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Deubiquitinases and cancer: A snapshot

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ABSTRACT

Ubiquitination is the vital system for controlling protein degradation and regulation of basic cellular processes. Deubiquitinases (DUBs) are emerging as an important regulator of several pathways related to cancer and other diseases. Their ability to detach ubiquitin from the target substrate and regulation of signaling makes it potential target to treat cancer and other fatal diseases. In the current review, we are trying to summarize deubiquitination, and their role in cancer and potential small molecules DUBs inhibitors which can be used as drugs for cancer treatment.

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1. Introduction

Proteins are vital for the structure and function of the cells, and the regulation of protein synthesis is a prime aspect of cellular metabolism. About 30% of mammalian proteins are short lived, have very short half-life of less than 10 min and are rapidly degraded after translation (Schubert et al., 2000). Such a high level of protein degradation requires a dedicated system to regulate the selective unwanted protein degradation. Ubiquitin-proteasome system (UPS) has emerged as a key supervisor of protein function and stability. UPS has many vital roles in eukaryotic cellular processes including cell cycle progression, stress response, signal transduction, DNA repair, control of transcription factor activity and membrane trafficking (Coux et al., 1996; Hershko and

Ciechanover, 1998; Ciechanover et al., 2000a; Ciechanover, 2006; Welchman et al., 2005). Ubiquitin plays an important role to degrade proteins through proteasome targeting as well as by direct sorting to the lysosome. Ubiquitin is a small eukaryotic polypeptide which marks unwanted or damaged proteins for degradation, and the proteasome, is a large molecule breaks down protein into smaller peptides, to be used in other anabolic processes (D'Arcy et al., 2015). More than 80% of proteins are degraded by UPS and that is why it has emerged as an important player in the regulation of various cellular processes (Rock et al., 1994). UPS plays a pivotal role in the pathogenesis of many human diseases like cancer and neurodegenerative disorders (Ciechanover et al., 2000b).

The process of ubiquitination is a multi-step process ultimately leading to the covalent modification of a protein substrate with small molecule ubiquitin. There are three types of ubiquitination: 1) mono-ubiquitination in which single ubiquitin is attached to target 2) multi ubiquitination or poly-mono-ubiquitination where several single ubiquitin are attached to target proteins 3) poly ubiq-

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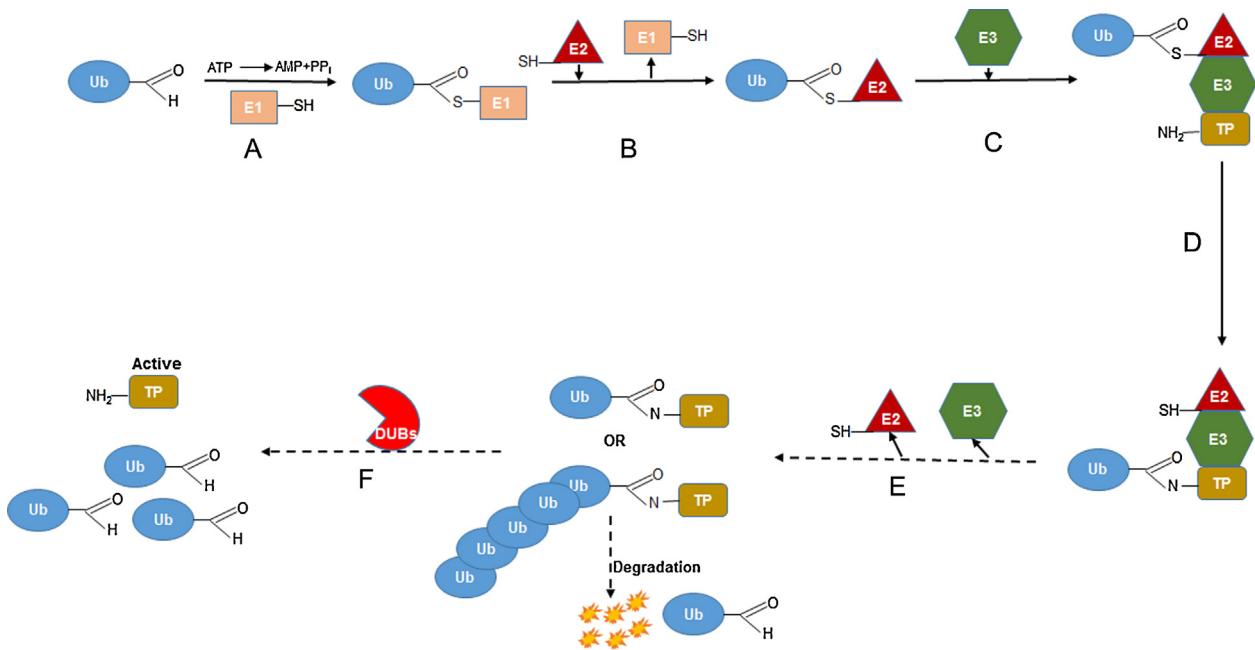


Fig. 1. Overview of ubiquitination and deubiquitination. TP; target protein, Ub; ubiquitin, E1, E2, and E3; different classes of ubiquitin, DUBs; deubiquinases.

