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CAML regulates Bim-dependent thymocyte death

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Abstract

Appropriate control of apoptosis during T lymphocyte differentiation is critical for destruction of T cells bearing potentially autoreactive or useless immuno-receptors, and for survival of those T cells bearing antigen receptors that may recognize foreign proteins. Despite the well-established importance of thymocyte survival, the exact signals regulating thymocyte apoptosis have not been fully elucidated. Here, we show that thymocytes lacking the endoplasmic reticulum (ER) protein CAML failed to undergo normal T cell development and exhibited dramatically increased rates of apoptosis. *In vitro*, CAML-deficient thymocytes accumulated high levels of reactive oxygen species (ROS) and underwent abnormally accelerated death in response to several cytotoxic stimuli, including treatment with etoposide, cytokine deprivation or Fas ligation. Although neither p53 deletion nor loss of Fas rescued the survival and continued development of CAML-deficient thymocytes, removal of the pro-apoptotic BH3-only Bcl-2 family member Bim significantly restored their survival. This work reveals CAML to be a critically important regulator of ROS- and Bim-dependent thymocyte death.

Keywords and Abbreviations

CAML: Calcium modulating cyclophilin ligand; ROS: reactive oxygen species; Bim: Bcl-2 interacting mediator of cell death

Introduction

Cell survival during T cell development is a carefully controlled process. Early in development, signals through the pre-T cell receptor (pre-TCR, composed of TCR β and pT α) promote proliferation, survival and differentiation of immature (CD4–8–) progenitors.

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Conflict of Interest Disclosure

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Further rearrangement of the α chain of the TCR completes the assembly of the T cell antigen receptor, and thymocytes are then subjected to stringent selection based on its specificity. Cells bearing a TCR with low affinity for self-peptide bound to a self-MHC molecule receive a survival signal and undergo continued differentiation, a process termed positive selection. Thymocytes with receptors that have high affinity for self-peptide MHC complexes undergo apoptosis in a process called negative selection, while those cells that are unable to bind any ligand undergo death by neglect (1). The developing thymocyte has a window of opportunity during which to sample a variety of ligand/MHC complexes and receive a positively selecting signal before it undergoes programmed cell death, but the signals maintaining thymocyte viability and the pathways that control cell survival or death in response to these interactions are not well understood.

Mammalian cells, including thymocytes, have two distinct, albeit ultimately converging, pathways of apoptosis, the 'extrinsic' ('death receptor'-controlled) pathway and the 'intrinsic' (also called 'mitochondrial' or 'Bcl-2-regulated) pathway (2). The 'extrinsic' pathway is activated by certain members of the TNF family of ligands that bind to so-called 'death receptors' (members of the TNF receptor family with an intracellular 'death domain') on the cell surface, triggering a cascade of intracellular events that result in cell death (2). Conversely, the 'intrinsic' apoptotic pathway is regulated by the balance between pro- and anti-apoptotic members of the Bcl-2 family. The pro-survival Bcl-2 family members – Bcl-2, Bcl-x_L, Bcl-w, Mcl-1 and A1 – share with each other up to 4 regions of homology (BH: Bcl-2 Homology regions) and are essential for cell survival functioning in a cell type-specific manner. There are two pro-apoptotic subgroups within the Bcl-2 family. Bak, Bax and possibly also Bok have 3 BH domains and share surprising structural similarity with their pro-survival relatives but are essential (in a redundant manner) for downstream events in apoptosis, including mitochondrial outer membrane permeabilization (MOMP) and consequent activation of the caspase cascade. BH3-only proteins, on the other hand, share with each other and the remainder of the Bcl-2 family only the BH3 domain and they are critical for initiation of apoptosis signaling, functioning in a cell death stimulus- and cell type-specific manner (2).

Bcl-2 interacting mediator of cell death (Bim) is a BH3-only Bcl-2 family member that is transcriptionally and/or post-translationally induced in response to diverse apoptotic stimuli, such as cytokine deprivation or deregulated calcium flux, in a broad range of hematopoietic, epithelial and neuronal cell types (3–5). Experiments with gene-targeted mice showed that Bim is required for the death of thymocytes in response to growth factor withdrawal, calcium flux, taxol treatment, as well as strong T cell receptor ligation (6, 7). Bim is also critical for the apoptosis of peripheral T cells during the contraction phase of an acute immune response (8–10). As a potent mediator of cell death, Bim is highly regulated both transcriptionally and post-translationally. Bim is found in three major isoforms that have differing levels of pro-apoptotic activity (4) and are each regulated by various post-translational mechanisms, including phosphorylation (11), proteosomal degradation (12) and sequestration to the microtubular dynein motor complex (13). While much is known about Bim regulation, questions still remain as to which signaling pathways directly inhibit or promote Bim activity.

CAML is a transmembrane protein that localizes to the endoplasmic reticulum and small cytoplasmic vesicles (14, 15). It is ubiquitously expressed and evolutionarily highly conserved in all vertebrates. The exact cellular function of CAML is still unknown, but it has been implicated in diverse cell signaling processes, including calcium signaling (14), protein trafficking (16) and chromosome segregation (17). A CAML conditional knockout mouse in which CAML was deleted relatively late during thymic T cell development has been described (18). In that study, using CAML^{fl/fl} mice expressing the jmlckcre transgene, it was observed that CAML-deficient cells experienced increased T cell receptor stimulation, but that the overall processes of positive and negative selection were intact (19). While previous work suggested that CAML-deficient cells were more susceptible to apoptosis in response to T cell receptor stimulation compared to control cells, there was no direct evidence that CAML-deficient CD4⁺CD8⁺ (double positive; DP) thymocytes exhibit abnormally high rates of apoptosis. Furthermore, it was not examined how CAML may interact with apoptotic signaling pathways.

The current study represents the first description of mice in which CAML is deleted during the DN2/DN3 stage of development, considerably earlier than the timing of excision reported for the *caml*^{fl/fl} mice expressing the jmlckcre transgene. In our study, we observed severe reductions in thymocyte subsets, beginning at the double negative (DN) 4 stage of development, which was due to increased apoptosis and did not result from a defect in cell proliferation. The loss of cellularity was dependent upon the presence of Bim, but did not require T cell receptor ligands (peptide/MHC complexes), p53 or Fas signaling.

Results

Loss of CAML early in T-cell development leads to a severe reduction in thymocyte numbers due to abnormally increased apoptosis

To determine the role of CAML in early T cell development, we crossed mice bearing loxP-flanked *Caml* alleles (CAML^{fl/fl}) to cwLckCre mice, which express the Cre recombinase prior to the DN3 stage of thymocyte development (20). The cwLckCre-CAML^{fl/fl} mice showed a severe depletion of total thymocytes (Figure 1A) in comparison to littermate control CAML^{fl/fl} mice. To investigate whether early T cell development was impaired by CAML deletion, we examined lineage marker (Mac-1, GR-1, CD8 α , TCR β , TCR $\gamma\delta$, CD3 ϵ , B220, CD19, Ter119, and NK1.1) negative thymocytes for expression of c-kit and CD25 (Figure 1B). Quantitation of the earliest subsets of T cells: (ETPs: lin⁻, c-kit⁺, CD25⁻), DN2 (lin⁻, c-kit⁺, CD25⁺) and DN3 (lin⁻, c-kit⁻, and CD25⁺) indicated that the numbers of these cells were similar between cwLckCre-CAML^{fl/fl} and control CAML^{fl/fl} mice. However, the numbers of DN4 cells (lin⁻, c-kit⁻, CD25⁻) were reduced by ~50% in cwLckCre-CAML^{fl/fl} compared to CAML^{fl/fl} mice (Figure 1C). By staining thymocytes for expression of CD4 and CD8, we found that loss of CAML caused an 80–90% reduction in the percentages and total numbers of double-positive (CD4⁺CD8⁺, DP), CD4 single positive (CD4⁺CD8⁻, SP) as well as CD8 SP (CD4⁻CD8⁺) subsets compared to littermate controls (Figure 1D–E).

