



## Metagenomics: A new horizon in cancer research



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### ABSTRACT

Metagenomics has broadened the scope of targeting microbes responsible for inducing various types of cancers. About 16.1% of cancers are associated with microbial infection. Metagenomics is an equitable way of identifying and studying micro-organisms within their habitat. In cancer research, this approach has revolutionized the way of identifying, analyzing and targeting the microbial diversity present in the tissue specimens of cancer patients. The genomic analyses of these micro-organisms through next generation sequencing techniques invariably facilitate in recognizing the microbial population in biopsies and their evolutionary relationships with each other. In this review an attempt has been made to generate current metagenomic view on cancer microbiota. Different types of micro-organisms have been found to be linked to various types of cancers, thus, contributing significantly in understanding the disease at molecular level.

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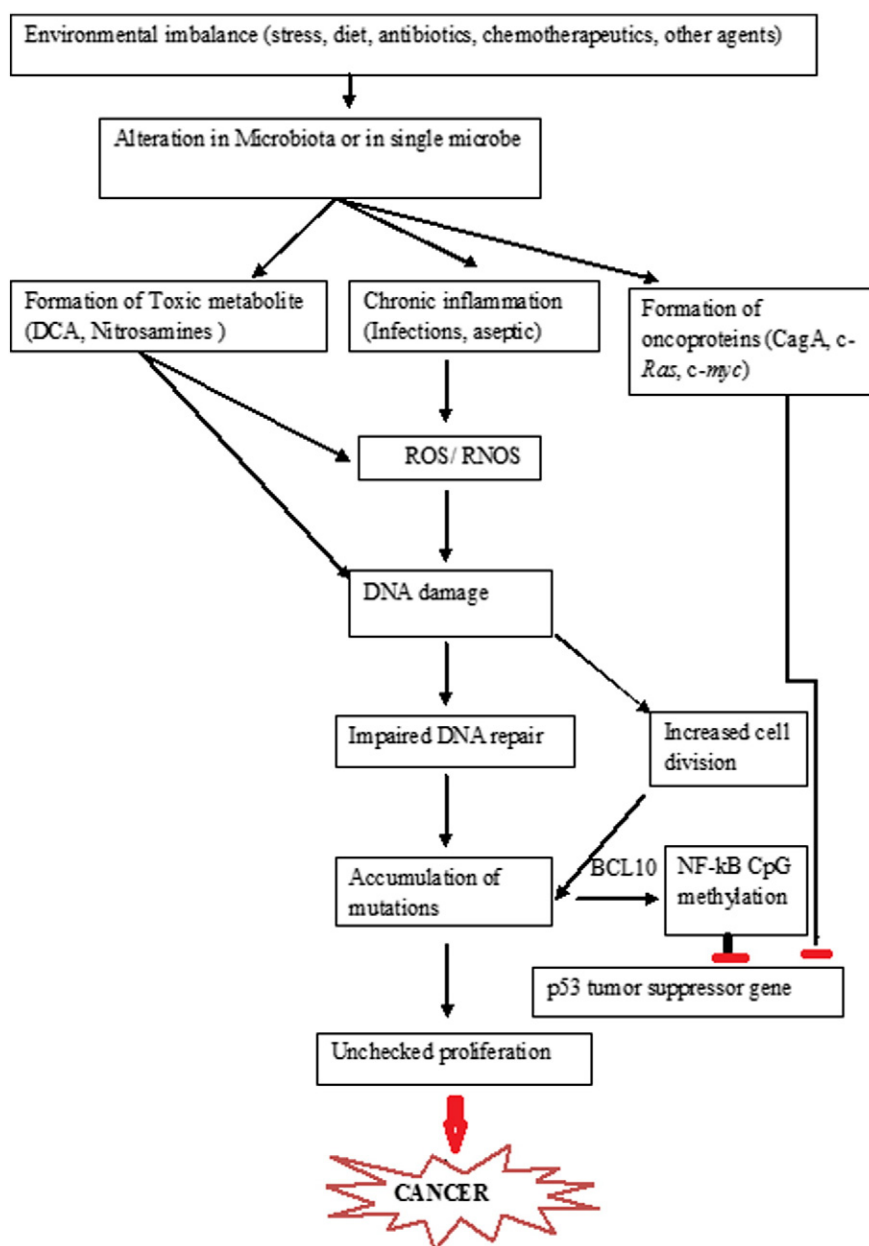
### Introduction

Cancer is a major health concern in the developed and developing countries. On the World Cancer Day (February 4, 2014), the International Agency for Research on Cancer (IARC) published a worldwide report on cancer in 2012 which estimated about 14.1 million new cancer cases, 8.2 million deaths due to cancer and 32.6 million people living with cancer (International Agency for Research on Cancer, WHO [Internet],

2012). Combined effects of several factors such as genetic, environmental, life style can lead to cancer. One such factor for causing cancer is the cancer induced by microbes which estimates about 16.1% of the total cancer burden globally (De Martel et al., 2012). (See Fig. 1.)

Several studies on microbes and cancers showed distinct associations of various viruses with different types of cancers. Human papilloma virus (HPV) causes cervical cancers (Hausen, 1996) whereas *Helicobacter pylori* induce gastric cancers and Mucosa-associated lymphoid tissue (MALT) lymphoma (Cover and Blaser, 2009). Hepatitis B and C viruses are responsible for Hepatocellular carcinoma (Raza et al., 2007) and Merkel Cell Polyomavirus cause Merkel cell carcinoma (Feng et al., 2008), which is a rare type of skin cancer. Epstein–Barr virus (EBV) has been found to be responsible for Nasopharyngeal carcinoma (NPC), Burkitt's lymphoma,

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**Fig. 1.** Mechanism of oncogenesis induced due to change in the microbiota (Chang and Parsonnet, 2010). In this figure, environmental imbalances result in alteration of normal microbiota, subsequent formation of toxic metabolites, chronic inflammation, oncoproteins. The generation of free radicals followed by DNA damage and loss of function of p53 tumor suppressor gene, result in uncontrolled proliferation of cells and formation of cancer. DCA – Deoxycholic acid; BCL10 – B cell lymphoma/leukemia 10 protein; NF-kB – Nuclear Factor kappa B; ROS/RNS – Reactive oxygen species/Reactive nitrogen species.

Hodgkin's lymphoma and to some extent to cause HIV-positive Central nervous system (CNS) lymphomas, hypopharyngeal and laryngeal tumors (Goldenberg et al., 2004).

