

Guest editorial:

HIGHLIGHT REPORT: CARDIOTOXICITY SCREENING

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Recently, Bin Zhao and colleagues from the Chinese Academy of Science in Beijing and University of California, Sacramento, have published a review about identification of cardiotoxic compounds by rapid screening methods (Li et al., 2015). Cardiotoxicity is one of the most frequent causes for withdrawal of drugs from the market and also during drug development (MacDonald and Robertson, 2009; Apostolakis et al., 2013; Li et al., 2015). Totally, 81 drugs have been taken from the market between 1990 and 2013 (Li et al., 2015). Other drugs, such as the antidiabetic rosiglitazone have been amended with cautionary notes informing about possible cardiotoxicity (Sager et al., 2015). Therefore, it is of high relevance to establish screening methods for identification of cardiotoxic compounds and exclude them already during an early stage of drug development.

A relatively high number of withdrawn drugs have been shown to cause arrhythmia by blocking the hERG channel, a potassium channel responsible for repolarizing the cardiac action potential (Li et al., 2015). The likelihood to block hERG increases with high lipophilicity, the presence of a positively charged nitrogen atom and the absence of negatively charged oxygen atoms (Villoutreix and Taboureau, 2015). Meanwhile, software is available to identify possible interactions with hERG channel functions (re-

viewed in Li et al., 2015). The authors recommend to first apply these *in silico* methods, followed by *in vitro* screening which also includes non-hERG drug targets. For this purpose, simple approaches, such as fluorescent imaging plate reader based assays and Ca²⁺ dye technologies represent convenient initial steps. Also, human pluripotent stem cells (hPSC) derived cardiomyocytes have been introduced and several gene and microRNA cardiotoxicity markers have been identified (Chaudhari et al., 2015, 2016). Currently, large efforts are undertaken to establish alternative methods for toxicity testing, particularly in the fields of liver (Godoy et al., 2009, 2013, 2015; Grinberg et al., 2014), kidney (Yang et al., 2014; Bulacio and Torres 2015; Gong et al., 2015) and neurotoxicity (Rempel et al., 2015; Shinde et al., 2015; Balmer et al., 2014; Krug et al., 2013), which are often supported by mathematical modeling (Drasdo et al., 2014; Vartak et al., 2015; Ghallab et al., 2015; Schliess et al., 2014; Hoehme et al., 2010). The here discussed review of Li and colleagues gives a practical and helpful overview over currently available *in silico* and *in vitro* technologies for cardiotoxicity testing and critically discusses their possibilities as well as limitations.

REFERENCES

- Apostolakis S, Oeff M, Tebbe U, Fabritz L, Breithardt G, Kirchhof P. Flecainide acetate for the treatment of atrial and ventricular arrhythmias. *Expert Opin Pharmacother.* 2013;14:347-57.
- Balmer NV, Klima S, Rempel E, Ivanova VN, Kolde R, Weng MK, et al. From transient transcriptome responses to disturbed neurodevelopment: role of histone acetylation and methylation as epigenetic switch between reversible and irreversible drug effects. *Arch Toxicol.* 2014;88:1451-68.
- Bulacio RP, Torres AM. Time course of organic anion transporter 5 (Oat5) urinary excretion in rats treated with cisplatin: a novel urinary biomarker for early detection of drug-induced nephrotoxicity. *Arch Toxicol.* 2015;89:1359-69.
- Chaudhari U, Nemade H, Wagh V, Gaspar JA, Ellis JK, Srinivasan SP, et al. