Contents lists available at ScienceDirect

Saudi Journal of Biological Sciences

journal homepage: www.sciencedirect.com

Original article

Determination of phylogenetic relationships among methicillin-resistant Staphylococcus aureus recovered from infected humans and Companion Animals

Hassan A. Hemeg

Department of Medical Laboratory Technology, College of Applied Sciences, Taibah University, P.O. Box 344, Al-Madinah Al-Munawarah, Saudi Arabia

ARTICLE INFO

Article history: Received 18 December 2020 Revised 9 January 2021 Accepted 11 January 2021 Available online 20 January 2021

Keywords: S. aureus Phylogenetic relationships Methicillin-resistant Staphylococcus aureus Companion animals Dogs Cats

ABSTRACT

Companion animals carry different microorganism of severely public health hazard for human; the kindness relation and contact between humans and companion animals may the route in the transmission of most zoonotic bacteria, including Methicillin-Resistant Staphylococcus aureus (MRSA). Therefore, the current study investigate the companion animals mainly dogs and cat as a reservoir for MRSA and the genetic similarity between the recovered strains of MRSA from such companion animals and their owners. One hundred swabs were collected under aseptic condition from companion animals and seventy swabs were collected from nasal and soft tissue of the infected owners in contact. All samples were examined with standard microbiological techniques, antimicrobial sensitivity, molecular typing and genetic finger printing using RAPD-PCR to determine the genetic finger printing of the recovered strains from humans and companion animals. The prevalence of the MRSA was higher in dog's swabs than human swabs. Dog swabs showed a rate of (44.4%), cat's revealed (27.3%), while the owner swabs could detect (42.8%). The antibiotics profiles were 69.2% and all MRSA strains were positive for mecA gene (100%), while only 25 strains (38.5%) were positive for Panton Valentine Leukocidin (PVL gene). Phylogenetic tree revealed 4 clusters with complete genetic relatedness and higher identity between the strains recovered from humans and companion animals. Our results revealed that there is great similarity between the recovered strains, indicating that pets play an important role in colonization and transmitting MRSA to humans, and vice versa.

© 2021 The Author(s). Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Animals are the natural reservoirs of microorganism's complex populations. Staphylococcus is one of the most opportunistic bacteria frequently isolated genera from animals and human beings (Vitale et al., 2006; Aklilu et al., 2012). Staphylococcus aureus is a colonizing organism for skin and upper respiratory tract of an innocuous component of the commensal flora and causes invasive infection in both human and animals (Benito et al., 2016). It presented exclusively in humans, and also found in apparent healthy and diseases companion animals (dogs and cats) that considered

Peer review under responsibility of King Saud University.

Production and hosting by Elsevier ELSEVIER

E-mail address: hasanhemeg@hotmail.com

https://doi.org/10.1016/j.sjbs.2021.01.017

1319-562X/© 2021 The Author(s). Published by Elsevier B.V. on behalf of King Saud University. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).









The kindness relation and contact between humans and companion animals may the route in the transmission of most zoonotic bacteria, including MRSA. Companion animals are potential sources of public health concern of MRSA for human. The incidence of MRSA transmission phenomenon is unclear between people and companion animals (Bergstrom et al., 2012; Bierowiec et al., 2016).

family members (Aires-de-Sousa 2017; Bierowiec et al., 2019).

Many authors suggested that humans are responsible for the initial colonization MRSA (Oehler et al., 2009; Smith and Pearson 2011) as they attract the infections with MRSA by Several exposures to hospitals and healthcare settings (Baptiste et al., 2005; Oehler et al., 2009). At the same times the isolates recovered from companion animals were often the same that recovered from infected humans (Weese et al., 2006; Faires et al., 2009). Indicating that there are recurrent infections and transmission of MRSA from infected outpatient's clinics (owners) to companion animals, and colonization of such strains occurs in such animal which become carriers and infect human cohabitants.