uitination where substrate is attached with poly-ubiquitin chains (D'Arcy et al., 2015; Jentsch and Schlenker, 1995; Di Fiore et al., 2003; Lander et al., 2012). Ubiquitin has ≥ 76 conserved amino acid residues that covalently attach through a peptide bond between the carboxyl glycine residues at 76 position of ubiquitin to the amino groups of lysine residues in target proteins. The process of ubiquitination depends on the consecutive activity of three distinct enzymes, ubiquitin-activating enzyme (E1), ubiquitin-conjugating enzyme (E2) and ubiquitin-ligase or E3 ubiquitin ligase (E3) (fig. 1). In the first step, ubiquitin is activated by the E1 in the presence of ATP, forming a thio-ester bond between the carboxyl-terminal glycine residue of ubiquitin and the active site cysteine of the E1 enzyme. Once activated, ubiquitin is transferred from E1 to a cysteine residue of E2 ubiquitin carrier proteins. Substrate specificity is mediated by E3 ligases, which bind target substrates and coordinate the covalent attachment of ubiquitin. Two distinct families of E3 ligases exist, the HECT domain family that receives ubiquitin from the E2 ligase forming an ubiquitin-E3 intermediate, and the RING finger family of E3 ligases that form a molecular bridge between the E2 ligase and target proteins (D'Arcy et al., 2015; Ross et al., 2015; Voges et al., 1999; Pickart and Eddins, 2004). Once ubiquitin-E2, E3 and substrate complex is formed, ubiquitin binds to the substrate which is mediated by E3 ligase. In the subsequent process E2 and E3 are removed from complex and substrate protein will be either degraded or alternatively substrated could be free from ubiquitin through deubiquitinases and remain active (fig. 1).

2. Deubiquitination, deubiquitinases and cancer

Protein deubiquitination is reverse process of ubiquitination and performed by deubiquitinases or deubiquitinating enzymes (DUBs), which help in removal of ubiquitin from target proteins and involve in ubiquitin maturation, recycling and editing (Pföh et al., 2015; Kim et al., 2003; Amerik and Hochstrasser, 2004; Nijman et al., 2005a; Reyes-Turcu and Wilkinson, 2009; Clague et al., 2013; Tyagi et al., 2015a). The function of deubiquitinases however, are rescuing the protein which is marked for degradation, and releasing ubiquitin from target proteins substrate. Dubs are playing an important role in cell growth, apoptosis and cancer

(Pföh et al., 2015; Hu et al., 2005; Avvakumov et al., 2006; Renatus et al., 2006) and are associated with 26S proteasome to rescue ubiquitin chain before the degradation of the substrate protein. These free polyubiquitin chains are processed by other DUBs to restore the ubiquitin in the cell (Pföh et al., 2015). Approximately hundred human DUBs are discovered till date and these are divided into four classes; (1) ubiquitin specific proteases (USP), (2) ubiquitin C-terminal hydrolases (UCH), (3) ovarian tumor proteases (OTU), (4) Josephins and the Jab1/MPN/MOV34 metalloenzymes (JAMM). USPs are the major DUBs and are associated with most of the cancers and play an important role in regulation of various pathways, for example in Fanconi Anemia (FA). FA is a genomic instability syndrome characterized by bone marrow failure, developmental abnormalities and increased probability of cancers (Kee and D'Andrea, 2010). There are many reports, based on clinical investigation indicated that FA is a chromosomal instability disorder, and cells from FA patients accumulate DNA damage at an increased rate. Mono-ubiquitination of FANCD2 and FANCI is a vital event in FA pathway, downstream of this pathway, it interacts with FAND1, FANCN, FANCJ and BRCA1 (Knipscheer et al., 2009). USP1/UAF1 is known to deubiquitinases FANCD2 and FANCI. Disruption of USP1/UAF1 complex promote level of FANCD2/FANCI ubiquitination and DNA repair defects, suggesting a failure in the completion of FA pathway (Nijman et al., 2005b; Smogorzewska et al., 2007). ELG1 (Enhanced Levels of Genomic Instability) are recently found associated with USP1/USF1 and may play a role in the successful completion of the FA pathway (Lee et al., 2010; Yang et al., 2011; Shkedy et al., 2015).

Ubiquitin C-terminal hydrolase-L1 (UCLL1), is one of the most explored DUBs which is involved in neurodegenerative disorders such as Parkinson disease. It has been shown that expression of neurons of neuroendocrine system and gonads is unregulated in non-small cell lung cancer (Hibi et al., 1998; Hibi et al., 1999), oesophageal cancer (Takase et al., 2003) invasive colorectal cancer (Yamazaki et al., 2002) and pancreatic cancer (Tezel et al., 2000). Overexpression of the UCHL-1 has also been related with tumor progression, size, invasiveness and apoptosis in breast cancer (Wang et al., 2008).

Table 1
Deubiquitinases and their mechanism of action.

DUBs	Pathway	DUBs	Pathway
USP1	Fanconi anemia, DNA replication (Nijman et al., 2005b; Huang et al., 2006)	USP20	Deubiquitinates and stabilizes HIF1- α (Li et al., 2005)
USP2	Regulation through androgen synthesis and fatty acid synthetase (Graner et al., 2004)	USP21	Deubiquitinates histone H2A, activating transcription (Nakagawa et al., 2008)
USP3	Cell cycle regulation (Nicassio et al., 2007)	USP39, USP44	Involve in mitotic spindle checkpoint (van Leuken et al., 2008; Stegmeier et al., 2007)
USP4	Regulate WNT signaling pathway (Zhao et al., 2009)	OTUB1	Interacts with estrogen receptor alpha and negatively regulates ER- α -mediated transcription (Kee and D'Andrea, 2010)
USP5	Cell cycle regulation (Dayal et al., 2009)	A20	Negatively regulates NF- κ B signaling (Sacco et al., 2010)
USP6	Actin remodeling (Masuda-Robens et al., 2003)	Cezanne	Dowun-regulates NF- κ B signaling (Enesa et al., 2008)
USP7	Regulation of p53 (Sacco et al., 2010)	TRABID	Positive regulator of WNT signaling required for TCF-mediated transcription (Tran et al., 2008)
USP8	Stabilization of ESCRT components (Huang et al., 2009; Schweitzer et al., 2007; Xu et al., 2009)	POH1	Regulates ErbB2 ubiquitination (Liu et al., 2009)
USP9X	Regulation of AMPK kinase family	UCHL	Mechanism unknown
USP11	Interact with BRCA2 and stabilizes I κ B	UCHL5	Regulates TGF β signaling (Wicks et al., 2005)

Table 2
Deubiquitinases involved in different types of cancer.

