	CC, cohort	RR	0.91 (0.82, 1.01)	Random	0.455	0.22	No
Li et al, 2014 <sup>42</sup>	Ovarian cancer	10	CC, cohort	OR	0.963 (0.86, 1.08)	Random	0.116	0.406	No
Tang et al, 2014 <sup>43</sup>	Bladder cancer	6	CC, cohort	RR	0.95 (0.82, 1.10)	Random	0	NR	No
Tang et al, 2014 <sup>43</sup>	Bladder cancer	4	CC, cohort	RR	0.60 (0.17, 2.08)		0.61	NR	No
Ge et al, 2013 <sup>44</sup>	Esophageal cancer	2	CC, cohort	OR	0.75 (0.64, 0.88)	Fixed	0	0.114-0.962	Yes
Xu et al, 2013 <sup>45</sup>	Colorectal adenoma	8	CC	RR	0.87 (0.67, 1.13)	Random	0.44	NR	No
Chen et al, 2013 <sup>46</sup>	Prostate cancer	5	CC, cohort	OR	0.93 (0.86, 1.01)	Random	0.18	NR	No
Chen et al, 2013 <sup>46</sup>	Prostate cancer	9	CC, cohort	OR	0.97 (0.88, 1.07)	Random	0	NR	No
Myung et al, 2011 <sup>47</sup>	Cervical neoplasm	5	CC	OR	0.54 (0.39, 0.75)	Fixed	0.044	NR	Yes
Ilic et al, 2011 <sup>48</sup>	Prostate cancer	3	RCT	RR	0.67 (0.36, 1.23)	Random	0	0.859	No
Veloso et al, 2009 <sup>49</sup>	Total cancer	9	Cohort	OR/RR	0.99 (0.94, 1.05)	NR	NR	NR	No
Veloso et al, 2009 <sup>49</sup>	Total cancer	14	Nested CC	OR/RR	0.87 (0.77, 0.99)	NR	NR	NR	Yes
Veloso et al, 2009 <sup>49</sup>	Total cancer	24	CC	OR/RR	0.76 (0.64, 0.91)	NR	NR	NR	Yes
Etminan et al, 2004 <sup>50</sup>	Prostate cancer	10	CC, cohort	RR	0.89 (0.81, 0.98)	Random	NR	NR	Yes
Etminan et al, 2004 <sup>50</sup>	Prostate cancer	7	CC, cohort	RR	0.74 (0.59, 0.92)	Random	NR	NR	Yes

Abbreviations: N, number of meta-analyses; RCT, randomized controlled trial; CC, case-control; CI, confidence interval; OR, odds ratio; RR, relative risk; NR, not reported.



**Figure 2.** (A) Resveratrol, (B) Modulation of cell signaling pathways by resveratrol.

protein phosphorylation and inducing p21 and p53 protein expression.<sup>62</sup> Effective expression of these proteins provides the cell ample time to halt the cell cycle at various checkpoints and repair the DNA damage.

The antitumor activities of resveratrol are also attributed to various mechanisms by which it affects different signal transduction pathways responsible for cell growth, division, apoptosis, angiogenesis, and metastasis.<sup>53</sup> A study by Aziz et al observed that resveratrol inhibits the activation of phosphatidylinositol 3'-kinase/Akt, which ultimately results in modulations in the Bcl-2 family proteins, promoting the apoptosis of human prostate carcinoma LNCaP cells.<sup>63</sup> Resveratrol also induces apoptosis by modulating the mitogen-activated protein kinase pathway (MAPK)<sup>64</sup> and inhibiting NF-kB activation.<sup>65</sup> Similar studies noted that resveratrol alters pathways like Wnt/ $\beta$ -catenin, NF-kB, and AKT/GSK-3 $\beta$ /Snail, which play a significant role in cancer metastasis.<sup>66,67</sup> Specifically, resveratrol has been shown to inhibit colon cancer's invasion and metastasis by reversing

epithelial-mesenchymal transition via the AKT/GSK-3 $\beta$ /Snail signaling pathway.<sup>67</sup>

### *Immunomodulatory Effects of Resveratrol*

Resveratrol can function as an immunomodulator in a variety of pathways. Increased splenic proliferation, which leads to a larger cell population of T and B cells, is one of the mechanisms promoted by resveratrol. In a study by Lai et al, mice treated with resveratrol showed a significantly higher spleen index than mice treated with levamisole or control.<sup>68</sup> In separate in vivo experiments, comparable increases in IL-2, TNF- $\alpha$ , and NF-kB were noted, although PGE2 production was suppressed. When resveratrol was used, production of TNF- $\alpha$  gradually decreased up to 25  $\mu$ M, with the highest production reported at about 12.5  $\mu$ M PGE2.<sup>69</sup> Antitumor cytokines are primarily produced by activated CD8<sup>+</sup> T cells. Breast tumor-derived T cells treated with resveratrol showed a significant rise in the CD8<sup>+</sup>/CD4<sup>+</sup> ratio—a necessary ratio for

a longer survival rate for cancer patients—without causing any discernible change in CD4<sup>+</sup> levels.<sup>70</sup> Resveratrol promotes the release of anticancer cytokines such as IFN- $\gamma$  and inhibits the release of TGF- $\beta$ . It is also observed that resveratrol stimulates the polarization of CD4<sup>+</sup> T cells toward anticancer cells and decreases infiltration and polarization of immunosuppressive cells.<sup>71</sup> These immunomodulatory properties of resveratrol make it a suitable candidate for immunotherapy. Still, its poor bioavailability and low potency have made it difficult to be used in clinical settings. Nonetheless, new methods such as combination therapy and nanotechnology are being studied to overcome the limitations.

### Resveratrol Human Studies

A study by Honari et al. compiled the clinical trials conducted to evaluate the association of resveratrol with cancer incidence risk.<sup>72</sup> (Table 2) Currently, the potential chemopreventive effect of resveratrol is being investigated in a phase II clinical trial in colorectal polyp prevention in high-risk individuals in England and Wales. In the COLO-PREVENT trial, individuals with high-risk colon polyp will be randomized as placebo, low and high-dose resveratrol, and treated with 5 mg and 1g of resveratrol daily. Several endpoints will be measured to determine the efficacy of resveratrol in cancer chemoprevention.<sup>73</sup>

### Sulforaphane

Sulforaphane is a naturally occurring plant compound of the isothiocyanate family found in cruciferous vegetables such as broccoli, cabbage, cauliflower, and kale (Figure 3). It is not always present in its active form but is formed when myrosinase transforms its precursor glucoraphanin after any damage to the plant. Plants produce sulforaphane as a defense mechanism against pathogens, insects, and

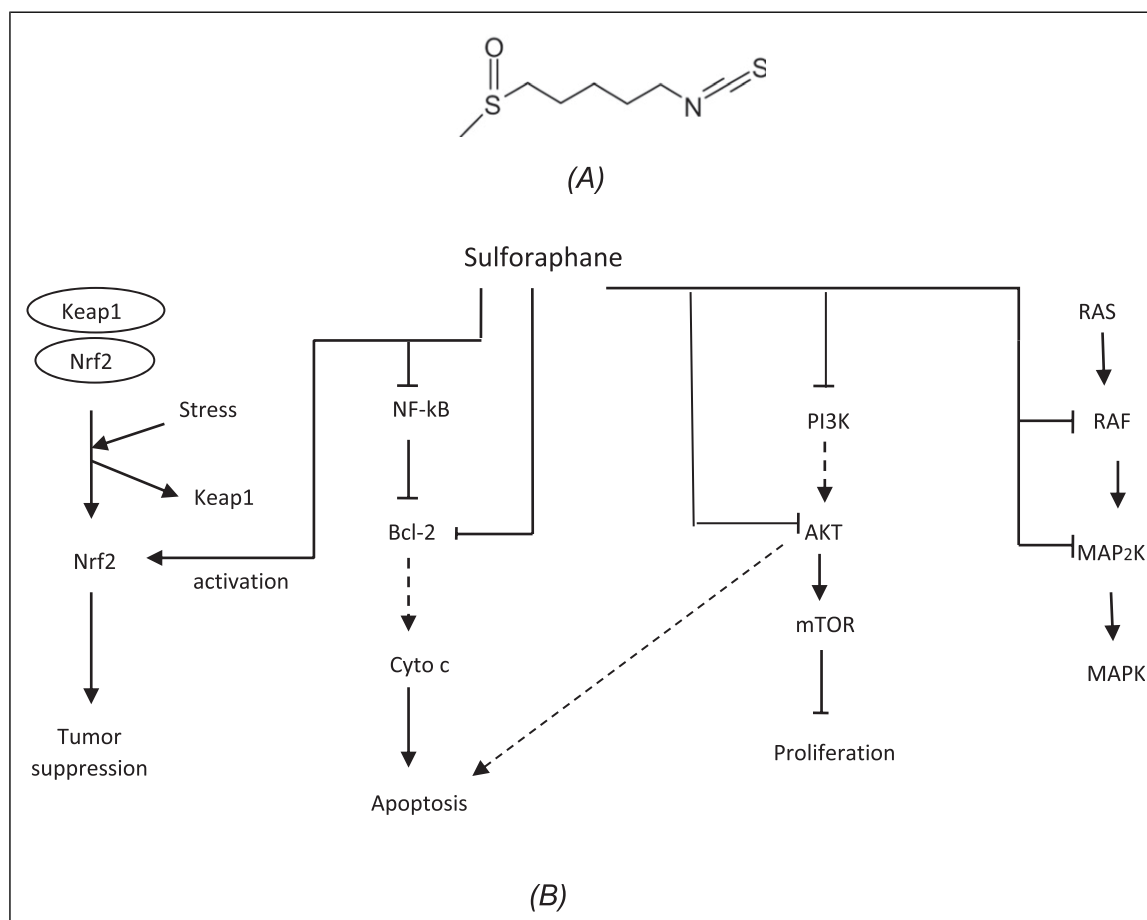
herbivores.<sup>81</sup> Numerous studies highlight sulforaphane's health benefits, including protection against cardiovascular diseases,<sup>82</sup> diabetic effects,<sup>83</sup> deterioration of brain health,<sup>84</sup> and gastrointestinal malfunctions.<sup>85</sup> Sulforaphane is also labeled a chemopreventive phytonutrient due to its cancer-preventive properties in vitro and in vivo.<sup>86</sup> Evidence suggests that sulforaphane intake is linked with lowering the risk of colon,<sup>87</sup> lung,<sup>88</sup> and prostate cancer.<sup>89</sup>

Sulforaphane acts as a tumor suppressor agent by promoting the transcription of Nrf2, a known tumor suppressor protein. It also displays apoptotic potential by blocking NF- $\kappa$ B, leading to Bcl-2 inactivation or direct inactivation of Bcl-2, triggering a series of steps that activate caspase 3, necessary for apoptosis. Moreover, sulforaphane subdues the PI3K/AKT pathway by hampering the PI3K and AKT. This disruption by inhibition of AKT stimulates the apoptotic pathways and disturbs mTOR, which is necessary for tumor survival and proliferation. Sulforaphane impedes the MAPK pathway by targeting multiple points: RAF and MAP2K. MAPK pathway is a crucial oncogenic pathway vital for cell proliferation (Figure 3B).<sup>90-92</sup>

The chemopreventive activity of sulforaphane is due to the inhibition of phase I enzymes responsible for activating pro-carcinogens and the promotion of phase II enzymes crucial in mutagen elimination.<sup>93</sup> It also alters cancer-related events like cell cycle, cell death, angiogenesis, metastasis, and invasion. The apoptotic action of sulforaphane is related to affecting the expression of apoptosis-related genes like p53, p21, Cdk2, Bax,<sup>94</sup> modulating the Bcl-2 family proteins,<sup>95</sup> activating caspases, and modulating cyclins and Cdks.<sup>96</sup> It is also noted that sulforaphane prevents proliferation, angiogenesis, and metastasis, the crucial steps in developing a full-blown cancer. A case in point is provided by an in vitro study conducted by Carrasco-Pozo et al, where only 10  $\mu$ M

**Table 2.** Resveratrol Clinical Studies.

Author and Year	Type of Cancer	Dose of Resveratrol	Duration of Study	Number of Patients	Results
Howells et al, 2011 <sup>74</sup>	Colorectal cancer	5g/day	14 days	6	Increased the cleaved caspase-3 in malignant hepatic tissue
Patel et al, 2010 <sup>75</sup>	Colorectal cancer	0.5 or 1g/day	8 days	20	Decreased expression of Ki-67
Nguyen et al, 2009 <sup>76</sup>	Colorectal cancer	20 or 80 mg/day	14 days	8	Inhibited the wnt pathway
Kjaer et al, 2015 <sup>77</sup>	Prostate cancer	150 mg or 1000 mg/day	4 months	66	Serum levels of androgen decreased
Paller et al, 2015 <sup>78</sup>	Prostate cancer	4000 mg/day	4 months	14	Safe
Popat et al, 2013 <sup>79</sup>	Multiple myeloma	5 g/day	21 days	24	Unsafe and no positive effects recorded in relapsed patients
Zhu et al, 2012 <sup>80</sup>	Breast cancer	5 or 50 mg twice a day	3 months	39	Reduced the methylation of RASSF-1 $\alpha$



**Figure 3.** (A) Sulforaphane, (B) Modulation of cell signaling pathways by sulforaphane.

sulforaphane could suppress the increased proliferation rate of lymph node carcinoma of the prostate (LnCaP) cells containing stimulated androgen receptors and prostate-specific antigen (PSA).<sup>97</sup> A similar *in vitro* study by Bertl et al, conducted using immortalized human microvascular endothelial HMEC-1 cells, suggested that sulforaphane significantly decreases microcapillaries' formation, inhibits capillary-like tubes on the basement membrane matrix, and hinders cell migration.<sup>98</sup>

Sulforaphane's apoptotic, antiproliferative, and anti-metastatic properties are intricately linked to its ability to modulate various signal transduction pathways involved in cancer development and progression. Extensive studies have shown that sulforaphane regulates numerous oncogenic signaling pathways like the Nrf2 - Keap1 pathway, NF- $\kappa$ B pathway, Akt pathway, signal transducer and activator of transcription 3 and 5, and other survival proteins.<sup>99</sup> Administration of sulforaphane inhibited the proliferation of CRC cells by upregulating the expression of UDP glucuronosyltransferase 1A (UGT1A) in CRC cells via extracellular signal-regulated kinase/Nuclear factor erythroid 2-related factor 2 (ERK/Nrf2) signaling pathway.<sup>87</sup> Moreover, sulforaphane inhibited the activities of sonic hedgehog (Shh), smoothened

(Smo), Glioma-associated oncogene homolog 1 (Gli1), and Polyhomeotic-like protein 3 (PHC3) in CD133<sup>+</sup> lung cancer cells by modulating the Shh signaling pathways and PHC3 to constrain the self-renewal of lung cancer stem cells.<sup>88</sup>

### Immunomodulatory Effects of Sulforaphane

Sulforaphane is a potent immunomodulator. It inhibits the NF- $\kappa$ B signaling pathway, which thwarts the expression of proinflammatory cytokines such as TNF- $\alpha$ , COX-2, IL-1 $\beta$ , and IL-18.<sup>100</sup> Similarly, it enhances the production of IL-2 and IFN- $\gamma$  along with the proliferation of splenocytes, bone marrow cells, and thymocytes by stimulating various mitogens such as concanavalin A, phytohaemagglutinin, pokeweed mitogen, and lipopolysaccharide.<sup>101</sup> Similar attributes related to Nrf2 activation by sulforaphane enhanced cytoprotection by mitigating DNA damage and suppressing lipogenesis.<sup>102</sup> It also induces ROS-mediated upregulation of NKG2D ligands, increasing the susceptibility of tumor cells to NK cell-mediated lysis.<sup>103</sup> Furthermore, sulforaphane increases dendritic cell stimulatory capacity by modulating regulatory molecules, JAK/STAT3 - and microRNA - signaling, suggesting the potential for immunotherapy.<sup>104</sup>



However, sulforaphane's effects on cancer cells are intricate because, in primary human T cells, it acts in a pro-oxidative manner, potentially hindering T cell activation and their effector functions.<sup>105</sup> This dual nature of sulforaphane on cancer cells suggests that even though it shows promise in cancer immunotherapy, its combination with T cell-mediated therapy requires serious considerations to avoid any mishaps in the treatment.

### Sulforaphane Human Studies

In 2023, a systematic study was carried out to analyze the randomized clinical trials conducted to test the efficacy and safety of sulforaphane against cancer. (Table 3) Among the 8 clinical trials identified, 4 were conducted to detect the efficacy of the phytochemical against prostate cancer, 2 against breast cancer, 1 against melanoma, and 1 against pancreatic cancer. While some histological markers and vital genes were positively affected by sulforaphane, there was substantial clinical and methodological heterogeneity. Although not statistically significant, sulforaphane improved the overall survival of pancreatic cancer patients. Further clinical trials with robust experimental designs and dosing regimens are necessary to determine the clinical efficacy of this compound as an anticancer agent.<sup>106</sup>

A 2021 paper by Kaiser et al evaluated several small clinical trials to assess the chemoprotective role of sulforaphane against various cancers.<sup>115</sup> Most of these trials tested whether the compound could inhibit type I enzymes, promote type II

enzymes, repair DNA damage breaks, decrease deacetylase activity, and upregulate and downregulate important cancer biomarkers. Although around twenty preliminary clinical trials demonstrated positive outcomes, larger clinical trials with more robust primary and secondary endpoints need to be included.

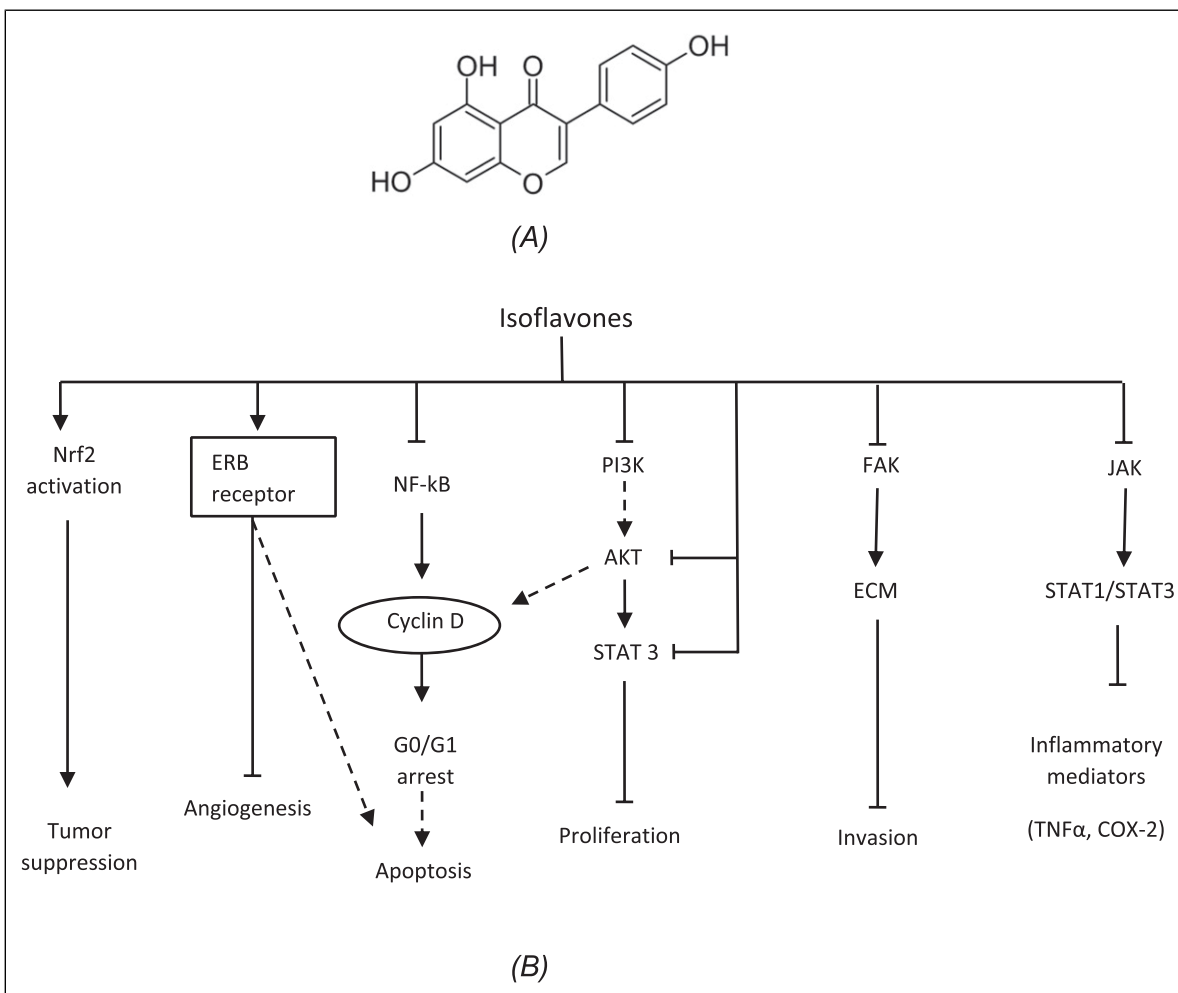
### Isoflavones

Isoflavones are naturally occurring flavonoids produced by the bean family members (Figure 4). Legumes, grains, and some vegetables contain isoflavones, but soybeans are the richest source of isoflavones. Some of the major isoflavones in soybeans are genistein, daidzein, and glycitein, ranging from 1.2 to 4.2 mg/g dry weight.<sup>116</sup> Isoflavones are considered phytoestrogens as they have estrogen-like properties and are derived from plants.<sup>117</sup> These phenolic plant compounds exert estrogenic/anti-estrogenic effects, influencing hormonal balance. Initially present in inactive forms in plants as glycosides, isoflavones are later converted to active aglycone forms by gut bacteria in the intestine.<sup>118</sup>

Isoflavones have numerous potential health benefits, particularly concerning hormone regulation, and are used to treat hormone-dependent health conditions like menopause, breast cancer, and CVD.<sup>119</sup> Isoflavone maintains healthy endothelium and prevents endothelial cell dysfunction by inducing nitric oxide production to protect from CVD.<sup>120</sup> Isoflavones' increased nitric oxide bioavailability also imparts antihypertensive effects by reducing endothelial cell oxidative stress or modulating vascular ion channel activity.<sup>121</sup> One study by Guo et al highlights the ability of isoflavones to exert estrogen-

**Table 3.** Sulforaphane Clinical Studies.

Author and Year	Type of Cancer	Dose of Sulforaphane	Duration of Study	Number of Patients	Results
Zhang et al, 2020 <sup>107</sup>	Prostate cancer	100 $\mu$ mol twice a day	4 - 8 weeks	98	No significant difference in HDAC activity or prostate tissue biomarkers was observed
Traka et al, 2019 <sup>108</sup>	Prostate cancer	Broccoli soup 300 mL/week	1 year	61	Changes in gene expression and associated oncogenic pathways were attenuated
Cipolla et al, 2015 <sup>109</sup>	Prostate cancer	60 mg for 6 months and 2 months without treatment	8 months	78	Managed biochemical recurrences in prostate cancer after radical prostatectomy
Traka et al, 2008 <sup>110</sup>	Prostate cancer	400 g broccoli	12 months	20	Changed signaling pathways associated with inflammation and carcinogenesis in the prostate
Visvanathan., 2018 <sup>111</sup>	Breast cancer	100 $\mu$ mol once a day	14 days	34	The mean proliferative rate was changed
Atwell et al, 2015 <sup>112</sup>	Breast cancer	2 pills of glucoraphanin thrice a day	2 - 8 weeks	54	No changes in breast tissue tumor biomarkers were observed
Tahata et al, 2018 <sup>113</sup>	Melanoma	100 or 200 $\mu$ mol once daily	28 days	17	Proinflammatory cytokines were decreased and tumor suppressor decorin was increased
Lozanovski et al, 2020 <sup>114</sup>	Pancreatic cancer	90 mg sulforaphane + 180 mg glucoraphanin per day	1 year	40	No statistically significant results were observed



**Figure 4.** (A) Genistein, (B) Modulation of cell signaling pathways by isoflavones.

like effects in modulating the Nrf2 signaling pathway, which improved atherosclerosis and oxidative stress in *in vivo* and *in vitro* experiments.<sup>122</sup> Isoflavones also prevent diabetes by altering several cellular pathways crucial for maintaining glucose homeostasis. They help prevent type 2 diabetes by modulating nuclear receptor activity and non-receptor signaling on the cells necessary to maintain glucose homeostasis.<sup>123</sup> Since isoflavones have a similar chemical structure to endogenous estrogens, they interact with intracellular estrogen receptors, reducing the accumulation of lipids and the distribution of adipose tissue, thereby helping reduce obesity. They inhibit adipogenesis and lipogenesis by interacting with various transcription factors and upstream signaling molecules, aiding in maintaining metabolism and balance to treat obesity.<sup>124</sup>