Now, the population prevalence of MRSA transmission to others is relatively high. Therefore, studying of MRSA infection in cats and dogs is important to understanding public hazard of MRSA. Several studies showed that MRSA clones circulating in cats and dogs are similar to the ones identified in humans and belong mostly to house-contact clones (HC-MRSA). This is clinically important since HC-MRSA isolates usually carry more virulence genes, and also more resistance genes than MRSA originating from companion animals (Cuny et al., 2015; Haenni et al., 2015; Loeffler et al., 2010).

The current study investigate the companion animals mainly dogs and cat as a reservoir for MRSA and their owners or persons in contact with them and to investigate the genetic similarity between the MRSA recovered from the companion animals like and their owners using RAPD-PCR.

2. Materials and methods

2.1. Samples

One hundred swabs were collected from companion animals clinically suffered from otitis media and upper respiratory diseases; cats (55 nasal and ear swabs) and dogs (45 nasal and ear swabs). All animals lived in households in close contact with their owners. Seventy swabs from nostril, pharynx, and infected tissues were collected from the owners suffering from upper respiratory tract and pyogenic infections. All samples were taken under the acceptance of the owners.

2.2. Isolation and identification of Staphylococcus species from samples

All swabs were subcultured in Mannitol Salt Agar plates (BBL, United Kingdom) then incubated for 24 h. The pure colony was identified by Gram staining morphology, and traditionally method biotyping (catalase, coagulase tube test and sugars fermentation test) (Mutters et al., 2016).

2.3. Susceptibility testing

All *S. aureus* detected at culture and identification step of samples were subjected to antimicrobial susceptibility test using the disk diffusion method (Price et al., 2012). Eleven antibiotics of veterinary and/or human interest were examined: penicillin G, cefoxitin, kanamycin, gentamicin, tetracycline, erythromycin, spiramycin, lincomycin, chloramphenicol, enrofloxacin and vancomycin (Oxoid, United Kingdom). The result was interpretated according to CLSI (CLSI 2015).

2.4. Molecular typing

The multi-drug resistance isolates (MRD) were examined using multiplex PCR for simultaneous detection of 16S rRNA of *Staphylococcus aureus*, *mecA* and Panton–Valentine leukocidin (PVL) genes reported by Moussa et al. (2012).

Using three primer pairs, one of them specific to 16S rRNA of *S. aureus* "Staph756F & Staph750R primers" which amplify 756 base pair fragments specific of *S. aureus*, the second one amplify 433 bp fragments and specific for PVL gene (Luk-PV-1 and Luk-PV-2 primers) reported by McClure et al. (2006a, 2006b). The third pair amplify 1399 base pair fragments specific for *mecA* gene (mecA F and mecA R primers) reported by Weller (1999). The reaction mixtures and the PCR condition were carried out according to Moussa et al. (2012). Finger printing of MRSA by RAPD- PCR had been carried out according to the methods reported by (Mehndiratta and Bhalla 2012) using short size primers and the PCR product have

been separated by electrophoresis on 1.5% agarose gel (Applichem, Germany, GmbH). The gel was photographed by a gel documentation system (Alpha Innotech, Biometra) and the data was analyzed through computer software.

3. Results

Positive microbiological data for *S. aureus* reveals Gram positive cocci arranged in clusters, golden yellow pin point colony, no effervescence of gas at catalase test, clotting rabbit plasma with coagulase test and ferment glucose.

Dog swabs show a rate of 44.4% (20/45), while, cats samples reveal 27.3% (15/55). The owner swabs detect 30 out of 70 with incidence 42.8%. All 65 *S. aureus* were identified from nasal swabs except two isolated from ear swabs from cases of otitis media. All *S. aureus* were subjected to antimicrobial susceptibility test using the disk diffusion method against 11 antibiotics. Forty five *S. aureus* were MRD with ratio 69.2%. The MRD profiles were: 10 were resistant for (penicillin G, cefoxitin, kanamycin, and gentamicin); 15 were resistant for (erythromycin, spiramycin, lincomycin, chloramphenicol, enrofloxacin); 18 were resistance for (penicillin G, erythromycin, spiramycin, kanamycin, vancomycin); two *S. aureus* isolated from cases of otitis media were resistant to all used antibiotic.