In comparison to jmlckcre-CAML^{fl/fl} mice (19), the cwLckCre-CAML^{fl/fl} mice contained strikingly fewer thymocytes overall and within all subsets from the DN4 to the SP stages of

development. For example, CD4⁺CD8⁺ DP cells in *jmlckcre-CAML^{fl/fl}* thymi were reduced 2.2-fold compared to age-matched littermate *CAML^{fl/fl}* controls whereas in *cwLckCre-CAML^{fl/fl}* CD4⁺CD8⁺ DP cells were reduced 8.6-fold compared to *CAML^{fl/fl}* age-matched littermate controls. In the single positive populations, *jmlckcre-CAML^{fl/fl}* thymi had 7.9-fold fewer CD4⁺CD8⁻ SP cells compared to control littermates while the *cwLckCre-CAML^{fl/fl}* CD4⁺CD8⁻ SP cells had 19.4-fold fewer cells compared *CAML^{fl/fl}* thymi. Finally, in the CD4⁻CD8⁺ SP population, *jmlckcre-CAML^{fl/fl}* thymi had 5.2-fold fewer cells when compared to *CAML^{fl/fl}* age-matched littermate controls while *cwLckCre-CAML^{fl/fl}* CD4⁻CD8⁺ SP cells had 7.3-fold fewer cells compared to *CAML^{fl/fl}* age-matched littermate controls.

To ensure the efficiency of deletion of the floxed *CamI* alleles in *cwLckCre-CAML^{fl/fl}* cells, we used magnetic sorting to enrich for CD4⁻CD8⁻CD44⁻CD25⁻ (DN4) or CD4⁺CD8⁺ cells. Western blotting revealed that *cwLckCre-CAML^{fl/fl}* cells had a >90% reduction in CAML expression compared to *CAML^{fl/fl}* control cells (Fig 1F).

We next examined whether the loss of thymocytes in *cwLckCre-CAML^{fl/fl}* mice was due to increased cell death or decreased proliferation. We examined the number of apoptotic cells by staining with Annexin V and propidium iodide. A larger percentage of cells in each major thymocyte subset stained positive for Annexin V and propidium iodide in *cwLckCre-CAML^{fl/fl}* cells compared to *CAML^{fl/fl}* littermates (Figure 2A). As CAML has been implicated as a regulator of the mitotic spindle checkpoint and CAML-deficient cells were reported to exhibit defects in cellular division (17), it was important to determine whether CAML is essential for mitosis in thymocytes. To determine whether CAML loss was accompanied by defective proliferation, we injected mice with 2-bromodeoxyuridine (BrdU) and then removed thymi after 90 min. Thymocytes were surface stained with antibodies to CD4 and CD8 to define the four major thymocyte subsets and intra-cellularly with an antibody to BrdU to measure the rate of incorporation of this thymidine analog into dividing cells. Cells of all thymocyte subsets from *cwLckCre-CAML^{fl/fl}* showed increased proliferation compared to control *CAML^{fl/fl}* littermates (Figure 2B). Thus, we conclude that loss of CAML early in T cell development decreases the ability of pre-T cells to survive, resulting in reduced thymic numbers beginning at the DN4 stage of development (and possibly a compensatory increase in cell cycling). Additionally, the increase in proliferation in DN and DP subsets indicates that *cwLckCre-CAML^{fl/fl}* cells have intact pre-TCR signaling and that lack of a signal through the pre-TCR cannot account for the loss of the DP *cwLckCre-CAML^{fl/fl}* cells.

CAML is critical for thymocyte survival after exposure to a specific subset of apoptotic stimuli

To further investigate the differences in survival between control and CAML-deficient thymocytes, we subjected *cwLckCre-CAML^{fl/fl}* and control *CAML^{fl/fl}* cells to a variety of apoptotic stimuli. We observed that in response to treatment with phorbol ester, ionomycin, thapsigargin, staurosporine, dexamethasone, or hydrogen peroxide, cell death was comparable between control and CAML-deficient cells (Figure 3). In contrast, *cwLckCre-CAML^{fl/fl}* thymocytes were significantly more sensitive than control cells to certain other

stimuli, including cytokine deprivation, treatment with etoposide (even 100-fold lower concentrations) or stimulation of the death receptor Fas (Figure 3).

P53 is not required for the enhanced apoptosis caused by CAML-deficiency

Cell death following treatment with the topo-II inhibitor etoposide is dependent upon p53 and its transcriptional induction of the pro-apoptotic BH3-only protein Puma (21). To investigate whether the enhanced apoptosis in CAML-deficient thymocytes might be due to abnormally increased activation of p53, we crossed cwLckCre-CAML^{fl/fl} mice to p53^{-/-} animals. However, cwLckCre-CAML^{fl/fl};p53^{-/-} mice showed no increase in thymocyte numbers compared to cwLckCre-CAML^{fl/fl} controls (Figure S1), indicating that p53 is not required for CAML-deficiency induced thymocyte death *in vivo*.

Fas is not required for the enhanced apoptosis caused by CAML-deficiency

Since CAML-deficient thymocytes exhibited abnormally increased sensitivity to Fas-induced apoptosis, we explored whether Fas is critical for the abnormal thymocyte reduction caused by loss of CAML. Fas-mediated apoptosis was dependent upon caspase activity, as treatment with ZVAD rescued both cwLckCre-CAML^{fl/fl} and CAML^{fl/fl} cells from this death (Figure 4A–B). Next, we analyzed Fas surface expression on cwLckCre-CAML^{fl/fl} and CAML^{fl/fl} littermates and found that the levels were comparable (Figure 4C). The increased sensitivity to Fas stimulation was unexpected, since evidence suggests that Fas does not play a role in the developmentally programmed death of thymocytes (22). Thus, we considered two possible scenarios: one in which the Fas receptor had been inappropriately activated in the absence of CAML, and another in which the Fas-sensitivity of cwLckCre-CAML^{fl/fl} cells reflected a deficiency of a central survival pathway within the cells. To distinguish between these two possibilities, cwLckCre-CAML^{fl/fl} mice were crossed to Fas deficient *Fas*^{lpr/lpr} mice (23). Interestingly, loss of Fas did not cause a rescue of thymic cellularity in cwLckCre-CAML^{fl/fl}; *Fas*^{lpr/lpr} mice (Figure 4D, compare 3rd and 4th columns). This supports the notion that Fas does not mediate the abnormally enhanced *in vivo* death of cwLckCre-CAML^{fl/fl} thymocytes, but instead suggests that these cells have a more general aberration in survival that renders them more sensitive to Fas stimulation *in vitro* plus other death stimuli.

Others have used cwLckCre mice for multiple analyses and have not reported nonspecific toxicity due to thymic Cre expression. Nonetheless, to rule out a possible cytotoxic effect (24) of Cre in our mice, we also analyzed cwLckCre-CAML^{+/+} mice for overall thymic numbers and sensitivity to Fas-mediated apoptosis. As seen in Figure 4E–F, CAML loss, and not the *cre* transgene, was responsible for the loss of thymic cellularity and the abnormally increased sensitivity to Fas-mediated apoptosis.