Isoflavones show tumor suppression ability by activating the Nrf2 protein. Genistein is observed to bind readily with ER $\beta$  receptors and thwart oncogenic progression in breast cancer by triggering apoptosis and restraining angiogenesis. The G0/G1 arrest of tumor cells is achieved by blocking NF-kB and AKT, which are crucial for upregulating the expression

of cyclin D. Tumor cells undergo a DNA repair process in this quiescent stage and die off by apoptosis if the damage is irreversible. A three-way obstruction of PI3K, AKT, and STAT3 helps impede tumor proliferation. Isoflavones downregulate FAK, which is necessary to form focal adhesions between tumor cells and ECM required for migration, thereby containing the invasion. Furthermore, isoflavones modulate the JAK/STAT pathway by hindering the phosphorylation of STAT1 and STAT3, which reduces the production of inflammatory mediators like TNF- $\alpha$  and COX-2 (Figure 4B).<sup>125-127</sup>

Soy isoflavones, like genistein, exhibit notable anti-cancer properties against breast cancer,<sup>119</sup> prostate cancer,<sup>125</sup> cervical cancer,<sup>128</sup> ovarian cancer,<sup>129</sup> lung cancer,<sup>130</sup> and renal cancer.<sup>131</sup> This chemopreventive nature of genistein is associated with its anti-inflammatory,<sup>118</sup> anti-oxidant,<sup>132</sup> anti-angiogenic,<sup>133</sup> anti-metastatic,<sup>134</sup> and anti-proliferative properties.<sup>135</sup> Isoflavones are observed to suppress prostate cancer cell growth by disrupting the expression of 2 copper transporter genes, CTR1 and ATP7A. Since copper levels are increased drastically in cancers, isoflavones cause pro-oxidant

signaling by targeting endogenous copper, leading to ROS-mediated cell death.<sup>136</sup> After 50  $\mu$ M genistein treatment, superoxide anion is generated, which quickly converts into hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) before forming hydroxyl (HO-) through the oxidation of reduced copper. This accumulation of ROS induces irreversible DNA damage, causing cell death in breast cancer cells.<sup>137</sup>

Genistein downregulates cytokine-induced signal transduction events in the immune cells to work as an anti-inflammatory agent.<sup>118</sup> The anti-inflammatory effects of isoflavones are further demonstrated by a human trial where markers of inflammation (IL-18 and C-reactive protein) were decreased, and plasma nitric oxide levels were increased when postmenopausal women with metabolic syndrome were fed a soy nut diet (340 mg isoflavones/100 g soy nut) for 8 weeks.<sup>138</sup> Another study illustrated the anti-oxidative properties of isoflavones by observing a significant decrease in thiobarbituric acid reactive substances (TBARS) in plasma, liver, and brain by 33%, 18%, and 12%, respectively, after treatment with 2.5 mg/kg body weight isoflavones for 5 weeks.<sup>139</sup> The anti-angiogenic activity of genistein is caused by inducing apoptosis in VEGF-loaded endothelial cells, attributed to inhibition of MMP-2,-9 production and activity. Additionally, the study showed that genistein exposure decreased the activation of JNK and p38 induced by VEGF.<sup>133</sup> A similar study observed genistein's anti-angiogenic and anti-metastatic effects by inhibiting c-erbB-2, MMP-2, and MMP-9 in breast carcinoma.<sup>140</sup>

Isoflavones demonstrate chemoprevention by modulating various signaling pathways responsible for causing full-blown cancer. These pathways may be related to apoptosis, cell proliferation, inflammation, and hormone regulation. The activation of both the nuclear transcription factor NF- $\kappa$ B and Akt signaling pathway is inhibited by genistein, which is known to regulate the balance between cell survival and apoptosis.<sup>140</sup> A study by Xu et al noted the promotion of apoptosis in genistein-treated lung cancer A549 cells by modulating the IMPDH2/AKT1 pathway.<sup>130</sup> Genistein is also shown to arrest the MCF-7 and MDA-MB-231 breast cancer cells in the G<sub>0</sub>/G<sub>1</sub> phase, thereby inducing an apoptotic pathway.<sup>134</sup> It has been demonstrated that isoflavones bind to ER $\alpha$  and ER $\beta$ , with a higher affinity towards ER $\beta$  receptors, to exert various anti-estrogenic, anti-apoptotic, and anti-inflammatory properties.<sup>126</sup> Genistein alters the PI3K/Akt pathway to induce differentiation in breast cancer stem cells by interactions with ER + cells.<sup>141</sup> The abundance of such evidence provides ample ground for further investigation of the chemopreventive abilities of this phytochemical.

### *Immunomodulatory Effects of Isoflavones*

While isoflavones have a variety of physiological actions, the majority of them suppress NF- $\kappa$ B pathways and limit the

generation of pro-inflammatory cytokines such as IL-6, TNF- $\alpha$ , IL-1 $\beta$ , and IFN- $\gamma$ .<sup>142</sup> One of the most physiologically active isoflavones in soy, genistein, inhibits the growth of cancer cells by p-JNK, SGK1, Wnt, Akt, and Shh signaling pathways. According to a study by Jian Qin et al, the HCT-116 cell group treated with genistein at 50 and 100  $\mu$ M showed a substantial decrease in Akt and SGK1 mRNA expression compared to normal cells.<sup>143</sup> Similar suppression of the p-MEK, p-ERK, p-JNK, and Shh pathways was observed in independent investigations by Qi Zhang and K. Li.<sup>144,145</sup> Another frequent soy component, daidzein, enhances the BMP-2/Smads pathway while downregulating the expression of the STAT3 and STAT5 signaling pathways.<sup>146,147</sup> Additionally, when isoflavones were applied to malignant cells, the CD4+/CD8 + ratio was considerably reduced.<sup>148</sup> Genistein is also being studied as an adjuvant in combination therapy with other drugs. One such study conducted by Liu et al to observe the effect of a genistein and cisplatin combination on CaSki human cervical cancer cells showed enhanced anticancer effects of cisplatin in CaSki cells. It was noted that the p-ERK1/2 and Bcl2 expression levels were decreased by 37% and 69%, respectively, and cleaved caspase-3 and p53 expression levels were increased by 115% and 304%, respectively, in the cisplatin plus genistein group as compared to that in cisplatin group.<sup>128</sup> Isoflavones' significant immunomodulatory properties make them a promising candidate for cancer therapy.

### *Isoflavones Human Studies*

Given that isoflavones act on hormone receptors, it would make sense to test the role of isoflavones in the chemoprevention of hormonal cancers such as breast cancer and prostate cancer. Several clinical trials (7 cohort studies and 17 case-control studies) have been conducted to test the chemopreventive role of isoflavones against breast cancer. A systematic review was conducted to look at dose-dependent chemoprevention (Table 4A). This study noted a statistically significant protective effect of isoflavone intake on breast cancer in the case-control studies, while no such effect was seen in the cohort studies. It also showed that below the intake of 10 mg/day, isoflavones did not affect breast cancer risk. In different statistical models, dose-dependent reduction in breast cancer risk was found to be dependent on isoflavone intake.<sup>149</sup> Another systematic review analyzed the role of isoflavones in prostate cancer treatment and chemoprevention in high-risk individuals. (Table 4B) Eight randomized control trials were identified. Six recruited men were diagnosed with prostate cancer, while 2 recruited men were at high risk of prostate cancer. No significant difference in the prostate serum antigen level and sex steroid endpoints was observed between the treatment and non-treatment groups. Two studies that tested the chemopreventive role of isoflavone against prostate

**Table 4A.** Analyses of Isoflavone Intake and the Risk of Breast Cancer.

Study Design	No. of Studies	OR (95%CI)	p heterogeneity	I <sup>2</sup> (%)	p for Interaction
Case-control	17	0.62 (0.50, 0.76)	0.000	83.8	0.000
Cohort	7	0.94 (0.86, 1.02)	0.178	32.7	
Isoflavones highest intake < 10 mg/d	6	1.01 (0.94, 1.08)	0.452	0.0	0.000
Isoflavones highest intake ≥ 10 mg/d	18	0.63 (0.53, 0.75)	0.000	81.4	

**Table 4B.** Isoflavones Clinical Studies Against Prostate Cancer.

Author and Year	Type of Cancer	Dose of Isoflavones	Duration of Study	Number of Patients	Results
White et al, 2010 <sup>151</sup>	Prostate cancer	450 mg genistein + 300 mg daidzein daily	6 - 12 months	53	PSA levels were not lowered
Kumar et al, 2007 <sup>152</sup>	Prostate cancer	80 mg per day	12 weeks	50	Modulation of serum steroid hormone levels was not observed
Kumar et al, 2010 <sup>153</sup>	Prostate cancer	40 mg, 60 mg and 80 mg per day	30 ± 3 days	44	Changes in serum sex hormone-binding globulin, PSA, and percentage of tissue Ki-67 were not statistically significant
Lazarevic et al, 2011 <sup>154</sup>	Prostate cancer	30 mg synthetic genistein per day	3 to 6 weeks	40	The level of serum PSA was reduced
Miyanaaga et al, 2012 <sup>155</sup>	Men with rising prostate-specific antigen	60 mg per day	12 months	153	No difference in PSA levels was observed. The incidence of cancer was lower in the isoflavone groups as compared to the placebo group
Dalais et al, 2004 <sup>156</sup>	Prostate cancer	117 mg per day	22 - 27 days	28	PSA levels and free/total PSA levels reduced
Hamilton-Reeves et al, 2007 <sup>157</sup>	Men with a high risk of prostate cancer or with low-grade prostate cancer	107 mg/day or <6 mg/day or 0 mg/day	6 months	58(at 3 months)/55	AR expression in the prostate was suppressed
Hamilton-Reeves et al, 2007 <sup>158</sup>	Men with a high risk of prostate cancer or with low-grade prostate cancer	107 mg/day or <6 mg/day or 0 mg/day	6 months	58(at 3 months)/55	Prostate tissue biomarkers were not altered but less prostate cancer is detected after 6 months of isoflavone consumption irrespective of dose
Kumar et al, 2004 <sup>159</sup>	Prostate cancer	40 mg, 60 mg and 80 mg per day	12	59	Serum PSA and free testosterone were altered

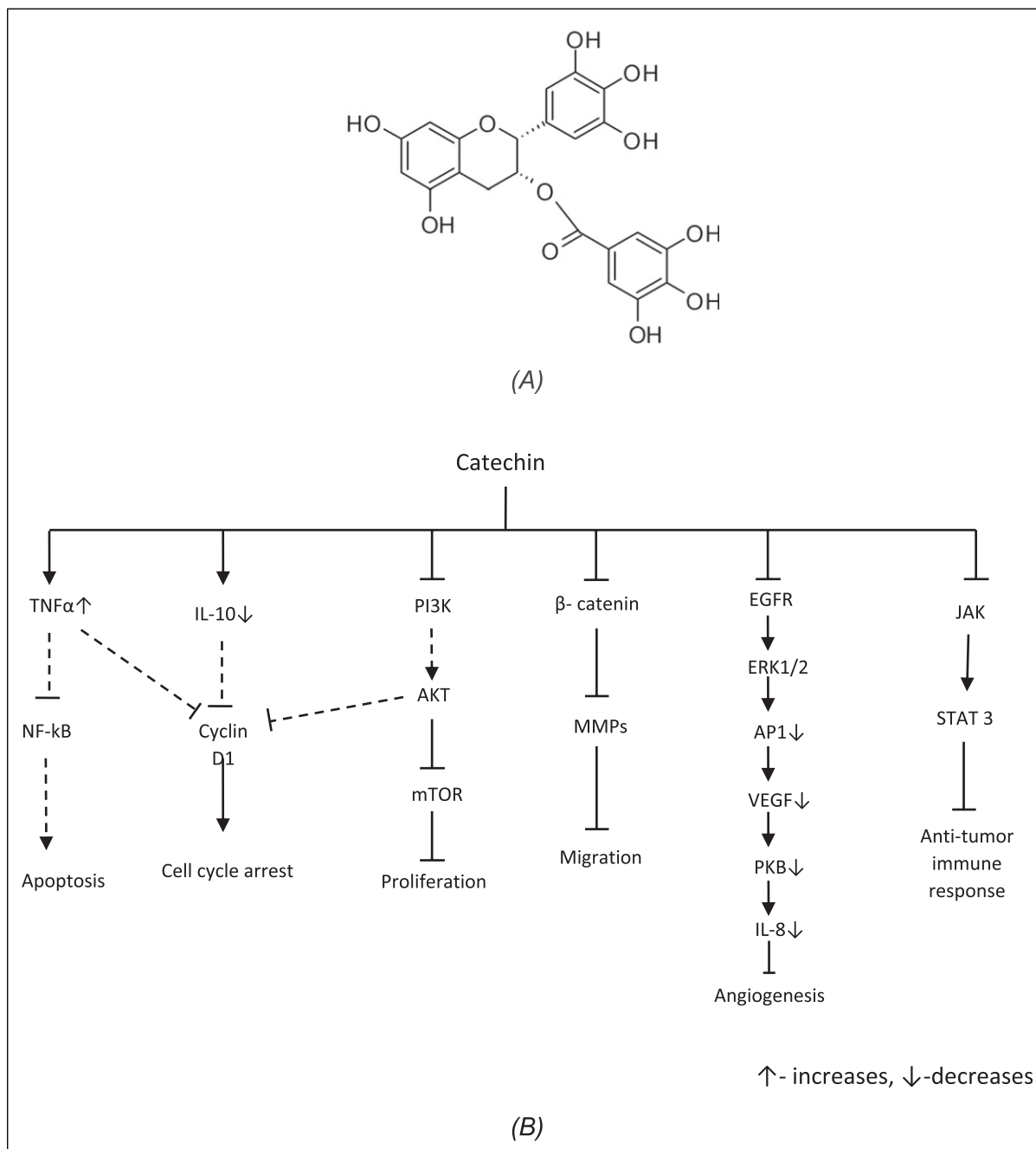
cancer found a positive role of the phytochemical in cancer prevention.<sup>150</sup>

## Catechins

Catechins are naturally occurring powerful antioxidants found primarily in green tea, cocoa, and berries. Classified as flavonols, catechins include compounds like epicatechin(EC), epigallocatechin(EGC), and epigallocatechin gallate(EGCG) (Figure 5). Their concentration in a typical brewed green tea beverage (250 mL) may vary from 50-100 mg depending upon preparation methods.<sup>160</sup> Catechins are found to exert beneficial effects in neurodegenerative diseases, CVD, cancer, and diabetes.<sup>161</sup> They enhance nitric oxide production, reduce LDL cholesterol levels, and inhibit platelet aggregation to benefit vascular endothelium.<sup>162</sup> It is

observed that catechins can prevent diabetes by alleviating ER stress and promoting anti-inflammatory pathways.<sup>163</sup> Similarly, Brown adipose tissue (BAT) thermogenesis is noted in mice, suggesting catechin's anti-obesity properties.<sup>164</sup> Additionally, catechins exert significant anti-inflammatory properties to prevent inflammatory bowel disease by regulating the activation and deactivation of inflammation-related oxidative stress-related cell signaling pathways such as NF-kB, MAPKs, and STAT1/3.<sup>165</sup> One of the most potent anti-cancer and anti-inflammatory catechins is EGCG.<sup>166</sup> EGCG is observed to be efficient in preventing prostate cancer, breast cancer,<sup>167</sup> lung cancer,<sup>168</sup> gastric cancer,<sup>169</sup> lymphomas,<sup>170</sup> and leukemia.<sup>171</sup>

Catechin affects anti-inflammatory signals to promote cell cycle arrest and apoptosis. It increases TNF- $\alpha$ , which decreases NF-kB activity, causing apoptosis. Another



**Figure 5.** (A) Epigallocatechin gallate, (B) Modulation of cell signaling pathways by catechins.

anti-inflammatory signal, IL-10 is lessened, thereby down-regulating the cyclin D1, leading to cell cycle arrest. TNF- $\alpha$  increment also has the same effect on cyclin D1. Catechin inhibits the PI3K/AKT pathway, causing cell cycle arrest and reduced proliferation by decreasing cyclin D1 and blocking mTOR. It modulates the Wnt signaling pathway by degrading  $\beta$ -catenin, halting the transcription of MMPs, and obstructing the migration of tumor cells. One major mechanism by which catechin curbs tumor angiogenesis is blocking the EGFR, which downregulates a series of proteins, namely,

ERK1/2, AP1, VEGF, PKB, and IL-8, halting angiogenesis. Moreover, catechin modulates the JAK/STAT pathway by limiting STAT3 phosphorylation, causing a rise in antitumor immune response (Figure 5B).<sup>172,173</sup>

Catechins work as both pro- and antioxidants to prevent cancer. In low doses, they act as antioxidants, scavenging the DNA-damaging ROS. They also chelate transition metals to prevent them from catalyzing oxidation reactions. Alternatively, catechins function as pro-oxidants at higher concentrations, triggering apoptosis and ferroptosis.<sup>174,175</sup> Many

articles note that EGCG's anticancer abilities are due to its pro-oxidant nature, which promotes the Fenton reaction that generates radicals. EGCG elevates ROS levels to induce cell cycle arrest and apoptosis. It induces mitochondrial damage, ROS level elevation, DNA damage, and JNK activation to induce apoptosis in pancreatic cells,<sup>176</sup> glioblastoma cells,<sup>177</sup> and lung cancer.<sup>178</sup>

Numerous studies have suggested that EGCG exerts significant efficacy against various forms of cancer by the mechanism of apoptosis and inhibition of proliferation and metastasis. EGCG alters both the caspase-dependent (intrinsic) pathway and death receptor (extrinsic) pathway by modulation of Bcl-2 family proteins<sup>170</sup> and downregulation of Mcl-1 and XIAP<sup>171</sup> to induce programmed cell death. In addition, EGCG also achieves apoptosis by the inhibition of TNF- $\alpha$  activity.<sup>179</sup> The key antitumor mechanism of EGCG is attributed to its ability to suppress metalloproteinase activity. Many studies claim matrix metalloproteinases aid in tumor progression by allowing tumor invasion and metastasis.<sup>180</sup> A study on hepatocellular carcinoma cells (HCCLM6) demonstrates that the antimetastatic activity of the EGCG is due to its ability to inhibit MMP-2 and MMP-9 activity.<sup>181</sup> EGCG may either directly bind to the MMP-2 and MMP-9 to inhibit their activities<sup>182</sup> or alter some signaling pathways, including NF- $\kappa$ B,<sup>183</sup> MAPK/ERK,<sup>184</sup> and PI3K/Akt<sup>185</sup> to inhibit MMP expression. Similar mechanisms involving the modulation of several signaling pathways are responsible for EGCG's antiproliferative abilities. One study puts forward the idea that EGCG lowers the expression of phosphorylated Akt (p-Akt) and phosphorylated mTOR (p-mTOR) through PTEN to modulate the PI3K/Akt/mTOR pathway, resulting in the decreased proliferation and apoptosis of pancreatic cancer cells linked with the expression of PTEN.<sup>186</sup> Another study demonstrates that EGCG subdues the proliferation of gastric cancer cells by silencing the wnt/ $\beta$ -catenin signaling pathway.<sup>169</sup> A similar study suggests that EGCG can reduce the proliferation and invasiveness of breast tumors by blocking the Wnt pathway with the help of the HBPI gene. The downregulation of the Wnt pathway increases the expression of G1 regulators, c-MYC, and cyclin D1 genes, causing a reduction in invasive and migratory properties of the tumor.<sup>187</sup>

### *Immunomodulatory Effects of Catechins*

Catechins exert strong immunomodulatory effects on cancer cells. It is observed that the proinflammatory cytokines generated by malignant cells, such as IL-1 $\beta$ , IL-6, and TNF- $\alpha$ , dramatically decrease after being treated with catechins.<sup>188</sup> Catechins can also inhibit the NF- $\kappa$ B pathway to reduce the amount of proinflammatory cytokines. A study by Mackenzie et al found that NF- $\kappa$ B binding activity was suppressed dose-dependently in Jurkat cells pretreated with catechins, with the lowest binding occurring at 17.2  $\mu$ M concentration of catechin.<sup>189</sup> Rawangkan et al suggested use of green tea catechin as an immune checkpoint inhibitor allowing cancer prevention

and treatment based on the findings that EGCG partially restores T cell activity by inhibition of PD-L1/PD-1 signaling.<sup>190</sup> Catechins have shown potential to be used in combination therapy with other chemotherapeutic drugs for immunotherapy. A case in point is provided by the observation that catechins limit toxicity induced by a chemotherapeutic drug, irinotecan, by reducing its side effects, such as diarrhea, neutropenia, leucopenia, and non-alcoholic fatty liver disease. The same study also noted that it works as an effective anticancer agent as it mitigates the histopathology of colon adenocarcinoma.<sup>191</sup>

### *Catechin Human Studies*

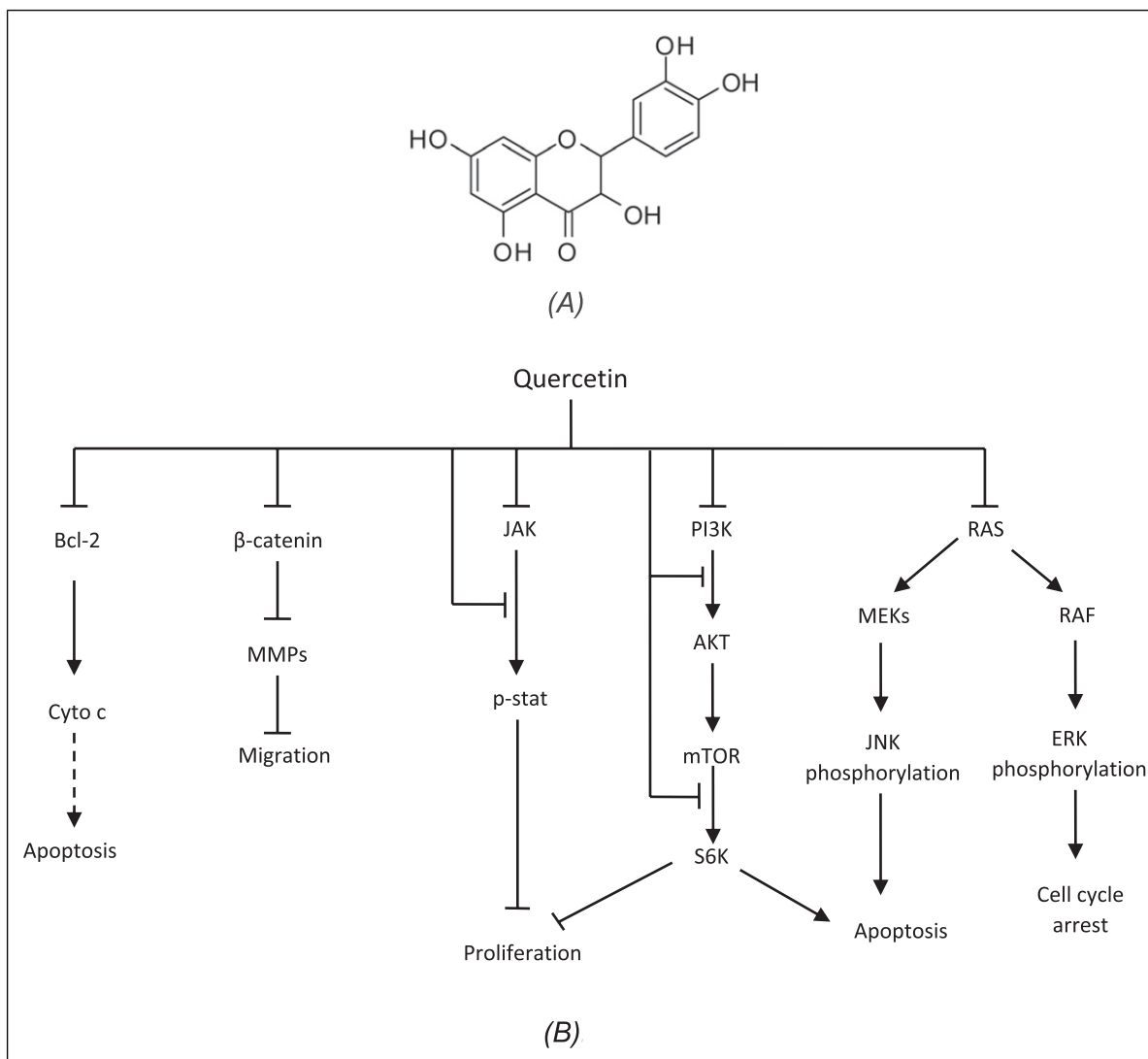
Several clinical trials have been conducted that highlight the chemopreventive effects of catechins. One such study in Shanghai, China, established a clear interrelationship between green tea consumption and colorectal cancer risk. The study demonstrated that each additional 2 g of dry green tea leaves consumed daily correlated with reduced risk, with a confidence interval of 0.78-0.99 and a *P*-value of .03.<sup>192</sup> A systematic review also showed a positive effect of green tea consumption on prostate cancer chemoprevention.<sup>193</sup> (Table 5) Currently, a phase II clinical trial is being conducted to evaluate the bioavailability, safety, and efficacy of green tea catechins in men with low to intermediate-grade prostate cancers. In this study, 3 capsules of Sunphenon will be administered twice a day containing a total of 405 mg green tea to the participants for 24 months.<sup>194</sup> It is on the foundation of this and plenty of other studies that the possibilities of comprehensive research on the chemopreventive characteristics of this phytochemical thrive.

### *Quercetin*

Quercetin is a plant flavonol belonging to the flavonoid group of polyphenols (Figure 6). It is found in many fruits, vegetables, and grains including capers, red onions, kale, and shallots. It has anti-oxidant and anti-inflammatory potential, showing great promise in treating and containing chronic health conditions, including CVDs and cancers.<sup>198</sup> Constant administration of quercetin decreases vascular smooth muscle cells (VSMC) and hampers the progression of CVDs. It also reduces NADPH oxidase, ROS, superoxide anion, and free radicals to exert anti-inflammatory and anti-hypertensive properties.<sup>199</sup> At the same time, it enhances oral glucose tolerance and pancreatic  $\beta$ -cell function while suppressing the activity of  $\alpha$ -glucosidase and DPP-IV enzymes to impart protective effects against type-2 diabetes mellitus.<sup>200</sup> The anti-inflammatory and hypoglycemic nature of quercetin helps prevent obesity by decreasing fat deposition.<sup>201</sup> Quercetin also demonstrates potential chemopreventive properties against breast cancer,<sup>202</sup> liver cancer,<sup>203</sup> colon cancer,<sup>204</sup> cervical cancer,<sup>205</sup> blood cancer, prostate cancer, and lung cancer.<sup>206</sup>

**Table 5.** Catechins Clinical Studies Against Prostate Cancer.

Author and Year	Dose of Catechins	Duration of Study	Number of Patients	Results
Kumar et al, 2015 <sup>195</sup>	400 mg EGCG/day	1 year	97	No reduction in the likelihood of prostate cancer in men with baseline HGPIN or ASAP was observed
Bettuzzi et al, 2006 <sup>196</sup>	600 mg green tea catechins/day	1 year	60	Levels of PSA were reduced, and lower urinary tract symptoms were also reduced
Brausi et al, 2008 <sup>197</sup>	600 mg green tea catechins/day	30 months	22	A reduction of almost 80% was observed in prostate cancer diagnosis

**Figure 6.** (A) Quercetin, (B) Modulation of cell signaling pathways by quercetin.

Quercetin triggers apoptotic pathways through multiple routes to safeguard against cancer. It can phosphorylate p53 or inhibit Bcl-2 to release cytochrome c and induce apoptosis through a series of steps. Quercetin conducts cell cycle arrest and apoptosis by phosphorylating ERK and JNK in the MAPK pathway. Disrupting the PI3K/AKT

pathway by phosphorylating PI3K, AKT, and S6K contributes to its antiproliferative and apoptotic effects. It modulates the Wnt/ $\beta$ -catenin pathway by obstructing the nuclei's  $\beta$ -catenin translocation and hampers the production of MMPs necessary for tumor cell migration. Quercetin also impedes the JAK/STAT pathway by halting

the formation of p-STAT to promote its antiproliferative nature (Figure 6B).<sup>206,207</sup>

The combination of antioxidant, anti-inflammatory, apoptotic, antiproliferative, anti-angiogenic, and anti-metastatic effects of quercetin collectively endows it with chemopreventive qualities. Quercetin boasts antioxidant quality, thanks to its ability to hunt down and scavenge ROS such as peroxynitrite and hydroxyl radicals.<sup>208</sup> Rac1 is a GTPase that promotes cell migration and invasion by the production of ROS. Quercetin is found to target Rac1 with high affinity to make a stable complex and modulate the Rac1-p66Shc pathway to control ROS generation within the cells.<sup>209</sup> Likewise, its anti-inflammatory ability allows it to inhibit inflammatory enzymes and mediators to help prevent cancer. In vitro studies have shown reduced levels of inflammatory mediators, such as NO-synthase, COX-2, and CRP, in human hepatocyte-derived Chang liver cell lines after treatment with quercetin. The same study observed that quercetin blocks NF- $\kappa$ B activation resulting in the downregulation of pro-inflammatory genes.<sup>210</sup> Furthermore, another in vitro study discovered that quercetin lessens the levels of other inflammatory mediators like TNF- $\alpha$ , IL-6, and IL-1 $\beta$  in RAW264.7 macrophages by inhibiting the PI3K/Akt signaling pathway.<sup>211</sup> The apoptotic nature of quercetin is due to its ability to arrest the cell cycle, activate caspase proteases, and modify signaling pathways. Quercetin causes a G2 phase arrest in HPV-positive human cervical cancer-derived cells triggering apoptosis.<sup>212</sup> In human T-cell acute lymphoblastic leukemia Jurkat clones (J/Neo cell lines), quercetin is found to activate caspase-3 and caspase-9 to induce apoptosis in a dose-dependent manner.<sup>213</sup>

Numerous studies have emphasized quercetin's apoptotic potential through its ability to modulate signaling pathways like NF- $\kappa$ B/I $\kappa$ B, p38 MAPK, Bcl-2/Bax,<sup>214</sup> PI3K/Akt/mTOR, Wnt/ $\beta$ -catenin,<sup>215</sup> and FOXO3a.<sup>216</sup> Inhibition of angiogenesis by quercetin treatment was achieved by targeting the VEGFR-2 mediated angiogenesis pathway, suppressing the expression of the downstream regulatory factor AKT, and restraining tumor growth.<sup>217,218</sup> Another study demonstrated quercetin's anti-invasive and anti-metastatic effects on lung cancer cells by inhibiting the Snail-dependent Akt activation pathway. This study further observed that the expression of N-cadherin, vimentin, ADAM9, and MMPs-related proteins were notably downregulated and the expression of E-cadherin was significantly increased after quercetin treatment, which helped contain the invasiveness of the lung cancer cells.<sup>219</sup> Similarly, in a different study, the administration of 100  $\mu$ M quercetin in CD44<sup>+</sup>/CD24<sup>-</sup> CSCs resulted in the restriction of proliferation of both cells to a great extent.<sup>202</sup>

### Immunomodulatory Effects of Quercetin

Quercetin functions as an immunomodulator for malignant cells. It acts as an anti-inflammatory agent by decreasing the levels of pro-inflammatory cytokines such as TNF- $\alpha$ , IL- $\beta$ ,

and IL-6 in a dose- and time-dependent manner.<sup>220</sup> Quercetin administration causes rapid NK cell proliferation as shown by a study where 100  $\mu$ g/mL of quercetin administration dramatically enhanced the proliferation of the NK-92 cell line.<sup>221</sup> Additionally, quercetin promotes the susceptibility of cancer cells to NK cell-mediated killing by inducing natural killer group 2D (NKG2D) ligands and suppressing heat shock protein (HSP70).<sup>222</sup> However, the low bioavailability of quercetin makes it challenging to use it alone in clinical settings which highlights the need for nano-formulation developments. It was observed, in a study, that nanoparticle-delivered quercetin has enhanced antitumor potential compared to quercetin alone.<sup>223</sup> Quercetin, when combined with conventional cancer therapies, acts as a sensitizer and protects non-cancer cells from the side effects of chemotherapy and radiotherapy.<sup>224</sup> Zhang et al best summarized the use of quercetin in immunotherapy where they introduced a codelivery of nano-formulated quercetin and alantolactone, and observed the promotion of antitumor immune response through synergistic immunogenic cell death in microsatellite-stable colorectal cancer.<sup>225</sup>

### Quercetin Human Studies

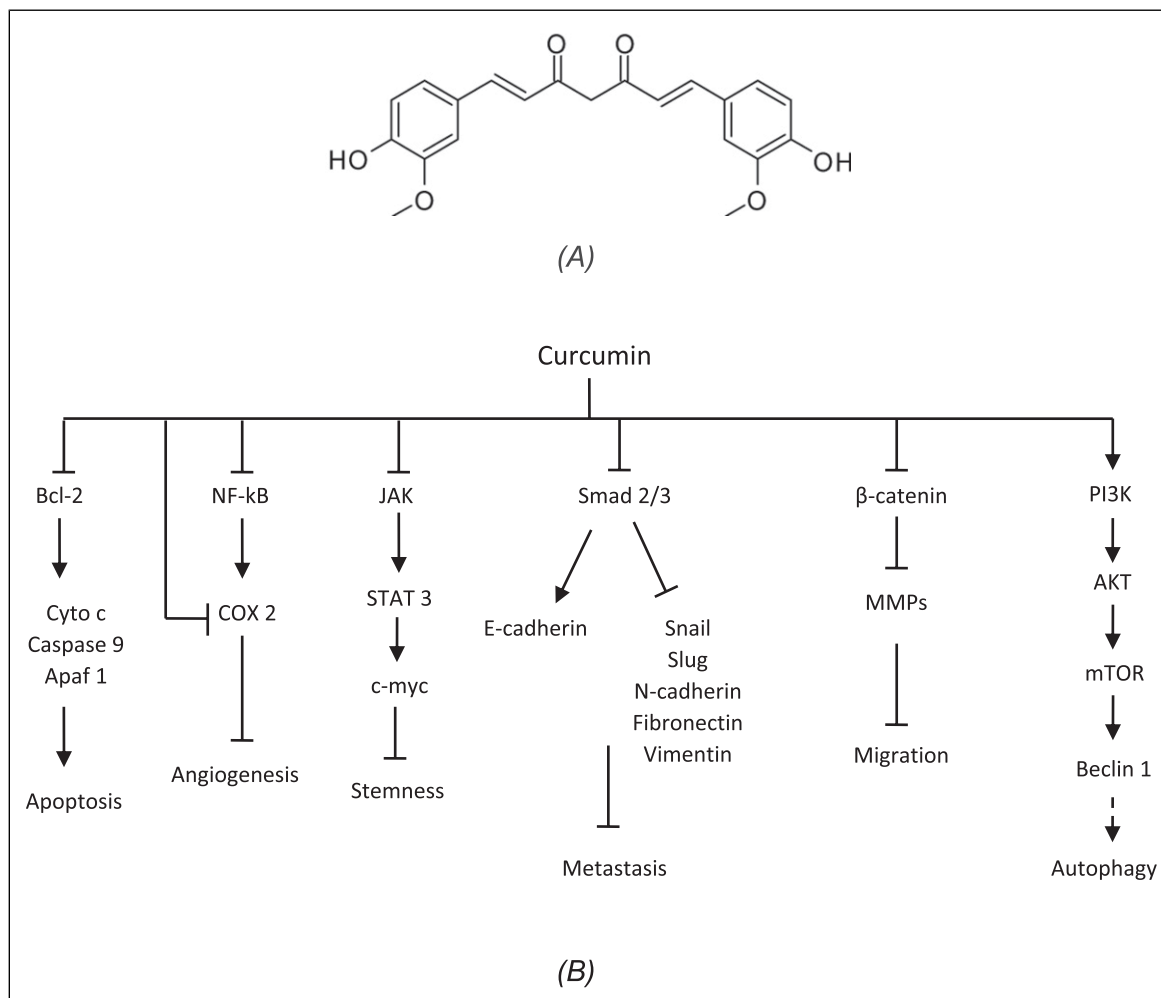
Clinical trial data on quercetin against cancer is not abundant. A clinical trial, involving 433 men with primary prostate cancer and 538 population-based controls, to study the effects of quercetin on prostate cancer revealed the potential chemopreventive attributes of quercetin. The results of the study showed a 27% reduction in the risk of prostate cancer from an intake of at least 24  $\mu$ g of quercetin.<sup>226</sup> This clinical trial outcome hinted at promising chemotherapeutic and chemopreventive prospects of quercetin however, further clinical trials are required to confirm the results.

### Curcumin

Curcumin is a bright yellow-colored polyphenol belonging to a group of natural compounds called curcuminoids (Figure 7). It is the primary bioactive compound in turmeric, with a concentration reaching up to 31.4 mg/g in pure turmeric powder.<sup>227</sup> Curcumin exhibits many therapeutic qualities including antioxidant,<sup>228</sup> anti-inflammatory,<sup>229</sup> chemopreventive, and immunity-enhancing properties.<sup>230</sup> A lower occurrence of cancer is associated with a dietary intake of curcumin.<sup>231</sup> The risk and incidence of various forms of cancer such as breast cancer,<sup>232</sup> lung cancer,<sup>233</sup> prostate cancer,<sup>234</sup> brain cancer,<sup>235</sup> pancreatic cancer,<sup>236</sup> and endometrial cancer,<sup>237</sup> is reduced by curcumin.

Curcumin inhibits Bcl-2 and releases cytochrome c, Caspase 9, and Apaf 1 to induce apoptosis. It can suppress angiogenesis by inhibiting the NF- $\kappa$ B pathway and inflammatory mediators like COX-2. It modulates the JAK/STAT pathway by obstructing JAK to restrict the stemness of the tumor. Curcumin enhances E-cadherin levels while





**Figure 7.** (A) Curcumin, (B) Modulation of cell signaling pathways by curcumin.

reducing the levels of N-cadherin, vimentin, fibronectin, snail, and slug through the repression of the TGF- $\beta$ /Smad2/3 pathway to check metastasis. It modulates the Wnt signaling pathway by degrading  $\beta$ -catenin, halting the transcription of MMPs, and obstructing the migration of tumor cells. Moreover, curcumin can alter PI3K/AKT/mTOR pathway to increase the production of key proteins (e.g. Beclin 1) involved in autophagy (Figure 7B).<sup>238,239</sup>

The antioxidant potential of curcumin comes from its ability to scavenge free radicals such as ROS and RNS,<sup>240</sup> and modulate the activity of GSH, catalase, and SOD enzymes necessary for the neutralization of free radicals.<sup>241</sup> Working as an anti-inflammatory agent, curcumin reduces the expression levels of pro-inflammatory cytokines: TNF- $\alpha$ , IL-6, and IL-1 $\beta$  by inhibiting the NF-kB signaling pathway.<sup>242</sup> The study further concluded that curcumin inhibits the TGF- $\beta$ 1/Smads signaling pathway by reducing mRNA expression levels of fibrotic factors  $\alpha$ -SMA, Smad2/3, and TGF- $\beta$  to exert anti-inflammatory effects. Scientific research has established curcumin as an apoptotic, anti-angiogenic, anti-proliferative,

and anti-metastatic phytochemical. Curcumin achieves apoptosis via various mechanisms, including ROS-mediated pathway,<sup>243</sup> downregulation of apoptosis suppressor proteins,<sup>234</sup> and alteration in signaling pathways such as induction of FOXO1 and inhibition of the PI3K/Akt pathway.<sup>236</sup> Meanwhile, an in vivo study claims curcumin suppresses angiogenesis by downregulating VEGF, CD31, and  $\alpha$ SMC expression levels.<sup>244</sup> Another similar study using human ovarian cancer cell lines SKOV3ip1, HeyA8, and HeyA8-MDR in athymic mice revealed that oral intake of 500 mg curcumin per kg body weight was the ideal dose to suppress NF-kB and signal transducers and activators of transcription 3 (STAT3) activation and to mitigate angiogenic cytokine expression.<sup>245</sup> Curcumin modulates cell cycle and signaling pathways to subdue proliferation and perform apoptosis of the tumor cells. G2 phase accumulation of head and neck cancer SCC-9 cells was achieved by downregulating the PI3K/Akt/mTOR pathway that arrested the cell cycle on G2/M transition.<sup>246</sup> In addition, similar effects of curcumin in the modulation of signaling pathways were

seen in an experiment involving non-small cell lung cancer cells (NSCLC) where tumor proliferation was inhibited and apoptosis was regulated by the upregulation of microRNA-192-5p (miR-192-5p) and the downregulation of the PI3K/Akt signaling pathway.<sup>233</sup> It is a well-known fact that epithelial-mesenchymal transition is involved in cancer progression and metastasis, and curcumin acts as an anti-metastatic substance by blocking the same. Curcumin achieves this by altering several expression pathways, including inhibiting c-Met expression,<sup>247</sup> upregulating NKD2, which downregulates the Wnt signaling pathway,<sup>248</sup> and phosphorylating Smad2/3, which suppresses the TGF- $\beta$ 2 signaling pathway.<sup>249</sup> Furthermore, curcumin was shown to decrease N-cadherin, twist, snail, and vimentin, while increasing E-cadherin in colorectal cancer SW480 cells, suggesting that it could suppress the EMT process by inhibiting CDX2/Wnt3a/ $\beta$ -catenin pathway.<sup>250</sup>

### Immunomodulatory Effects of Curcumin

Curcumin has been investigated frequently as an immunomodulator against malignant cells. Curcumin downregulates pro-inflammatory cytokines such as IL-1 and TNF- $\alpha$ <sup>251</sup> whose activation mostly depends on the NF- $\kappa$ B pathway.<sup>252</sup> Curcumin inhibits IKK to downregulate the NF- $\kappa$ B pathway. According to a study by Shih et al curcumin treatment of CT26 cell lines resulted in a dose-dependent decrease in the expression of PD-L1 and NF- $\kappa$ B.<sup>253</sup> Additionally, curcumin can repolarize cancer-associated macrophages into a tumoricidal form and can provoke NK cells to target tumor cells and cancer stem cells indirectly.<sup>254</sup> Challenges similar to those of other phytochemicals, like poor solubility and bioavailability, have made clinical application of curcumin a daunting task. Curcumin is currently being studied as a chemo-sensitizing agent and shows potential to be used as an adjuvant to cancer immunotherapy by enhancing the immune system's ability to combat tumors.<sup>255</sup> Farghadani et al highlight the chemo-sensitizing ability of curcumin in a study where they noted that curcumin enhanced the anticancer efficacy of chemotherapy drugs like doxorubicin, paclitaxel, 5-fluorouracil, and cisplatin in breast cancer.<sup>256</sup>

### Curcumin Human Studies

A systematic review of RCTs designed to study the anticancer effects of curcumin in the last 5 years concluded that while curcumin has positive results in terms of cancer research, it is still not an out-and-out therapeutic agent that can be used alone to treat cancer.<sup>257</sup> (Table 6) However, a favorable safety profile of curcumin has been established that has opened possibilities for more sophisticated and continued study on the potential anti-tumor abilities of the compound.

### Luteolin

Luteolin is a flavone, belonging to a group of plant compounds called flavonoids (Figure 8). It is a yellow-colored compound

found primarily in celery, thyme, parsley, chamomile tea, broccoli, carrots, and several other foods. This polyphenol protects plants from UV radiation, fluctuating temperatures, insects, and microorganisms.<sup>265</sup> Luteolin exhibits several pharmacological properties, including antioxidant, anti-inflammatory,<sup>266</sup> neuroprotective,<sup>267</sup> and analgesic effects.<sup>268</sup> Numerous studies demonstrate the health benefits of luteolin in protection against Alzheimer's disease,<sup>269</sup> CVDs,<sup>270</sup> and various cancers. Luteolin's anticancer effects have been observed across a variety of cancers, ranging from breast cancer to colon cancer<sup>271</sup> and lung cancer<sup>272</sup> to gastric cancer.<sup>273</sup>

Luteolin can trigger apoptosis by regulating both extrinsic and intrinsic pathways. It modulates the extrinsic apoptotic pathway by promoting the expression of death receptors (Fas, DR4, DR5), which induce apoptosis by caspase cascade induction. It can regulate the intrinsic apoptotic pathway by inhibiting antiapoptotic proteins such as Bcl-2. The inhibition of Bcl-2 is also linked to the activation of p53 as a result of DNA damage by ROS. Luteolin suppresses Ras, Raf, and MEK1/2, thereby impeding the expression of ERK1/2 and cyclin B1 to thwart cell cycle progression and angiogenesis. Moreover, it also disrupts the PI3K and AKT in the PI3K/AKT pathway to suppress GSK3 $\beta$ , downregulating cyclin D1 and causing cell cycle arrest. The inhibition of AKT also suppresses the mTOR obstructing cell cycle progression and angiogenesis. Luteolin also modulates the Wnt/ $\beta$ -catenin pathway, causing inhibition of GSK3 $\beta$  and  $\beta$ -catenin, and resulting in the inhibition of EMT (Figure 8B).<sup>274</sup>

Chemoprevention by luteolin is a multifaceted approach that acts at multiple points by obstructing invasion, metastasis, angiogenesis, cell cycle regulation, and induction of apoptosis. The upregulation of pro-apoptotic proteins such as Bax and Caspase 3, combined with the downregulation of the anti-apoptotic protein Bcl-2, enhances luteolin's apoptotic potential.<sup>274</sup> Luteolin can scavenge and neutralize ROS and protect the cells from lipid peroxidation and DNA damage. A recent study by Fernando et al demonstrated that luteolin scavenged intracellular ROS dose-dependently. They observed that ROS decreased by 31% at 0.625, 51% at 1.250, 58% at 2.500, 68% at 5.000, and 75% at 10.000  $\mu$ g/ml luteolin concentration in lung fibroblast cells.<sup>275</sup> Cell cycle regulation is another mechanism by which luteolin safeguards against cancer. Luteolin causes cell cycle arrest and halts the proliferation of tumor cells by downregulating cyclin D1 and Survivin, and upregulating p21.<sup>276</sup> The antimetastatic character of luteolin is due to its ability to suppress epithelial-mesenchymal transition, a critical process of cancer metastasis. Several studies in breast cancer cells have highlighted this potential of luteolin.<sup>273,277</sup> A case in point is provided by a study conducted by Cao et al, where they revealed that luteolin significantly inhibited YAP/TAZ activity by promoting YAP/TAZ degradation in Triple-Negative Breast Cancer (TNBC) cells to suppress their migration.<sup>277</sup>

**Table 6.** Curcumin Clinical Studies.

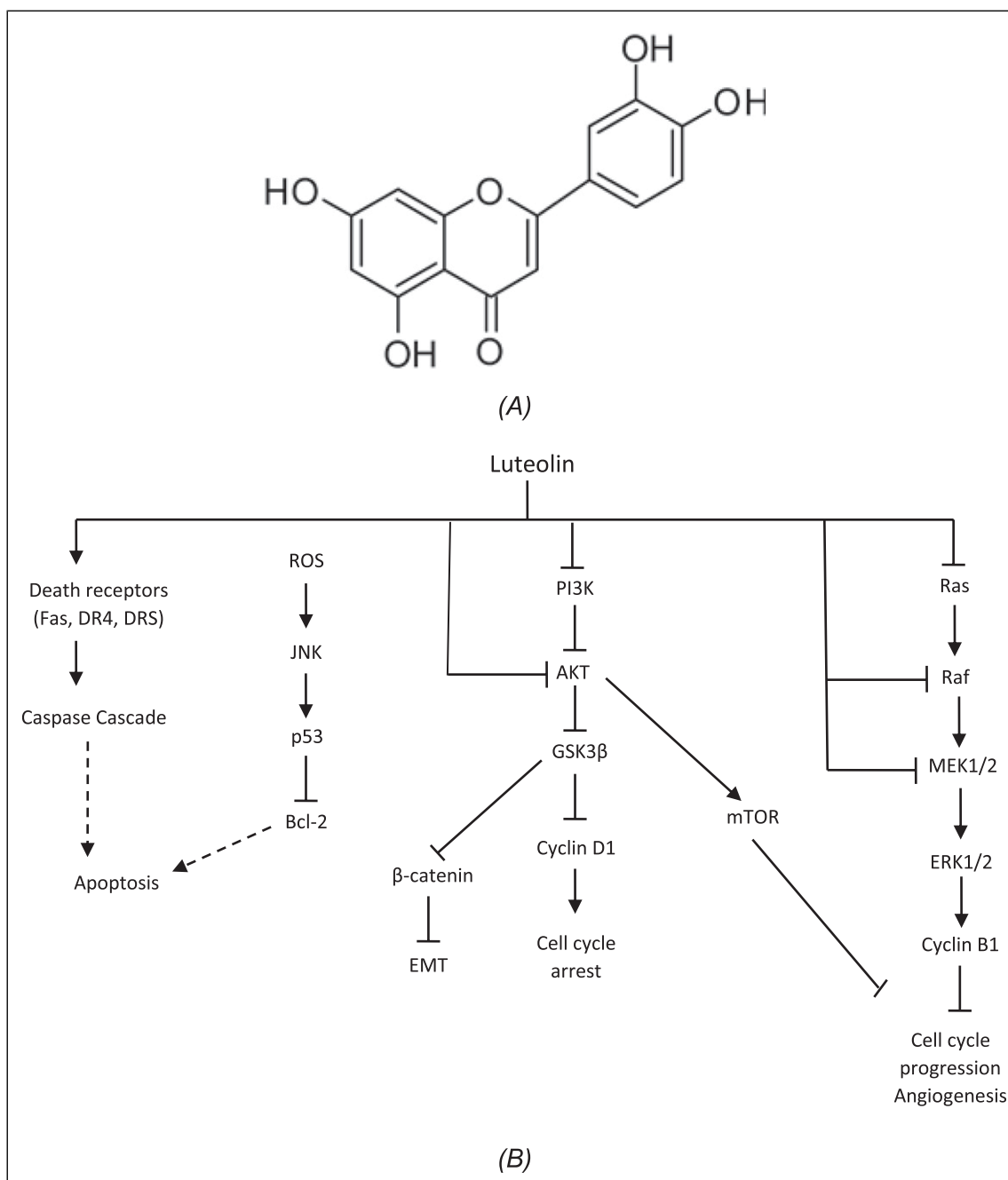
Author and Year	Type of Cancer	Dose of Curcumin	Duration of Study	No. of Participants	Results
Gunther et al, 2022 <sup>258</sup>	Locally advanced rectal cancer (LARC)	4 g twice a day	13 years	22	The addition of curcumin to CRT did not increase the pathologic complete response (pCR) rate
Santosa et al, 2022 <sup>259</sup>	Myeloma	8g/day	16 weeks	33	Overall remission was improved and NF-kB, VEGF, TNF- $\alpha$ , and IL-6 levels were decreased.
Passildas-Jahanmohan et al, 2021 <sup>260</sup>	Metastatic castration-resistant prostate cancer (mCRPC)	6 g/day for 7 days every 3 weeks	18 weeks	50	Adding curcumin to mCRPC patients' treatment strategies was not found to be efficacious
Saghatelian et al, 2020 <sup>261</sup>	Breast cancer	300 mg once a week for 12 weeks	23 weeks	150	Objective response rate (ORR) and overall physical performance was found to be higher
Howells et al, 2019 <sup>262</sup>	Colorectal cancer	2g/day	24 weeks	27	Curcumin is a safe and tolerable adjunct to folinic acid/5-fluorouracil/oxaliplatin chemotherapy (FOLFOX)
Choi et al, 2019 <sup>263</sup>	Prostate cancer	1440 mg/day for 6 months	36 months	97	PSA elevation was suppressed but curcumin did not significantly affect the overall off-treatment duration of intermittent androgen deprivation (IAD)
Kuriakose et al, 2016 <sup>264</sup>	Oral leukoplakia	3.6 g/day for 6 months	12 months	223	Curcumin was well tolerated and demonstrated a significant and durable clinical response

Luteolin modulates numerous cell signaling pathways to disrupt tumor initiation and progression. It can inhibit the growth, migration, and inhibition of SW620 and SW480 colon cancer cells by acting on the IL-6/STAT3 pathway.<sup>278</sup> A study on A549 lung cancer cells revealed that luteolin restricts tumor migration by suppressing focal adhesion development and limiting the FAK-Src signaling.<sup>279</sup> Another research reported the inhibition of viability, migration, angiogenesis, and invasion in vascular endothelial cells of NSCLC via miR-133a-3p upregulation.<sup>280</sup> This study also noted that luteolin decreased purine-rich element-binding protein B (PURB) and showed the inhibitory effects of the compound on tumors by miR-133a-3p/PURB-mediated MAPK and PI3K/Akt pathways.<sup>280</sup> Moreover, luteolin also inhibits YAP/TAZ activity by modulating the Hippo signaling pathway to suppress the migration of TNBC cells.<sup>277</sup> The effects of luteolin on the modulation of the PI3K/AKT signaling pathway to inhibit cancer have been studied extensively. One such research demonstrated that luteolin could contain the proliferation of melanoma cells and trigger apoptosis by reducing MMP-2 and MMP-9 expression through the mediation of the PI3K/AKT pathway.<sup>281</sup> A recent study also observed the ability of luteolin to modulate the PI3K/AKT pathway in HeLa cells. The findings suggest that luteolin suppresses proliferation and promotes apoptosis by altering AKT/mTOR/PI3K and MAPK pathways.<sup>282</sup>

### Immunomodulatory Effects of Luteolin

Luteolin has garnered attention in cancer immunotherapy because of its immunomodulatory effects. It suppresses

inflammation by modulating cytokine secretion involved in immune responses. Luteolin promotes NK cell secretion of type I cytokines, especially IL-2 and IFN- $\gamma$ , critical for providing antitumor immunity.<sup>283</sup> A similar study on the immunomodulatory effects of luteolin on lung cancer observed the increment of NK cell cytotoxicity and granule secretion at 12.5  $\mu\text{g/ml}$  and 25  $\mu\text{g/ml}$  luteolin concentrations.<sup>284</sup> As stated earlier, luteolin has antioxidant and anticancer properties along with its immunomodulatory properties, making it a potential candidate for cancer immunotherapy. A review by Shang et al explores the possibility of using nanocarriers as a delivery system to enhance solubility, circulation time, and targeting ability of luteolin, addressing its poor bioavailability.<sup>285</sup> Modifying luteolin nanocrystals with SDS enhanced the bioavailability of luteolin by 3.48-fold, according to a study by Liu et al.<sup>286</sup> Luteolin is also being studied as an adjunct in cancer therapy. In a study, luteolin has been shown to increase the antitumor activity of oxaliplatin in colorectal carcinoma. When 50 mg/kg BW/day luteolin was administered with 10 mg/kg BW/day oxaliplatin 3 times a week for 3 weeks in mice implanted with HCT116 colorectal carcinoma, a synergistic suppression of tumors was observed.<sup>287</sup> Another recent research by Chen et al notes that luteolin effectively inhibits cervical tumor proliferation and growth when combined with Asiatic acid.<sup>288</sup> Evidence and research such as this provide a foundation to explore the use of luteolin in cancer therapy, either alone or in combination with other drugs.



**Figure 8.** (A) Luteolin, (B) Modulation of cell signaling pathways by luteolin.

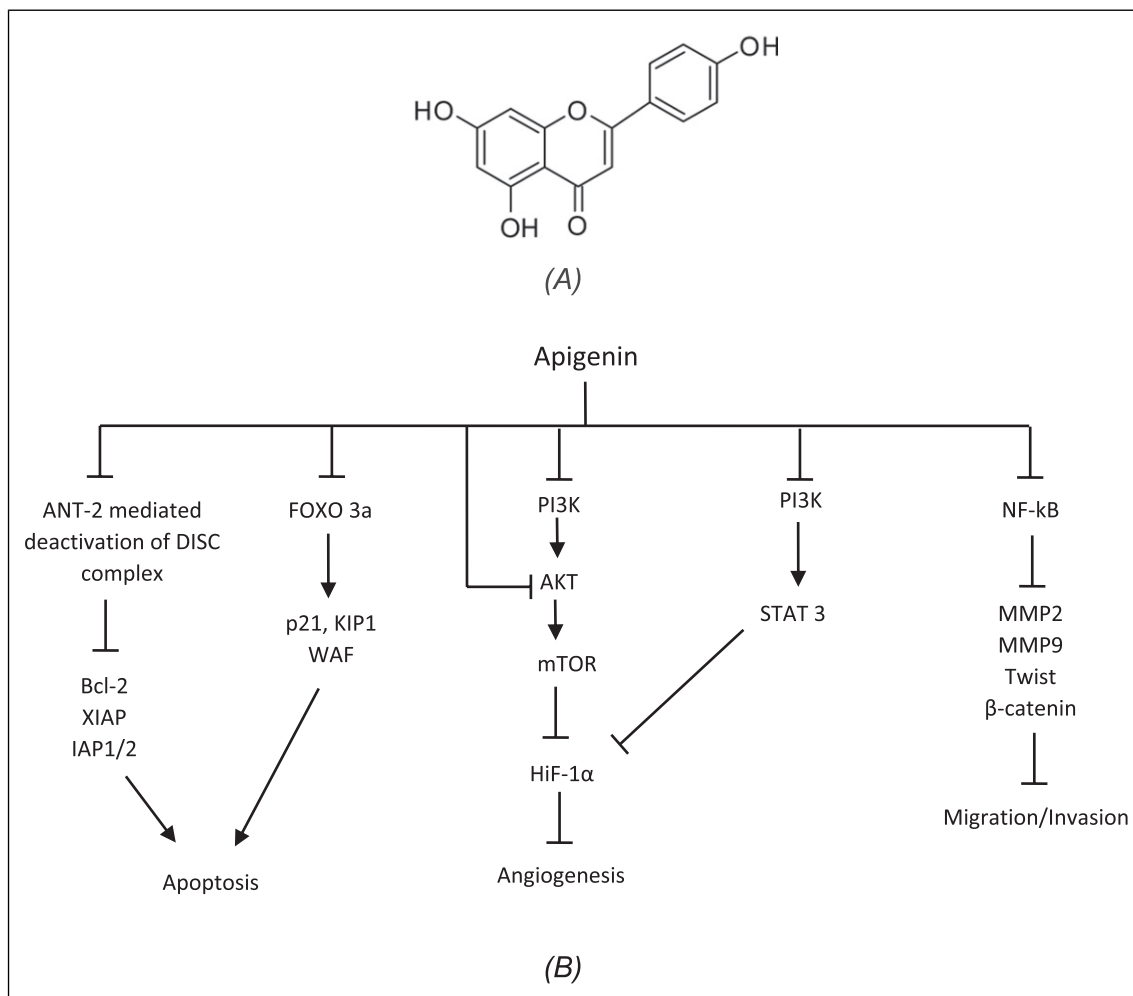
### Luteolin Human Studies

While multiple preclinical studies involving in vivo and in vitro research techniques have already demonstrated the potential anticancer properties of luteolin, including its ability to induce apoptosis, inhibit proliferation, and disrupt cell migration, these findings are yet to be translated to clinical research.<sup>273,274,289</sup> One of the primary reasons for the inability to conduct clinical trials is the poor bioavailability of luteolin, mainly due to its low solubility

in water.<sup>290</sup> Another factor contributing to its poor bioavailability is the excessive glucuronidation of luteolin by enzymes such as uridine diphosphate glucuronosyltransferases 1 As (UGT1As).<sup>291</sup>

### Apigenin

Apigenin is a naturally occurring yellow-colored phytochemical of the flavone class found in various plants and vegetables (Figure 9). Dried chamomile flowers have up to



**Figure 9.** (A) Apigenin, (B) Modulation of cell signaling pathways by apigenin.

5000  $\mu\text{g/g}$  of apigenin.<sup>292</sup> Parsley, celery, spinach, grapefruits, etc. are other sources of apigenin. This flavonoid has attracted significant attention recently owing to its beneficial effects in ameliorating diverse health complications. Apigenin has various biological and pharmacological properties, such as antioxidant, anti-inflammatory, neuroprotective, and cardioprotective abilities.<sup>293</sup> It decreases oxidative stress and neuroinflammation to protect against neurodegenerative diseases.<sup>294</sup> Apigenin shows beneficial effects against various forms of cancer, including breast,<sup>295</sup> prostate,<sup>296</sup> colorectal,<sup>297</sup> and hepatocellular carcinoma.<sup>298</sup>

Apigenin induces apoptosis through the regulation of the TRAIL signaling pathway by preventing ANT-2-mediated deactivation of the DISC complex, resulting in the downregulation of antiapoptotic proteins such as Bcl-2, XIAP, and IAP 1/2. It also blocks the downstream activation of transcription factors, such as FOXO3a, that promote the expression of pro-apoptotic proteins like p21, KIP1, and WAF, thereby triggering apoptosis. Apigenin obstructs the phosphorylation of PI3K, AKT, and STAT3, causing the downregulation of HIF-1 $\alpha$  and VEGF to contain angiogenesis.

Moreover, it suppresses the NF-kB signaling pathway to thwart the expression of genes involved in cell migration and invasion, such as MMP-2, MMP-9, Twist1, and  $\beta$ -catenin (Figure 9B).<sup>299,300</sup>

Apigenin monitors the hallmarks of cancer by inducing apoptosis and autophagy, and inhibiting cell proliferation, angiogenesis, and metastasis. The antioxidant mechanism of apigenin helps in cancer prevention by the inhibition of oxidant enzymes, modulation of redox signaling pathways, and free radical scavenging.<sup>301</sup> Apigenin can regulate the cell cycle at various cell cycle checkpoints to obstruct cell proliferation. It can halt the cell cycle at the G2 phase by reducing mRNA and protein levels of the key regulators that control the G2-M transition in prostate cancer cells.<sup>302</sup> Research by Bao et al concluded that apigenin also arrests renal cell carcinoma (RCC) at the G2-M transition by reducing the expression levels of cyclin A, B1, D3, and E.<sup>303</sup> Apigenin induces apoptosis in cancer cells by triggering both intrinsic and extrinsic pathways. It can trigger apoptosis intrinsically by altering mitochondrial membrane potential leading to the release of cytochrome c and activating a caspase cascade or by impeding

the levels of anti-apoptotic proteins, such as Bcl-2 and Bcl-xL, while increasing pro-apoptotic proteins like Bax.<sup>304,305</sup> Extrinsicly, apigenin induces apoptosis by upregulating the mRNA expressions of caspase-3, caspase-8, and TNF- $\alpha$ .<sup>305</sup> Furthermore, it may induce other cell death pathways besides apoptosis, such as autophagic cell death, ferroptosis, necroptosis, etc., by inducing endogenous ROS generation.<sup>306</sup>

Modulation of signaling pathways is a critical process in cancer chemoprevention by apigenin. Apigenin alters numerous signaling routes to disrupt cancer initiation, proliferation, angiogenesis, and metastasis processes. It inhibits AKT phosphorylation, a key regulator of the cell cycle, growth, and survival, by suppressing AKT function or directly repressing PI3K activity by blocking its ATP-binding site.<sup>307</sup> Apigenin causes cell cycle arrest and apoptosis of human prostate cancer cells by inhibiting the PI3K/AKT/FOXO signaling pathway.<sup>308</sup> A mixture of apigenin and chrysin suppressed the activity of the p38 MAPK/Akt pathway to impede the proliferation, migration, and invasion of colorectal cancer cells.<sup>309</sup> Another recent study by Naponelli et al highlighted the role of apigenin in suppressing tumor angiogenesis by targeting HIF-1 $\alpha$ /HIF signaling pathways.<sup>310</sup> The same study also noted that apigenin inhibits multiple signaling pathways--Wnt/ $\beta$ -catenin, PI3K-AKT, and Hippo-YAP/TAZ to inhibit EMT, a key step for cancer metastasis.<sup>310</sup> Similarly, vitexin, a glycosylated form of apigenin, reduced the stemness of human endometrial cancer by downregulating Oct4 and Nanog through the inhibition of the PI3K/AKT pathway.<sup>311</sup> Moreover, apigenin reduces the expression of STAT3 and JAK2 to prevent tumor growth and proliferation by triggering apoptosis in HER2-overexpressing breast cancer cells.<sup>312,313</sup>

### *Immunomodulatory Effects of Apigenin*

Apigenin modulates the immunological response via multiple routes to boost immunity. It enhances anti-tumor immunity by stimulating natural killer cells and CD8<sup>+</sup> T lymphocytes while suppressing immunosuppressive cells like regulatory T cells and myeloid-derived suppressor cells.<sup>314</sup> It enhances the ability of NK cells to kill pancreatic cancer cells by upregulating the expression of NKG2D, perforin, and granzyme B.<sup>315</sup> Apigenin's immunomodulatory effects in cancer are not limited to the stimulation and enhancement of the proliferative ability of immune cells, as it can also modulate immune checkpoint pathways in cancer. A recent study aiming to highlight the immunomodulatory potential of apigenin in triple-negative breast cancer (TNBC) concluded that it overcomes the inhibitory effects of PD-L1 inhibitors by halting the AKT/mTOR pathway.<sup>316</sup> This finding underscores the potential for developing combination therapies and personalized treatment strategies to boost the immune response in combating cancer. Combination treatments of apigenin and other chemotherapeutic agents are being studied *in vivo* and *in vitro*. One such study observed that apigenin sensitizes

leukemia cells to the topoisomerase inhibitors, etoposide and cyclophosphamide, by promoting apoptosis and reducing ATP levels.<sup>317</sup> Apigenin also enhanced the action of BTK inhibitor abivertinib by the downregulation of BclX through the suppression of PI3K/p-AKT in diffuse large B-cell lymphoma cell lines.<sup>318</sup> A recent study by Nimal et al also noted that apigenin combined with vorinostat showed anticancer activities by modulating the expression of epigenetic and apoptotic regulators, leading to reduced cell migration and increased cell death in TNBC models.<sup>319</sup> In light of these findings, apigenin has gathered much attention from the medical community as a possible cancer chemopreventive agent.

### *Apigenin Human Studies*

Chemoprevention using apigenin has not been successful because no clinical trial data have been published on the compound to date. Hence, there is a need to conduct clinical trials in controlled settings to identify and label the chemopreventive attributes of apigenin accurately. Nonetheless, the success in preclinical settings surely gives a foreground for designing and developing clinical trials.

## **Limitations of Chemopreventive Compounds in Cancer Therapy and Future Directions**

Humans have been using phytochemicals as healing potions since they were hunter-gatherers. While phytochemicals are generally non-toxic and provide multiple health benefits, their use in cancer chemoprevention has not been fully established despite tremendous research. Not a single food product has been labeled a chemopreventive drug through clinical trials so far (Table 7).

Early epidemiological studies showed that micronutrient deficiency results in cancer risk. There was a higher risk of esophageal cancer in populations of northern China, central Asia, and northern Iran that depended on cereals for diet and consumed very few fruits and vegetables. With US-China cooperation, the idea that micronutrients prevent cancer was tested in large clinical trials in the 1980s. A factorial design that tested the role of retinol, zinc, riboflavin, niacin, ascorbate, molybdenum, alpha-tocopherol, beta-carotene, and selenium was conducted. The study showed that a diet consisting of beta-carotene, alpha-tocopherol, and selenium reduced the rate of gastric and total cancer mortality. The effect was especially pronounced in the young population.<sup>320</sup>

Carotene and alpha-tocopherol are vitamins, whose role has been extensively tested through clinical trials. In a factorial design, the role of the 2 chemicals in preventing lung cancer was tested in smokers with a high risk of lung cancer in the 1980s and surprisingly, the  $\beta$ -carotene group has a higher incidence of lung cancer. However, a secondary endpoint, the risk of prostate cancer, was reduced in the alpha-tocopherol

**Table 7.** Summary of Phytochemicals and Their Corresponding Molecular Targets and Therapeutic Use.

Compound	Natural Sources	Therapeutic Use	Effects/Molecular Targets	Ref. No.
Lycopene	Tomato Watermelon Red guava Papaya	Prostate cancer	<ul style="list-style-type: none"> <li>• Activates TP53 and induces apoptosis</li> <li>• Alters PI3K/Akt, NF-<math>\kappa</math>B, AKT/mTOR, MAPKs, and JAK/STATs signaling pathways to prevent tumor growth</li> <li>• Inhibits VEGF to check angiogenesis</li> </ul>	15,17-22
		Breast cancer	<ul style="list-style-type: none"> <li>• Acts synergistically with quinacrine to inhibit Wnt-TCF signaling</li> </ul>	
		Gastric cancer, pancreatic cancer	<ul style="list-style-type: none"> <li>• Anti-inflammatory effect</li> </ul>	
Resveratrol	Red grapes Berries	Skin cancer	<ul style="list-style-type: none"> <li>• Scavenges and quenches ROS to protect against lipid peroxidation and DNA damage</li> </ul>	53,55,59,63,64
		Lung cancer	<ul style="list-style-type: none"> <li>• Reduces Rb phosphorylation and induces p21 and p53 protein expression</li> <li>• Causes S phase arrest to repair DNA damage</li> </ul>	
		Prostate cancer	<ul style="list-style-type: none"> <li>• Inhibits PI3K/Akt pathway and modulates Bcl-2 family proteins, promoting apoptosis</li> </ul>	
		Colon cancer	<ul style="list-style-type: none"> <li>• Reverses epithelial-mesenchymal transition via the AKT/GSK-3<math>\beta</math>/Snail signaling pathway to inhibit invasion and metastasis</li> </ul>	
Sulforaphane	Broccoli Cabbage Cauliflower Kale	Prostate cancer	<ul style="list-style-type: none"> <li>• Suppresses the proliferation of cancer cells containing stimulated androgen receptors and prostate-specific antigen (PSA)</li> </ul>	88,89,98
		Colorectal cancer	<ul style="list-style-type: none"> <li>• Inhibits proliferation by upregulating the expression of UGT1A in CRC cells via the ERK/Nrf2 signaling pathway</li> </ul>	
		Lung cancer	<ul style="list-style-type: none"> <li>• Modulates sonic hedgehog pathways and PHC3 to inhibit the activities of Shh, Smo, Gli1, and PHC3 in lung cancer cells to deprive them of self-renewal ability</li> </ul>	
Isoflavones (Genistein)	Soyabean Chickpea	Breast cancer	<ul style="list-style-type: none"> <li>• Inhibits c-erbB-2, MMP-2, and MMP-9 to contain angiogenesis and metastasis of breast cancer cells</li> <li>• Arrests cancer cells in the G0/G1 phase and causes apoptosis</li> <li>• Alters the PI3K/Akt pathway to induce differentiation in breast cancer stem cells by interactions with ER + cells</li> </ul>	126,127,129,131,135
		Lung cancer	<ul style="list-style-type: none"> <li>• Modulates IMPDH2/AKT1 pathway to promote apoptosis</li> </ul>	
		Cervical cancer	<ul style="list-style-type: none"> <li>• Enhances anticancer effects of cisplatin by altering p-ERK1/2, Bcl2, cleaved caspase 3, and p53 expression levels</li> </ul>	
Catechins (Epigallocatechin gallate)	Green tea	Hepatocellular carcinoma	<ul style="list-style-type: none"> <li>• Inhibits MMP-2 and MMP-9 activity to halt metastasis.</li> </ul>	170-172,182,187
		Pancreatic cancer	<ul style="list-style-type: none"> <li>• Decreases proliferation and conducts apoptosis by modulation of PI3K/Akt/mTOR pathway due to lowered p-Akt and p-mTOR expression</li> </ul>	
		Gastric cancer	<ul style="list-style-type: none"> <li>• Silences wnt/<math>\beta</math>-catenin pathway to impede proliferation</li> </ul>	
		Breast cancer	<ul style="list-style-type: none"> <li>• Downregulates the wnt pathway by increasing the expression of G1 regulators, c-MYC, and cyclin D1 genes to hinder tumor proliferation and invasiveness</li> </ul>	
		Lymphoma	<ul style="list-style-type: none"> <li>• Induces apoptosis via caspase-dependent pathway and Bcl-2 family protein modulation</li> </ul>	

(continued)