Cancer	Deubiquitinase involved
Lung	USP2, USP4, USP5, USP8, USP18, A20, USP33, CYLD, OTUB1 (Sacco et al., 2010; Komander et al., 2009; Dikic et al., 2009)
Prostate	USP1, USP33, BRCC36 (Sacco et al., 2010; Dikic et al., 2009)
Brain	USP1, USP2, USP3, USP4, USP5, USP6, USP8, A20, USP18, USP22, USP33, USP39, USP44, CYLD, AMSH-LP, MYSM, POH1, UCHL, UCHL5, OUTB1, TRABID, A20 (Renatus et al., 2006; Kee and D'Andrea, 2010; Dikic et al., 2009)
Leukemia	USP1, USP3, USP5, USP8, USP9X, USP20, USP21, USP33, USP44, CYLD, AMSH, POH1 (Renatus et al., 2006; Dikic et al., 2009)
Colon	USP2, USP5, USP7, USP9X, USP21, USP22, AMSH, BRC36, POH1, UCHL1 (Renatus et al., 2006; Dikic et al., 2009)
Ovarian	USP1, USP5, USP18, USP39, CYLD, BAP1, UCHL1, UCH25, A20 (Renatus et al., 2006; Dikic et al., 2009)
Breast	USP7, USP15, USP33, POH1, UCHL5 (Sacco et al., 2010)
Pancreas	USP11, USP39 (Sacco et al., 2010)
Kidney	BAP1 (Dikic et al., 2009)
Gastric	USP1, USP44, UCHL1 (Sacco et al., 2010)
Fanconi Anemia	USP1, UAF1 (Dikic et al., 2009)
Liver	USP1, USP4, USP5, USP15, USP21, AMSH, AMSH-LP, CEZANNE, TRABD (Sacco et al., 2010)

Mechanism of action and potential role of different DUBs in the different cancers is well established (D'Arcy et al., 2015; Pföh et al., 2015; Sacco et al., 2010) and shown in Tables 1 and 2.

3. Therapeutic potential of DUBs for the treatment of cancer

Accumulation of genetic mutations and aberrant signaling of various growth and survival related pathways in cancer cells (Tyagi et al., 2015b; Tyagi et al., 2015c; Srivastava et al., 2015; Arora et al., 2015; Tyagi et al., 2014) leads to the clinical diversity and therapeutic resistance. However, advance understanding of the complex biology of cancer cells (Tyagi and Ghosh, 2011; Tyagi et al., 2011; Bhardwaj et al., 2014; Deshmukh et al., 2015) and the involvement of deubiquitinating enzymes in cancer (Schubert et al., 2000; Ciechanover, 2006; Pföh et al., 2015) reveals the therapeutic potential of DUBs for the treatment of cancer. As mentioned in Table 2, DUBs are playing an important role in different type of cancers. More extensive studies are needed in this area of research to explore the detailed mechanism and target of DUBs. Study and designing the targeted small molecule DUB inhibitors, will most probably a new therapeutics for different types of cancers. DUBs have been identified in most of the cancer (Table 2) and targeting them can be an effective way to treat, diagnose and prevent the disease.

DUBs are involved in cell cycle regulation and DNA damage pathways and since, in cancer and different stress conditions all the proteins which regulate cell cycle and DNA damage/repair are either up/down regulated (Singh et al., 2013, 2011; Kumar and de Massy, 2010; Gupta et al., 2010), it would be interesting to investigate the potential role of DUBs in cell cycle and DNA

damage/repair. In most of the cancers, tumor suppresser genes are degraded by ubiquitin which can be rescued by application of controlled mechanism of enhancing deubiquitinases in the tumor cells, to prevent degradation of tumor suppresser proteins. Development of small molecules which inhibits DUBs would be a great idea to target cancer cells and it will be facilitated by the development of suitable high throughput screening. Two molecules were identified that inhibit the SARS coronavirus DUBs papain like protease (PLpro) (Ratia et al., 2008; Ghosh et al., 2009). Some small molecules inhibit USP7 and USP8 (Daviet and Colland, 2008). Compound like G5 and F6 were reported as total DUBs inhibitor (Aleo et al., 2006) along with USP2, USP7 and SENP2 deSUMOylase (Aleo et al., 2006; Fontanini et al., 2009; Nicholson et al., 2008). b-AP15 was identified as an USP14 and UCHL5 inhibitor at a concentration of about 5 μ M (D'Arcy et al., 2011). AM146, RA-14, RAMB1, RA-9 and WP1130 are other molecules which inhibit many DUBs including USPs and UCH and induce accumulation of polyubiquitinated proteins (Anchoori et al., 2011; Issaenko and Amerik, 2012; Kapuria et al., 2010). WP1130 in combination with bortezomib had showed antitumor activity in mantle cell lymphoma (Pham et al., 2010). Several other small molecules have been reported as DUBs inhibitors like Eeyarestatin (Fiebiger et al., 2004), Velcade (Bold, 2004) and Kyprolis (Steele, 2013).