Loss of Bim partially rescues thymocyte survival in CAML-deficient mice

The pro-apoptotic BH3-only protein Bim has been shown to be essential for the initiation of several pathways that induce apoptosis in thymocytes (7), including cytokine withdrawal and calcium flux. We therefore crossed *bim*^{-/-} mice with the conditional CAML knockout mice to test whether Bim is required for the reduction in cell numbers observed in cwLckCre-CAML^{fl/fl} thymi. Interestingly, cwLckCre-CAML^{fl/fl}; *bim*^{-/-} thymi showed a

significant increase in the number of DP thymocytes. While *cwLckCre-CAML^{fl/fl}* mice had only 13% of the numbers of double positive cells compared to *CAML^{fl/fl}* animals, *cwLckCre-CAML^{fl/fl};bim^{-/-}* mice had 46% of the numbers of these thymocytes compared to *CAML^{fl/fl};bim^{-/-}* littermate controls (Figure 5A–B), indicating that Bim must contribute to the enhanced death of CAML-deficient thymocytes.

Given that Bim appeared responsible for a significant portion of thymocyte death elicited by loss of CAML, we examined whether Bim expression is abnormal in CAML-deficient cells. We first evaluated total cellular levels of Bim in *CD4⁺CD8⁺* double positive cells and found it to be comparable or slightly diminished in the absence of CAML when compared to control cells (Fig. 5C). In addition to total cellular levels, Bim is also regulated by intracellular localization (13), including subcellular localization to the mitochondrial outer membrane, and by association with pro-survival Bcl-2 family members (25). We found that in CAML-deficient cells, Bim was reproducibly increased in the pellet fraction of cells lysed with 1% TritonX-100 (TX-100) and modestly increased in isolated mitochondria (Fig. 5D), indicating that loss of CAML causes mislocalization of Bim.

T cell receptor interaction with peptide/MHC complexes is not required for the enhanced apoptosis of CAML-deficient thymocytes

Bim is essential for thymocyte apoptosis caused by strong TCR stimulation and critical for negative selection of autoreactive thymocytes. Since we previously reported that *jmLckCre-CAML^{fl/fl}* T lymphocytes exhibit abnormally increased T cell receptor stimulation (19), it appeared likely that augmented TCR signaling might be responsible for their abnormally increased apoptosis. Others have demonstrated the role of TCR signaling in DP thymocyte killing in MINK-deficient mice by removing class I and II major histocompatibility antigens (26). In similar fashion, we examined whether MHC/T cell receptor interaction was required for the enhanced death of *cwLckCre-CAML^{fl/fl}* thymocytes by crossing *cwLckCre-CAML^{fl/fl}* mice with MHC class II- (*Ab⁰*) (27) and MHC class I-deficient (*β2 microglobulin^{-/-}*) mice (28). In contrast to thymocytes lacking MINK in which MHC deficiency rescued normal double positive development, loss of MHC did not rescue the survival of DP thymocytes in *cwLckCre-CAML^{fl/fl}* mice (Figure 6A–B). This indicates that T cell receptor/MHC interactions are not required for the abnormally increased death of CAML-deficient pre-T cells.

Abnormally elevated levels of Reactive Oxygen Species (ROS) in *cwLckCre-CAML^{fl/fl}* mice

ROS have been reported to induce Bim-dependent death in T cells (29). Since loss of Bim increases thymocyte numbers in the absence of CAML, we compared ROS levels between CAML-deficient and control thymocytes by intracellular staining with DCFDA. *cwLckCre-CAML^{fl/fl}* *CD4⁺CD8⁺* thymocytes were reproducibly found to have elevated ROS levels (Figure 7A–B) in comparison to cells from littermate control animals. ROS in *cwLckCre-CAML^{fl/fl}* cells appeared to be of mitochondrial origin, as verified by staining with the mitochondrial-localizing ROS dye MitoSox (30) (Figure 7C).

To ensure that DCFDA staining was indeed labeling reactive oxygen species, we administered the antioxidant N-acetyl-cysteine (NAC) to mice to reduce ROS, removed

thymi and then stained the cells with DCFDA. As expected, treatment of mice with NAC reduced the high ROS levels found in thymocytes from CAML-deficient mice (Figure 7D). Additionally, we treated cells *in vitro* with inhibitors of ROS generation: diphenyleneiodonium chloride (DPI), an inhibitor of NADPH oxidase, and carbonyl cyanide-p-trifluoromethoxyphenyl-hydrazone (FCCP), a mild decoupling agent that reduces mitochondrial ROS when used at low concentrations (31). We found that both reagents reduced ROS levels to a greater extent in the cwLckCre-CAML^{fl/fl} cells than within CAML^{fl/fl} cells (7E).

Abnormally increased ROS levels could be a cause of death or simply a downstream consequence of programmed cell death. To distinguish between these possibilities, we treated cwLckCre-CAML^{fl/fl} and CAML^{fl/fl} cells with NAC during stimulation of cells with anti-Fas antibody. Although addition of NAC had little effect on normal thymocytes, NAC-treated cwLckCre-CAML^{fl/fl} cells were significantly protected from Fas-induced apoptosis (Figure 7F), indicating that ROS are required for the abnormally increased sensitivity of CAML-deficient thymocytes to Fas-mediated death. To determine whether NAC could also diminish the sensitivity of cwLckCre-CAML^{fl/fl} cells to other apoptotic stimuli, we added NAC to cells immediately after isolation from the thymus and cultured them for 6, 12, and 24 h in simple medium (to mimic cytokine deprivation). Remarkably, NAC was able to augment the viability of cwLckCre-CAML^{fl/fl} thymocytes cultured without cytokines at each time point (Figure 7G).

Discussion

Here, we establish an essential role for CAML in maintaining the survival of developing thymocytes. CAML does not protect thymocytes indiscriminately against all cytotoxic insults; rather, it protects them from Fas stimulation, etoposide treatment and cytokine deprivation but not from treatment with phorbol ester, ionomycin, thapsigargin, staurosporine, dexamethasone or hydrogen peroxide. Our data suggest that CAML does not prevent TCR or p53-mediated death, but instead protects cells from Bim-induced death, most likely by blocking the production of reactive oxygen species. Although previous studies suggested a connection between CAML and cell survival, the role of CAML in this process has been a mystery. CAML interacts with K7 (32), a viral protein important for maintaining cell viability during infection, and E3-6.7K, a TACI viral homologue (33) that was shown to increase survival of cells exposed to extrinsic apoptotic stimuli (34).

What is the molecular mechanism by which CAML potentiates thymocyte survival? Several important steps of this process are revealed by this work. First, CAML-deficient cells have enhanced levels of ROS. ROS have been implicated in many types of cell death, including activation-induced apoptosis of T cells (35) and increasing the susceptibility of cells to extrinsic death stimuli (36). The decreased redox potential of CAML-deficient cells may render them more susceptible to Fas ligation and cytokine deprivation, as evidenced by the ability of the antioxidant NAC to reduce the susceptibility of CAML-deficient cells to Fas stimulation and cytokine deprivation. The ability of NAC to inhibit this enhanced death also provides evidence that the high ROS levels are not a consequence of cellular demolition but

rather than the ROS generated in CAML-deficient cells contribute to increased apoptosis signaling.