The studying, analyzing and interpreting of the microbial linkage to cancer has been revolutionized in the emergent era of metagenomics. It is an equitable way of studying culture independent micro-organisms which includes the study of their structures, functions and interactions with their habitat (Handelsman et al, 2007). Various recent researches on cancer due to infection have been explored on the light of genomic analysis of microorganisms residing in the cancerous tissue specimens.

The detailed genomic analysis of microbiota of Colorectal Carcinoma (CRC) reported the presence of various *Fusobacterium* spp. and also species from *Campylobacter* and *Leptotrichia* genera (Castellarin et al., 2012; Kostic et al., 2012; Warren et al., 2013). Further, the metagenomic analysis on the prostate secretions showed the presence of microorganisms belonging to Proteobacteria phylum (Smelov et al., 2014).

Metagenomics approach has provided a new way of treating and preventing microbe associated cancers. This review aims to provide extensive studies on the metagenomic approaches concerning microbes induced cancer.

#### Historical perspectives of microbes inducing cancer

The association of microbes with cancer is not a new fact. The famous experiment of Plymouth Rock hen by Francis Peyton Rous in 1911 evidently proved this fact and subsequently named the virus as Rous sarcoma virus (Rous, 1910, 1911). Rous for his notable tumor inducing RNA virus discovery awarded him Nobel Prize in Medicine in 1966. After this discovery of Rous, the 1930s experienced extensive researches on mammalian tumor virus (Becsei-Kilborn, 2010; Vogt, 1996). In 1964, Anthony Epstein, Bert Achong and Yvonne Barr identified EBV particles in Burkitt's lymphoma cell line derived from African

**Table 1**  
List of microbe associated cancer.

Microbes	Induced cancer	References
Human Papilloma virus	Cervical cancer	Hausen (1996)
<i>Helicobacter pylori</i>	Gastric cancer, MALT lymphoma, Oral cancer	Cover and Blaser (2009); Dayama et al. (2011)
Epstein–Barr virus	Nasopharyngeal carcinoma	Goldenberg et al. (2004)
Merkel Cell Polyomavirus	Merkel cell carcinoma	Feng et al. (2008)
Hepatitis B and C viruses	Hepatocellular carcinoma	Raza et al. (2007)
Human cytomegalo virus	Glioblastoma multiforme	Ranganathan et al. (2012); Soroceanu et al. (2011)
Simian Virus 40	Brain cancer	Pagano et al. (2004)
<i>Streptococcus anginosus</i>	Esophageal cancer, Head and neck cancer	Tateda et al. (2000)
<i>Salmonella typhi</i>	Gall bladder cancer, Cholangiocarcinoma	Lazcano-Ponce et al. (2001); Wistuba and Gazdar (2004); Robbins et al. (1988)
<i>Tropheryma whippelii</i>	Extraintestinal lymphoma, Lymphoma and gastric adenocarcinoma	Cadenas et al. (1999); Gillen et al. (1993)
Herpes virus (Kaposi's sarcoma associated Herpes virus)	Kaposi's sarcoma	Ueda (2012)
<i>Mycoplasma penetrans</i>	Kaposi's sarcoma	Barete et al. (2000); Tamburini et al. (2007); Wang et al. (1993)
<i>Mycoplasma tuberculosis</i>		
<i>Chlamydia trachomatis</i>	Epithelial ovarian carcinoma	Quirk and Kupinski (2001)
<i>Chlamydia pneumoniae</i>	Lung carcinoma	Kocazeybek (2003); Koyi et al. (2001); Mager (2006)
<i>Chlamydia psittaci</i>	Ocular lymphoma	Ferreri et al. (2004)

patients by the use of electron microscope (Epstein et al., 1964). The discovery of Harald Zur Hausen on the prevalence of highly risk HPV genotypes in patients' specimens suffering from cervical cancer and strong association of HPV strains with cervical cancer awarded him Nobel Prize in 2008 (Hausen, 1996, 2002). Table 1 contains the list of micro-organisms which are susceptible for causing cancer.

#### Metagenomics and novel sequencing strategies to target diseases

The complete genome sequencing studies of many pathogenic microorganisms have been enhanced with the outbreak of knowledge about these organisms (Monaghan and Barrett, 2006) and generated plethora of informations about the diseases caused by these organisms. Further, these studies on the micro-organisms contributed in selecting potential antibacterial targets (Sakharkar et al., 2004), pathogenesis (Field et al., 2004; Polissi and Soria, 2005) and antibiotic resistance (Black and Hodgson, 2005). Elucidation on the liaison between the genome sequencing and the microbiome encouraged better understanding of the diseases.

Based on a report by the United States National Research Council committee, "The New Science of Metagenomics: Revealing the Secrets of Our Microbial Planet", metagenomics has revolutionized the research in microbiology and opened up a window for exploring previously unknown world of micro-organisms and their diversity (Handelsman et al., 2007). It has by-passed the need of isolation and lab cultivation of individual species (Chen and Pachter, 2005) and provided the surplus knowledge about the microbial communities and the practical applications from new medical approaches to alternative sources of energy (Jurkowski et al., 2007).

About 99% of the micro-organisms remain uncultured and cannot be demonstrated fully by conventional laboratory culture based techniques (Qin et al., 2010). In metagenomics approach the 16S rRNA gene sequence of bacterial chromosome having highly conserved and variable sequences are used for characterizing microbial communities in diverse conditions (Turnbaugh et al., 2008).

Splendid researches have been carried out on metagenomic analyses of the airborne and soil microbes (Daniel, 2005; Delmont et al., 2011; Yooseph et al., 2013). In recent years, the human microbiota which includes micro-organisms of gut, oral cavity and skin (Grice and Segre, 2012) is much a talked research subject where the role of the microbiome interacting with the human body associated with the development of immune function, disease causation, cancer incidence and defense mechanism against pathogens is vastly

studied (Hannigan and Grice, 2013). Moreover, the progress in development of sequencing tools and techniques has helped in investigating human microbiome in various sites of the body (Mardis, 2008).

Recent advancements in the sequence technology have encouraged the microbial genome study and analyses. The next generation sequencing has simplified the process of detection of viral diversity in clinical samples without having prior sequence information (Radford et al., 2012).

#### Metagenomics and human microbiome

The human intestinal microbiota harbors at least a trillion bacterial cells per gram of feces (Gill et al., 2006). The human gut is the vital center of the body controlling physiology, metabolism, nutrition and immune function. The dysbiosis in the gut microbiome may have direct connection with gastrointestinal conditions such as Crohn's disease, inflammatory bowel disease and obesity (Guinane and Cotter, 2013; Peterson et al., 2008; Tamboli et al., 2004). In a recent study, it has been found that the disruption in the gut microbiota also leads to age related alterations in human (Rampelli et al., 2013).