**Table 7.** (continued)

Compound	Natural Sources	Therapeutic Use	Effects/Molecular Targets	Ref. No.
Quercetin	Capers Red onions Kale Shallots	Cervical cancer	<ul style="list-style-type: none"> <li>Reduces inflammatory mediators such as NO-synthase, COX-2, and CRP</li> <li>Blocks NF-κB activation causing downregulation of pro-inflammatory genes</li> <li>Causes G2 phase arrest to trigger apoptosis</li> </ul>	207,211,213-215
		Leukemia	<ul style="list-style-type: none"> <li>Activates caspase-3 and caspase-9 to induce apoptosis</li> </ul>	
		Hepatocellular carcinoma	<ul style="list-style-type: none"> <li>Modulates signaling pathways like NF-κB/ικB, p38 MAPK, Bcl-2/Bax, resulting in apoptosis</li> </ul>	
		Lung cancer	<ul style="list-style-type: none"> <li>Inhibits Snail-dependent Akt activation pathway</li> <li>Contains tumor's invasiveness by attenuating the expression of N-cadherin, vimentin, ADAM9, and MMPs-related proteins and upregulating the expression of E-cadherin</li> </ul>	
Curcumin	Turmeric	Liver cancer	<ul style="list-style-type: none"> <li>Suppresses angiogenesis by downregulating VEGF, CD31, and αSMC expression levels</li> </ul>	234,247-249,251
		Ovarian cancer	<ul style="list-style-type: none"> <li>Inhibits NF-κB and signal transducers and activators of transcription 3 activation</li> <li>Mitigates angiogenic cytokine expression</li> </ul>	
		Head and Neck cancer	<ul style="list-style-type: none"> <li>Induces apoptosis by G2 phase accumulation of tumor cells via repression of PI3K/Akt/mTOR pathway</li> </ul>	
		Lung cancer	<ul style="list-style-type: none"> <li>Prevents tumor proliferation and regulates apoptosis by upregulating miR-192-5p and downregulating the PI3K/Akt signaling pathway.</li> </ul>	
		Colorectal cancer	<ul style="list-style-type: none"> <li>Reduces N-cadherin, twist, snail, and vimentin, and increases E-cadherin in tumor cells to suppress the EMT process by inhibiting the CDX2/Wnt3a/β-catenin pathway</li> </ul>	
Luteolin	Celery Thyme Parsley Chamomile tea	Colon cancer	<ul style="list-style-type: none"> <li>Modulates the IL-6/STAT3 pathway</li> </ul>	280-284
		Lung cancer	<ul style="list-style-type: none"> <li>Restricts the FAK-Src signaling</li> <li>Upregulates miR-133a-3p and downregulates PURB to modulate MAPK and PI3K/AKT pathways</li> </ul>	
		Breast cancer	<ul style="list-style-type: none"> <li>Inhibits YAP/TAZ activity by altering the Hippo signaling pathway to suppress the migration of TNBC cells</li> </ul>	
		Melanoma	<ul style="list-style-type: none"> <li>Limits migration and triggers apoptosis by reducing MMP-2 and MMP-9 expression through the mediation of the PI3K/AKT pathway</li> </ul>	
Apigenin	Chamomile flowers Parsley Celery Spinach Grapefruits	Prostate cancer	<ul style="list-style-type: none"> <li>Halts the cell cycle at G2/M transition by reducing mRNA and protein levels.</li> <li>Causes cell cycle arrest and apoptosis by inhibiting the PI3K/AKT/FOXO signaling pathway</li> </ul>	299,300,305,311,312,314
		Breast cancer	<ul style="list-style-type: none"> <li>Reduces the expression of STAT3 and JAK2 to prevent tumor growth and proliferation by triggering apoptosis</li> </ul>	
		Endometrial cancer	<ul style="list-style-type: none"> <li>Downregulates the expression of Oct4 and Nanog by inhibiting the PI3K/AKT pathway</li> </ul>	
		Colorectal cancer	<ul style="list-style-type: none"> <li>A mixture of apigenin and chrysin suppresses the activity of the p38 MAPK/Akt pathway to impede proliferation, migration, and invasion</li> </ul>	

group. Another clinical trial was launched in the US in the 1980s and 1990s to study the effect of selenium in the chemoprevention of basal cell or squamous cell carcinoma. While these cancer risks were not reduced, overall cancer risk was reduced. The Selenium and Vitamin E Cancer Prevention Trial (SELECT) was conducted later using selenium and Vitamin E

in a 2 × 2 factorial design to test their efficacy in preventing cancer. The results came out negative.<sup>320</sup>

The reasons why these phytochemical nutrients failed in clinical trials may be severalfold. In an already nutritionally well-nourished population, the nutrients might not prevent cancer. Additionally, these trials used very high



doses of the nutrients, which might lead to untoward consequences. Moreover, the preventive effects might be prominent in early life and the population sample in these studies did not select young subjects. The selection of agent may have been inappropriate leading to negative results.<sup>320</sup>

Besides nutritive phytochemicals, non-nutritive phytochemicals have also been tested in human experiments. While some experiments show positive results, others show negative results. The studies give very inconsistent results. Lifestyle factors, genetic polymorphisms, and other interfering factors may reduce the power of epidemiological studies compared to animal studies where the results are often promising. The inconsistencies may lie in the interpretation of cell lines and animal studies. In animal studies, very high levels of phytochemicals are used to get results, while in human intervention, much lower doses are used which might not give the same result. Differences in the bioavailability of agents in animal studies vs human studies might lead to result discrepancies. It is not always that high doses are required for effect. For example, for resveratrol, the lowest dose suppressed cancer much better than the higher dose.<sup>320</sup>

The positive results shown by *in vitro* studies and animal models are obtained using a much higher dose of these compounds than the amount gained from a normal diet. This makes it difficult to acquire the same health benefits as the *in vitro* studies promised. One needs to take them in larger concentrations to compensate for their poor solubility and inability to penetrate the plasma membrane. Individual genetics, metabolic patterns, and genetic diversity due to cultural and geographic locations make it even more challenging to pinpoint these compounds' bioavailability accurately. Using non-nutritive phytochemicals, in the long run, might be toxic if the dosage is very high. The presumed advantage of phytochemicals due to their nontoxicity would then be negated. Especially if the chemicals have to be taken for a long duration as chemopreventive agents, this aspect would come to the forefront.

Some phytochemicals impart benefits in certain forms of cancer while increasing the risk associated with other forms of cancer at the same time. For instance, isoflavones, although beneficial in breast and lung cancer, increase the risk associated with advanced forms of prostate cancer.<sup>321</sup> Flavonoids work as mutagens, pro-oxidants, and inhibitors of drug-metabolizing enzymes in addition to being an anticancer agent.<sup>322</sup> The lack of convincing evidence and ambiguous findings from the clinical trials have made it even more daunting to label specific phytochemicals with their cancer-mitigating abilities.

For most of the non-nutritive phytochemicals, there is a plethora of cell line and animal studies but very few high-powered human intervention trials. This can be attributed to a lack of resources as well. Furthermore, some of the large clinical trials using beta-carotene, alpha-tocopherol, and selenium were not very successful. Based on these results, the

community is reluctant to spend a lot of time and resources conducting trials with other phytochemicals unless very promising and consistent results are derived from smaller clinical trials. It is very easy to choose a phytochemical of interest, select a population, plan dosage, and conduct a large clinical trial but the result has to be worth the effort.

One possible suggestion for conducting a worthwhile large-scale clinical trial is to combine the usage of multiple non-nutritive phytochemicals of different chemical classes and different targets. The trial can measure not only the effects on cancer chemoprevention but also the effects on other chronic diseases and health parameters. Evaluating the effect of combination phytochemicals is by no means an easy task. First, you have to start with 2 phytochemicals and measure their synergistic and antagonistic effects. Once you start adding third or fourth chemicals, the number of combinations increases tremendously. Analyzing synergism and antagonism in *in vivo* studies of all combinations might be very time-consuming and not feasible (perspective, a positive cocktail effect of bioactive components in the diet). One option is to try the hit-and-trial method with multiple chemicals at appropriate dosages determined by a panel of experts. After carrying out *in vivo* studies, bioavailability and toxicity studies in humans can be carried out using multiple chemicals at designated dosages. If no untoward effects are observed then efficacy studies can be conducted.

Alternatively, to test if combinations of phytochemicals have a synergistic chemopreventive effect, a randomized control trial where different dietary phytochemical index foods can be supplied to treatment vs control groups can be conducted and cancer risk measured. The dietary phytochemical index measures the amount of calories obtained from plant food, excluding potatoes. To devise a more accurate scale that measures total phytochemicals consumed, a different scale that also takes into account phytochemicals consumed from low-calorie food such as green tea can be used. Given the beneficial effects of a high phytochemical diet on cardiovascular and chronic diseases, the use of a high phytochemical diet should be promoted. If further chemopreventive effects on cancer were observed due to the intake of phytochemicals, the promotion of a phytochemical diet would get priority.<sup>323</sup>

Biomarker discovery, so that endpoints can be properly measured, is crucial for cancer chemoprevention trial design. The success of cardiovascular chemoprevention has largely depended on finding biomarkers such as lipid levels and hypertension. In cases where large clinical trials using thousands of individuals are not possible, smaller clinical trials using high-risk individuals can be designed.<sup>11</sup>

## Conclusion

Cancer poses a severe public health concern globally. As the scientific community shifts its attention toward a preventive approach, phytochemicals can have a pivotal role in the fight

against cancer. With proven therapeutic properties, many phytochemicals have been used in our households since ancient times. However, dosage, potential side effects, and negative interactions of these chemopreventive compounds had been overlooked previously, which somewhat decreases their true potential. As more research and findings surface, the use of phytochemicals in cancer prevention is becoming more evident. Clinical and pre-clinical studies of phytochemicals show encouraging results in cancer prevention. While having immense potential for the battle against cancer, these phytochemicals have limitations in dosage and bioavailability, as described in the limitations section. It is only through the evidence-based promotion of phytochemicals from studies with robust methodologies that a preventive approach against cancer can be effectively implemented and widely accepted in the medical fraternity.

## Abbreviations

IARC	International Agency for Research on Cancer
NSAID	Non-steroidal Anti-inflammatory Drugs
VEGF	Vascular Endothelial Growth Factor
TGFβ	Transforming Growth Factor-β
ROS	Reactive Oxygen Species
CRC	Colorectal Cancer
CVD	Cardiovascular Diseases
EGCG	Epigallocatechin Gallate
BAT	Brown Adipose Tissue
VSMC	Vascular Smooth Muscle Cells
RNS	Reactive Nitrogen Species
NSCLC	Non-Small Cell Lung Cancer
TNBC	Triple-Negative Breast Cancer
PURB	Purine-rich element-binding Protein B
NK	Natural Killer
SDS	Sodium Dodecyl Sulfate
RCC	Renal Cell Carcinoma
SELECT	Selenium and Vitamin E Cancer Prevention Trial

## Acknowledgments

We are deeply grateful to Mr. Sandesh Gaudel for his valuable insights into the immunomodulatory effects of phytochemicals. His guidance and thoughtful input significantly enriched this work. We would also like to express our sincere gratitude to Ms. Shristina Shrestha and Ms. Ruchie Shrees for their meticulous proofreading; their language suggestions and edits were instrumental in refining the article. Lastly, special thanks to Mr. Nirav Lekhak for his creative assistance in developing the figures and for his helpful language edits.

## Author Contributions

NL and HKB conceptualized the paper. NL wrote on the preclinical aspects. HKB wrote on the clinical aspects.

## Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

## Funding

The author(s) received no financial support for the research, authorship, and/or publication of this article.

## Ethical Statement

This manuscript is a review article and does not involve a research protocol requiring approval by the relevant institutional review board or ethics committee.

## ORCID iDs

Nitish Lekhak  <https://orcid.org/0009-0009-6370-0729>

Hitesh Kumar Bhattarai  <https://orcid.org/0000-0002-7147-1411>

## References

1. Das S, Kundu M, Jena BC, Mandal M. Causes of cancer: physical, chemical, biological carcinogens, and viruses. *Biomater 3D Tumor Model*. 2020;607-641. doi:10.1016/B978-0-12-818128-7.00025-3
2. Cancer today. (n.d.). Retrieved August 25, 2024, from: <https://gco.iarc.fr/today/en/fact-sheets-populations>
3. Falzone L, Salomone S, Libra M. Evolution of cancer pharmacological treatments at the turn of the third millennium. *Front Pharmacol*. 2018;9(NOV):1300. doi:10.3389/FPHAR.2018.01300
4. Miller KD, Siegel RL, Lin CC, et al. Cancer treatment and survivorship statistics, 2016. *CA Cancer J Clin*. 2016;66(4):271-289. doi:10.3322/CAAC.21349
5. Zhou Y, Zheng J, Li Y, et al. Natural polyphenols for prevention and treatment of cancer. *Nutrients*. 2016;8(8):515. doi:10.3390/NU8080515
6. Shanbhag VKL. Lycopene in cancer therapy. *J Pharm BioAllied Sci*. 2016;8(2):170-171. doi:10.4103/0975-7406.171740
7. Alavi M, Farkhondeh T, Aschner M, Samarghandian S. Resveratrol mediates its anti-cancer effects by Nrf2 signaling pathway activation. *Cancer Cell Int*. 2021;21(1):579. doi:10.1186/S12935-021-02280-5/TABLES/1
8. Slavin JL. Mechanisms for the impact of whole grain foods on cancer risk. *J Am Coll Nutr*. 2000;19(3 Suppl):300S-307S. doi:10.1080/07315724.2000.10718964
9. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;144(5):646-674. doi:10.1016/J.CELL.2011.02.013
10. Ranjan A, Ramachandran S, Gupta N, et al. Role of phytochemicals in cancer prevention. *Int J Mol Sci*. 2019;20(20):4981. doi:10.3390/IJMS20204981
11. Steward WP, Brown K. Cancer chemoprevention: a rapidly evolving field. *Br J Cancer*. 2013;109(1):1-7. doi:10.1038/bjc.2013.280

12. Khan UM, Sevindik M, Zarrabi A, et al. Lycopene: food sources, biological activities, and human health benefits. *Oxid Med Cell Longev*. 2021;2021:2713511. doi:10.1155/2021/2713511
13. Unlu NZ, Bohn T, Francis DM, Nagaraja HN, Clinton SK, Schwartz SJ. Lycopene from heat-induced cis-isomer-rich tomato sauce is more bioavailable than from all-trans-rich tomato sauce in human subjects. *Br J Nutr*. 2007;98(1):140-146. doi:10.1017/S0007114507685201
14. Kapała A, Szlendak M, Motacka E. The anti-cancer activity of lycopene: a systematic review of human and animal studies. *Nutrients*. 2022;14(23):5152. doi:10.3390/NU14235152/S1
15. Khalaf RA, Awad M. Lycopene as a potential bioactive compound: chemistry, extraction, and anticancer prospective. *Curr Cancer Drug Targets*. 2023;23(8):634-642. doi:10.2174/1568009623666230131124236
16. Long Y, Paengkoum S, Lu S, et al. Physicochemical properties, mechanism of action of lycopene and its application in poultry and ruminant production. *Front Vet Sci*. 2024;11:1364589. doi:10.3389/FVETS.2024.1364589/BIBTEX
17. Chen J, O'Donoghue A, Deng Y-F, Zhang B, Kent F, O'Hare T. The effect of lycopene on the PI3K/Akt signalling pathway in prostate cancer. *Anti Cancer Agents Med Chem*. 2014;14(6):800-805. doi:10.2174/1871520614666140521121317
18. Obermüller-Jevic UC, Olano-Martin E, Corbacho AM, et al. Lycopene inhibits the growth of normal human prostate epithelial cells in vitro. *J Nutr*. 2003;133(11):3356-3360. doi:10.1093/JN/133.11.3356
19. Bai B, Chen Q, Jing R, et al. Molecular basis of prostate cancer and natural products as potential chemotherapeutic and chemopreventive agents. *Front Pharmacol*. 2021;12:738235. doi:10.3389/FPHAR.2021.738235/BIBTEX
20. Yang CM, Yen YT, Huang CS, Hu ML. Growth inhibitory efficacy of lycopene and  $\beta$ -carotene against androgen-independent prostate tumor cells xenografted in nude mice. *Mol Nutr Food Res*. 2011;55(4):606-612. doi:10.1002/MNFR.201000308
21. Preet R, Mohapatra P, Das D, et al. Lycopene synergistically enhances quinacrine action to inhibit Wnt-TCF signaling in breast cancer cells through APC. *Carcinogenesis*. 2013;34(2):277-286. doi:10.1093/CARCIN/BGS351
22. Nakanishi R, Shimizu T, Kumagai K, Takai A, Marusawa H. Genetic pathogenesis of inflammation-associated cancers in digestive organs. *Pathogens*. 2021;10(4):453. doi:10.3390/PATHOGENS10040453
23. Puah B, Jalil J, Attiq A, Kamisah Y. New insights into molecular mechanism behind anti-cancer activities of lycopene. *Molecules*. 2020;26(13):3888. doi:10.3390/molecules26133888
24. Jiang LN, Liu YB, Li BH. Lycopene exerts anti-inflammatory effect to inhibit prostate cancer progression. *Asian J Androl*. 2019;21(1):80-85. doi:10.4103/AJA.AJA\_70\_18
25. Sui J, Guo J, Pan D, et al. The efficacy of dietary intake, supplementation, and blood concentrations of carotenoids in cancer prevention: insights from an umbrella meta-analysis. *Foods*. 2023;13(9):1321. doi:10.3390/foods13091321
26. Yin L, Yan H, Chen K, et al. Diet-derived circulating antioxidants and risk of digestive system tumors: a mendelian randomization study. *Nutrients*. 2022;14(16):3274. doi:10.3390/NU14163274
27. Aune D, Keum N, Giovannucci E, et al. Dietary intake and blood concentrations of antioxidants and the risk of cardiovascular disease, total cancer, and all-cause mortality: a systematic review and dose-response meta-analysis of prospective studies. *Am J Clin Nutr*. 2018;108(5):1069-1091. doi:10.1093/ajcn/nqy097
28. Psaltopoulou T, Ntanas-Stathopoulos I, Tsilimigras DI, Tzanninis IG, Gavriatopoulou M, Sergentanis TN. Micronutrient intake and risk of hematological malignancies in adults: a systematic review and meta-analysis of cohort studies. *Nutr Cancer*. 2018;70(6):821-839. doi:10.1080/01635581.2018.1490444
29. He J, Gu Y, Zhang S. Vitamin A and breast cancer survival: a systematic review and meta-analysis. *Clin Breast Cancer*. 2018;18(6):e1389-e1400. doi:10.1016/J.CLBC.2018.07.025
30. Cataño JG, Trujillo CG, Caicedo JI, et al. [Efficacy of lycopene intake in primary prevention of prostate cancer: a systematic review of the literature and meta-analysis.]. *Arch Esp Urol*. 2018;71(2):187-197.
31. Chen F, Hu J, Liu P, Li J, Wei Z, Liu P. Carotenoid intake and risk of non-Hodgkin lymphoma: a systematic review and dose-response meta-analysis of observational studies. *Ann Hematol*. 2017;96(6):957-965. doi:10.1007/s00277-016-2898-1
32. Panic N, Nedovic D, Pastorino R, Boccia S, Leoncini E. Carotenoid intake from natural sources and colorectal cancer: a systematic review and meta-analysis of epidemiological studies. *Eur J Cancer Prev*. 2017;26(1):27-37. doi:10.1097/CEJ.0000000000000251
33. Rowles JL, Ranard KM, Smith JW, An R, Erdman JW. Increased dietary and circulating lycopene are associated with reduced prostate cancer risk: a systematic review and meta-analysis. *Prostate Cancer Prostatic Dis*. 2017;20(4):361-377. doi:10.1038/PCAN.2017.25
34. Chen J, Jiang W, Shao L, Zhong D, Wu Y, Cai J. Association between intake of antioxidants and pancreatic cancer risk: a meta-analysis. *Int J Food Sci Nutr*. 2016;67(7):744-753. doi:10.1080/09637486.2016.1197892
35. Zhou Y, Wang T, Meng Q, Zhai S. Association of carotenoids with risk of gastric cancer: a meta-analysis. *Clin Nutr*. 2016;35(1):109-116. doi:10.1016/j.clnu.2015.02.003
36. Abar L, Vieira A, Aune D, et al. Blood concentrations of carotenoids and retinol and lung cancer risk: an update of the WCRF-AICR systematic review of published prospective studies. *Cancer Med*. 2016;5(8):2069-2083. doi:10.1002/cam4.676
37. Wang X, Yang HH, Liu Y, Zhou Q, Chen ZH. Lycopene consumption and risk of colorectal cancer: a meta-analysis of observational studies. *Nutr Cancer*. 2016;68(7):1083-1096. doi:10.1080/01635581.2016.1206579
38. Huang X, Gao Y, Zhi X, Ta N, Jiang H, Zheng J. Association between vitamin A, retinol and carotenoid intake and

- pancreatic cancer risk: evidence from epidemiologic studies. *Sci Rep*. 2016;6:38936. doi:10.1038/srep38936
39. Leoncini E, Nedovic D, Panic N, Pastorino R, Edefonti V, Boccia S. Carotenoid intake from natural sources and head and neck cancer: a systematic review and meta-analysis of epidemiological studies. *Cancer Epidemiol Biomarkers Prev*. 2015;24(7):1003-1011. doi:10.1158/1055-9965.EPI-15-0053
40. Wang Y, Cui R, Xiao Y, Fang J, Xu Q. Effect of carotene and lycopene on the risk of prostate cancer: a systematic review and dose-response meta-analysis of observational studies. *PLoS One*. 2015;10(9):e0137427. doi:10.1371/JOURNAL.PONE.0137427
41. Chen P, Zhang W, Wang X, et al. Lycopene and risk of prostate cancer: a systematic review and meta-analysis. *Medicine*. 2015;94(33):e1260. doi:10.1097/MD.0000000000001260
42. Li X, Xu J. Meta-analysis of the association between dietary lycopene intake and ovarian cancer risk in postmenopausal women. *Sci Rep*. 2014;4:4885. doi:10.1038/srep04885
43. Tang JE, Wang RJ, Zhong H, Yu B, Chen Y. Vitamin A and risk of bladder cancer: a meta-analysis of epidemiological studies. *World J Surg Oncol*. 2014;12:130. doi:10.1186/1477-7819-12-130
44. Ge X, Xing M, Yu L, Shen P. Carotenoid intake and esophageal cancer risk: a meta-analysis. *Asian Pac J Cancer Prev*. 2013;14(3):1911-1918. doi:10.7314/APJCP.2013.14.3.1911
45. Xu X, Yu E, Liu L, et al. Dietary intake of vitamins A, C, and E and the risk of colorectal adenoma: a meta-analysis of observational studies. *Eur J Cancer Prev*. 2013;22(6):529-539. doi:10.1097/CEJ.0B013E328364F1EB
46. Chen J, Song Y, Zhang L. Lycopene/tomato consumption and the risk of prostate cancer: a systematic review and meta-analysis of prospective studies. *J Nutr Sci Vitaminol*. 2013;59(3):213-223. doi:10.3177/JNSV.59.213
47. Myung S, Ju W, Kim S, Kim H, Korean Meta-analysis KORMA Study Group. Vitamin or antioxidant intake (or serum level) and risk of cervical neoplasm: a meta-analysis. *BJOG*. 2011;118(11):1285-1291. doi:10.1111/j.1471-0528.2011.03032.x
48. Ilic D, Forbes KM, Hammad C. Lycopene for the prevention of prostate cancer. *Cochrane Database Syst Rev*. 2011;2011(12):CD008007. doi:10.1002/14651858.CD008007.PUB2
49. Musa-Veloso K, Card JW, Wong AW, Cooper DA. Influence of observational study design on the interpretation of cancer risk reduction by carotenoids. *Nutr Rev*. 2009;67(9):527-545. doi:10.1111/J.1753-4887.2009.00225.X
50. Etmiman M, Takkouche B, Caamaño-Isorna F. The role of tomato products and lycopene in the prevention of prostate cancer: a meta-analysis of observational studies. *Cancer Epidemiol Biomarkers Prev*. 2004;13(3):340-345. doi:10.1158/1055-9965.340.13.3
51. Williams LD, Burdock GA, Edwards JA, Beck M, Bausch J. Safety studies conducted on high-purity trans-resveratrol in experimental animals. *Food Chem Toxicol*. 2009;47(9):2170-2182. doi:10.1016/J.FCT.2009.06.002
52. Ndiaye M, Philippe C, Mukhtar H, Ahmad N. The grape antioxidant resveratrol for skin disorders: promise, prospects, and Challenges. *Arch Biochem Biophys*. 2011;508(2):164-170. doi:10.1016/J.ABB.2010.12.030
53. Ko JH, Sethi G, Um JY, et al. The role of resveratrol in cancer therapy. *Int J Mol Sci*. 2017;18(12):2589. doi:10.3390/IJMS18122589
54. Lee YJ, Lee GJ, Yi SS, et al. Cisplatin and resveratrol induce apoptosis and autophagy following oxidative stress in malignant mesothelioma cells. *Food Chem Toxicol*. 2016;97:96-107. doi:10.1016/J.FCT.2016.08.033
55. Ren B, Kwah MXY, Liu C, et al. Resveratrol for cancer therapy: Challenges and future perspectives. *Cancer Lett*. 2021;515:63-72. doi:10.1016/J.CANLET.2021.05.001
56. Bishayee A. Cancer prevention and treatment with resveratrol: from rodent studies to clinical trials. *Cancer Prev Res*. 2009;2(5):409-418. doi:10.1158/1940-6207.CAPR-08-0160/338955/P/CANCER-PREVENTION-AND-TREATMENT-WITH-RESVERATROL
57. Sgambato A, Ardito R, Faraglia B, Boninsegna A, Wolf FI, Cittadini A. Resveratrol, a natural phenolic compound, inhibits cell proliferation and prevents oxidative DNA damage. *Mutat Res*. 2001;496(1-2):171-180. doi:10.1016/S1383-5718(01)00232-7
58. Leonard SS, Xia C, Jiang BH, et al. Resveratrol scavenges reactive oxygen species and effects radical-induced cellular responses. *Biochem Biophys Res Commun*. 2003;309(4):1017-1026. doi:10.1016/j.bbrc.2003.08.105
59. Attia SM. Influence of resveratrol on oxidative damage in genomic DNA and apoptosis induced by cisplatin. *Mutat Res*. 2012;741(1-2):22-31. doi:10.1016/J.MRGENTOX.2011.10.008
60. Liao PC, Ng LT, Lin LT, Richardson CD, Wang GH, Lin CC. Resveratrol arrests cell cycle and induces apoptosis in human hepatocellular carcinoma Huh-7 cells. *J Med Food*. 2010;13(6):1415-1423. doi:10.1089/JMF.2010.1126
61. Wolter F, Akoglu B, Clausnitzer A, Stein J. Downregulation of the cyclin D1/Cdk4 complex occurs during resveratrol-induced cell cycle arrest in colon cancer cell lines. *J Nutr*. 2001;131(8):2197-2203. doi:10.1093/JN/131.8.2197
62. Kim YA, Lee WH, Choi TH, Rhee SH, Park KY, Choi YH. Involvement of p21WAF1/CIP1, pRB, Bax, and NF-kappaB in induction of growth arrest and apoptosis by resveratrol in human lung carcinoma A549 cells. *Int J Oncol*. 2003;23(4):1143-1149. doi:10.3892/IJO.23.4.1143/HTML
63. Aziz MH, Nihal M, Fu VX, Jarrard DF, Ahmad N. Resveratrol-caused apoptosis of human prostate carcinoma LNCaP cells is mediated via modulation of phosphatidylinositol 3'-kinase/Akt pathway and Bel-2 family proteins. *Mol Cancer Ther*. 2006;5(5):1335-1341. doi:10.1158/1535-7163.MCT-05-0526
64. Parekh P, Motiwale L, Naik N, Rao KVK. Downregulation of cyclin D1 is associated with decreased levels of p38 MAP kinases, Akt/PKB and Pak1 during chemopreventive effects of