Second, the death of thymocytes lacking CAML requires the pro-apoptotic BH3-only Bcl-2 family member Bim. Bim has been shown to be critical for negative selection of autoreactive thymocytes and immature B cells, and for cytokine-withdrawal induced death of a broad range of cell types (6), and Bim is induced by TCR or BCR signaling (7, 37) and ROS (38). In contrast to our previous study of more mature single positive thymocytes that showed abnormally augmented TCR signaling as a consequence of CAML-deficiency (19), we did not find evidence for TCR signal dependence of the enhanced death of CAML-deficient thymocytes, although Bim is clearly required for the excess thymocyte death. Ca^{2+} signaling also regulates Bim (39), but we do not suspect that CAML-deficient cells have abnormal Ca^{2+} influx, because there was no defect in pro-T cell proliferation, which also requires Ca^{2+} signaling. Moreover, CAML-deficient thymocytes had a normal content of intracellular free Ca^{2+} (data not shown). We examined how Bim might be activated to promote the destruction of CAML-deficient cells and found that Bim was not increased at the protein level, nor did it show altered migration on SDS-PAGE, which would suggest a difference in phosphorylation. However, in the absence of CAML, Bim was localized more strongly to the pellet fraction of cells lysed with 1% Triton lysis buffer. Although we have not conclusively shown that CAML directly alters Bim localization, in light of CAML's role in trafficking the EGFR (16), it is tempting to speculate that CAML may play a role in preventing Bim from moving to the mitochondria.

Precisely how CAML acts to block ROS induction and Bim activation is not yet clear. However, CAML has previously been shown to interact with several protein kinases, including EGFR (16), Lck (19) and Hck (unpublished data). Modulation of the activity or intracellular location of one or more such kinases may therefore constitute one of the critical functions of CAML. Indeed, in preliminary studies, we have identified a physical interaction between the serine-threonine kinase Raf-1 and CAML (Figure S2). Notably, Raf has been implicated in signaling pathways that inhibit apoptosis (40), and dominant negative mutants of Raf block thymocyte development (41). Moreover, Raf-1 was shown to regulate cellular levels of reactive oxygen species and is thought to thereby control cellular responses to cytokine-withdrawal (31). Importantly for this study, it has been shown that pharmacological blockade of Raf signaling kills certain tumor cells via Bim induction (42). Raf-1 has also been reported protect cells from external apoptotic stimuli, including Fas ligation (43), a stimulus to which CAML-deficient thymocytes are extremely sensitive. It is therefore tempting to speculate that CAML might regulate the survival function of Raf-1. Future studies will be required to determine whether the Raf-CAML interaction participates in the pro-survival role of CAML in this cell type.

Materials and methods

Mice

All experiments used 3–6 week old gender-matched littermate controls that were on a mixed C57BL/6x129/SvJ background. Dr. David Tran developed CAML^{fl/fl} mice (19), which were generated using 129/SvJ ES cells. cwlcckre (C57BL/6NTac-TgN(Lck-Cre) stock number:

004197) mice were purchased from Taconic. Bim-deficient (B6.129S1-Bcl2l1^{tm1.1Ast}/J stock number: 004525) and LPR (B6.MRL-Fas^{lpr}/J stock number: 000482) mice (both backcrossed onto a C57BL/6 background for >20 generations) were purchased from Jackson Labs. p53-deficient mice were a gift from Dr. Jan VanDeurson. MHC-deficient (β 2microglobulin^{-/-} and class II MHC^{-/-}) mice were a gift from Dr. Chella David. All animal experiments were performed according to IACUC guidelines and were approved by the IACUC.

Reagents

Ionomycin (407952) and thapsigargin (586005) were purchased from Calbiochem. PMA (13139-019) was purchased from Gibco. Etoposide (E1383), hydrogen peroxide (349887), Diphenyleiiodonium chloride (DPI D2926), carbonyl cyanide-p-trifluoromethoxyphenyl-hydrazine (FCCP C2920) and N-acetyl cysteine (NAC A9165) were purchased from Sigma. Jo2 (554254) hamster anti-mouse Fas antibody was purchased from BD Pharmingen.

Flow Cytometry

Labeled anti-CD4 (clone RM4-5), anti-CD8 (clone 53-6.7), anti-c-kit (clone 2B8) and anti-CD25 (clone 3C7) antibodies, propidium iodide (#51-66211E) and Cy5-conjugated Annexin V (#559934) were obtained from BD Pharmingen. Cocktails of lineage marker-specific antibodies were a generous gift from Dr. Virginia Shapiro and consisted of FITC-labeled antibodies specific for Mac-1 (CD11b), GR-1 (Ly6G/C), CD8 α , TCR β , TCR $\gamma\delta$, CD3 ϵ , B220, CD19, Ter119, and NK1.1. DCFDA (C2938) was purchased from Invitrogen. Mitosox (M36008) was purchased from Molecular Probes. Analyses were performed on BD FACS Calibur and Accuri C6 flow cytometers.

Cell Survival Assays

Thymi were removed from mice and tweezed into a single cell suspension. Two million thymocytes were cultured for 16 h in 24 well plates with the indicated reagents. Cells were then stained with anti-CD4 and anti-CD8 antibodies, Annexin V-Cy5 and PI to assess DP thymocyte viability.

Immunoprecipitation and Western Blotting

CD4+CD8+ double positive cells were isolated using Miltenyi MACS multisort kit. Cells were lysed for 5 min with TX-100 lysis buffer on ice (1% Triton X-100, 20 mM HEPES [pH 7.4], 5 mM NaCl, 5 mM MgCl₂, 1 mM EGTA, 1 mM EDTA, 1 mM Iodoacetamide, 100 μ M Na₂VO₄, 1 mM PMSF, 10 μ g/ml leupeptin, and 45 μ g/ml aprotinin). Proteins were precipitated with 5 μ g of antigen-specific antibodies: CAML (polyclonal rabbit serum raised against amino acids 1–189 of CAML), and Raf-1 (BD Pharmingen 610152), or isotype-matched control antibodies (BD Pharmingen). Western blots were probed for CAML (polyclonal rabbit serum raised against amino acids 1–189 of CAML), Raf-1 (BD Pharmingen 610152), actin (Millipore C4), and Bim (Cell Signaling 2819).