The human oral cavity like that of human gut, is the dwelling place of hundreds of bacterial species, some of them being the cause of oral diseases like dental caries, periodontal diseases (Marsh, 2010). The oral metagenomic study in Wang et al. (2013) depicted the presence of *Streptococcus* sp., *Haemophilus* sp., *Rothia* sp. and *Capnocytophaga* sp. in the periodontal swab samples and the distinct presence of *Prevotella* sp. forming 14.4–44.7% of the bacterial communities in periodontal disease plaque samples.

Like gut and oral cavity, human skin also harbors varied types of microbes. Disruption in the skin microbiota is responsible for incidence of dermatological diseases (Hannigan and Grice, 2013). The sputum samples of patients suffering from Cystic fibrosis, a genetic disease, have been analyzed for metagenome study, revealed that higher abundance of *S. maltophilia* (41% to 90%) whereas lower incidence of *P. aeruginosa* (<1%) (Lim et al., 2014).

#### Metagenomic studies on different types of cancers

##### Gastric carcinoma

The correlations between the microbiota and occurrence of cancers in the liver and gastrointestinal have been identified majorly in the recent investigations (De Martel et al., 2012). The gut associated microbes

have been found to be the component for gastrointestinal cancers and the metagenomic profiling of gastrointestinal biopsies revealed the presence of *H. pylori* (Zheng et al., 2011). Low pH in the gastric secretions favors the growth of bacteria and the production of carcinogenic N-nitrosamine compounds. The studies on animal models also confirm the role of gastric microbiota in the development of gastric cancer (Wang et al., 2000). The predominant microbial population in the gastric cancer patients was found to be *Veillonella*, *Haemophilus*, *Streptococcus*, *Lactobacillus*, *Prevotella* and *Neisseria* spp. (Dicksved et al., 2009). The significant difference in the gastric microbiota of patients with *H. pylori* infection and control group without infection revealed the role of *H. pylori* in interfering with the composition of gastric microbiome (Maldonado-Contreras et al., 2010). Eun and his colleagues used 454-high throughput sequencer to confirm the presence of *Helicobacter* spp. in gastric mucosa of patients with gastric cancer and also reported significant difference in the microbiota of patients with chronic gastritis and intestinal metaplasia (Eun et al., 2014).

#### Colorectal carcinoma

Every year, approximately 1.2 million individuals worldwide are diagnosed with Colorectal Carcinoma (CRC) (Dejea et al., 2013) and the colon is highly exposed to a diverse class of micro-organisms (Warren et al., 2013). Various researches on gut associated microbes, their interactions and incidence of intestinal cancers unravel the accountability of the intestinal microbiome in inducing cancer in human gut. The structural variation in gut microbiota has been found to be responsible for the progression of the CRC (Candela et al., 2014). In 1951, McCoy and Mason put forward the prevalence of *Streptococcus bovis/galloyticus* in colonic carcinoma, which was traditionally considered to be involved in infectious endocarditis (McCoy and Mason, 1951) until in 1974 when *Streptococcus bovis* was recognized to be associated with the colorectal cancer (Savage, 1977).

The genomic analyses of the tissue samples infected with CRC, carried out by various researchers have been seen to be overpopulated with *Fusobacterium* spp. especially *F. nucleatum*, *F. mortiferum*, *F. necrophorum* along with significant co-occurrence of various anaerobes viz. *Campylobacter* and *Leptotrichia* genera (Castellarin et al., 2012; Kostic et al., 2012; Warren et al., 2013). Although these Gram negative anaerobes are specific to the oral cavity but *Fusobacterium* and *Campylobacter* are genetically diverged in the tumor tissues from their oral counterparts (Warren et al., 2013). *Fusobacterium* spp. also has linkage to inflammatory bowel disease including ulcerative colitis and Crohn's disease (Neut et al., 2002; Strauss et al., 2011) establishing a relatedness of inflammatory bowel disease with CRC.

In another study on CRC, the distinct population of *C. leptum*, *C. coccoides*, *Bacteroides/Prevotella*, *Lactobacillus/Leuconostoc/Pediococcus* spp., *Bifidobacterium* spp., *E. coli*, and *Faecalibacterium prausnitzii* species have been observed in the tumor samples of patients with colon cancer compared to normal individuals (Sobhani et al., 2011).

#### Cervical cancer

Cervical cancer, the cancer of the cervix, is the third most common type of cancer of women worldwide with the high mortality rates of 88% in the developing countries (Arbyn et al., 2011). A healthy vaginal microbiota that comprises mainly *Lactobacillus* spp. contributes to women health by lowering pH in vagina through production of lactic acid (Lee et al., 2013). Bacterial Vaginosis (BV) causes the loss of indigenous *Lactobacillus* spp. and the over-growth of anaerobic bacteria instigating imbalance in vaginal microbiota associated with various detrimental health issues like vaginal discharge syndrome, poor pregnancy outcomes, pelvic inflammatory disease, post-operative wound infections and endometritis after elective abortions (Mancuso et al., 2011; Martin, 2012; Ness et al., 2005). The incidence of BV makes an individual to be susceptible more to HPV infection (Gillet et al., 2011).

Persistent HPV infection is the central causative agent for the development and progression of cervical cancer to a higher grade (Koshiol et al., 2008) and only few types of HPV infections possess the risk of cervical cancer (Castellsagué, 2008). HPV can be classified into two types with high-risk type giving rise to cervical carcinoma and low-risk type producing benign warts (Hausen, 2002). There are more than 200 HPV types with 90% of sequence identity in the L1 major capsid region with almost 118 types have their full genome sequenced (Bernard et al., 2010). About 40 types of HPV give rise to anogenital warts with 12 types probably carcinogenic to humans and 12 types possibly carcinogenic to humans as classified by International Agency for Research on Cancer (IARC) (Bouvard et al., 2009). Moreover, HPV-16 and HPV-18 are considered as the most oncogenic types and are almost present in 71% of cases with cervical cancer (De Sanjose et al., 2010).