- resveratrol in liver cancer cells. *Exp Toxicol Pathol*. 2011; 63(1–2):167–173. doi:10.1016/J.ETP.2009.11.005
65. Benitez DA, Hermoso MA, Pozo-Guisado E, Fernández-Salguero PM, Castellón EA. Regulation of cell survival by resveratrol involves inhibition of NF kappa B-regulated gene expression in prostate cancer cells. *Prostate*. 2009;69(10):1045–1054. doi:10.1002/PROS.20953
66. Farooqi AA, Khalid S, Ahmad A. Regulation of cell signaling pathways and miRNAs by resveratrol in different cancers. *Int J Mol Sci*. 2018;19(3):652. doi:10.3390/IJMS19030652
67. Yuan L, Zhou M, Huang D, et al. Resveratrol inhibits the invasion and metastasis of colon cancer through reversal of epithelial-mesenchymal transition via the AKT/GSK-3 $\beta$ /Snail signaling pathway. *Mol Med Rep*. 2022;20(6):2783–2795. doi:10.3892/MMR.2022.12870
68. Schwager J, Richard N, Widmer F, Raederstorff D. Resveratrol distinctively modulates the inflammatory profiles of immune and endothelial cells. *BMC Complement Altern Med*. 2017;17(1):309. doi:10.1186/S12906-017-1823-Z/FIGURES/8
69. Lai X, Pei Q, Song X, et al. The enhancement of immune function and activation of NF- $\kappa$ B by resveratrol-treatment in immunosuppressive mice. *Int Immunopharmacol*. 2016;33:42–47. doi:10.1016/j.intimp.2016.01.028
70. Choi YJ, Yang KM, Kim SD, et al. Resveratrol analogue HS-1793 induces the modulation of tumor-derived T cells. *Exp Ther Med*. 2012;3(4):592–598. doi:10.3892/etm.2012.472
71. Chen L, Musa AE. Boosting immune system against cancer by resveratrol. *Phytother Res*. 2021;35(10):5514–5526. doi:10.1002/ptr.7189
72. Honari M, Shafabakhsh R, Reiter RJ, Mirzaei H, Asemi Z. Resveratrol is a promising agent for colorectal cancer prevention and treatment: focus on molecular mechanisms. *Cancer Cell Int*. 2019;19:180. doi:10.1186/s12935-019-0906-y
73. Brown K, Theofanous D, Britton RG, et al. Resveratrol for the management of human health: how far have we come? A systematic review of resveratrol clinical trials to highlight gaps and opportunities. *Int J Mol Sci*. 2024;25(2):747. doi:10.3390/IJMS25020747/S1
74. Howells LM, Berry DP, Elliott PJ, et al. Phase I randomized, double-blind pilot study of micronized resveratrol (SRT501) in patients with hepatic metastases—Safety, pharmacokinetics, and pharmacodynamics. *Cancer Prev Res*. 2011;4(9):1419–1425.
75. Patel KR, Brown VA, Jones DJ, et al. Clinical pharmacology of resveratrol and its metabolites in colorectal cancer patients. *Cancer Res*. 2010;70(19):7392–7399.
76. Nguyen AV, Martinez M, Stamos MJ, et al. Results of a phase I pilot clinical trial examining the effect of plant-derived resveratrol and grape powder on Wnt pathway target gene expression in colonic mucosa and colon cancer. *Cancer Manag Res*. 2009;1:25–37.
77. Kjaer TN, Omstrup MJ, Poulsen MM, et al. Resveratrol reduces the levels of circulating androgen precursors but has no effect on, testosterone, dihydrotestosterone, PSA levels or prostate volume. A 4-month randomised trial in middle-aged men. *Prostate*. 2015;75(12):1255–1263. doi:10.1002/pros.23006
78. Paller CJ, Rudek MA, Zhou XC, et al. A phase I study of muscadine grape skin extract in men with biochemically recurrent prostate cancer: safety, tolerability, and dose determination. *Prostate*. 2015;75(14):1518–1525. doi:10.1002/pros.23024
79. Popat R, Plesner T, Davies F, et al. A phase 2 study of SRT501 (resveratrol) with bortezomib for patients with relapsed and or refractory multiple myeloma. *Br J Haematol*. 2013;160(5):714–717. doi:10.1111/bjh.12154
80. Zhu W, Qin W, Zhang K, et al. Trans-resveratrol alters mammary promoter hypermethylation in women at increased risk for breast cancer. *Nutr Cancer*. 2012;64(3):393–400. doi:10.1080/01635581.2012.654926
81. Janczewski Ł. Sulforaphane and its bifunctional analogs: synthesis and biological activity. *Molecules*. 2022;27(5):1750. doi:10.3390/MOLECULES27051750
82. Bai Y, Wang X, Zhao S, Ma C, Cui J, Zheng Y. Sulforaphane protects against cardiovascular disease via Nrf2 activation. *Oxid Med Cell Longev*. 2015;2015:407580. doi:10.1155/2015/407580
83. Tian S, Li X, Wang Y, Lu Y. The protective effect of sulforaphane on type II diabetes induced by high-fat diet and low-dosage streptozotocin. *Food Sci Nutr*. 2021;9(2):747–756. doi:10.1002/FSN3.2040
84. Klomparens EA, Ding Y. The neuroprotective mechanisms and effects of sulforaphane. *Brain Circ*. 2019;5(2):74–83. doi:10.4103/BC.BC\_7\_19
85. Houghton CA. The rationale for sulforaphane favourably influencing gut homeostasis and gut–organ dysfunction: a clinician’s hypothesis. *Int J Mol Sci*. 2023;24(17):13448. doi:10.3390/IJMS241713448
86. Jiang X, Liu Y, Ma L, et al. Chemopreventive activity of sulforaphane. *Drug Des Devel Ther*. 2018;12:2905–2913. doi:10.2147/DDDT.SI00534
87. Hao Q, Wang M, Sun NX, et al. Sulforaphane suppresses carcinogenesis of colorectal cancer through the ERK/Nrf2-UDP glucuronosyltransferase 1A metabolic axis activation. *Oncol Rep*. 2020;43(4):1067–1080. doi:10.3892/OR.2020.7495
88. Wang F, Sun Y, Huang X, et al. Sulforaphane inhibits self-renewal of lung cancer stem cells through the modulation of sonic Hedgehog signaling pathway and polyhomeotic homolog 3. *Amb Express*. 2021;11(1):121. doi:10.1186/S13568-021-01281-X
89. Mordecai J, Ullah S, Ahmad I. Sulforaphane and its protective role in prostate cancer: a mechanistic approach. *Int J Mol Sci*. 2023;24(8):6979. doi:10.3390/IJMS24086979
90. Asif Ali M, Khan N, Kaleem N, et al. Anticancer properties of sulforaphane: current insights at the molecular level. *Front Oncol*. 2023;13:1168321. doi:10.3389/FONC.2023.1168321/BIBTEX
91. Jabbarzadeh Kaboli P, Afzalipour Khoshkbejari M, Mohammadi M, et al. Targets and mechanisms of sulforaphane

- derivatives obtained from cruciferous plants with special focus on breast cancer – contradictory effects and future perspectives. *Biomed Pharmacother.* 2020;121:109635. doi:10.1016/J.BIOPHA.2019.109635
92. Su X, Jiang X, Meng L, Dong X, Shen Y, Xin Y. Anticancer activity of sulforaphane: the epigenetic mechanisms and the Nrf2 signaling pathway. *Oxid Med Cell Longev.* 2018;2018:5438179. doi:10.1155/2018/5438179
  93. Tortorella SM, Royce SG, Licciardi Pv., Karagiannis TC. Dietary sulforaphane in cancer chemoprevention: the role of epigenetic regulation and HDAC inhibition. *Antioxid Redox Signal.* 2015;22(16):1382-1424. doi:10.1089/ARS.2014.6097
  94. Wang Y, Wu H, Dong N, et al. Sulforaphane induces S-phase arrest and apoptosis via p53-dependent manner in gastric cancer cells. *Sci Rep.* 2021;11(1):2504. doi:10.1038/s41598-021-81815-2
  95. Kan SF, Wang J, Sun GX. Sulforaphane regulates apoptosis- and proliferation-related signaling pathways and synergizes with cisplatin to suppress human ovarian cancer. *Int J Mol Med.* 2018;42(5):2447-2458. doi:10.3892/IJMM.2018.3860
  96. Parnaud G, Li PF, Cassar G, et al. Mechanism of sulforaphane-induced cell cycle arrest and apoptosis in human colon cancer cells. *Nutr Cancer.* 2004;48(2):198-206. doi:10.1207/S15327914NC4802\_10
  97. Carrasco-Pozo C, Tan KN, Rodriguez T, Avery VM. The molecular effects of sulforaphane and capsaicin on metabolism upon androgen and Tip60 activation of androgen receptor. *Int J Mol Sci.* 2019;20(21):5384. doi:10.3390/IJMS20215384
  98. Bertl E, Bartsch H, Gerhäuser C. Inhibition of angiogenesis and endothelial cell functions are novel sulforaphane-mediated mechanisms in chemoprevention. *Mol Cancer Ther.* 2006;5(3):575-585. doi:10.1158/1535-7163.MCT-05-0324
  99. Ullah MF. Sulforaphane (SFN): an isothiocyanate in a cancer chemoprevention paradigm. *Medicines.* 2015;2(3):141-156. doi:10.3390/MEDICINES2030141
  100. Bessler H, Djaldetti M. Broccoli and human health: immunomodulatory effect of sulforaphane in a model of colon cancer. *Int J Food Sci Nutr.* 2018;69(8):946-953. doi:10.1080/09637486.2018.1439901
  101. Thejass P, Kuttan G. Augmentation of natural killer cell and antibody-dependent cellular cytotoxicity in BALB/c mice by sulforaphane, a naturally occurring isothiocyanate from broccoli through enhanced production of cytokines IL-2 and IFN-gamma. *Immunopharmacol Immunotoxicol.* 2006;28(3):443-457. doi:10.1080/08923970600928049
  102. Palliyaguru DL, Yang L, Chartoumpakis DV, et al. Sulforaphane diminishes the formation of mammary tumors in rats exposed to 17 $\beta$ -estradiol. *Nutrients.* 2020;12(8):2282. doi:10.3390/nu12082282
  103. Amin PJ, Shankar BS. Sulforaphane induces ROS mediated induction of NKG2D ligands in human cancer cell lines and enhances susceptibility to NK cell mediated lysis. *Life Sci.* 2015;126:19-27. doi:10.1016/J.LFS.2015.01.026
  104. Wang Y, Petrikova E, Gross W, et al. Sulforaphane promotes dendritic cell stimulatory capacity through modulation of regulatory molecules, JAK/STAT3- and MicroRNA-signaling. *Front Immunol.* 2020;11:589818. doi:10.3389/FIMMU.2020.589818/PDF
  105. Liang J, Hänsch GM, Hübner K, Samstag Y. Sulforaphane as anticancer agent: a double-edged sword? Tricky balance between effects on tumor cells and immune cells. *Adv Biol Regul.* 2019;71:79-87. doi:10.1016/J.JBIOR.2018.11.006
  106. ElKhalifa D, Al-Ziftawi N, Awaisu A, Alali F, Khalil A. Efficacy and tolerability of sulforaphane in the therapeutic management of cancers: a systematic review of randomized controlled trials. *Front Oncol.* 2023;13:1251895. doi:10.3389/FONC.2023.1251895/FULL
  107. Zhang Z, Garzotto M, Davis EW, et al. Sulforaphane bioavailability and chemopreventive activity in men presenting for biopsy of the prostate gland: a randomized controlled trial. *Nutr Cancer.* 2020;72(1):74-87. doi:10.1080/01635581.2019.1619783
  108. Traka MH, Melchini A, Coode-Bate J, et al. Transcriptional changes in prostate of men on active surveillance after a 12-mo glucoraphanin-rich broccoli intervention—Results from the Effect of Sulforaphane on prostate CAncer PrEvention (ESCAPE) randomized controlled trial. *Am J Clin Nutr.* 2019;109(4):1133-1144. doi:10.1093/ajcn/nqz012
  109. Cipolla BG, Mandron E, Lefort JM, et al. Effect of sulforaphane in men with biochemical recurrence after radical prostatectomy. *Cancer Prev Res.* 2015;8(8):712-719. doi:10.1158/1940-6207.CAPR-14-0459/36746/AM/EFFECT-OF-SULFORAPHANE-IN-MEN-WITH-BIOCHEMICAL
  110. Traka M, Gasper AV, Melchini A, et al. Broccoli consumption interacts with GSTM1 to perturb oncogenic signalling pathways in the prostate. *PLoS One.* 2008;3(7):e2568. doi:10.1371/journal.pone.0002568
  111. Study results | study to evaluate the effect of sulforaphane in broccoli sprout extract on breast tissue | ClinicalTrials.gov. (n.d.). Retrieved August 23, 2024, from: <https://clinicaltrials.gov/study/NCT00982319?tab=results>
  112. Atwell L, Zhang Z, Mori M, Farris P, et al. Sulforaphane bioavailability and chemopreventive activity in women scheduled for breast biopsy. *Cancer Prev Res.* 2015;8:1184. Retrieved August 23, 2024, from: <https://aacrjournals.org/cancerpreventionresearch/article-abstract/8/12/1184/50399>
  113. Tahata S, Singh S, Lin Y, Hahm E, et al. Evaluation of bio-distribution of sulforaphane after administration of oral broccoli sprout extract in melanoma patients with multiple atypical nevi. *Cancer Prev Res.* 2018;11:429. Retrieved August 23, 2024, from: <https://aacrjournals.org/cancerpreventionresearch/article-abstract/11/7/429/275112>
  114. Lozanovski VJ, Polychronidis G, Gross W, et al. Broccoli sprout supplementation in patients with advanced pancreatic cancer is difficult despite positive effects—results from the POWDER pilot study. *Invest New Drugs.* 2020;38(3):776-784. doi:10.1007/S10637-019-00826-Z
  115. Kaiser AE, Baniyadi M, Giansiracusa D, et al. Sulforaphane: a broccoli bioactive phytochemical with cancer

- preventive potential. *Cancers*. 2021;13(19):4796. doi:10.3390/CANCERS13194796
116. Kaufman PB, Duke JA, Briellmann H, Boik J, Hoyt JE. A comparative survey of leguminous plants as sources of the isoflavones, genistein and daidzein: implications for human nutrition and health. *J Altern Complement Med*. 2007;3(1):7-12. doi:10.1089/ACM.1997.3.7
117. Kurzer MS, Xu X. Dietary phytoestrogens. *Annu Rev Nutr*. 1997;17:353-381. doi:10.1146/ANNUREV.NUTR.17.1.353
118. Yu J, Bi X, Yu B, Chen D. Isoflavones: anti-inflammatory benefit and possible caveats. *Nutrients*. 2016;8(6):361. doi:10.3390/NU8060361
119. Gómez-Zorita S, González-Arceo M, Fernández-Quintela A, Eseberri I, Trepiana J, Portillo MP. Scientific evidence supporting the beneficial effects of isoflavones on human health. *Nutrients*. 2020;12(12):1-25. doi:10.3390/NU12123853
120. Yamagata K. Soy isoflavones inhibit endothelial cell dysfunction and prevent cardiovascular disease. *J Cardiovasc Pharmacol*. 2019;74(3):201-209. doi:10.1097/FJC.0000000000000708
121. Maaliki D, Shaito AA, Pintus G, El-Yazbi A, Eid AH. Flavonoids in hypertension: a brief review of the underlying mechanisms. *Curr Opin Pharmacol*. 2019;45:57-65. doi:10.1016/J.COPH.2019.04.014
122. Guo J, Ma J, Cai K, et al. Isoflavones from semen sojæe preparatum improve atherosclerosis and oxidative stress by modulating Nrf2 signaling pathway through estrogen-like effects. *Evid Based Complement Alternat Med*. 2022;2022:4242099. doi:10.1155/2022/4242099
123. Kuryłowicz A. The role of isoflavones in type 2 diabetes prevention and treatment—A narrative review. *Int J Mol Sci*. 2020;22(1):218. doi:10.3390/IJMS22010218
124. Wang S, Wang Y, Pan MH, Ho CT. Anti-obesity molecular mechanism of soy isoflavones: weaving the way to new therapeutic routes. *Food Funct*. 2017;8(11):3831-3846. doi:10.1039/C7FO01094J
125. van der Eecken H, Joniau S, Berghen C, Rans K, de Meerleer G. The use of soy isoflavones in the treatment of prostate cancer: a focus on the cellular effects. *Nutrients*. 2023;15(23):4856. doi:10.3390/NU15234856
126. Sotoca Covaleda AM, van den Berg H, Vervoort J, et al. Influence of cellular ER $\alpha$ /ER $\beta$  ratio on the ER $\alpha$ -agonist induced proliferation of human T47D breast cancer cells. *Toxicol Sci*. 2008;105(2):303-311. doi:10.1093/TOXSCI/KFN141
127. Tuli HS, Tuorkey MJ, Thakral F, et al. Molecular mechanisms of action of genistein in cancer: recent advances. *Front Pharmacol*. 2019;10:1336. doi:10.3389/FPHAR.2019.01336/BIBTEX
128. Liu H, Lee G, Lee JI, Ahn TG, Kim SA. Effects of genistein on anti-tumor activity of cisplatin in human cervical cancer cell lines. *Obstet Gynecol Sci*. 2019;62(5):322-328. doi:10.5468/OGS.2019.62.5.322
129. Sahin K, Yenice E, Bilir B, et al. Genistein prevents development of spontaneous ovarian cancer and inhibits tumor growth in hen model. *Cancer Prev Res*. 2019;12(3):135-146. doi:10.1158/1940-6207.CAPR-17-0289/254530/AM/GENISTEIN-PREVENTS-DEVELOPMENT-OF-SPONTANEOUS
130. Xu H, Ma H, Zha L, Li Q, Pan H, Zhang L. Genistein promotes apoptosis of lung cancer cells through the IMPDH2/AKT1 pathway. *Am J Transl Res*. 2022;14(10):7040-7051. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC9641449/>
131. Li E, Zhang TAO, Sun X, et al. Sonic hedgehog pathway mediates genistein inhibition of renal cancer stem cells. *Oncol Lett*. 2019;18(3):3081-3091. doi:10.3892/OL.2019.10657
132. Record IR, Dreosti IE, McInerney JK. The antioxidant activity of genistein in vitro. *J Nutr Biochem*. 1995;6(9):481-485. doi:10.1016/0955-2863(95)00076-C
133. Yu X, Zhu J, Mi M, Chen W, Pan Q, Wei M. Anti-angiogenic genistein inhibits VEGF-induced endothelial cell activation by decreasing PTK activity and MAPK activation. *Med Oncol*. 2012;29(1):349-357. doi:10.1007/S12032-010-9770-2
134. Alatawi FS, Faridi U. Anticancer and anti-metastasis activity of 1,25 dihydroxycholecalciferols and genistein in MCF-7 and MDA-MB-231 breast cancer cell lines. *Heliyon*. 2023;9(11):e21975. doi:10.1016/J.HELIYON.2023.E21975
135. Choi EJ, Kim GH. Antiproliferative activity of daidzein and genistein may be related to ER $\alpha$ /c-erbB-2 expression in human breast cancer cells. *Mol Med Rep*. 2013;7(3):781-784. doi:10.3892/MMR.2013.1283
136. Farhan M, Oirdi ME, Aatif M, Nahvi I, Muteeb G, Alam MW. Soy isoflavones induce cell death by copper-mediated mechanism: understanding its anticancer properties. *Molecules*. 2023;28(7):2925. doi:10.3390/molecules28072925
137. Ullah MF, Ahmad A, Zubair H, et al. Soy isoflavone genistein induces cell death in breast cancer cells through mobilization of endogenous copper ions and generation of reactive oxygen species. *Mol Nutr Food Res*. 2011;55(4):553-559. doi:10.1002/MNFR.201000329
138. Azadbakht L, Kimiagar M, Mehrabi Y, Esmaillzadeh A, Hu FB, Willett WC. Soy consumption, markers of inflammation, and endothelial function: a cross-over study in postmenopausal women with the metabolic syndrome. *Diabetes Care*. 2007;30(4):967-973. doi:10.2337/DC06-2126
139. Yoon GA, Park S. Antioxidant action of soy isoflavones on oxidative stress and antioxidant enzyme activities in exercised rats. *Nutr Res Pract*. 2014;8(6):618-624. doi:10.4162/NRP.2014.8.6.618
140. Sarkar FH, Li Y. Mechanisms of cancer chemoprevention by soy isoflavone genistein. *Cancer Metastasis Rev*. 2002;21(3-4):265-280. doi:10.1023/A:1021210910821
141. Liu Y, Zou T, Wang S, et al. Genistein-induced differentiation of breast cancer stem/progenitor cells through a paracrine mechanism. *Int J Oncol*. 2016;48(3):1063-1072. doi:10.3892/IJO.2016.3351
142. Kim I. Current perspectives on the beneficial effects of soybean isoflavones and their metabolites for humans. *Antioxidants*. 2021;10(7):1064. doi:10.3390/antiox10071064
143. Qin J, Chen JX, Zhu Z, Teng JA. Genistein inhibits human colorectal cancer growth and suppresses MiR-95, Akt and

- SGK1. *Cell Physiol Biochem*. 2015;35(5):2069-2077. doi:10.1159/000374013
144. Li K, Hong S, Lin S, Chen K. Genistein inhibits the proliferation, migration and invasion of the squamous cell carcinoma cells via inhibition of MEK/ERK and JNK signalling pathways. *J BUON*. 2020;25(2):1172-1177.
145. Zhang Q, Cao S, Wang Q, et al. Genistein inhibits nasopharyngeal cancer stem cells through sonic hedgehog signaling. *Phytother Res*. 2019;33(10):2783-2791. doi:10.1002/ptr.6464
146. Kim C, Lee S, Yang WM, et al. Formononetin-induced oxidative stress abrogates the activation of STAT3/5 signaling axis and suppresses the tumor growth in multiple myeloma pre-clinical model. *Cancer Lett*. 2018;431:123-141. doi:10.1016/j.canlet.2018.05.038
147. Yu B, Tang DZ, Li SY, Wu Y, Chen M, Chen M. Daidzein promotes proliferation and differentiation in osteoblastic OCT1 cells via activation of the BMP-2/Smads pathway. *Pharmazie*. 2017;72(1):35-40. doi:10.1691/PH.2017.6502
148. Lesinski GB, Reville PK, Mace TA, et al. Consumption of soy isoflavone enriched bread in men with prostate cancer is associated with reduced proinflammatory cytokines and immunosuppressive cells. *Cancer Prev Res*. 2015;8(11):1036-1044. doi:10.1158/1940-6207.CAPR-14-0464/36723/AM/CONSUMPTION-OF-SOY-ISOFILAVONE-ENRICHED-BREAD-IN
149. Yang J, Shen H, Mi M, Qin Y. Isoflavone consumption and risk of breast cancer: an updated systematic review with meta-analysis of observational studies. *Nutrients*. 2023;15(10):2402. doi:10.3390/NU15102402/S1
150. van Die MD, Bone KM, Williams SG, Pirotta MV. Soy and soy isoflavones in prostate cancer: a systematic review and meta-analysis of randomized controlled trials. *BJU Int*. 2014;113(5b):E119-E130. doi:10.1111/BJU.12435
151. DeVere White RW, Tsodikov A, Stapp EC, Soares SE, Fujii H, Hackman RM. Effects of a high dose, aglycone-rich soy extract on prostate-specific antigen and serum isoflavone concentrations in men with localized prostate cancer. *Nutr Cancer*. 2010;62(8):1036-1043. doi:10.1080/01635581.2010.492085
152. Kumar NB, Krischer JP, Allen K, et al. A phase II randomized, placebo-controlled clinical trial of purified isoflavones in modulating steroid hormones in men diagnosed with localized prostate cancer. *Nutr Cancer*. 2007;59(2):163-168. doi:10.1080/01635580701432678
153. Kumar N, Kang L, Pow-Sang J, et al. Results of a randomized phase I dose-finding trial of several doses of isoflavones in men with localized prostate cancer: administration prior to radical. *J Soc Integr Oncol*. 2010;8:3-13. Retrieved August 23, 2024, from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3277948/>
154. Lazarevic B, Boezelijn G, Diep LM, et al. Efficacy and safety of short-term genistein intervention in patients with localized prostate cancer prior to radical prostatectomy: a randomized, placebo-controlled, double-blind phase 2 clinical trial. *Nutr Cancer*. 2011;63(6):889-898. doi:10.1080/01635581.2011.582221
155. Miyanaga N, Akaza H, Hinotsu S, et al. Prostate cancer chemoprevention study: an investigative randomized control study using purified isoflavones in men with rising prostate-specific antigen. *Cancer Sci*. 2012;103(1):125-130. doi:10.1111/j.1349-7006.2011.02120.x
156. Dalais FS, Meliala A, Wattanapenpaiboon N, et al. Effects of a diet rich in phytoestrogens on prostate-specific antigen and sex hormones in men diagnosed with prostate cancer. *Urology*. 2004;64(3):510-515. doi:10.1016/j.urology.2004.04.009
157. Hamilton-Reeves JM, Rebello SA, Thomas W, Slaton JW, Kurzer MS. Isoflavone-rich soy protein isolate suppresses androgen receptor expression without altering estrogen receptor-beta expression or serum hormonal profiles in men at high risk of prostate cancer. *J Nutr*. 2007;137(7):1769-1775. doi:10.1093/jn/137.7.1769
158. Hamilton-Reeves JM, Rebello SA, Thomas W, Kurzer MS, Slaton JW. Effects of soy protein isolate consumption on prostate cancer biomarkers in men with HGPIN, ASAP, and low-grade prostate cancer. *Nutr Cancer*. 2007;60(1):7-13. doi:10.1080/01635580701586770
159. Kumar NB, Cantor A, Allen K, et al. The specific role of isoflavones in reducing prostate cancer risk. *Prostate*. 2004;59(2):141-147. doi:10.1002/PROS.10362
160. Jówko E. Green tea catechins and sport performance. *Antioxidant Sport Nutrition*. 2015;123-140. doi:10.1201/b17442-8
161. Bernatoniene J, Kopustinskiene DM. The role of catechins in cellular responses to oxidative stress. *Molecules*. 2018;23(4):965. doi:10.3390/MOLECULES23040965
162. Maternia J, Miszczyk K, Maj P, et al. Effect of green tea on the cardiovascular system. *J Educ Health Sport*. 2023;46(1):201-215. doi:10.12775/JEHS.2023.46.01.014
163. Wen L, Wu D, Tan X, et al. The role of catechins in regulating diabetes: an update review. *Nutrients*. 2022;14(21):4681. doi:10.3390/nu14214681
164. Basu T, Selman A, Reddy AP, Reddy PH. Current status of obesity: protective role of catechins. *Antioxidants*. 2023;12(2):474. doi:10.3390/antiox12020474
165. Fan FY, Sang LX, Jiang M, McPhee DJ. Catechins and their therapeutic benefits to inflammatory bowel disease. *Molecules*. 2017;22(3):484. doi:10.3390/MOLECULES22030484
166. Musial C, Kuban-Jankowska A, Gorska-Ponikowska M. Beneficial properties of green tea catechins. *Int J Mol Sci*. 2020;21(5):1744. doi:10.3390/IJMS21051744
167. Wang C, Bai M, Sun Z, et al. Epigallocatechin-3-gallate and cancer: focus on the role of microRNAs. *Cancer Cell Int*. 2023;23(1):241. doi:10.1186/S12935-023-03081-8/TABLES/2
168. Zhong Z, Dong Z, Yang L, Chen X, Gong Z. Inhibition of proliferation of human lung cancer cells by green tea catechins is mediated by upregulation of let-7. *Exp Ther Med*. 2012;4(2):267-272. doi:10.3892/ETM.2012.580
169. Yang C, Du W, Yang D. Inhibition of green tea polyphenol EGCG(-)-epigallocatechin-3-gallate on the proliferation of gastric cancer cells by suppressing canonical wnt/ $\beta$ -catenin signalling pathway. *Int J Food Sci Nutr*. 2016;67(7):818-827. doi:10.1080/09637486.2016.1198892



170. Wang J, Xie Y, Feng Y, et al. (-)-Epigallocatechingallate induces apoptosis in B lymphoma cells via caspase-dependent pathway and Bcl-2 family protein modulation. *Int J Oncol.* 2015;46(4):1507-1515. doi:10.3892/IJO.2015.2869
171. della Via FI, Alvarez MC, Basting RT, Saad STO. The effects of green tea catechins in hematological malignancies. *Pharmaceuticals.* 2023;16(7):1021. doi:10.3390/PH16071021
172. Li XX, Liu C, Dong SL, et al. Anticarcinogenic potentials of tea catechins. *Front Nutr.* 2022;9:1060783. doi:10.3389/FNUT.2022.1060783/BIBTEX
173. Oh JW, Muthu M, Pushparaj SSC, Gopal J. Anticancer therapeutic effects of green tea catechins (GTCs) when integrated with antioxidant natural components. *Molecules.* 2023;28(5):2151. doi:10.3390/MOLECULES28052151
174. Lambert JD, Elias RJ. The antioxidant and pro-oxidant activities of green tea polyphenols: a role in cancer prevention. *Arch Biochem Biophys.* 2010;501(1):65-72. doi:10.1016/J.ABB.2010.06.013
175. Wang Y, Qi H, Liu Y, et al. The double-edged roles of ROS in cancer prevention and therapy. *Theranostics.* 2021;11(10):4839-4857. doi:10.7150/thno.56747
176. Takada M, Nakamura Y, Koizumi T, et al. Suppression of human pancreatic carcinoma cell growth and invasion by epigallocatechin-3-gallate. *Pancreas.* 2002;25(1):45-48.
177. Das A, Banik N, Ray SK. Flavonoids activated caspases for apoptosis in human glioblastoma T98G and U87MG cells but not in human normal astrocytes. *Cancer.* 2010;116(1):164-176. doi:10.1002/cncr.24699
178. Li G, Chen Y, Hou Z, Xiao H, Jin H. Pro-oxidative activities and dose-response relationship of (-)-epigallocatechin-3-gallate in the inhibition of lung cancer cell growth: a comparative study in vivo. *Carcinogenesis.* 2010;31:902. Retrieved August 20, 2024, from: <https://academic.oup.com/carcin/article-abstract/31/5/902/2477437>
179. Okabe S, Ochiai Y, Aida M, et al. Mechanistic aspects of green tea as a cancer preventive: effect of components on human stomach cancer cell lines. *Jpn J Cancer Res.* 1999;90(7):733-739. doi:10.1111/J.1349-7006.1999.TB00808.X
180. de Almeida LGN, Thode H, Eslambolchi Y, et al. Matrix metalloproteinases: from molecular mechanisms to physiology, pathophysiology, and pharmacology. *Pharmacol Rev.* 2022;74(3):712-768. doi:10.1124/PHARMREV.121.000349
181. Zhang Y, Owusu L, Duan W, et al. Anti-metastatic and differential effects on protein expression of epigallocatechin-3-gallate in HCCLM6 hepatocellular carcinoma cells. *Int J Mol Med.* 2013;32(4):959-964. doi:10.3892/IJMM.2013.1446
182. Saeki K, Hayakawa S, Nakano S, et al. In vitro and in silico studies of the molecular interactions of epigallocatechin-3-O-gallate (EGCG) with proteins that explain the health benefits of green tea. *Molecules.* 2018;23(6):1295. doi:10.3390/MOLECULES23061295
183. Luo KW, Wei C, Lung WY, et al. EGCG inhibited bladder cancer SW780 cell proliferation and migration both in vitro and in vivo via down-regulation of NF- $\kappa$ B and MMP-9. *J Nutr Biochem.* 2017;41:56-64. doi:10.1016/J.JNUTBIO.2016.12.004
184. Won HR, Lee P, Oh SR, Kim YM. Epigallocatechin-3-Gallate suppresses the expression of TNF- $\alpha$ -induced MMP-1 via MAPK/ERK signaling pathways in human dermal fibroblasts. *Biol Pharm Bull.* 2021;44(1):18-24. doi:10.1248/BPB.B20-00304
185. Tanabe H, Suzuki T, Ohishi T, Isemura M, Nakamura Y, Unno K. Effects of epigallocatechin-3-gallate on matrix metalloproteinases in terms of its anticancer activity. *Molecules.* 2023;28(2):525. doi:10.3390/MOLECULES28020525
186. Liu S, Xu ZL, Sun L, et al. (-)-Epigallocatechin-3-gallate induces apoptosis in human pancreatic cancer cells via PTEN. *Mol Med Rep.* 2016;14(1):599-605. doi:10.3892/MMR.2016.5277
187. Kim J, Zhang X, Rieger-Christ KM, et al. Suppression of Wnt signaling by the green tea compound (-)-epigallocatechin 3-gallate (EGCG) in invasive breast cancer cells. Requirement of the transcriptional repressor HBP1. *J Biol Chem.* 2006;281(16):10865-10875. doi:10.1074/JBC.M513378200
188. Ganeshpurkar A, Saluja A. Immunomodulatory effect of rutin, catechin, and hesperidin on macrophage function. *Indian J Biochem Biophys.* 2020;57(1):58-63. doi:10.56042/IJBB.V57I1.31775
189. Mackenzie GG, Carrasquedo F, Delfino JM, Keen CL, Fraga CG, Oteiza PI. Epicatechin, catechin, and dimeric procyanidins inhibit PMA-induced NF-kappaB activation at multiple steps in Jurkat T cells. *Faseb J.* 2004;18(1):167-169. doi:10.1096/fj.03-0402fje
190. Rawangkan A, Wongsirisin P, Namiki K, et al. Green tea catechin is an alternative immune checkpoint inhibitor that inhibits PD-L1 expression and lung tumor growth. *Molecules.* 2018;23(8):2071. doi:10.3390/molecules23082071
191. Borah G, Bharali MK. Green tea catechins in combination with irinotecan attenuates tumorigenesis and treatment-associated toxicity in an inflammation-associated colon cancer mice model. *J Egypt Natl Canc Inst.* 2021;33(1):17. doi:10.1186/s43046-021-00074-4
192. Yang G, Zheng W, Xiang YB, et al. Green tea consumption and colorectal cancer risk: a report from the Shanghai Men's Health Study. *Carcinogenesis.* 2011;32(11):1684-1688. doi:10.1093/CARCIN/BGR186
193. Guo Y, Zhi F, Chen P, et al. Green tea and the risk of prostate cancer: a systematic review and meta-analysis. *Medicine.* 2017;96(13):e6426. doi:10.1097/MD.00000000000006426
194. Study Details | Clinical Trial of Green Tea Catechins in Men on Active Surveillance | ClinicalTrials.gov. n.d. Retrieved August 24, 2024, from: <https://clinicaltrials.gov/study/NCT04300855>
195. Kumar N, Pow-Sang J, Egan K, et al. Randomized, placebo-controlled trial of green tea catechins for prostate cancer prevention. *Cancer Prev Res.* 2015;8:879. Retrieved August 24, 2024, from: <https://aacrjournals.org/cancerpreventionresearch/article-abstract/8/10/879/113064>
196. Bettuzzi S, Brausi M, Rizzi F, Castagnetti G, Peracchia G, Corti A. Chemoprevention of human prostate cancer by oral administration of green tea catechins in volunteers

- with high-grade prostate intraepithelial neoplasia: preliminary report from a one-year proof-of-principle study. *Cancer Res.* 2006;66:1234. Retrieved August 24, 2024, from: <https://aacrjournals.org/cancerres/article-abstract/66/2/1234/526429>
197. Brausi M, Rizzi F, Bettuzzi S. Chemoprevention of human prostate cancer by green tea catechins: two years later. A follow-up update. *Eur Urol.* 2008;54:472-473. Retrieved August 24, 2024, from: <https://www.compagniadelleerbe.com/inglese/assets/brausi-et-al-eur-urol-2008.pdf>
  198. Aghababaei F, Hadidi M. Recent advances in potential health benefits of quercetin. *Pharmaceuticals.* 2023;16(7):1020. doi:10.3390/PH16071020
  199. Nazari-Khanamiri F, Ghasemnejad-Berenji M. Quercetin and heart health: from molecular pathways to clinical findings. *J Food Biochem.* 2022;2023(1):1-9. doi:10.1155/2023/8459095
  200. Ansari P, Choudhury ST, Seidel V, et al. Therapeutic potential of quercetin in the management of type-2 diabetes mellitus. *Life.* 2022;12(8):1146. doi:10.3390/life12081146
  201. Wang Y, Li Z, He J, Zhao Y. Quercetin regulates lipid metabolism and fat accumulation by regulating inflammatory responses and glycometabolism pathways: a review. *Nutrients.* 2023;16(8):1102. doi:10.3390/nu16081102
  202. Wang R, Yang L, Li S, et al. Quercetin inhibits breast cancer stem cells via downregulation of aldehyde dehydrogenase 1A1 (ALDH1A1), chemokine receptor type 4 (CXCR4), mucin 1 (MUC1), and epithelial cell adhesion molecule (EpCAM). *Med Sci Monit.* 2018;24:412-420. doi:10.12659/MSM.908022
  203. Sethi G, Rath P, Chauhan A, et al. Apoptotic mechanisms of quercetin in liver cancer: recent trends and advancements. *Pharmaceutics.* 2023;15(2):712. doi:10.3390/PHARMACEUTICS15020712
  204. Bhatiya M, Pathak S, Jothimani G, Duttaroy AK, Banerjee A. A comprehensive study on the anti-cancer effects of quercetin and its epigenetic modifications in arresting progression of colon cancer cell proliferation. *Arch Immunol Ther Exp.* 2023; 71(1):6-16. doi:10.1007/S00005-023-00669-W/FIGURES/7
  205. Ali A, Kim MJ, Kim MY, et al. Quercetin induces cell death in cervical cancer by reducing O-GlcNAcylation of adenosine monophosphate-activated protein kinase. *Anat Cell Biol.* 2018; 51(4):274-283. doi:10.5115/ACB.2018.51.4.274
  206. Lotfi N, Yousefi Z, Golabi M, et al. The potential anti-cancer effects of quercetin on blood, prostate and lung cancers: an update. *Front Immunol.* 2023;14:1077531. doi:10.3389/FIMMU.2023.1077531/BIBTEX
  207. Asgharian P, Tazekand AP, Hosseini K, et al. Potential mechanisms of quercetin in cancer prevention: focus on cellular and molecular targets. *Cancer Cell Int.* 2022;22:257. doi:10.1186/s12935-022-02677-w
  208. Boots AW, Haenen GRMM, Bast A. Health effects of quercetin: from antioxidant to nutraceutical. *Eur J Pharmacol.* 2008;585(2-3):325-337. doi:10.1016/J.EJPHAR.2008.03.008
  209. Baba RA, Mir HA, Mokhdomi TA, Bhat HF, Ahmad A, Khanday FA. Quercetin suppresses ROS production and migration by specifically targeting Rac1 activation in gliomas. *Front Pharmacol.* 2024;15:1318797. doi:10.3389/fphar.2024.1318797
  210. García-Mediavilla V, Crespo I, Collado PS, et al. The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. *Eur J Pharmacol.* 2007;557(2-3): 221-229. doi:10.1016/J.EJPHAR.2006.11.014
  211. Zhang J, Li H, Wang W, Li H. Assessing the anti-inflammatory effects of quercetin using network pharmacology and in vitro experiments. *Exp Ther Med.* 2022;23(4):301. doi:10.3892/ETM.2022.11230
  212. Clemente-Soto AF, Salas-Vidal E, Milan-Pacheco C, Sánchez-Carranza JN, Peralta-Zaragoza O, González-Maya L. Quercetin induces G2 phase arrest and apoptosis with the activation of p53 in an E6 expression-independent manner in HPV-positive human cervical cancer-derived cells. *Mol Med Rep.* 2019;19(3):2097-2106. doi:10.3892/MMR.2019.9850
  213. Ha EJ, Kim KY, Kim CE, Jun DY, Kim YH. Enhancement of quercetin-induced apoptosis by cotreatment with autophagy inhibitor is associated with augmentation of BAK-dependent mitochondrial pathway in Jurkat T cells. *Oxid Med Cell Longev.* 2019;2019:7989276. doi:10.1155/2019/7989276
  214. Wang R, Zhang H, Wang Y, Song F, Yuan Y. Inhibitory effects of quercetin on the progression of liver fibrosis through the regulation of NF-κB, p38 MAPK, and Bcl-2/Bax signaling. *Int Immunopharmacol.* 2017;47:126-133. doi:10.1016/J.INTIMP.2017.03.029
  215. Shahbaz M, Naeem H, Momal U, et al. Anticancer and apoptosis inducing potential of quercetin against a wide range of human malignancies. *Int J Food Prop.* 2023;26(1):2590-2626. doi:10.1080/10942912.2023.2252619
  216. Nguyen LT, Lee YH, Sharma AR, et al. Quercetin induces apoptosis and cell cycle arrest in triple-negative breast cancer cells through modulation of Foxo3a activity. *Korean J Physiol Pharmacol.* 2017;21(2):205-213. doi:10.4196/KJPP.2017.21.2.205
  217. Pratheeshkumar P, Budhraj A, Son YO, et al. Quercetin inhibits angiogenesis mediated human prostate tumor growth by targeting VEGFR-2 regulated AKT/mTOR/P70S6K signaling pathways. *PLoS One.* 2012;7(10):e47516. doi:10.1371/JOURNAL.PONE.0047516
  218. Balakrishnan S, Bhat FA, Raja Singh P, et al. Gold nanoparticle-conjugated quercetin inhibits epithelial-mesenchymal transition, angiogenesis and invasiveness via EGFR/VEGFR-2-mediated pathway in breast cancer. *Cell Prolif.* 2016;49(6):678-697. doi:10.1111/CPR.12296
  219. Chang JH, Lai SL, Chen WS, et al. Quercetin suppresses the metastatic ability of lung cancer through inhibiting Snail-dependent Akt activation and Snail-independent ADAM9 expression pathways. *Biochim Biophys Acta Mol Cell Res.* 2017;1864(10):1746-1758. doi:10.1016/J.BBAMCR.2017.06.017

220. Cho SY, Park SJ, Kwon MJ, et al. Quercetin suppresses proinflammatory cytokines production through MAP kinases and NF- $\kappa$ B pathway in lipopolysaccharide-stimulated macrophage. *Mol Cell Biochem*. 2003;243(1–2):153–160. doi:10.1023/A:1021624520740/METRICS
221. Hasyam AMO, Nor NM, Adnan LHM, et al. Effects of apigenin, luteolin, and quercetin on the natural killer (NK-92) cells proliferation: a potential role as immunomodulator. *Sains Malays*. 2021;50(3):821–828. doi:10.17576/JSM-2021-5003-22
222. Bae JH, Kim JY, Kim MJ, et al. Quercetin enhances susceptibility to NK cell-mediated lysis of tumor cells through induction of NKG2D ligands and suppression of HSP70. *J Immunother*. 2010;33(4):391–401. doi:10.1097/CJI.0B013E3181D32F22
223. Men K, Duan X, Wei X-W, et al. Nanoparticle-delivered quercetin for cancer therapy. *Anti Cancer Agents Med Chem*. 2014;14(6):826–832. doi:10.2174/1871520614666140521122932
224. Neuwirthová J, Gál B, Smilek P, Urbánková P. Potential of the flavonoid quercetin to prevent and treat cancer - current status of research. *Klin Onkol*. 2018;31(3):184–190. doi:10.14735/AMKO2018184
225. Zhang J, Shen L, Li X, Song W, Liu Y, Huang L. Nanoformulated codelivery of quercetin and alantolactone promotes an antitumor response through synergistic immunogenic cell death for microsatellite-stable colorectal cancer. *ACS Nano*. 2019;13:12511–12524. doi:10.1021/ACS.NANO.9B02875
226. McCann SE, Ambrosone CB, Moysich KB, et al. Intakes of selected nutrients, foods, and phytochemicals and prostate cancer risk in Western New York. *Nutr Cancer*. 2005;53:33–41.
227. Tayyem RF, Heath DD, Al-Delaimy WK, Rock CL. Curcumin content of turmeric and curry powders. *Nutr Cancer*. 2006;55(2):126–131. doi:10.1207/S15327914NC5502\_2
228. Jakubczyk K, Drużga A, Katarzyna J, Skonieczna-żydecka K. Antioxidant potential of curcumin—A meta-analysis of randomized clinical trials. *Antioxidants*. 2020;9(11):1–13. doi:10.3390/ANTIOX9111092
229. Peng Y, Ao M, Dong B, et al. Anti-inflammatory effects of curcumin in the inflammatory diseases: status, limitations and countermeasures. *Drug Des Devel Ther*. 2021;15:4503–4525. doi:10.2147/DDDT.S327378
230. Gupta SC, Patchva S, Aggarwal BB. Therapeutic roles of curcumin: lessons learned from clinical trials. *AAPS J*. 2013;15(1):195–218. doi:10.1208/S12248-012-9432-8
231. Johnson IT, Williamson G, Musk SRR. Anticarcinogenic factors in plant foods: a new class of nutrients? *Nutr Res Rev*. 1994;7(1):175–204. doi:10.1079/NRR19940011
232. Hu S, Xu Y, Meng L, Huang L, Sun H. Curcumin inhibits proliferation and promotes apoptosis of breast cancer cells. *Exp Ther Med*. 2018;16(2):1266–1272. doi:10.3892/ETM.2018.6345
233. Jin H, Qiao F, Wang Y, Xu Y, Shang Y. Curcumin inhibits cell proliferation and induces apoptosis of human non-small cell lung cancer cells through the upregulation of miR-192-5p and suppression of PI3K/Akt signaling pathway. *Oncol Rep*. 2015;34(5):2782–2789. doi:10.3892/OR.2015.4258
234. Dorai T, Gehani N, Katz A. Therapeutic potential of curcumin in human prostate cancer-I. curcumin induces apoptosis in both androgen-dependent and androgen-independent prostate cancer cells. *Prostate Cancer Prostatic Dis*. 2000;3(2):84–93. doi:10.1038/SJ.PCAN.4500399
235. Bojko A, Cierniak A, Adameczyk A, Ligeza J. Modulatory effects of curcumin and tyrphostins (AG494 and AG1478) on growth regulation and viability of LN229 human brain cancer cells. *Nutr Cancer*. 2015;67(7):1170–1182. doi:10.1080/01635581.2015.1073764
236. Zhao Z, Li C, Xi H, Gao Y, Xu D. Curcumin induces apoptosis in pancreatic cancer cells through the induction of forkhead box O1 and inhibition of the PI3K/Akt pathway. *Mol Med Rep*. 2015;12(4):5415–5422. doi:10.3892/MMR.2015.4060
237. Jahanbakhshi F, Maleki Dana P, Badehnoosh B, et al. Curcumin anti-tumor effects on endometrial cancer with focus on its molecular targets. *Cancer Cell Int*. 2021;21(1):120. doi:10.1186/S12935-021-01832-Z/FIGURES/1
238. Yang J, Huang Y, Zhou D, et al. Effects and mechanisms of curcumin for the prevention and management of cancers: an updated review. *Antioxidants*. 2022;11(8):1481. doi:10.3390/antiox11081481
239. Yu C, Yang B, Najafi M. Targeting of cancer cell death mechanisms by curcumin: implications to cancer therapy. *Basic Clin Pharmacol Toxicol*. 2021;129(6):397–415. doi:10.1111/bcpt.13648
240. Menon VP, Sudheer AR. Antioxidant and anti-inflammatory properties of curcumin. *Adv Exp Med Biol*. 2007;595:105–125. doi:10.1007/978-0-387-46401-5\_3
241. Chiorcea-Paquim AM. Electrochemical sensing of curcumin: a review. *Antioxidants*. 2023;12(12):2029. doi:10.3390/ANTIOX12122029
242. Li N, Liu TH, Yu JZ, et al. Curcumin and curcuminol inhibit NF- $\kappa$ B and TGF- $\beta$ 1/smads signaling pathways in CSE-treated RAW246.7 cells. *Evid Based Complement Alternat Med*. 2019;2019:3035125. doi:10.1155/2019/3035125
243. Zhou GZ, Li AF, Sun YH, Sun GC. A novel synthetic curcumin derivative MHMM-41 induces ROS-mediated apoptosis and migration blocking of human lung cancer cells A549. *Biomed Pharmacother*. 2018;103:391–398. doi:10.1016/J.BIOPHA.2018.04.086
244. Tian S, Liao L, Zhou Q, et al. Curcumin inhibits the growth of liver cancer by impairing myeloid-derived suppressor cells in murine tumor tissues. *Oncol Lett*. 2021;21(4):286. doi:10.3892/OL.2021.12547
245. Lin YG, Kunnumakkara AB, Nair A, et al. Curcumin inhibits tumor growth and angiogenesis in ovarian carcinoma by targeting the nuclear factor-kappaB pathway. *Clin Cancer Res*. 2007;13(11):3423–3430. doi:10.1158/1078-0432.CCR-06-3072
246. Borges GA, Elias ST, Amorim B, et al. Curcumin downregulates the PI3K-AKT-mTOR pathway and inhibits growth