Mitochondrial Isolation

CD8⁺ thymocytes (96–98% pure for CD4⁺CD8⁺ double positive cells) were isolated using Miltenyi MACS CD8 isolation beads. Mitochondria isolation protocol was followed according to a protocol modified from (44). 60×10^6 cells were lysed in 10 mM Hepes-KOH, pH 7.9, 10 mM KCl, 1.5 mM MgCl₂, 0.2 mM PMSF, 1 mM DTT, and 0.6% NP-40. After 45 minutes on ice, lysates were centrifuged 300g for 1 minute to remove the nuclear fraction. Supernatant was transferred to a new tube and centrifuged at 700g for 10 minutes and subsequently 17,000g for 40 minutes. A brown mitochondrial pellet was observed after this final centrifugation. The supernatant, a cytosolic fraction, was removed and the mitochondrial pellet was lysed in 10 uL of the lysis buffer described above. Lysates were loaded onto a 15% SDS-PAGE gel, transferred, and probed with Bim-specific (Cell Signaling 2819) and actin-specific (Millipore C4) antibodies.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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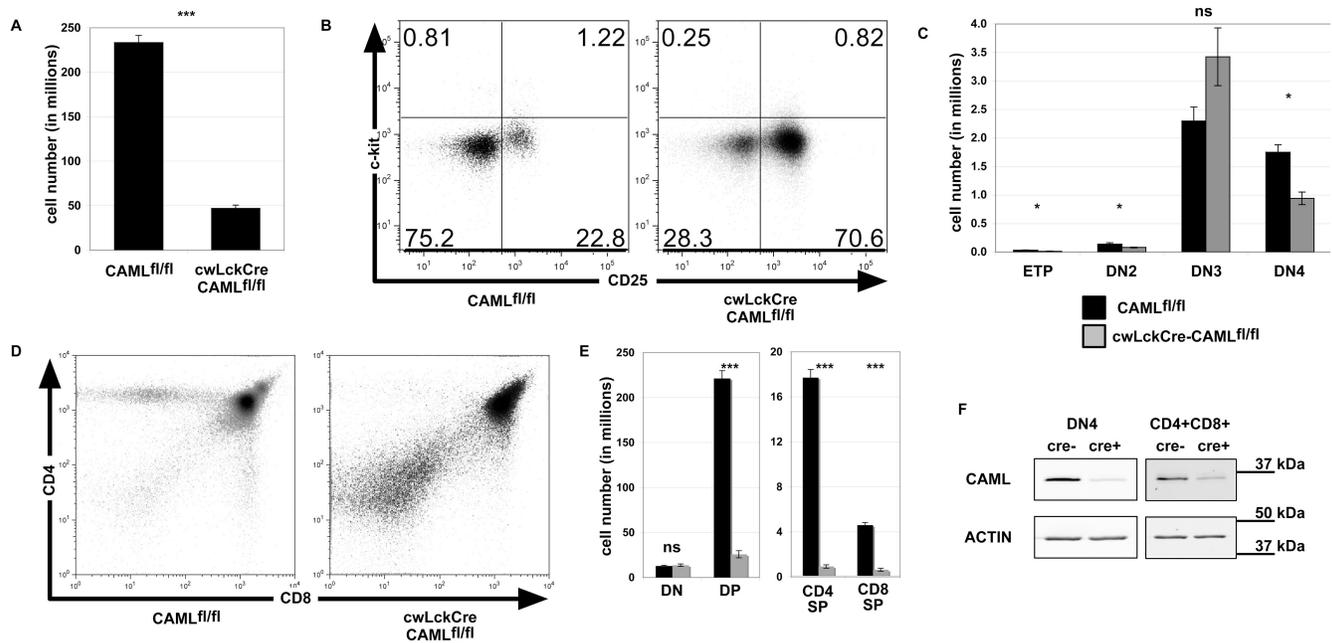


Figure 1. cwLckCre-CAML^{fl/fl} mice exhibit a defect in thymocyte development, beginning at the DN4 stage

(A) Thymocytes from three-six week old cwLckCre-CAML^{fl/fl} mice or littermate controls were tweezed into single cell suspensions and counted. n=50+ for each genotype. (B) Cells were stained for surface expression of lineage markers, CD44, and CD25. Lineage marker (Mac-1 (CD11b), GR-1 (Ly6G/C), CD8 α , TCR β , TCR $\gamma\delta$, CD3 ϵ , B220, CD19, Ter119, and NK1.1) -negative cells were analyzed for c-kit and CD25 expression, as shown in the representative flow cytometry profiles. (C) Lineage-negative subsets were quantified. ETP (lin⁻, c-kit⁺, CD25⁻), DN2 (lin⁻, c-kit⁺, CD25⁺), DN3 (lin⁻, c-kit⁻, and CD25⁺), and DN4 (lin⁻, c-kit⁻ and CD25⁻) n=5 for each genotype. (D) Total thymocytes were analyzed for expression of CD4 and CD8, as shown in these representative flow cytometry profiles. (E) CD4 and CD8 subsets were quantified. n=17 for each genotype. Error bars represent standard error of the mean. (F) Total thymocytes were sorted using Miltenyi MACS beads based on expression of CD4, CD8, CD44, and CD25. CD4⁻CD8⁻CD44⁻CD25⁻ (DN4) and CD4⁺CD8⁺ cells were isolated and lysed with 1% Triton lysis buffer. Western blots were probed for CAML and actin (loading control) levels.

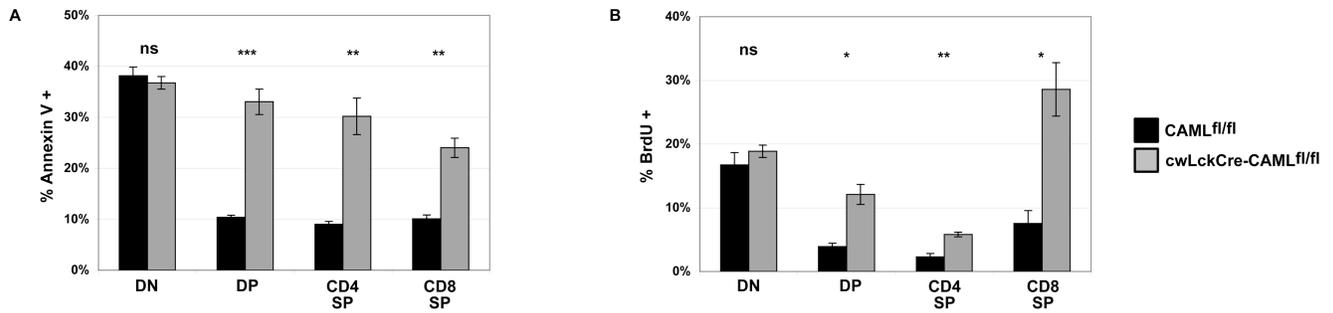


Figure 2. Abnormally increased apoptosis and increased proliferation are evident in cwLckCre-CAML^{fl/fl} thymocytes

(A) Freshly isolated thymocytes were stained with anti-CD4 plus anti-CD8 antibodies plus Annexin V-Cy5 and propidium iodide to determine the percentage of apoptotic cells in each of the four major thymocyte subsets. Data from one representative of three independent experiments is shown. n=5 of each genotype. (B) Mice were injected intraperitoneally with 15 μ g BrdU. After 90 min, thymocytes were isolated and stained for surface expression of CD4 and CD8 and intra-cellularly for uptake of BrdU. n=3 of each genotype. Error bars represent standard error of the mean. * p<.05 ** p<.005 *** p<.001

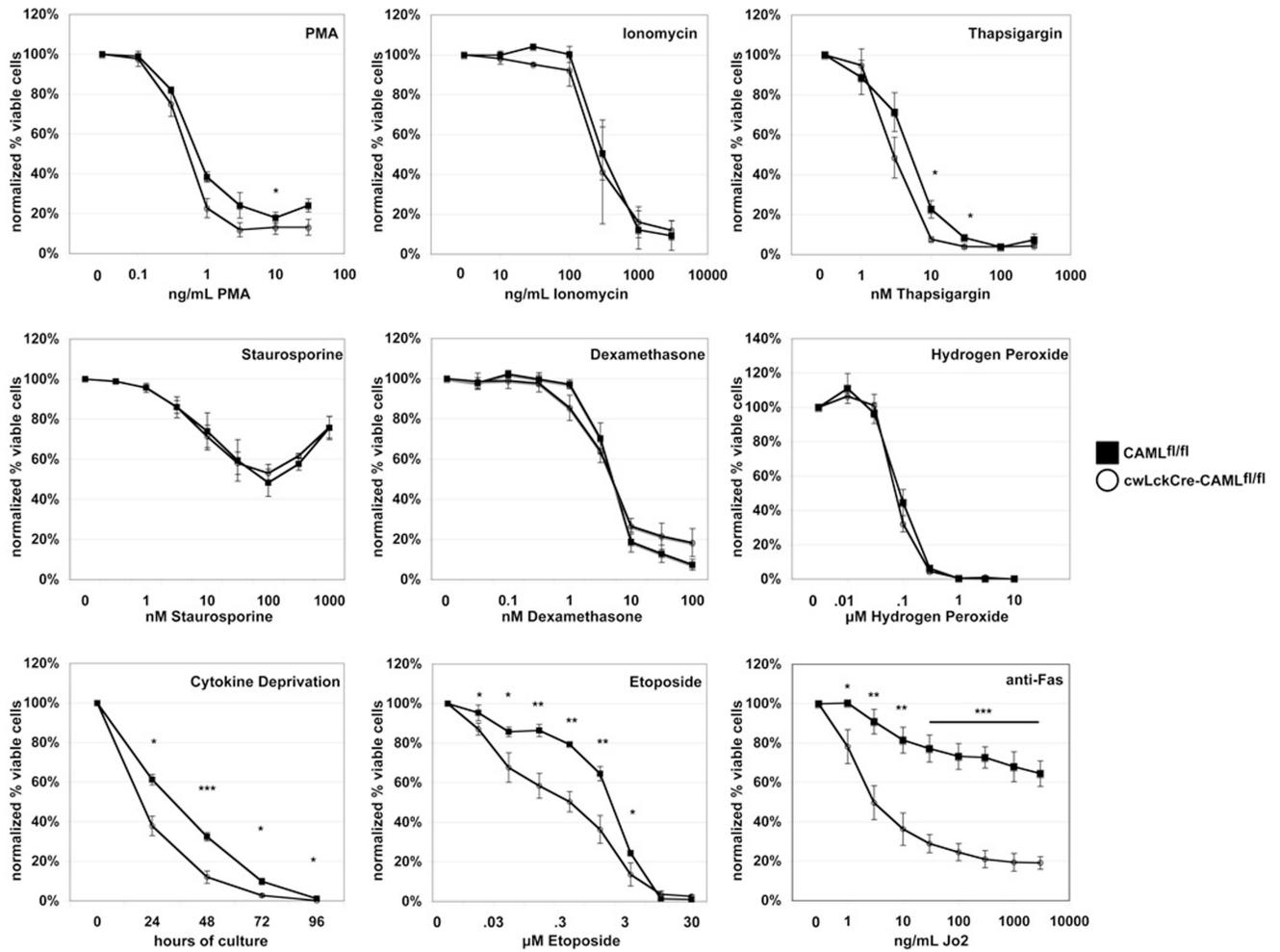


Figure 3. cwLckCre-CAML^{fl/fl} CD4⁺CD8⁺ thymocytes show abnormally increased susceptibility to a subset of apoptotic stimuli

Total thymocytes (1×10^6) were cultured for 14 h with graded concentrations of the indicated reagents. Cells were stained with anti-CD4 plus anti-CD8 antibodies, Annexin V-Cy5 and propidium iodide to quantify cell survival. The average of three to five independent experiments is shown. Error bars represent standard error of the mean. Cytokine deprivation: thymocytes (1×10^6) were cultured for four days in regular medium in the absence of additional exogenous cytokines. Cell survival was examined daily as described above. One representative of three independent experiments is shown. Error bars represent standard error of the mean. * $p < .05$ ** $p < .005$ *** $p < .001$

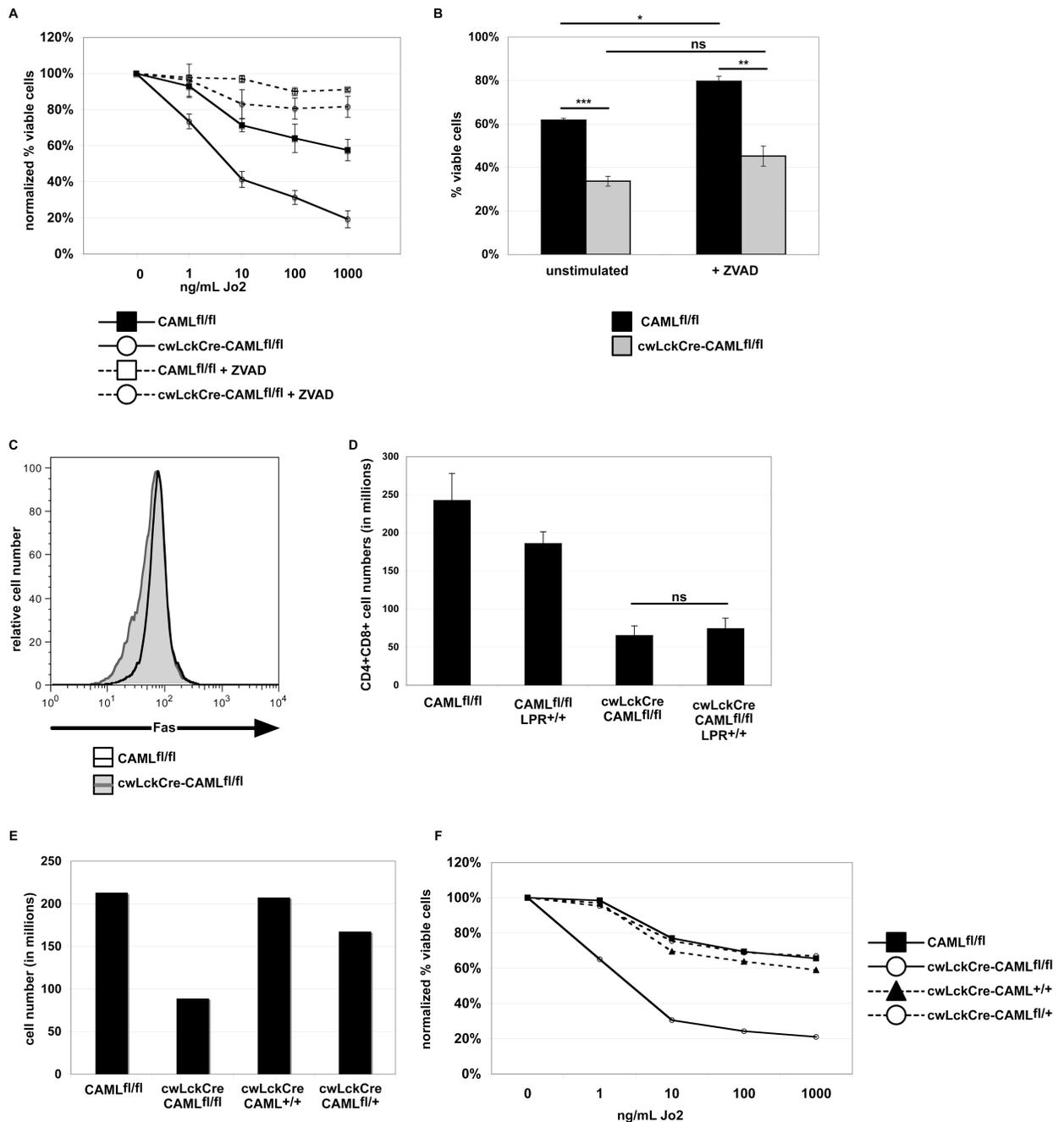


Figure 4. Fas is not essential for the abnormally increased apoptosis of cwLckCre-CAML^{fl/fl} thymocytes

(A) Caspase inhibition protects cwLckCre-CAML^{fl/fl} thymocytes against Fas-induced apoptosis. Thymocytes (1×10^6) were cultured for 14 h with graded concentrations of anti-mouse Fas antibody with or without 50 μ M ZVAD. Cells were stained with anti-CD4 plus anti-CD8 antibodies, Annexin V-Cy5 and propidium iodide to quantify cell survival. The average of three independent experiments is shown. (B) Non-normalized viability of thymocytes after an overnight incubation with or without 50 μ M ZVAD. Cell survival was quantified as described above. One representative of three independent experiments is

shown. (C) Thymocytes from *cwLckCre-CAML^{fl/fl}* mice express normal levels of Fas. Freshly isolated thymocytes from *cwLckCre-CAML^{fl/fl}* mice or littermate controls were stained for surface expression of CD4, CD8, and Fas. The histogram shows Fas surface expression on CD4⁺CD8⁺ cells. One representative of three independent experiments is shown. (D) Numbers of CD4⁺CD8⁺ thymocytes in *cwLckCre-CAML^{fl/fl}* and *cwLckCre-CAML^{fl/fl} lpr/lpr* mice. Error bars represent standard error of the mean. (E) Deletion of CAML, and not cre expression, is responsible for the decreased thymic cellularity and increased Fas susceptibility. Thymi from three to six week old mice *CAML^{fl/fl} cwLckCre-CAML^{fl/fl}* *cwLckCre-CAML^{+/+}* *cwLckCre-CAML^{fl/+}* were tweezed into single cell suspensions and counted. (F) Thymocytes (1×10^6) were cultured for 14 h with graded concentrations of anti-mouse Fas antibody (Jo2). Cells were stained with anti-CD4 plus anti-CD8 antibodies, Annexin V-Cy5 and propidium iodide to quantify cell survival. The average of two independent experiments is shown.

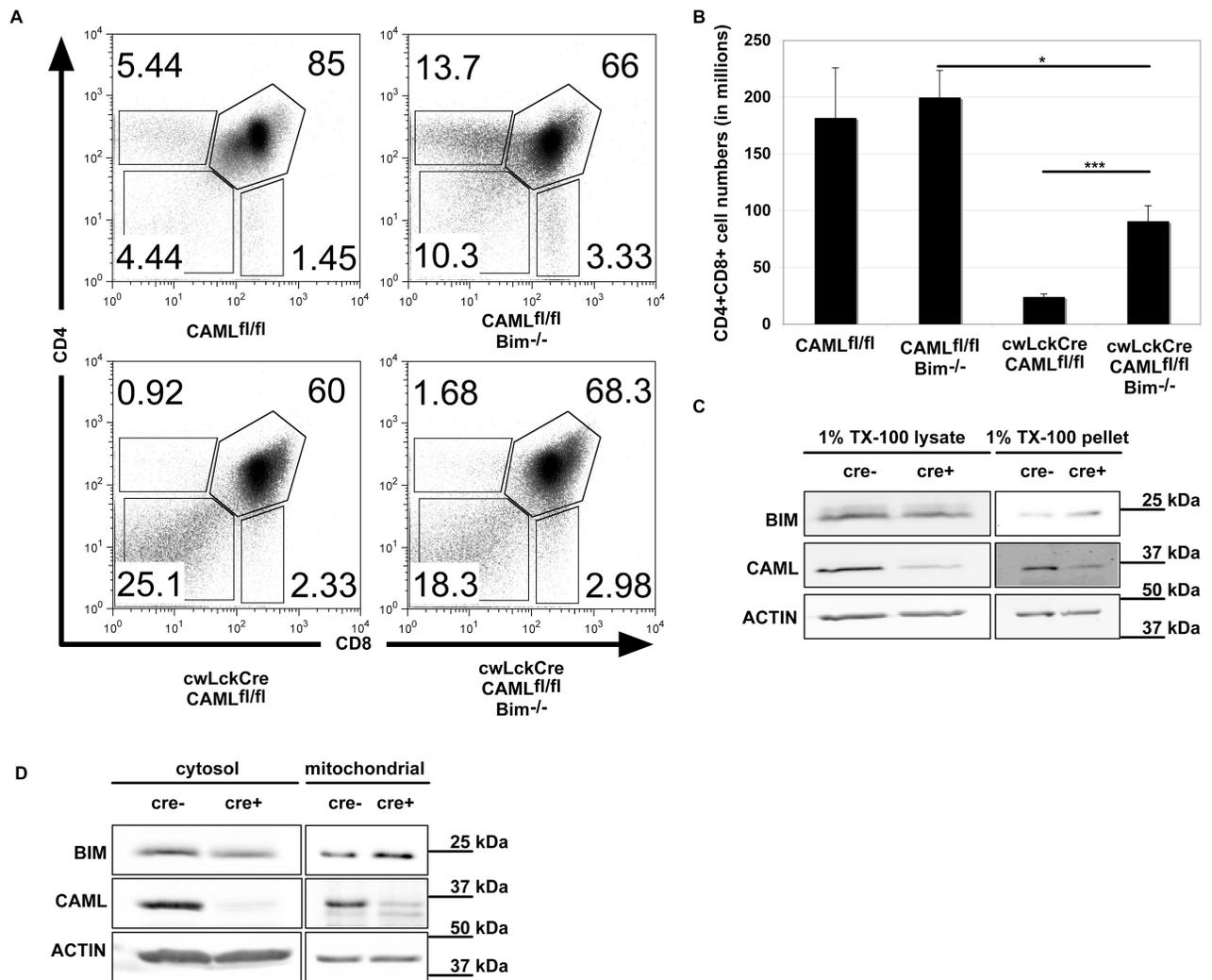


Figure 5. Bim deficiency significantly rescues intra-thymic T cell development in cwLckCre-CAML^{fl/fl}

(A) CD4/CD8 thymic profiles of freshly isolated thymocytes, comparing cwLckCre-CAML^{fl/fl} and cwLckCre-CAML^{fl/fl} Bim^{-/-} mice. (B) Quantification of thymocyte subsets defined by CD4 and CD8 expression (CD4⁻8⁻, CD4⁺8⁺, CD4⁺8⁻, CD4⁻8⁺). Error bars represent standard error of the mean. Total numbers of thymocytes are as follows: Bim^{+/+} CAML^{fl/fl}: 220×10^6 cells, Bim^{-/-} CAML^{fl/fl}: 312×10^6 cells, Bim^{+/+} cwLckCre-CAML^{fl/fl}: 40×10^6 cells and Bim^{-/-} cwLckCre-CAML^{fl/fl}: 136×10^6 cells. (C) CD4⁺8⁺ cells were isolated and lysed in 1% TX-100 lysis buffer. Lysates were centrifuged at 17,000 g for 10 minutes and pellets were suspended in Laemmli Sample buffer. Western blots show Bim levels in the supernatant and pellet fractions. (D) Mitochondria were isolated from CD4⁺8⁺ thymocytes and examined for levels of Bim.