A comparison study between HPV positive and HPV negative women evaluated by Gao et al. (2013) exhibited an association between vaginal microbiota and the HPV infection. This study unveiled significant presence of *Lactobacillus* spp., including *L. gallinarum*, *L. iners* and *L. gasseri* in all women and *L. gasseri* and *Gardnerella vaginalis* being predominant in women with HPV positive. In another study of detecting diversity in vaginal microbiome among Korean twins by Lee et al. (2013) showed a lowered population of *Lactobacillus* spp. and notable presence of *Fusobacteria* spp., including *Sneathia* spp. in HPV positive cases.

#### Oral carcinoma

The oral microbiota also plays dramatic effect on the human health. The microbes residing in the oral cavity are often detrimental and give rise to different oral diseases including oral cancers. A well equipped knowledge about the normal microbiota present in the healthy person and the change in the microbial population in the diseased state is the primary requirement (Lazarevic et al., 2009) for targeting the disease. Mager et al. (2005) suggested the fact that salivary micro-organisms can be used as the diagnostic marker for oral cancers and found *Prevotella melaninogenica* and *Campylobacter gingivalis* of the Bacteroidetes and *Streptococcus mitis* of the Firmicutes in the Oral Squamous Cell Carcinoma (OSCC). The first metagenomic investigation of the microbiome dwelling in the oral cavity has been carried out to a single sample of healthy individual by next generation sequencing (Xie et al., 2010). Belda-Ferre et al. (2011) used 454-pyrosequencing for studying oral metagenome in various health conditions. The saliva of patients with OSCC mainly comprised large population of Firmicutes and Bacteroidetes and unclassified bacteria and a relatively small percentage of *Mycoplasma* spp. (Tenericutes) were detected by using V4–V5 16S rDNA based 454-parallel DNA sequencing (Pushalkar et al., 2012, 2011).

#### Brain cancer

Glioblastoma multiforme (GBM) is a most aggressive and malignant, grade 4 type of brain cancer arising from the glial cells or their precursors within the central nervous system with poor prognosis (Holland, 2000; Zhu and Parada, 2002). The polyomavirus simian virus 40 (SV40), a potent oncogenic DNA virus (Butel and Lednický, 1999), has been found to be associated with the brain cancers (Pagano et al., 2004). In animal models SV40 has been found to induce primary brain neoplasia along with the other types of cancers such as malignant mesotheliomas, bone tumors and systemic lymphomas (Butel and Lednický, 1999). Ranganathan et al. showed a close association of Human Cytomegalo Virus (HCMV) with the brain cancers. The genomic analysis using quantitative real time PCR of the GBM samples found to comprise all the regions of the genome of HCMV (Ranganathan et al., 2012). A recent study imparted the presence of the CMV sequences and the viral gene expression in most GBM cases (Dziurzynski et al., 2012). A contradictory metagenomic analyses of GBM specimens using High Throughput Sequencing (HTS) and analyses with the multistage computational



pipeline of patient GBM biopsies proclaimed absence of any known viruses despite earlier studies that concluded the strong association of virus with GBM (Cosset et al., 2014). Further, Cosset et al. (2014) revealed the presence of non-specific interferon like pattern analogous to antiviral gene expression.

### Skin cancer

The skin that acts as the protective barrier of our body from the external environment comprises large pool of diverse class of micro-organisms. Imbalance in the normal microbial population in the skin may give rise to skin neoplasia. The substantial research investigations confirm the explicit role of micro-organisms even in the non-infectious skin diseases, such as atopic dermatitis, rosacea, psoriasis and acne (Paulino et al., 2006; Till et al., 2000). Gao et al. (2007) used the 16S rRNA sequencing technique for predicting the composition of skin microbiota. Mathieu et al. explored the skin microbiome and conveyed the presence of *Corynebacterium*, *Staphylococcus* and *Propionibacterium* as the dominant skin colonizing taxa (Mathieu et al., 2013), on the other hand, another investigation of the metagenomic analyses of human skin lesions revealed the skin prevalence of 97% of HPV sequences by employing HTS (Bzhalava and Dillner, 2013). A most significant presence of Merkel Cell Polyomavirus (MCPyV) in an aggressive neuro-endocrine skin cancer has been detected (Feng et al., 2008; Shuda et al., 2008) but its co-occurrence in the normal healthy skin surface aroused the questionability of its relatedness towards skin carcinoma (Foulongne et al., 2010; Schowalter et al., 2010; Wieland et al., 2009).

### Conclusion

Metagenomics has widened the scope of studying micro-organisms within their habitat. The recent advancements in the field of next generation sequencing techniques have also boosted up this approach. A significant percentage of cancers are induced due to micro-organisms, so metagenomics studies may facilitate the cancer researches by identifying microbes responsible for causing cancer. Finding out potential targets in cancer research will enhance the remedial measures and expand the horizon of cancer research. Thus, future studies are encouraged to explore cancer microbiota for further different types of cancers.

### References

Arbyn, M., Castellsague, X., De Sanjose, S., Bruni, L., Saraiya, M., Bray, F., Ferlay, J., 2011. World-wide burden of cervical cancer in 2008. *Ann. Oncol.* 22 (12), 2675–2686.

Barete, S., Calvez, V., Mouquet, C., Barrou, B., Kreis, H., Dantal, J., Dorent, R., Durand, F., Dimitrov, Y., Dupin, N., Marcelin, A.G., Piette, J.C., Bitker, M.O., Frances, C., 2000. Clinical features and contribution of virological findings to the management of Kaposi sarcoma in organ-allograft recipients. *Arch. Dermatol.* 136 (12), 1452–1458.

Becsei-Kilborn, E., 2010. Scientific discovery and scientific reputation: the reception of Peyton Rous' discovery of the chicken sarcoma virus. *J. Hist. Biol.* 43 (1), 111–157.

Belda-Ferre, P., Alcaraz, L.D., Cabrera-Rubio, R., Romero, H., Simón-Soro, A., Pignatelli, M., Mira, A., 2011. The oral metagenome in health and disease. *ISME J.* 6 (1), 46–56.

Bernard, H.U., Burk, R.D., Chen, Z., van Doorslaer, K., Hausen, H.Z., de Villiers, E.M., 2010. Classification of papillomaviruses (PVs) based on 189 PV types and proposal of taxonomic amendments. *Virology* 401 (1), 70–79.

Black, M.T., Hodgson, J., 2005. Novel target sites in bacteria for overcoming antibiotic resistance. *Adv. Drug Deliv. Rev.* 57 (10), 1528–1538.

Bouvard, V., Baan, R., Straif, K., Grosse, Y., Secretan, B., El Ghissassi, F., Benbrahim-Tallaa, L., Guha, N., Freeman, C., Galichet, L., Coglian, V., 2009. A review of human carcinogens—part B: biological agents. *Lancet Oncol.* 10 (4), 321–322.

Butel, J.S., Lednický, J.A., 1999. Cell and molecular biology of simian virus 40: implications for human infections and disease. *J. Natl. Cancer Inst.* 91 (2), 119–134.

Bzhalava, D., Dillner, J., 2013. Bioinformatics for viral metagenomics. *J. Datamining Genomics Proteomics* 4, 134.

Cadenas, F., Sánchez-Lombráña, J.L., Pérez, R., Lomo, F.J., Madrigal, R.B., Vivas, S., Rodrigo, L., 1999. Persistent leucocytosis as initial manifestation of Whipple's disease and development of gastric cancer in the follow up. *Rev. Esp. Enferm. Dig.* 91 (11), 785–788.

Candela, M., Turroni, S., Biagi, E., Carbonero, F., Rampelli, S., Fiorentini, C., Brigidi, P., 2014. Inflammation and colorectal cancer, when microbiota–host mutualism breaks. *World J. Gastroenterol.* 20 (4), 908.

Castellari, M., Warren, R.L., Freeman, J.D., Dreolini, L., Krzywinski, M., Strauss, J., Barnes, R., Watson, P., Allen-Vercoe, E., Moore, R.A., Holt, R.A., 2012. *Fusobacterium nucleatum* infection is prevalent in human colorectal carcinoma. *Genome Res.* 22 (2), 299–306.

Castellsagué, X., 2008. Natural history and epidemiology of HPV infection and cervical cancer. *Gynecol. Oncol.* 110 (3), S4–S7.

Chang, A.H., Parsonnet, J., 2010. Role of bacteria in oncogenesis. *Clin. Microbiol. Rev.* 23, 837–857.

Chen, K., Pachter, L., 2005. Bioinformatics for whole-genome shotgun sequencing of microbial communities. *PLoS Comput. Biol.* 1 (2), e24.

Cosset, É., Petty, T.J., Dutoit, V., Cordey, S., Padioleau, I., Otten-Hernandez, P., Farinelli, L., Kaiser, L., Bruyère-Cerdan, P., Tirefor, D., Preynat-Seauve, O., 2014. Comprehensive metagenomic analysis of glioblastoma reveals absence of known virus despite antiviral-like type I interferon gene response. *Int. J. Cancer* 135 (6), 1381–1389.

Cover, T.L., Blaser, M.J., 2009. *Helicobacter pylori* in Health and Disease. *Gastroenterology* 136 (6), 1863–1873.

Daniel, R., 2005. The metagenomics of soil. *Nat. Rev. Microbiol.* 3 (6), 470–478.

Dayama, A., Srivastava, V., Shukla, M., Singh, R., Pandey, M., 2011. *Helicobacter pylori* and oral cancer: possible association in a preliminary case control study. *Asian Pac. J. Cancer Prev.* 12 (5), 1333–1336.

De Martel, C., Ferlay, J., Franceschi, S., Vignat, J., Bray, F., Forman, D., Plummer, M., 2012. Global burden of cancers attributable to infections in 2008: a review and synthetic analysis. *Lancet Oncol.* 13 (6), 607–615.

De Sanjose, S., Quint, W.G., Alemany, L., Geraets, D.T., Klaustermeier, J.E., Lloveras, B., Tous, S., Felix, A., Bravo, L.E., Shin, H.R., Puras, A., 2010. Human papillomavirus genotype attribution in invasive cervical cancer: a retrospective cross-sectional worldwide study. *Lancet Oncol.* 11 (11), 1048–1056.

Dejea, C., Wick, E., Sears, C.L., 2013. Bacterial oncogenesis in the colon. *Future Microbiol.* 8 (4), 445–460.

Delmont, T.O., Robe, P., Cecillon, S., Clark, I.M., Constancias, F., Simonet, P., Hirsch, P.R., Vogel, T.M., 2011. Accessing the soil metagenome for studies of microbial diversity. *Appl. Environ. Microbiol.* 77 (4), 1315–1324.

Dicksved, J., Lindberg, M., Rosenquist, M., Enroth, H., Jansson, J.K., Engstrand, L., 2009. Molecular characterization of the stomach microbiota in patients with gastric cancer and in controls. *J. Med. Microbiol.* 58, 509–516.

Dziurzynski, K., Chang, S.M., Heimberger, A.B., Kalejta, R.F., Dallas, S.R.M., Smit, M., Soroceanu, L., Cobbs, C.S., 2012. Consensus on the role of human cytomegalovirus in glioblastoma. *Neurooncol.* 14 (3), 246–255.

Epstein, M.A., Achong, B.G., Barr, Y.M., 1964. Virus particles in cultured lymphoblasts from Burkitt's lymphoma. *Lancet* 283 (7335), 702–703.

Eun, C.S., Kim, B.K., Han, D.S., Kim, S.Y., Kim, K.M., Choi, B.Y., Song, K.S., Kim, Y.S., Kim, J.F., 2014. Differences in gastric mucosal microbiota profiling in patients with chronic gastritis, intestinal metaplasia, and gastric cancer using pyrosequencing methods. *Helicobacter* 19, 407–416.

Feng, H., Shuda, M., Chang, Y., Moore, P.S., 2008. Clonal integration of a polyomavirus in human Merkel cell carcinoma. *Science* 319 (5866), 1096–1100.

Ferreri, A.J., Guidoboni, M., Ponzoni, M., De Conciliis, C., Dell'Oro, S., Fleischhauer, K., Caggiari, L., Lettini, A.A., Cin, E.D., Ieri, R., Freschi, M., Villa, E., Boiocchi, M., Dolcetti, R., 2004. Evidence for an association between *Chlamydia psittaci* and ocular adnexal lymphomas. *J. Natl. Cancer Inst.* 96 (8), 586–594.

Field, D., Hughes, J., Moxon, E.R., 2004. Using the genome to understand pathogenicity. In: Woodford, N., Johnson, A.P. (Eds.), *Genomics, Proteomics, and Clinical Bacteriology* vol. 266. Humana Press, NJ, pp. 261–287.

Foulongne, V., Kluger, N., Dereure, O., Mercier, G., Molès, J.P., Guillot, B., Segondy, M., 2010. Merkel cell polyomavirus in cutaneous swabs. *Emerg. Infect. Dis.* 16 (4), 685–687.

Gao, Z., Tseng, C.H., Pei, Z., Blaser, M.J., 2007. Molecular analysis of human forearm superficial skin bacterial biota. *Proc. Natl. Acad. Sci.* 104 (8), 2927–2932.