4. DUBs in immunotherapy

Nowadays immunotherapy is widely used to treat many diseases including cancer. Immunotherapy is the treatment that uses person's immune system to fight diseases and this can be either stimulate the immune system to work harder and cleverer or providing immune system machineries to escalation of immunity.

DUBs widely involve in the regulation of cell signaling and these signaling pathways are frequently altered in most of the cancers.

P^{53} is a well known tumor repressor protein involves in cell cycle control and frequently mutated in tumor cells (Harris and Levine, 2005). Many of the USPs involve in P^{53} regulation. USP7 and USP15 involve in the control of P^{53} -MDM2 pathway by regulating the stability of both P^{53} and MDM2 (Kon et al., 2010; Zou et al., 2014). USP2, USP4, USP5, USP10 and USP29 also involve in the regulation of P^{53} activity (D'Arcy et al., 2015). TNF- κB is another vital player in an immune response system which frequently deregulated and constitutively activated in cancer cells. Many DUBs like A20 and CYLD act as tumor repressor through their ability to downregulate TNF- κB signaling by acting on several components of the pathway (Harhaj and Dixit, 2012). USP21 and Cezanne inhibit TNF- κB activation by regulating ubiquitin level of RIPK1 (D'Arcy et al., 2015). Dr. Greenberg group from University of Pennsylvania showed that BRCC36 containing deubiquitinating complex BRISC which is also a sister protein complex of nuclear RAP80-BRCA1 complex, deubiquitinates type I interferon receptor (IFNAR1), resulting in its delayed lysosomal dependent degradation. They showed that BRISC deficient cells shows reduced inflammatory gene expression and BRISC deficient mice have an attenuated interferon response with a survival advantage from a LPS dependent septic shock (Zheng et al., 2013). All the above mentioned DUSs could be the potential candidate for immunotherapy.

5. Future perspectives

DUBs can be the potential target for treatment of many cancer or other fatal diseases but it needs to be investigated and explored. Small molecules inhibitors against DUBs could enhance the probability to treat various vital diseases. DUBs emerge as a promising target in the development of new disease specific treatment.

Conflict of interest

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References

- Aleo, E., Henderson, C.J., Fontanini, A., Solazzo, B., Brancolini, C., 2006. Identification of new compounds that trigger apoptosis-independent caspase activation and apoptosis. *Cancer Res.* 66, 9235–9244.
- Amerik, A.Y., Hochstrasser, M., 2004. Mechanism and function of deubiquitinating enzymes. *Biochim. Biophys. Acta* 1695, 189–207.
- Anchoori, R.K., Khan, S.R., Sueblinvong, T., Felthausser, A., Iizuka, Y., Gavioli, R., Destro, F., Isaksson, V.R., Peng, S., Roden, R.B., Bazzaro, M., 2011. Stressing the ubiquitin-proteasome system without 20S proteolytic inhibition selectively kills cervical cancer cells. *PLoS One* 6, e23888.
- Arora, S., Tyagi, N., Bhardwaj, A., Rusu, L., Palanki, R., Vig, K., Singh, S.R., Singh, A.P., Palanki, S., Miller, M.E., Carter, J.E., Singh, S., 2015. Silver nanoparticles protect human keratinocytes against UVB radiation-induced DNA damage and apoptosis: potential for prevention of skin carcinogenesis. *Nanomedicine* 11, 1265–1275.
- Avvakumov, G.V., Walker, J.R., Xue, S., Finerty Jr., P.J., Mackenzie, F., Newman, E.M., Dhe-Paganon, S., 2006. Amino-terminal dimerization, NRD1-rhodanese interaction, and inhibited catalytic domain conformation of the ubiquitin-specific protease 8 (USP8). *J. Biol. Chem.* 281, 38061–38070.
- Bhardwaj, A., Srivastava, S.K., Singh, S., Arora, S., Tyagi, N., Andrews, J., McClellan, S., Carter, J.E., Singh, A.P., 2014. CXCL12/CXCR4 signaling counteracts docetaxel-induced microtubule stabilization via p21-activated kinase 4-dependent activation of LIM domain kinase 1. *Oncotarget* 5, 11490–11500.
- Bold, R., 2004. Development of the proteasome inhibitor velcade (Bortezomib) by julian adams Ph.D., and michael kauffman, M.D., Ph.D. *Cancer Invest.* 22, 328–329.
- Ciechanover, A., Orian, A., Schwartz, A.L., 2000a. Ubiquitin-mediated proteolysis: biological regulation via destruction. *Bioessays* 22, 442–451.
- Ciechanover, A., Orian, A., Schwartz, A.L., 2000b. The ubiquitin-mediated proteolytic pathway: mode of action and clinical implications. *J. Cell Biochem.* 34, 40–51 (40–51).
- Ciechanover, A., 2006. The ubiquitin proteolytic system: from an idea to the patient bed. *Proc. Am. Thorac. Soc.* 3, 21–31.
- Clague, M.J., Barsukov, I., Coulson, J.M., Liu, H., Rigden, D.J., Urbe, S., 2013. Deubiquitylases from genes to organism. *Physiol. Rev.* 93, 1289–1315.
- Coux, O., Tanaka, K., Goldberg, A.L., 1996. Structure and functions of the 20S and 26S proteasomes. *Annu. Rev. Biochem.* 65, 801–847, 801–847.
- D'Arcy, P., Brnjic, S., Olofsson, M.H., Fryknas, M., Lindsten, K., De, C.M., Perego, P., Sadeghi, B., Hassan, M., Larsson, R., Linder, S., 2011. Inhibition of proteasome deubiquitinating activity as a new cancer therapy. *Nat. Med.* 17, 1636–1640.
- D'Arcy, P., Wang, X., Linder, S., 2015. Deubiquitinase inhibition as a cancer therapeutic strategy. *Pharmacol. Ther.* 147, 32–54.
- Daviet, L., Colland, F., 2008. Targeting ubiquitin specific proteases for drug discovery. *Biochimie* 90, 270–283.
- Dayal, S., Sparks, A., Jacob, J., Allende-Vega, N., Lane, D.P., Saville, M.K., 2009. Suppression of the deubiquitinating enzyme USP5 causes the accumulation of unanchored polyubiquitin and the activation of p53. *J. Biol. Chem.* 284, 5030–5041.
- Deshmukh, S.K., Srivastava, S.K., Bhardwaj, A., Singh, A.P., Tyagi, N., Marimuthu, S., Dyess, D.L., Dal, Z.V., Carter, J.E., Singh, S., 2015. Resistin and interleukin-6 exhibit racially-disparate expression in breast cancer patients, display molecular association and promote growth and aggressiveness of tumor cells through STAT3 activation. *Oncotarget* 6, 11231–11241.
- Di Fiore, P.P., Polo, S., Hofmann, K., 2003. When ubiquitin meets ubiquitin receptors: a signalling connection. *Nat. Rev. Mol. Cell Biol.* 4, 491–497.
- Dikic, I., Wakatsuki, S., Walters, K.J., 2009. Ubiquitin-binding domains – from structures to functions. *Nat. Rev. Mol. Cell Biol.* 10, 659–671.
- Enesa, K., Zakkar, M., Chaudhury, H., Luong, I.A., Rawlinson, L., Mason, J.C., Haskard, D.O., Dean, J.L., Evans, P.C., 2008. NF- κB suppression by the deubiquitinating enzyme Cezanne: a novel negative feedback loop in pro-inflammatory signaling. *J. Biol. Chem.* 283, 7036–7045.
- Fiebig, E., Hirsch, C., Vyas, J.M., Gordon, E., Ploegh, H.L., Tortorella, D., 2004. Dissection of the dislocation pathway for type I membrane proteins with a new small molecule inhibitor eeyarestatin. *Mol. Biol. Cell* 15, 1635–1646.
- Fontanini, A., Foti, C., Potu, H., Crivellato, E., Maestro, R., Bernardi, P., Demarchi, F., Brancolini, C., 2009. The isopeptidase inhibitor G5 triggers a caspase-independent necrotic death in cells resistant to apoptosis: a COMPARATIVE STUDY WITH THE PROTEASOME INHIBITOR BORTEZOMIB. *J. Biol. Chem.* 284, 8369–8381.
- Ghosh, A.K., Takayama, J., Aubin, Y., Ratia, K., Chaudhuri, R., Baez, Y., Sleeman, K., Coughlin, M., Nichols, D.B., Mulhearn, D.C., Prabhakar, B.S., Baker, S.C., Johnson, M.E., Mesecar, A.D., 2009. Structure-based design synthesis, and biological evaluation of a series of novel and reversible inhibitors for the severe acute respiratory syndrome-coronavirus papain-like protease. *J. Med. Chem.* 52, 5228–5240.
- Graner, E., Tang, D., Rossi, S., Baron, A., Migita, T., Weinstein, L.J., Lechpmann, M., Huesken, D., Zimmermann, J., Signoretti, S., Loda, M., 2004. The isopeptidase USP2a regulates the stability of fatty acid synthase in prostate cancer. *Cancer Cell* 5, 253–261.
- Gupta, V., Kalaiaras, P., Faheem, M., Singh, N., Iqbal, M.A., Bamezai, R.N., 2010. Dominant negative mutations affect oligomerization of human pyruvate kinase M2 isozyme and promote cellular growth and polyploidy. *J. Biol. Chem.* 285, 16864–16873.
- Harhaj, E.W., Dixit, V.M., 2012. Regulation of NF- κB deubiquitinases. *Immunol. Rev.* 246, 107–124.
- Harris, S.L., Levine, A.J., 2005. The p53 pathway: positive and negative feedback loops. *Oncogene* 24, 2899–2908.
- Hershko, A., Ciechanover, A., 1998. The ubiquitin system. *Annu. Rev. Biochem.* 67, 425–479, 425–479.
- Hibi, K., Liu, Q., Beaudry, G.A., Madden, S.L., Westra, W.H., Wehage, S.L., Yang, S.C., Heitmiller, R.F., Bertelsen, A.H., Sidransky, D., Jen, J., 1998. Serial analysis of gene expression in non-small cell lung cancer. *Cancer Res.* 58, 5690–5694.
- Hibi, K., Westra, W.H., Borges, M., Goodman, S., Sidransky, D., Jen, J., 1999. PGP9.5 as a candidate tumor marker for non-small-cell lung cancer. *Am. J. Pathol.* 155, 711–715.
- Hu, M., Li, P., Song, L., Jeffrey, P.D., Chenova, T.A., Wilkinson, K.D., Cohen, R.E., Shi, Y., 2005. Structure and mechanisms of the proteasome-associated deubiquitinating enzyme USP14. *EMBO J.* 24, 3747–3756.
- Huang, T.T., Nijman, S.M., Mirchandani, K.D., Galardy, P.J., Cohn, M.A., Haas, W., Gygi, S.P., Ploegh, H.L., Bernards, R., 2006. and A.D. D'Andrea: regulation of monoubiquitinated PCNA by DUB autocleavage. *Nat. Cell Biol.* 8, 339–347.
- Huang, X., Langelotz, C., Hetfeld-Pechoc, B.K., Schwenk, W., Dubiel, W., 2009. The COP9 signalosome mediates beta-catenin degradation by deneddylation and blocks adenomatous polyposis coli destruction via USP15. *J. Mol. Biol.* 391, 691–702.
- Issaenko, O.A., Amerik, A.Y., 2012. Chalcone-based small-molecule inhibitors attenuate malignant phenotype via targeting deubiquitinating enzymes. *Cell Cycle* 11, 1804–1817.