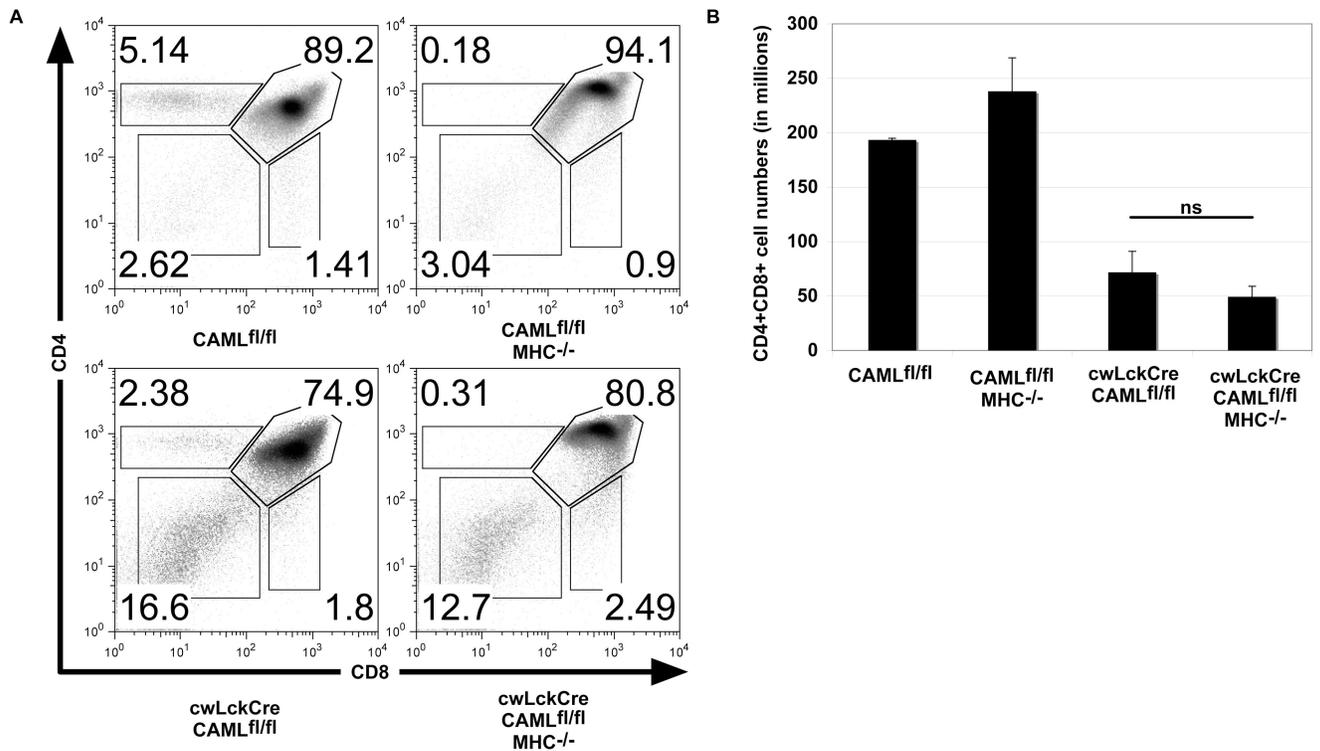


Figure 6. T cell receptor ligand (MHC) deficiency does not rescue the defect in thymocyte development in cwLckCre-CAML^{fl/fl} mice

(A) Representative flow cytometry profiles of anti-CD4 plus anti-CD8 antibody stained, freshly isolated thymocytes from CAML^{fl/fl} mice with or without the cwLckCre transgene that contained or lacked β 2-microglobulin and class II major histocompatibility (MHC) proteins (β 2-microglobulin^{-/-} and class II MHC^{-/-} mice are denoted MHC^{-/-}). (B) Total numbers of CD4⁺CD8⁺ thymocytes. Error bars represent standard error of the mean.

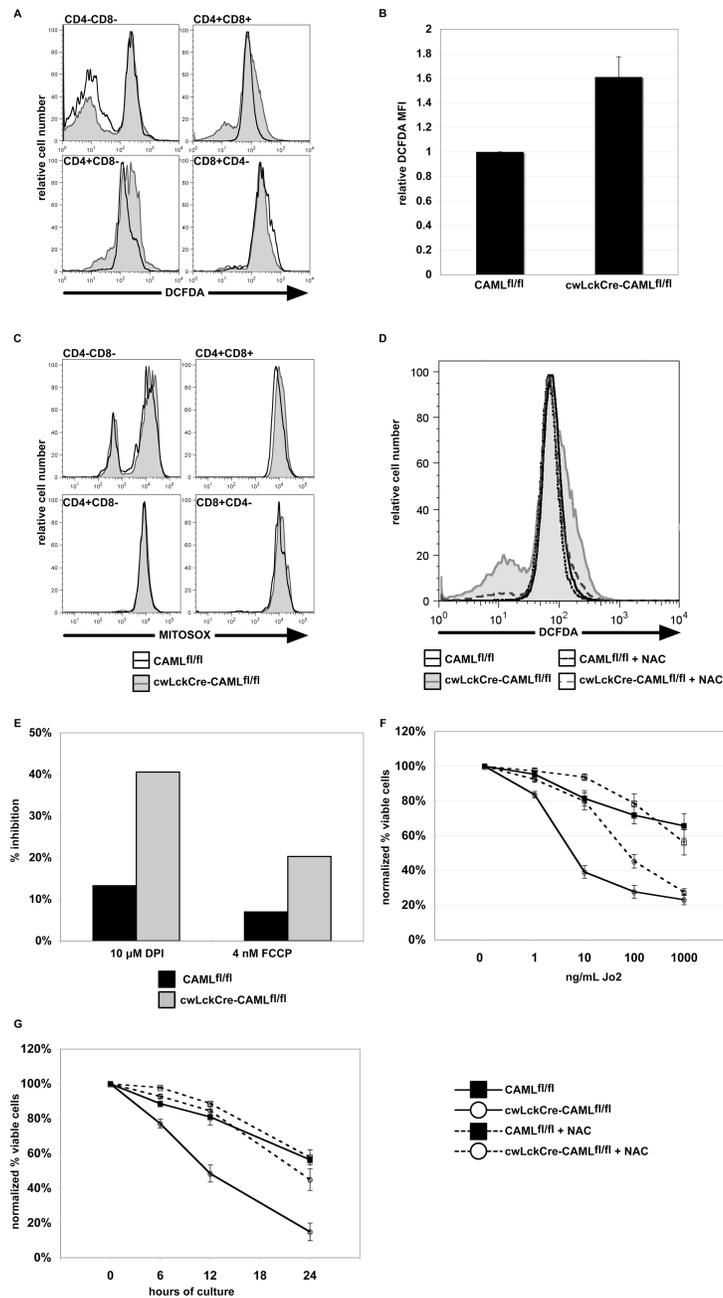


Figure 7. cwLckCre-CAML^{fl/fl} thymocytes contain abnormally increased levels of reactive oxygen species

(A) Freshly isolated thymocytes from cwLckCre-CAML^{fl/fl} mice and littermate controls were stained with anti-CD4 plus anti-CD8 antibodies and the reactive oxygen species (ROS)-specific dye 6-carboxy-2',7'-dichlorodihydrofluorescein diacetate, di(acetoxymethyl ester) (DCFDA). Histograms depicting ROS levels in each CD4/CD8 subset are shown. One representative of three independent experiments is shown. (B) Quantitation of the ROS staining in the CD4+CD8+ thymic sub-population. The average of three independent experiments is shown. (C) Freshly isolated thymocytes from cwLckCre-CAML^{fl/fl} mice and

littermate controls were stained with anti-CD4 plus anti-CD8 antibodies and the mitochondrial ROS-specific dye, Mitosox. Levels of mitochondrial ROS found in CD4⁻CD8⁻ and CD4⁺CD8⁺ thymic sub-populations are shown. One representative of three independent experiments is shown. (D) Mice of the indicated genotypes were fed NAC in their water at 10 mg/mL for two weeks. Thymocytes were removed and examined for levels of ROS via DCFDA staining. (E) Thymocytes were treated for 2 h with DPI plus FCCP and were then stained with DCFDA to determine percent inhibition of ROS levels. One representative experiment of two independent experiments is shown. (F) Antioxidant treatment reduces Fas-mediated apoptosis in cwLckCre-CAML^{fl/fl} thymocytes. Thymocytes (1×10^6) from mice of the indicated genotypes were cultured for 14 h with anti-mouse Fas antibody with or without N-acetyl cysteine, a potent antioxidant. Cells were stained with anti-CD4 plus anti-CD8 antibodies, Annexin V-Cy5 and propidium iodide to quantify cell survival. The average of three independent experiments is shown. Error bars represent standard error of the mean. DN double negative, DP double positive, SP single positive thymocytes. (G) Anti-oxidant treatment reduces cytokine deprivation induced apoptosis of cwLckCre-CAML^{fl/fl} thymocytes. Experimental set-up and analysis were the same as for (F) except neither serum nor anti-Fas antibody was added. One representative experiment of two is shown.