Gao, W., Weng, J., Gao, Y., Chen, X., 2013. Comparison of the vaginal microbiota diversity of women with and without human papillomavirus infection: a cross-sectional study. *BMC Infect. Dis.* 13 (1), 271.

Gill, S.R., Pop, M., DeBoy, R.T., Eckburg, P.B., Turnbaugh, P.J., Samuel, B.S., Gordon, J.I., Relman, D.A., Fraser-Liggett, C.M., Nelson, K.E., 2006. Metagenomic analysis of the human distal gut microbiome. *Science* 312 (5778), 1355–1359.

Gillen, C.D., Coddington, R., Monteith, P.G., Taylor, R.H., 1993. Extraintestinal lymphoma in association with Whipple's disease. *Gut* 34 (11), 1627–1629.

Gillet, E., Meys, J.F., Verstraelen, H., Bosire, C., De Sutter, P., Temmerman, M., Broeck, D.V., 2011. Bacterial vaginosis is associated with uterine cervical human papillomavirus infection: a meta-analysis. *BMC Infect. Dis.* 11 (1), 10.

Goldenberg, D., Benoit, N.E., Begum, S., Westra, W.H., Cohen, Y., Koch, W.M., Sidransky, D., Califano, J.A., 2004. Epstein-Barr virus in head and neck cancer assessed by quantitative polymerase chain reaction. *Laryngoscope* 114 (6), 1027–1031.

Grice, E.A., Segre, J.A., 2012. The human microbiome: our second genome. *Annu. Rev. Genomics Hum. Genet.* 13, 151.

Guinane, C.M., Cotter, P.D., 2013. Role of the gut microbiota in health and chronic gastrointestinal disease: understanding a hidden metabolic organ. *Ther. Adv. Gastroenterol.* 6 (4), 295–308.

Handelsman, J., Tiedje, J., Alvarez-Cohen, L., Ashburner, M., Cann, I.K.O., Delong, E.F., Schmidt, T.M., 2007. The new science of metagenomics: revealing the secrets of our microbial planet. *Nat. Res. Counc. Repub.* 13.

Hannigan, G.D., Grice, E.A., 2013. Microbial ecology of the skin in the era of metagenomics and molecular microbiology. *Cold Spring Harb. Perspect. Med.* 3 (12), a015362.

Hausen, H.Z., 1996. Papillomavirus infections—a major cause of human cancers. *Biochim. Biophys. Acta (BBA) Rev. Cancer* 1288 (2), F55–F78.

Hausen, H.Z., 2002. Papillomaviruses and cancer: from basic studies to clinical application. *Nat. Rev. Cancer* 2 (5), 342–350.

Holland, E.C., 2000. Glioblastoma multiforme: the terminator. *Proc. Natl. Acad. Sci.* 97 (12), 6242–6244.