- Jentsch, S., Schlenker, S., 1995. Selective protein degradation: a journey's end within the proteasome. *Cell* 82, 881–884.
- Kapuria, V., Peterson, L.F., Fang, D., Bornmann, W.G., Talpaz, M., Donato, N.J., 2010. Deubiquitinase inhibition by small-molecule WP1130 triggers aggresome formation and tumor cell apoptosis. *Cancer Res.* 70, 9265–9276.
- Kee, Y., D'Andrea, A.D., 2010. Expanded roles of the Fanconi anemia pathway in preserving genomic stability. *Genes Dev.* 24, 1680–1694.
- Kim, J.H., Park, K.C., Chung, S.S., Bang, O., Chung, C.H., 2003. Deubiquitinating enzymes as cellular regulators. *J. Biochem.* 134, 9–18.
- Knipscheer, P., Raschle, M., Smogorzewska, A., Enoui, M., Ho, T.V., Scharer, O.D., Elledge, S.J., Walter, J.C., 2009. The Fanconi anemia pathway promotes replication-dependent DNA interstrand cross-link repair. *Science* 326, 1698–1701.
- Komander, D., Clague, M.J., Urbe, S., 2009. Breaking the chains: structure and function of the deubiquitinases. *Nat. Rev. Mol. Cell Biol.* 10, 550–563.
- Kon, N., Kobayashi, Y., Li, M., Brooks, C.L., Ludwig, T., Gu, W., 2010. Inactivation of HAUSP invivo modulates p53 function. *Oncogene* 29, 1270–1279.
- Kumar, R., de Massy, Bernard, 2010. Initiation of meiotic recombination in mammals. *Genes* 1, 521–549.
- Lander, G.C., Estrin, E., Matyskiela, M.E., Bashore, C., Nogales, E., Martin, A., 2012. Complete subunit architecture of the proteasome regulatory particle. *Nature* 482, 186–191.
- Lee, K.Y., Yang, K., Cohn, M.A., Sikdar, N., D'Andrea, A.D., Myung, K., 2010. Human ELG1 regulates the level of ubiquitinated proliferating cell nuclear antigen (PCNA) through its interactions with PCNA and USP1. *J. Biol. Chem.* 285, 10362–10369.
- Li, Z., Wang, D., Messing, E.M., Wu, G., 2005. VHL protein-interacting deubiquitinating enzyme 2 deubiquitinates and stabilizes HIF-1alpha. *EMBO Rep.* 6, 373–378.
- Liu, H., Buus, R., Clague, M.J., Urbe, S., 2009. Regulation of ErbB2 receptor status by the proteasomal DUB POH1. *PLoS One* 4, e5544.
- Masuda-Robens, J.M., Kutney, S.N., Qi, H., Chou, M.M., 2003. The TRE17 oncogene encodes a component of a novel effector pathway for Rho GTPases Cdc42 and Rac1 and stimulates actin remodeling. *Mol. Cell Biol.* 23, 2151–2161.
- Nakagawa, T., Kajitani, T., Togo, S., Masuko, N., Ohdan, H., Hishikawa, Y., Koji, T., Matsuyama, T., Ikura, T., Muramatsu, M., Ito, T., 2008. Deubiquitylation of histone H2A activates transcriptional initiation via trans-histone cross-talk with H3K4 di- and trimethylation. *Genes Dev.* 22, 37–49.
- Nicassio, F., Corrado, N., Vissers, J.H., Areces, L.B., Bergink, S., Marteijn, J.A., Geverts, B., Houtsmuller, A.B., Vermeulen, W., Di Fiore, P.P., Citterio, E., 2007. Human USP3 is a chromatin modifier required for S phase progression and genome stability. *Curr. Biol.* 17, 1972–1977.
- Nicholson, B., Leach, C.A., Goldenberg, S.J., Francis, D.M., Kodrasov, M.P., Tian, X., Shanks, J., Sterner, D.E., Bernal, A., Mattern, M.R., Wilkinson, K.D., Butt, T.R., 2008. Characterization of ubiquitin and ubiquitin-like-protein isopeptidase activities. *Protein Sci.* 17, 1035–1043.
- Nijman, S.M., Luna-Vargas, M.P., Velds, A., Brummelkamp, T.R., Dirac, A.M., Sixma, T.K., Bernards, R., 2005a. A genomic and functional inventory of deubiquitinating enzymes. *Cell* 123, 773–786.
- Nijman, S.M., Huang, T.T., Dirac, A.M., Brummelkamp, T.R., Kerkhoven, R.M., D'Andrea, A.D., Bernards, R., 2005b. The deubiquitinating enzyme USP1 regulates the Fanconi anemia pathway. *Mol. Cell* 17, 331–339.
- Pförf, R., Lacdao, I.K., Saridakis, V., 2015. Deubiquitinases and the new therapeutic opportunities offered to cancer. *Endocr. Relat. Cancer* 22, T35–T54.
- Pham, L.V., Tamayo, A.T., Li, C., Bornmann, W., Priebe, W., Ford, R.J., 2010. Degrasyn potentiates the antitumor effects of bortezomib in mantle cell lymphoma cells in vitro and in vivo: therapeutic implications. *Mol. Cancer Ther.* 9, 2026–2036.