All MDR *S. aureus* were examined using multiplex PCR for simultaneous detection of 16S rRNA of *Staphylococcus aureus, mecA* and Panton–Valentine leukocidin (PVL) genes. The multiplex PCR could detect all the bacteriologically positive methicillin resistant *S. aureus* and all resistant strains for methicillin and oxacillin in few hours with 100% sensitivity and 100% specificity as shown in Fig. 1.

PVL genes were observed with only 25 strains (38.5%) as shown in Fig. 1.

RAPD – PCR of the recovered strains from human and companion using EP007, EP015, EP017, MN45 and KAY1 primers revealed characteristic RAPD profiles fingerprinting patterns based on the presence, size and the intensity of the amplified fragments. Moreover, it is noticed that the amplification reactions generated number of bands ranging from 3 to 10 bands with a molecular weight ranging from 180 up to 990 bp fragments as shown in Fig. 2. The majority of the examined MRSA had shared bands; but differs in their intensity. It's clear from the obtained data (Fig. 2) that the genetic profile of the strains recovered from humans and companions showed great similarity indicating the transmission of such strains between humans and companion animals.

The phylogenetic tree of the recovered MRSA from different species based on the similarity index between the strains, revealed 4 clusters. Most of the strains recovered from humans and companion were grouped in two clusters (3 and 4) and closely related



Fig. 1. Agarose gel electrophoresis showing multiplex PCR for simultaneous detection of 16S rRNA of *Staphylococcus aureus* (756 bp), *mecA* gene (1399 bp) and Panton–Valentine leukocidin (PVL) gene (733 bp).

H.A. Hemeg



Fig. 2. Agarose gel electrophoresis showing RAPD-PCR of MRSA recovered from Humans, dogs and cats using EP015, EP017, EP007, MN 45 and KAY1 short size primers.

to each other with high similarity (similarity index varied from 0.0800 up to 0.0900 between the strains). While clusters number one and number 2 contain only one strain as shown in Fig. 3.

4. Discussion

Staphylococcus aureus is a colonizing organism for skin and upper respiratory tract of an innocuous component of the commensal flora and causes invasive infection in both human and animals (Benito et al., 2016). The present work analyzed the relation of companion animals mainly dogs and cats in transmission of MRD *S. aureus* to their owners. The incidence of positive *S. aureus* were high in dogs then human and later in cats in comparison with the number of samples collected from each. The same results were documented by many authors (Vitale et al., 2006; Bierowiec et al., 2016; Aires-de-Sousa 2017). MRD means that *S. aureus* were resistant for at least 3 antibiotics, the collected data from the present research revealed 62.2% of *S. aureus* were MDR with different antibiotic profiles as expressed by Bierowiec et al. (2019).

The virulence of the CA-MRSA especially in severs soft tissues and skin infections and necrotizing infections is attributed to the PVL gene (Vitale et al., 2006; Bierowiec et al., 2016; Aires-de-Sousa 2017). *mecA* gene for MRD in *S. aureus* has been considered as a house keeping gene (Bierowiec et al., 2019; Benito et al., 2016). Therefore, multiplex PCR for simultaneous detection of 16S rRNA of *Staphylococcus aureus*, *mecA* and *Panton–Valentine leukocidin* (*PVL*) genes had been carried out.

The multiplex PCR could detect all the bacteriologically positive methicillin resistant *S. aureus* in few hours with 100% sensitivity and 100% specificity which confirm the conclusion of Moussa and



Fig. 3. Phylogenetic tree showing the genetic relatedness among MRSA recovered form humans and companion animals using EP015, EP017, EP007, MN 45 and KAY1 short size primers.

Shibl (2009) and Moussa et al. (2012). RAPD – PCR of the recovered strains from human and companion animals using EP007, EP015, EP017, MN45 and KAY1 primers revealed characteristic RAPD profiles fingerprinting patterns Fig. 2, Moreover, it is noticed that the amplification reactions generated number of bands similar in its molecular weight (shared bands) in all strains of humans and companion animals indicating the great similarity between The strains recovered from humans and pets (Loeffler et al., 2010; van Duijkeren, et al., 2011) however, these sharing bands differed in their intensity.