- International Agency for Research on Cancer (WHO) [Internet]. 2012. GLOBOCAN 2012: estimated cancer incidence, mortality and prevalence worldwide in 2012 [cited 2014 Sept 9]. [http://globocan.iarc.fr/Pages/fact\\_sheets/cancer.aspx](http://globocan.iarc.fr/Pages/fact_sheets/cancer.aspx) (Available from).
- Jurkowski, A., Reid, A.H., Labov, J.B., 2007. Metagenomics: a call for bringing a new science into the classroom (while it's still new). *CBE Life Sci. Educ.* 6 (4), 260–265.
- Kocazeybek, B., 2003. Chronic *Chlamydia pneumoniae* infection in lung cancer, a risk factor: a case–control study. *J. Med. Microbiol.* 52 (8), 721–726.
- Koshiol, J., Lindsay, L., Pimenta, J.M., Poole, C., Jenkins, D., Smith, J.S., 2008. Persistent human papillomavirus infection and cervical neoplasia: a systematic review and meta-analysis. *Am. J. Epidemiol.* 168 (2), 123–137.
- Kostic, A.D., Gevers, D., Pedamallu, C.S., Michaud, M., Duke, F., Earl, A.M., Ojesina, A.I., Jung, J., Bass, A.J., Tabernero, J., Baselga, J., Liu, C., Shivdasani, R.A., Ogino, S., Birren, B.W., Huttenhower, C., Garrett, W.S., Meyerson, M., 2012. Genomic analysis identifies association of *Fusobacterium* with colorectal carcinoma. *Genome Res.* 22 (2), 292–298.
- Koyi, H., Brandén, E., Gnarp, J., Gnarp, H., Steen, B., 2001. An association between chronic infection with *Chlamydia pneumoniae* and lung cancer. A prospective 2-year study note. *APMIS* 109 (9), 572–580.
- Lazarevic, V., Whiteson, K., Huse, S., Hernandez, D., Farinelli, L., Østerås, M., Schrenzel, J., François, P., 2009. Metagenomic study of the oral microbiota by Illumina high-throughput sequencing. *J. Microbiol. Methods* 79 (3), 266–271.
- Lazcano-Ponce, E.C., Miquel, J.F., Muñoz, N., Herrero, R., Ferricio, C., Wistuba, I.L., Alonso de Ruiz, P., Urista, G.A., Nervi, F., 2001. Epidemiology and molecular pathology of gallbladder cancer. *CA: A Cancer J. Clin.* 51 (6), 349–364.
- Lee, J.E., Lee, S., Lee, H., Song, Y.M., Lee, K., Han, M.J., Sung, J., Ko, G., 2013. Association of the vaginal microbiota with human papillomavirus infection in a Korean twin cohort. *PLoS ONE* 8 (5), e63514.
- Lim, Y.W., Evangelista, J.S., Schmieder, R., Bailey, B., Haynes, M., Furlan, M., Heather Maughan, H., Edwards, R., Rohwer, F., Conrad, D., 2014. Clinical insights from metagenomic analysis of sputum samples from patients with cystic fibrosis. *J. Clin. Microbiol.* 52, 425–437.
- Mager, D.L., 2006. Bacteria and cancer: cause, coincidence or cure? A review. *J. Transl. Med.* 4 (1), 14.
- Mager, D.L., Haffajee, A.D., Devlin, P.M., Norris, C.M., Posner, M.R., Goodson, J.M., 2005. The salivary microbiota as a diagnostic indicator of oral cancer: a descriptive, non-randomized study of cancer-free and oral squamous cell carcinoma subjects. *J. Transl. Med.* 3 (1), 27.
- Maldonado-Contreras, A., Goldfarb, K.C., Godoy-Vitorino, F., Karaoz, U., Contreras, M., Blaser, M.J., Brodie, E.L., Dominguez-Bello, M.G., 2010. Structure of the human gastric bacterial community in relation to *Helicobacter pylori* status. *ISME J.* 5, 574–579.
- Mancuso, M.S., Figueroa, D., Szychowski, J.M., Paden, M.M., Owen, J., 2011. Midtrimester bacterial vaginosis and cervical length in women at risk for preterm birth. *Am. J. Obstet. Gynecol.* 204 (4), 342, e1.
- Mardis, E.R., 2008. Next-generation DNA sequencing methods. *Annu. Rev. Genomics Hum. Genet.* 9, 387–402.
- Marsh, P.D., 2010. Microbiology of dental plaque biofilms and their role in oral health and caries. *Dent. Clin. N. Am.* 54 (3), 441–454.
- Martin, D.H., 2012. The microbiota of the vagina and its influence on women's health and disease. *Am. J. Med. Sci.* 343 (1), 2.
- Mathieu, A., Delmont, T.O., Vogel, T.M., Robe, P., Nalin, R., Simonet, P., 2013. Life on human surfaces: skin metagenomics. *PLoS ONE* 8 (6), e65288.
- McCoy, W.C., Mason, III, J.M., 1951. Enterococcal endocarditis associated with carcinoma of the sigmoid; report of a case. *J. Med. Assoc. State Ala.* 21 (6), 162.
- Monaghan, R.L., Barrett, J.F., 2006. Antibacterial drug discovery—then, now and the genomics future. *Biochem. Pharmacol.* 71 (7), 901–909.
- Ness, R.B., Kip, K.E., Hillier, S.L., Soper, D.E., Stamm, C.A., Sweet, R.L., Rice, P., Richter, H.E., 2005. A cluster analysis of bacterial vaginosis-associated microflora and pelvic inflammatory disease. *Am. J. Epidemiol.* 162 (6), 585–590.
- Neut, C., Bulois, P., Desreumaux, P., Membreé, J.M., Lederman, E., Gambiez, L., Cortot, A., Quandalle, P., van Kruijningen, H., Colombel, J.F., 2002. Changes in the bacterial flora of the neoterminal ileum after ileocolonic resection for Crohn's disease. *Am. J. Gastroenterol.* 97 (4), 939–946.
- Pagano, J.S., Blaser, M., Buendia, M.A., Damania, B., Khalili, K., Raab-Traub, N., Roizman, B., 2004. Infectious agents and cancer: criteria for a causal relation. *Semin. Cancer Biol.* 14 (6), 453–471.
- Paulino, L.C., Tseng, C.H., Strober, B.E., Blaser, M.J., 2006. Molecular analysis of fungal microbiota in samples from healthy human skin and psoriatic lesions. *J. Clin. Microbiol.* 44 (8), 2933–2941.
- Peterson, D.A., Frank, D.N., Pace, N.R., Gordon, J.I., 2008. Metagenomic approaches for defining the pathogenesis of inflammatory bowel diseases. *Cell Host Microbe* 3 (6), 417–427.
- Polissi, A., Soria, M.R., 2005. Functional genomics of bacterial pathogens: from post-genomics to therapeutic targets. *Mol. Microbiol.* 57 (2), 307–312.
- Pushalkar, S., Mane, S.P., Ji, X., Li, Y., Evans, C., Crasta, O.R., Morse, D., Meagher, R., Anup Singh, A., Saxena, D., 2011. Microbial diversity in saliva of oral squamous cell carcinoma. *FEMS Immunol. Med. Microbiol.* 61 (3), 269–277.
- Pushalkar, S., Ji, X., Li, Y., Estilo, C., Yegnanarayana, R., Singh, B., Li, X., Saxena, D., 2012. Comparison of oral microbiota in tumor and non-tumor tissues of patients with oral squamous cell carcinoma. *BMC Microbiol.* 12 (1), 144.
- Qin, J., Li, R., Raes, J., Arumugam, M., Burgdorf, K.S., Manichanh, C., Nielsen, T., Pons, N., Levenez, F., Yamada, T., Mende, D.R., Wang, J., 2010. A human gut microbial gene catalogue established by metagenomic sequencing. *Nature* 464 (7285), 59–65.
- Quirk, J.T., Kupinski, J.M., 2001. Chronic infection, inflammation, and epithelial ovarian cancer. *Med. Hypotheses* 57 (4), 426–428.
- Radford, A.D., Chapman, D., Dixon, L., Chantrey, J., Darby, A.C., Hall, N., 2012. Application of next-generation sequencing technologies in virology. *J. Gen. Virol.* 93 (9), 1853–1868.