- Pickart, C.M., Eddins, M.J., 2004. Ubiquitin: structures functions, mechanisms. *Biochim. Biophys. Acta* 1695, 55–72.
- Ratia, K., Pegan, S., Takayama, J., Sleeman, K., Coughlin, M., Balaji, S., Chaudhuri, R., Fu, W., Prabhakar, B.S., Johnson, M.E., Baker, S.C., Ghosh, A.K., Mesecar, A.D., 2008. A noncovalent class of papain-like protease/deubiquitinase inhibitors blocks SARS virus replication. *Proc. Natl. Acad. Sci. U. S. A.* 105, 16119–16124.
- Renatus, M., Parrado, S.G., D'Arcy, A., Eidhoff, U., Gerhardt, B., Hassiepen, U., Pierrat, B., Riedl, R., Vinzenz, D., Worpenberg, S., Kroemer, M., 2006. Structural basis of ubiquitin recognition by the deubiquitinating protease USP2. *Structure* 14, 1293–1302.
- Reyes-Turcu, F.E., Wilkinson, K.D., 2009. Polyubiquitin binding and disassembly by deubiquitinating enzymes. *Chem. Rev.* 109, 1495–1508.
- Rock, K.L., Gramm, C., Rothstein, L., Clark, K., Stein, R., Dick, L., Hwang, D., Goldberg, A.L., 1994. Inhibitors of the proteasome block the degradation of most cell proteins and the generation of peptides presented on MHC class I molecules. *Cell* 78, 761–771.
- Ross, J.M., Olson, L., Coppotelli, G., 2015. Mitochondrial and ubiquitin proteasome system dysfunction in ageing and disease: two sides of the same coin? *Int. J. Mol. Sci.* 1716 (8), 19458–19476.
- Sacco, J.J., Coulson, J.M., Clague, M.J., Urbe, S., 2010. Emerging roles of deubiquitinases in cancer-associated pathways. *IUBMB Life* 62, 140–157.
- Schubert, U., Anton, L.C., Gibbs, J., Norbury, C.C., Yewdell, J.W., Bennink, J.R., 2000. Rapid degradation of a large fraction of newly synthesized proteins by proteasomes. *Nature* 404, 770–774.
- Schweitzer, K., Bozko, P.M., Dubiel, W., Naumann, M., 2007. CSN controls NF-kappaB by deubiquitylation of IkappaBalpha. *EMBO J.* 26, 1532–1541.
- Shkedy, D., Singh, N., Shemesh, K., Amir, A., Geiger, T., Liefshitz, B., Harari, Y., Kupiec, M., 2015. Regulation of Elg1 activity by phosphorylation. *Cell Cycle* 0.
- Singh, N., Bhattacharya, S., Paul, J., 2011. Entamoeba invadens: dynamics of DNA synthesis during differentiation from trophozoite to cyst. *Exp. Parasitol.* 127, 329–333.
- Singh, N., Bhattacharya, A., Bhattacharya, S., 2013. Homologous recombination occurs in Entamoeba and is enhanced during growth stress and stage conversion. *PLoS One* 8, e74465.
- Smogorzewska, A., Matsuoka, S., Vinciguerra, P., McDonald III, E.R., Hurov, K.E., Luo, J., Ballif, S.P., Hofmann, K., D'Andrea, A.D., Elledge, S.J., 2007. Identification of the FANCI protein, a monoubiquitinated FANCD2 paralog required for DNA repair. *Cell* 129, 289–301.
- Srivastava, S.K., Bhardwaj, A., Arora, S., Tyagi, N., Singh, S., Andrews, J., McClellan, S., Wang, B., Singh, A.P., 2015. MicroRNA-345 induces apoptosis in pancreatic cancer cells through potentiation of caspase-dependent and –independent pathways. *Br. J. Cancer* 113, 660–668.
- Steele, J.M., 2013. Carfilzomib: a new proteasome inhibitor for relapsed or refractory multiple myeloma. *J. Oncol. Pharm. Pract.* 19, 348–354.
- Stegemeier, F., Rape, M., Draviam, V.M., Nalepa, G., Sowa, M.E., Ang, X.L., McDonald III, E.R., Li, M.Z., Hannan, G.J., Sorger, P.K., Kirschner, M.W., Harper, J.W., Elledge, S.J., 2007. Anaphase initiation is regulated by antagonistic ubiquitination and deubiquitination activities. *Nature* 446, 876–881.
- Takase, T., Hibi, K., Yamazaki, T., Nakayama, H., Taguchi, M., Kasai, Y., Ito, K., Akiyama, S., Nagasaka, T., Nakao, A., 2003. PGP9.5 overexpression in esophageal squamous cell carcinoma. *Hepatogastroenterology* 50, 1278–1280.
- Tezel, K., Nagasaka, T., Nakao, A., 2000. PGP9.5 as a prognostic factor in pancreatic cancer. *Clin. Cancer Res.* 6, 4764–4767.
- Tran, H., Hamada, F., Schwarzb-Romond, T., Bienz, M., 2008. Trabid: a new positive regulator of Wnt-induced transcription with preference for binding and cleaving K63-linked ubiquitin chains. *Genes Dev.* 22, 528–542.