- and progression in head and neck cancer cells. *Phytother Res.* 2020;34(12):3311-3324. doi:10.1002/PTR.6780
247. Ohnishi Y, Sakamoto T, Zhengguang L, et al. Curcumin inhibits epithelial-mesenchymal transition in oral cancer cells via c-Met blockade. *Oncol Lett.* 2020;19(6):4177-4182. doi:10.3892/OL.2020.11523
  248. Zhang Z, Chen H, Xu C, et al. Curcumin inhibits tumor epithelial-mesenchymal transition by downregulating the Wnt signaling pathway and upregulating NKD2 expression in colon cancer cells. *Oncol Rep.* 2016;35(5):2615-2623. doi:10.3892/OR.2016.4669
  249. Liu H, Mao Y, Xia B, et al. Curcumin inhibits proliferation and epithelial-mesenchymal transition in lens epithelial cells through multiple pathways. *BioMed Res Int.* 2020;2020:6061894. doi:10.1155/2020/6061894
  250. Chen T, Yang C, Xi Z, Chen F, Li H. Reduced caudal type homeobox 2 (CDX2) promoter methylation is associated with curcumin's suppressive effects on epithelial-mesenchymal transition in colorectal cancer cells. *Med Sci Monit.* 2020;26:e926443. doi:10.12659/MSM.926443
  251. Gorabi AM, Razi B, Aslani S, et al. Effect of curcumin on proinflammatory cytokines: a meta-analysis of randomized controlled trials. *Cytokine.* 2021;143:155541. doi:10.1016/j.cyto.2021.155541
  252. Gilmore TD. Introduction to NF-kappaB: players, pathways, perspectives. *Oncogene.* 2006;25(51):6680-6684. doi:10.1038/sj.onc.1209954
  253. Shih K, Chan H, Wu C, Chuang H. Curcumin enhances the abscopal effect in mice with colorectal cancer by acting as an immunomodulator. *Pharmaceutics.* 2023;15(5):1519. doi:10.3390/pharmaceutics15051519
  254. Mukherjee S, Baidoo JNE, Fried A, Banerjee P. Using curcumin to turn the innate immune system against cancer. *Biochem Pharmacol.* 2020;176:113824. doi:10.1016/J.BCP.2020.113824
  255. Paul S, Sa G. Curcumin as an adjuvant to cancer immunotherapy. *Front Oncol.* 2021;11:675923. doi:10.3389/FONC.2021.675923/PDF
  256. Farghadani R, Naidu R. Curcumin as an enhancer of therapeutic efficiency of chemotherapy drugs in breast cancer. *Int J Mol Sci.* 2022;23(4):2144. doi:10.3390/IJMS23042144
  257. de Waure C, Bertola C, Baccarini G, Chiavarini M, Mancuso C. Exploring the contribution of curcumin to cancer therapy: a systematic review of randomized controlled trials. *Pharmaceutics.* 2023;15(4):1275. doi:10.3390/PHARMACEUTICS15041275
  258. Gunther JR, Chadha AS, Guha S, et al. A phase II randomized double blinded trial evaluating the efficacy of curcumin with pre-operative chemoradiation for rectal cancer. *J Gastrointest Oncol.* 2022;13(6):2938-2950. doi:10.21037/JGO-22-259
  259. Santosa D, Suharti C, Riwanto I, et al. Curcumin as adjuvant therapy to improve remission in myeloma patients: a pilot randomized clinical trial. *Caspian J Intern Med.* 2022;13(2):375-384.
  260. Passildas-Jahanmohan J, Eymard J-C, Pouget M, et al. Multicenter randomized phase II study comparing docetaxel plus curcumin versus docetaxel plus placebo in first-line treatment of metastatic castration-resistant. *Cancer Med.* 2021;10(7):2332-2340. doi:10.1002/cam4.3806
  261. Saghateluyan T, Tananyan A, Janoyan N, et al. Efficacy and safety of curcumin in combination with paclitaxel in patients with advanced, metastatic breast cancer: a comparative, randomized, double-blind, placebo-controlled clinical trial. *Phytomedicine.* 2020;70:153218. doi:10.1016/j.phymed.2020.153218
  262. Howells LM, Iwujii COO, Irving GRB, et al. Curcumin combined with FOLFOX chemotherapy is safe and tolerable in patients with metastatic colorectal cancer in a randomized phase IIa trial. *J Nutr.* 2019;149(7):1133-1139. doi:10.1093/jn/nxz029
  263. Choi YH, Han DH, Kim SW, et al. A randomized, double-blind, placebo-controlled trial to evaluate the role of curcumin in prostate cancer patients with intermittent androgen deprivation. *Prostate.* 2019;79(6):614-621. doi:10.1002/pros.23766
  264. Kuriakose MA, Ramdas K, Dey B, et al. A randomized double-blind placebo-controlled phase IIB trial of curcumin in oral leukoplakia. *Cancer Prev Res.* 2016;9(8):683-691. doi:10.1158/1940-6207.CAPR-15-0390
  265. Nabil-Adam A, Elnosary ME, Ashour ML, et al. Flavonoids biosynthesis in plants as a defense mechanism: role and function concerning pharmacodynamics and pharmacokinetic properties. In: *Flavonoid Metabolism - Recent Advances and Applications in Crop Breeding*; 2023. doi:10.5772/INTECHOPEN.108637
  266. Xue JC, Yuan S, Meng H, et al. The role and mechanism of flavonoid herbal natural products in ulcerative colitis. *Biomed Pharmacother.* 2023;158:114086. doi:10.1016/J.BIOPHA.2022.114086
  267. Li R, Zhou Y, Zhang S, Li J, Zheng Y, Fan X. The natural (poly) phenols as modulators of microglia polarization via TLR4/NF-κB pathway exert anti-inflammatory activity in ischemic stroke. *Eur J Pharmacol.* 2022;914:174660. doi:10.1016/J.EJPHAR.2021.174660
  268. Ntalouka F, Tsrivakou A. Luteolin: a promising natural agent in management of pain in chronic conditions. *Front Pain Res.* 2023;4:1114428. doi:10.3389/FPAIN.2023.1114428/FULL
  269. Daily JW, Kang S, Park S. Protection against Alzheimer's disease by luteolin: role of brain glucose regulation, anti-inflammatory activity, and the gut microbiota-liver-brain axis. *Biofactors.* 2021;47(2):218-231. doi:10.1002/BIOF.170
  270. Pan Q, Liu Y, Ma W, Kan R, Zhu H, Li D. Cardioprotective effects and possible mechanisms of luteolin for myocardial ischemia-reperfusion injury: a systematic review and meta-analysis of preclinical evidence. *Front Cardiovasc Med.* 2022;9:685998. doi:10.3389/FCVM.2022.685998/FULL
  271. Monti E, Marras E, Prini P, Gariboldi MB. Luteolin impairs hypoxia adaptation and progression in human breast and colon cancer cells. *Eur J Pharmacol.* 2020;881:173210. doi:10.1016/J.EJPHAR.2020.173210
  272. Zheng H, Zhu X, Gong E, Lv Y, Li Y, Cai X. Luteolin suppresses lung cancer progression through targeting the circ\_0000190/miR-

- 130a-3p/notch-1 signaling pathway. *J Chemother.* 2023;35(4):330-342. doi:10.1080/1120009X.2022.2102303
273. Rocchetti MT, Bellanti F, Zadorozhna M, Fiocco D, Mangieri D. Multi-faceted role of luteolin in cancer metastasis: EMT, angiogenesis, ECM degradation and apoptosis. *Int J Mol Sci.* 2023;24(10):8824. doi:10.3390/IJMS24108824
274. Çetinkaya M, Baran Y. Therapeutic potential of luteolin on cancer. *Vaccines.* 2023;11(3):554. doi:10.3390/VACCINES11030554
275. Fernando PDSM, Ko DO, Piao MJ, Kang KA, Herath HMUL, Hyun JW. Protective effect of luteolin against oxidative stress-mediated cell injury via enhancing antioxidant systems. *Mol Med Rep.* 2024;30(1):121. doi:10.3892/MMR.2024.13244
276. Huang L, Jin K, Lan H. Luteolin inhibits cell cycle progression and induces apoptosis of breast cancer cells through down-regulation of human telomerase reverse transcriptase. *Oncol Lett.* 2019;17(4):3842-3850. doi:10.3892/ol.2019.10052
277. Cao D, Zhu GY, Lu Y, et al. Luteolin suppresses epithelial-mesenchymal transition and migration of triple-negative breast cancer cells by inhibiting YAP/TAZ activity. *Biomed Pharmacother.* 2020;129:110462. doi:10.1016/J.BIOPHA.2020.110462
278. Jiang J, Zhu F, Zhang H, et al. Luteolin suppresses the growth of colon cancer cells by inhibiting the IL-6/STAT3 signaling pathway. *J Gastrointest Oncol.* 2022;13(4):1722-1732. doi:10.21037/JGO-22-507
279. Masraksa W, Tanasawet S, Hutamekalin P, Wongtawatchai T, Sukketsiri W. Luteolin attenuates migration and invasion of lung cancer cells via suppressing focal adhesion kinase and non-receptor tyrosine kinase signaling pathway. *Nutr Res Pract.* 2020;14(2):127-133. doi:10.4162/NRP.2020.14.2.127
280. Pan J, Cai X, Zheng X, Zhu X, Feng J, Wang X. Luteolin inhibits viability, migration, angiogenesis and invasion of non-small cell lung cancer vascular endothelial cells via miR-133a-3p/purine rich element binding protein B-mediated MAPK and PI3K/Akt signaling pathways. *Tissue Cell.* 2022;75:101740. doi:10.1016/J.TICE.2022.101740
281. Yao X, Jiang W, Yu D, Yan Z. Luteolin inhibits proliferation and induces apoptosis of human melanoma cells: in vivo and in vitro by suppressing MMP-2 and MMP-9 through the PI3K/AKT pathway. *Food Funct.* 2019;10(2):703-712. doi:10.1039/C8FO02013B
282. Raina R, Pramodh S, Rais N, et al. Luteolin inhibits proliferation, triggers apoptosis and modulates Akt/mTOR and MAP kinase pathways in HeLa cells. *Oncol Lett.* 2021;21(3):192. doi:10.3892/OL.2021.12452
283. Oo AM, Nor MNM, Lwin OM, Simbak N, Mohd Adnan LH, Rao USM. Immunomodulatory effects of apigenin, luteolin, and quercetin through natural killer cell cytokine secretion. *J Appl Pharm Sci.* 2022;12(9):121-126. doi:10.7324/JAPS.2022.120914
284. Oo AM, Mohd Adnan LH, Nor NM, Simbak N, Ahmad NZ, Lwin OM. Immunomodulatory effects of flavonoids: an experimental study on natural-killer-cell-mediated cytotoxicity against lung cancer and cytotoxic granule secretion profile. *Proc Singapore Healthcare.* 2021;30(4):279-285. doi:10.1177/2010105820979006
285. Shang J, Yang J, Deng Q, Zhou M. Nano-scale drug delivery systems for luteolin: advancements and applications. *J Mater Chem B.* 2023;11(47):11198-11216. doi:10.1039/D3TB01753B
286. Liu J, Sun Y, Cheng M, et al. Improving oral bioavailability of luteolin nanocrystals by surface modification of sodium Dodecyl sulfate. *AAPS PharmSciTech.* 2021;22(3):133. doi:10.1208/S12249-021-02012-Y
287. Jang CH, Moon N, Lee J, Kwon MJ, Oh J, Kim JS. Luteolin synergistically enhances antitumor activity of oxaliplatin in colorectal carcinoma via AMPK inhibition. *Antioxidants.* 2022;11(4):626. doi:10.3390/ANTIOX11040626
288. Chen YH, Wu JX, Yang SF, Hsiao YH. Synergistic combination of luteolin and asiatic acid on cervical cancer in vitro and in vivo. *Cancers.* 2023;15(2):548. doi:10.3390/CANCERS15020548
289. Ganai SA, Sheikh FA, Baba ZA, Mir MA, Mantoo MA, Yattoo MA. Anticancer activity of the plant flavonoid luteolin against preclinical models of various cancers and insights on different signalling mechanisms modulated. *Phytother Res.* 2021;35(7):3509-3532. doi:10.1002/PTR.7044
290. Luo Y, Chen S, Zhou J, et al. Luteolin cocrystals: characterization, evaluation of solubility, oral bioavailability and theoretical calculation. *J Drug Deliv Sci Technol.* 2019;50:248-254. doi:10.1016/J.JDDST.2019.02.004
291. Wu W, Li K, Zhao C, Ran X, Zhang Y, Zhang T. A rapid HPLC-MS/MS method for the simultaneous determination of luteolin, resveratrol and their metabolites in rat plasma and its application to pharmacokinetic interaction studies. *J Chromatogr B Analyt Technol Biomed Life Sci.* 2022;1191:123118. doi:10.1016/J.JCHROMB.2022.123118
292. Qureshi MN, Stecher G, Bonn G. Determination of total polyphenolic compounds and flavonoids in Matricaria chamomella flowers. *Pak J Pharm Sci.* 2019;32:2163.
293. Azeem M, Khari H, Niazi S, et al. An overview of anti-inflammatory, antioxidant, anti-cancer, anti-hyperlipidemic, neuroprotective and muscle relaxant effects of natural flavonoid, apigenin; a review. *Biol Clin Sci Res J.* 2024;2024(1):644. doi:10.54112/BCSRJ.V202411.644
294. Bhratee A, Anand P, Singh S. Apigenin: exploring its neuroprotective potential in neurodegenerative disorders: mechanisms and promising therapeutic applications. *Pharmaspire.* 2023;15(04):257-263. doi:10.56933/PHARMASPIRE.2023.15138
295. Adel M, Zahmatkeshan M, Akbarzadeh A, et al. Chemotherapeutic effects of Apigenin in breast cancer: preclinical evidence and molecular mechanisms; enhanced bioavailability by nanoparticles. *Biotechnol Rep.* 2022;34:e00730. doi:10.1016/J.BTRE.2022.E00730
296. Ji X, Liu K, Li Q, et al. A mini-review of flavone isomers apigenin and genistein in prostate cancer treatment. *Front Pharmacol.* 2022;13:851589. doi:10.3389/FPHAR.2022.851589/BIBTEX
297. Cheng Y, Han X, Mo F, et al. Apigenin inhibits the growth of colorectal cancer through down-regulation of E2F1/3 by

- miRNA-215-5p. *Phytomedicine*. 2021;89:153603. doi:10.1016/J.PHYMED.2021.153603
298. Wang SM, Yang PW, Feng XJ, et al. Apigenin inhibits the growth of hepatocellular carcinoma cells by affecting the expression of microRNA transcriptome. *Front Oncol*. 2021;11:657665. doi:10.3389/FONC.2021.657665/FULL
  299. Fossatelli L, Maroccia Z, Fiorentini C, Bonucci M. Resources for human health from the plant kingdom: the potential role of the flavonoid apigenin in cancer counteraction. *Int J Mol Sci*. 2024;25(1):251. doi:10.3390/IJMS25010251/S1
  300. Javed Z, Sadia H, Iqbal MJ, et al. Apigenin role as cell-signaling pathways modulator: implications in cancer prevention and treatment. *Cancer Cell Int*. 2021;21(1):189. doi:10.1186/S12935-021-01888-X
  301. Kashyap P, Shikha D, Thakur M, Aneja A. Functionality of apigenin as a potent antioxidant with emphasis on bioavailability, metabolism, action mechanism and in vitro and in vivo studies: a review. *J Food Biochem*. 2021;46(4):e13950. doi:10.1111/JFBC.13950
  302. Hnit SST, Yao M, Xie C, et al. Apigenin impedes cell cycle progression at G2 phase in prostate cancer cells. *Discov Oncol*. 2022;13(1):44. doi:10.1007/S12672-022-00505-1
  303. Bao Y, Wu X, Jin X, et al. Apigenin inhibits renal cell carcinoma cell proliferation through G2/M phase cell cycle arrest. *Oncol Rep*. 2022;47(3):60. doi:10.3892/OR.2022.8271/DOWNLOAD
  304. Shukla S, Fu P, Gupta S. Apigenin induces apoptosis by targeting inhibitor of apoptosis proteins and Ku70–Bax interaction in prostate cancer. *Apoptosis*. 2014;19(5):883-894. doi:10.1007/S10495-014-0971-6
  305. Imran M, Aslam Gondal T, Atif M, et al. Apigenin as an anticancer agent. *Phytother Res*. 2020;34(8):1812-1828. doi:10.1002/PTR.6647
  306. Amini P, Moazamiyanfar R, Dakkali MS, et al. Induction of cancer cell death by apigenin: a review on different cell death pathways. *Mini Rev Med Chem*. 2023;23(14):1461-1478. doi:10.2174/1389557523666230119110744
  307. Tong X, Pelling JC. Targeting the PI3K/Akt/mTOR axis by apigenin for cancer prevention. *Anti Cancer Agents Med Chem*. 2013;13(7):971-978.
  308. Shukla S, Bhaskaran N, Babcook MA, Fu P, MacLennan GT, Gupta S. Apigenin inhibits prostate cancer progression in TRAMP mice via targeting PI3K/Akt/FoxO pathway. *Carcinogenesis*. 2014;35(2):452-460.
  309. Zhang X, Zhang W, Chen F, Lu Z. Combined effect of chrysin and apigenin on inhibiting the development and progression of colorectal cancer by suppressing the activity of P38-MAPK/AKT pathway. *IUBMB Life*. 2021;73(5):774-783. doi:10.1002/iub.2456
  310. Naponelli V, Rocchetti MT, Mangieri D. Apigenin: molecular mechanisms and therapeutic potential against cancer spreading. *Int J Mol Sci*. 2024;25(10):5569. doi:10.3390/IJMS25105569
  311. Liang C, Jiang Y, Sun L. Vitexin suppresses the proliferation, angiogenesis and stemness of endometrial cancer through the PI3K/AKT pathway. *Pharm Biol*. 2023;61(1):581-589. doi:10.1080/13880209.2023.2190774
  312. Seo HS, Ku JM, Choi HS, et al. Induction of caspase-dependent apoptosis by apigenin by inhibiting STAT3 signaling in HER2-overexpressing MDA-MB-453 breast cancer cells. *Anticancer Res*. 2014;34(6):2869-2882.
  313. Seo HS, Ku JMo., Choi HS, et al. Apigenin induces caspase-dependent apoptosis by inhibiting signal transducer and activator of transcription 3 signaling in HER2-overexpressing SKBR3 breast cancer cells. *Mol Med Rep*. 2015;12(2):2977-2984. doi:10.3892/MMR.2015.3698
  314. Huang J, Chen X, Chang Z, Xiao C, Najafi M. Boosting anti-tumour immunity using adjuvant apigenin. *Anti Cancer Agents Med Chem*. 2022;23(3):266-277. doi:10.2174/1871520622666220523151409
  315. Feng B, Chen L, Chen X, et al. Immunopotential effects of apigenin on NK cell proliferation and killing pancreatic cancer cells. *Int J Immunopathol Pharmacol*. 2023;37:3946320231161174. doi:10.1177/03946320231161174
  316. Samir A, Abdeltawab R, Stein U, Thabit S, el Tayebi HM. 42P Apigenin: an immunomodulatory nutraceutical overriding PD-L1 inhibitors by halting AKT/mTOR pathway in triple-negative breast cancer (TNBC). *ESMO Open*. 2023;8(1):101008. doi:10.1016/j.esmoop.2023.101008
  317. Mahbub AA, Le Maitre CL, Cross NA, Jordan-Mahy N. The effect of apigenin and chemotherapy combination treatments on apoptosis-related genes and proteins in acute leukaemia cell lines. *Sci Rep*. 2022;12(1):8858. doi:10.1038/s41598-022-11441-z
  318. Huang S, Yu M, Shi N, et al. Apigenin and Abivertinib, a novel BTK inhibitor synergize to inhibit diffuse large B-cell lymphoma in vivo and vitro. *J Cancer*. 2020;11(8):2123-2132. Retrieved August 15, 2024, from: <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7052937/>
  319. Nimal S, Kumbhar N, Rathore S, Naik N, Paymal S, Gacche RN. Apigenin and its combination with Vorinostat induces apoptotic-mediated cell death in TNBC by modulating the epigenetic and apoptotic regulators and related miRNAs. *Sci Rep*. 2024;14(1):1-28. doi:10.1038/s41598-024-60395-x
  320. Yang CS, Chen JX, Wang H, Lim J. Lessons learned from cancer prevention studies with nutrients and non-nutritive dietary constituents. *Mol Nutr Food Res*. 2016;60(6):1239-1250. doi:10.1002/mnfr.201500766
  321. Kurahashi N, Iwasaki M, Sasazuki S, et al. Soy product and isoflavone consumption in relation to prostate cancer in Japanese men. *Cancer Epidemiol Biomarkers Prev*. 2007;16(3):538-545. doi:10.1158/1055-9965.EPI-06-0517/343733/P/SOY-PRODUCT-AND-ISOFALVONE-CONSUMPTION-IN-RELATION
  322. Hodek P, Trefil P, Stiborová M. Flavonoids-potent and versatile biologically active compounds interacting with cytochromes P450. *Chem Biol Interact*. 2002;139(1):1-21. doi:10.1016/S0009-2797(01)00285-X
  323. Park K. The role of dietary phytochemicals: evidence from epidemiological studies. *Nutrients*. 2023;15(6):1371. doi:10.3390/NU15061371