Most of the strains recovered from humans and companion were grouped in two clusters (3 and 4) and closely related to each other with high similarity (similarity index varied from 0.0800 up to 0.0900 between the strains) indicating the great similarity between The strains recovered from humans and pets (van Duijkeren et al., 2008; Frank et al., 2009). While, clusters number one and number 2 contain only one strain recovered from humans as shown in Fig. 3.

5. Conclusion

The previously revealed data confirmed that companion animals like dogs and cats are reservoirs for *S. aureus* and source of public health hazard especially with MRD *S. aureus* in owners.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- Aires-de-Sousa, M., 2017. Methicillin-resistant Staphylococcus aureus among animals: current overview. Clin. Microbiol. Infect. 23, 373–380.
- Aklilu, E., Zakaria, Z., Hassan, L., Hui Cheng, C., 2012. Methicillin-resistant Staphylococcus aureus in companion animals. PLoS ONE 24 (78)), e43329.
- Baptiste, K.E., Williams, K., Willams, N.J., Wattret, A., Clegg, P.D., 2005. Methicillinresistant *Staphylococcus aureus* in companion animals. Emerg. Infect. Dis. 11 (12), 1942–1944.
- Benito, D., Aspiroz, C., Gilaberte, Y., Sanmartin, R., Hernandez-Martin, A., Alonso, M., Gomez, P., Lozano, C., Torres, C., 2016. Genetic lineages and antimicrobial resistance genotypesin *Staphylococcus aureus* from children with atopic dermatitis: detection of clonal complexes CC1, CC97 and CC398. J. Chemother. 28, 359–366.
- Bergstrom, K., Aspan, A., Landen, A., Johnston, C., Gronlund-Andersson, U., 2012. The first nosocomial outbreak of methicillin-resistant *Staphylococcus aureus* in horses in Sweden. Acta Vet. Scand. 54, 11.
- Bierowiec, K., Korzeniowska-Kowal, A., Wzorek, A., Rypuła, K., Gamian, A., 2019. Prevalence of *Staphylococcus* species colonization in healthy and sick cats. Biomed Res. Int. 4360525.
- Bierowiec, K., Ploneczka-Janeczko, K., Rypula, K., 2016. Is the colonisation of *Staphylococcus aureus* in pets associated with their close contact with owners?. PLOS ONE 11, e0156052.
- CLSI, 2015. Performance Standards for Antimicrobial Susceptibility Testing; Ninth Informational Supplement, NCCLS Document. National Committee for Laboratory Standard, Wayne
- Cuny, C., Abdelbary, M., Layer, F., Werner, G., Witte, W., 2015. Prevalence of the immune evasion gene cluster in *Staphylococcus aureus* CC398. Vet. Microbiol. 177, 219–223.
- Faires, M., Tater, K., Weese, J.S., 2009. An investigation of methicillin-resistant *Staphylococcus aureus* colonization in people and pets in the same household with an infected person or infected pet. J. Am. Vet. Med. Assoc. 235 (5), 540.
- Frank, L., Kania, S., Kirzeder, E., Eberlein, L., Bemis, D., 2009. Risk of colonization or gene transfer to owners of dogs with meticillin-resistant *Staphylococcus* pseudintermedius. Vet. Dermatol. 20 (5–6), 496.
- Haenni, M., Chatre, P., Dupieux, C., Metayer, V., Maillard, K., Bes, M., 2015. mecC positive MRSA in horses. J. Antimicrob. Chemother. 70, 3401–3402.
- Loeffler, A., Pfeiffer, D.U., Lindsay, J.A., Magalhaes, R.J., Lloyd, D.H., 2010. Prevalence of and risk factors for MRSA carriage in companion animals: a survey of dogs, cats and horses. Epidemiol. Infect. 14, 1–10.
- McClure, J.A., Conly, J.M., Lau, V., Elsayed, S., Louie, T., Hutchins, W., Zhang, K., 2006a. Novel multiplex PCR assay for detection of the staphylococcal virulence marker Panton-Valentine leukocidin genes and simultaneous discrimination of

H.A. Hemeg

methicillin-susceptible from -resistant staphylococci. J. Clin. Microbiol. 44, 1141-2114.