- Tyagi, N., Ghosh, P.C., 2011. Folate receptor mediated targeted delivery of ricin entrapped into sterically stabilized liposomes to human epidermoid carcinoma (KB) cells: effect of monensin intercalated into folate-tagged liposomes. *Eur. J. Pharm. Sci.* 43, 343–353.
- Tyagi, N., Rathore, S.S., Ghosh, P.C., 2011. Enhanced killing of human epidermoid carcinoma (KB) cells by treatment with ricin encapsulated into sterically stabilized liposomes in combination with monensin. *Drug Deliv.* 18, 394–404.
- Tyagi, N., Bhardwaj, A., Singh, A.P., McClellan, S., Carter, J.E., Singh, S., 2014. p-21 activated kinase 4 promotes proliferation and survival of pancreatic cancer cells through AKT- and ERK-dependent activation of NF-kappaB pathway. *Oncotarget* 5, 8778–8789.
- Tyagi, N., Tyagi, M., Pachauri, M., Ghosh, P.C., 2015a. Potential therapeutic applications of plant toxin-ricin in cancer: challenges and advances. *Tumour. Biol.*
- Tyagi, N., Marimuthu, S., Bhardwaj, A., Deshmukh, S.K., Srivastava, S.K., Singh, A.P., McClellan, S., Carter, J.E., Singh, S., 2015b. p-21 activated kinase 4 (PAK4) maintains stem cell-like phenotypes in pancreatic cancer cells through activation of STAT3 signaling. *Cancer Lett.* 10.
- Tyagi, A., Bhardwaj, S.K., Srivastava, S., Arora, S., Marimuthu, S.K., Deshmukh, A.P., Carter, J.E., Singh, S., 2015c. Development and characterization of a novel in vitro progression model for UVB-Induced skin carcinogenesis. *Sci. Rep.* 5, 13894, <http://dx.doi.org/10.1038/srep13894>.
- Voges, D., Zwickl, P., Baumeister, W., 1999. The 26S proteasome: a molecular machine designed for controlled proteolysis. *Annu. Rev. Biochem.* 68, 1015–1068, 1015–68.
- Wang, W.J., Li, Q.Q., Xu, J.D., Cao, X.X., Li, H.X., Tang, F., Chen, Q., Yang, J.M., Xu, Z.D., Liu, X.P., 2008. Over-expression of ubiquitin carboxy terminal hydrolase-L1 induces apoptosis in breast cancer cells. *Int. J. Oncol.* 33, 1037–1045.
- Welchman, R.L., Gordon, C., Mayer, R.J., 2005. Ubiquitin and ubiquitin-like proteins as multifunctional signals. *Nat. Rev. Mol. Cell Biol.* 6, 599–609.
- Wicks, S.J., Haros, K., Maillard, M., Song, L., Cohen, R.E., Dijke, P.T., Chantry, A., 2005. The deubiquitinating enzyme UCH37 interacts with Smads and regulates TGF-beta signalling. *Oncogene* 24, 8080–8084.
- Xu, M., Takanashi, M., Oikawa, K., Tanaka, M., Nishi, H., Isaka, K., Kudo, M., Kuroda, M., 2009. USP15 plays an essential role for caspase-3 activation during Paclitaxel-induced apoptosis. *Biochem. Biophys. Res. Commun.* 388, 366–371.
- Yamazaki, T., Hibi, K., Takase, T., Tezel, E., Nakayama, H., Kasai, Y., Ito, K., Akiyama, S., Nagasaka, T., Nakao, A., 2002. PGP9.5 as a marker for invasive colorectal cancer. *Clin. Cancer Res.* 8, 192–195.
- Yang, K., Moldovan, G.L., Vinciguerra, P., Murai, J., Takeda, S., D'Andrea, A.D., 2011. Regulation of the Fanconi anemia pathway by a SUMO-like delivery network. *Genes Dev.* 25, 1847–1858.
- Zhao, B., Schlesinger, C., Masucci, M.G., Lindsten, K., 2009. The ubiquitin specific protease 4 (USP4) is a new player in the Wnt signalling pathway. *J. Cell Mol. Med.* 13, 1886–1895.
- Zou, Q., Jin, J., Hu, H., Li, H.S., Romano, S., Xiao, Y., Nakaya, M., Zhou, X., Cheng, X., Yang, P., Lozano, G., Zhu, C., Watowich, S.S., Ullrich, S.E., Sun, S.C., 2014. USP15 stabilizes MDM2 to mediate cancer cell survival and inhibit antitumor T cell responses. *Nat. Immunol.* 15 (6), 562–570.
- van Leuken, R.J., Luna-Vargas, M.P., Sixma, T.K., Wolthuis, R.M., Medema, R.H., 2008. Usp39 is essential for mitotic spindle checkpoint integrity and controls mRNA-levels of aurora B. *Cell Cycle* 7, 2710–2719.
- Zheng, H., Gupta, V., Patterson-Fortin, J., Bhattacharya, S., Katlinski, K., Wu, J., Varghese, B., Carbone, C.J., Aressy, B., Fuchs, S.Y., Greenberg, R.A., 2013. A BRISC-SHMT complex deubiquitinates IFNAR1 and regulates interferon responses. *Cell Rep.* 5, 180–193.