- Rampelli, S., Candela, M., Turrioni, S., Biagi, E., Collino, S., Franceschi, C., O'Toole, P.W., Brigidi, P., 2013. Functional metagenomic profiling of intestinal microbiome in extreme ageing. *Aging (Albany NY)* 5 (12), 902.
- Ranganathan, P., Clark, P.A., Kuo, J.S., Salamat, M.S., Kalejta, R.F., 2012. Significant association of multiple human cytomegalovirus genomic loci with glioblastoma multiforme samples. *J. Virol.* 86 (2), 854–864.
- Raza, S.A., Clifford, G.M., Franceschi, S., 2007. World-wide variation in the relative importance of Razahepatitis B and C viruses in hepatocellular carcinoma: a systematic review. *Br. J. Cancer* 96, 1127–1134.
- Robbins, S., Chuang, V.P., Hersh, T., 1988. The development of hepatobiliary cancer in a carrier of *Salmonella typhus*. *Am. J. Gastroenterol.* 83 (6), 675–678.
- Rous, P., 1910. A transmissible avian neoplasm (Sarcoma of the common fowl). *J. Exp. Med.* 12 (5), 696–705.
- Rous, P., 1911. A sarcoma of the fowl transmissible by an agent separable from the tumor cells. *J. Exp. Med.* 13 (4), 397–411.
- Sakharkar, K.R., Sakharkar, M.K., Chow, V.T., 2004. A novel genomics approach for the identification of drug targets in pathogens, with special reference to *Pseudomonas aeruginosa*. *Silico Biol.* 4 (3), 355–360.
- Savage, D.C., 1977. Microbial ecology of the gastrointestinal tract. *Annu. Rev. Microbiol.* 31 (1), 107–133.
- Schwalter, R.M., Pastrana, D.V., Pumphrey, K.A., Moyer, A.L., Buck, C.B., 2010. Merkel cell polyomavirus and two previously unknown polyomaviruses are chronically shed from human skin. *Cell Host Microbe* 7 (6), 509–515.
- Shuda, M., Feng, H., Kwun, H.J., Rosen, S.T., Gjoerup, O., Moore, P.S., Chang, Y., 2008. T antigen mutations are a human tumor-specific signature for Merkel cell polyomavirus. *Proc. Natl. Acad. Sci.* 105 (42), 16272–16277.
- Smelov, V., Arroyo Mühr, L.S., Bzhalava, D., Brown, L.J., Komyakov, B., Dillner, J., 2014. Metagenomic sequencing of expressed prostate secretions. *J. Med. Virol.* 86, 2042–2048.
- Sobhani, I., Tap, J., Roudot-Thoraval, F., Roperch, J.P., Letulle, S., Langella, P., Corthier, G., Van Nhieu, J.T., Furet, J.P., 2011. Microbial dysbiosis in colorectal cancer (CRC) patients. *PLoS ONE* 6 (1), e16393.
- Soroceanu, L., Matlaf, L., Bezroukove, V., Harkins, L., Martinez, R., Greene, M., Soteropoulos, P., Cobbs, C.S., 2011. Human cytomegalovirus US28 found in glioblastoma promotes an invasive and angiogenic phenotype. *Cancer Res.* 71 (21), 6643–6653.
- Strauss, J., Kaplan, G.G., Beck, P.L., Rioux, K., Panaccione, R., DeVinney, R., Lynch, T., Allen-Vercoe, E., 2011. Invasive potential of gut mucosa-derived *Fusobacterium nucleatum* positively correlates with IBD status of the host. *Inflamm. Bowel Dis.* 17 (9), 1971–1978.
- Tamboli, C.P., Neut, C., Desreumaux, P., Colombel, J.F., 2004. Dysbiosis in inflammatory bowel disease. *Gut* 53 (1), 1–4.
- Tamburini, J., Grimaldi, D., Chiche, J.D., Braicaire, F., Bossi, P., 2007. Cytokine pattern in Kaposi's sarcoma associated with immune restoration disease in HIV and tuberculosis co-infected patients. *AIDS* 21 (14), 1980–1983.
- Tateda, M., Shiga, K., Saijo, S., Sone, M., Hori, T., Yokoyama, J., Matsuura, K., Takasaka, T., Miyagi, T., 2000. *Streptococcus anginosus* in head and neck squamous cell carcinoma: implication in carcinogenesis. *Int. J. Mol. Med.* 6 (6), 699–1402.
- Till, A.E., Goulden, V., Cunliffe, W.J., Holland, K.T., 2000. The cutaneous microflora of adolescent, persistent and late-onset acne patients does not differ. *Br. J. Dermatol.* 142 (5), 885–892.
- Turnbaugh, P.J., Hamady, M., Yatsunenkov, T., Cantarel, B.L., Duncan, A., Ley, R.E., Sogin, M.L., Jones, W.J., Roe, B.A., Affourtit, J.P., Egholm, M., Henrissat, B., Heath, A.C., Knight, R., Gordon, J.I., 2008. A core gut microbiome in obese and lean twins. *Nature* 457 (7228), 480–484.
- Ueda, K., 2012. Kaposi's sarcoma-associated herpes virus induced tumorigenesis; how viral oncogenic insults are evaded. *J. Blood Lymph* 2, 3.
- Vogt, P.K., 1996. Peyton Rous: homage and appraisal. *FASEB J.* 10 (13), 1559–1562.
- Wang, R.Y.H., Shih, J.W.K., Weiss, S.H., Grandinetti, T., Pierce, P.F., Lange, M., Alter, H.J., Wear, D.J., Davies, C.L., Mayur, R.K., Lo, S.C., 1993. *Mycoplasma penetrans* infection in male homosexuals with AIDS: high seroprevalence and association with Kaposi's sarcoma. *Clin. Infect. Dis.* 17 (4), 724–729.
- Wang, T.C., Dangler, C.A., Chen, D., Goldenring, J.R., Koh, T., Raychowdhury, R., Coffey, R.J., Ito, S., Varro, A., Dockray, G.J., Fox, J.G., 2000. Synergistic interaction between hypergastrinemia and *Helicobacter* infection in a mouse model of gastric cancer. *Gastroenterol.* 118, 36–47.
- Wang, J., Qi, J., Zhao, H., He, S., Zhang, Y., Wei, S., Zhao, F., 2013. Metagenomic sequencing reveals microbiota and its functional potential associated with periodontal disease. *Sci. Rep.* 3.
- Warren, R.L., Freeman, D.J., Pleasance, S., Watson, P., Moore, R.A., Cochrane, K., Allen-Vercoe, E., Holt, R.A., 2013. Co-occurrence of anaerobic bacteria in colorectal carcinomas. *Microbiome* 1 (1), 16.
- Wieland, U., Mauch, C., Kreuter, A., Krieg, T., Pfister, H., 2009. Merkel cell polyomavirus DNA in persons without Merkel cell carcinoma. *Emerg. Infect. Dis.* 15 (9), 1496.
- Wistuba, I.L., Gazdar, A.F., 2004. Gallbladder cancer: lessons from a rare tumour. *Nat. Rev. Cancer* 4 (9), 695–706.
- Xie, G., Chain, P.S.G., Lo, C.C., Liu, K.L., Gans, J., Merritt, J., Qi, F., 2010. Community and gene composition of a human dental plaque microbiota obtained by metagenomic sequencing. *Mol. Oral Microbiol.* 25 (6), 391–405.
- Yooseph, S., Andrews-Pfannkoch, C., Tenney, A., McQuaid, J., Williamson, S., Thiagarajan, M., Brame, D., Zeigler-Allen, L., Hoffman, J., Goll, J.B., Fadrosch, D., Glass, J., Adams, M.D., Friedman, R., Venter, J.C., 2013. A metagenomic framework for the study of airborne microbial communities. *PLoS ONE* 8 (12), e81862.
- Zheng, Z., Andersson, A.F., Ye, W., Nyrén, O., Normark, S., Engstrand, L., 2011. A method for metagenomics of *Helicobacter pylori* from archived formalin-fixed gastric biopsies permitting longitudinal studies of carcinogenic risk. *PLoS ONE* 6 (10), e26442.
- Zhu, Y., Parada, L.F., 2002. The molecular and genetic basis of neurological tumours. *Nat. Rev. Cancer* 2 (8), 616–626.