- McClure, J.A., Conly, J.M., Lau, V., Elsayed, S., Louie, T., Hutchins, W., 2006b. Novel multiplex PCR assay for detection of the Staphylococcal virulence marker Panton-Valentine leukocidin genes and simultaneous discrimination of methicillin-susceptible from - resistant staphylococci. J. Clin. Microbiol. 44, 1141–1144.
- Mehndiratta, P.L., Bhalla, P., 2012. Typing of Methicillin resistant *Staphylococcus aureus*: a technical review. Indian J. Med. Microbiol. 30 (1), 16–23. https://doi.org/10.4103/0255-0857.93015. PMID: 22361755.
- Moussa, I., Kabli, S.A., Hemeg, H.A., Al-Garni, S.M., Shibl, A.M., 2012. A novel multiplex PCR for molecular characterization of methicillin resistant *Staphylococcus aureus* recovered from Jeddah, Kingdom of Saudi Arabia. Indian J. Med. Microbiol. 30 (3), 296–301.
- Moussa, I.M., Shibl, A.M., 2009. Molecular characterization of methicillin-resistant *Staphylococcus aureus* recovered from outpatient clinics in Riyadh, Saudi Arabia. Saudi Med. J. 2009 (30), 611–617.
- Mutters, N.T., Bieber, C.P., Hauck, C., Reiner, G., Malek, V., Frank, U., 2016. Comparison of livestock-associated and health care-associated MRSA-genes, virulence, and resistance. Diagn. Microbiol. Infect. Dis. 86, 417–421.
- Oehler, R., Velez, A., Mizrachi, M., Lamarche, J., Gompf, S., 2009. Bite-related and septic syndromes caused by cats and dogs. Lancet Infect. Dis. 9 (7), 439.
- Price, L.B., Stegger, M., Hasman, H., Aziz, M., Larsen, J., Andersen, P.S., 2012. *Staphylococcus aureus* CC398: host adaptation and emergence of methicillin resistance in livestock. mBio 4. e00520-12.

- Smith, T.C., Pearson, N., 2011. The emergence of Staphylococcus aureus ST398. Vector Borne Zoonotic Dis. 11 (4), 327–339.
- van Duijkeren, E., Houwers, D.J., Schoormans, A., Broekhuizen-Stins, M.J., Ikawaty, R., 2008. Transmission of methicillin-resistant *Staphylococcus intermedius* between humans and animals. Vet. Microbiol. 128 (1–2), 213–215.
- van Duijkeren, E., Kamphuis, M., van der Mije, I.C., Laarhoven, L.M., Duim, B., 2011. Transmission of methicillin-resistant *Staphylococcus pseudintermedius* between infected dogs and cats and contact pets, humans and the environment in households and veterinary clinics. Vet. Microbiol. 150 (3–4), 338–343.
- Vitale, C.B., Gross, T.L., Weese, J.S., 2006. Methicillin-resistant *Staphylococcus aureus* in cat and owner. Emerg. Infect. Dis. 12 (12), 1998–2000.
- Weese, J.S., Dick, H., Willey, B.M., McGeer, A., Kreiswirth, B.N., 2006. Suspected transmission of methicillin-resistant *Staphylococcus aureus* between domestic pets and humans in veterinary clinics and in the household. Vet. Microbiol. 15 (115(1–3)), 148–155.
- Weller, T.M., 1999. The distribution of *mecA*, *mecR*1 and *mec*1 and sequence analysis of *mec*1 and the *mec* promoter region in Staphylococci expressing resistance to methicillin. J. Antimicrob. Chemother. 1999 (43), 15–22.

Further Reading

Loeffler, A., Lloyd, D.H., 2010. Companion animals: a reservoir for methicillinresistant *Staphylococcus aureus* in the community?. Epidemiol. Infect. 138, 595– 605.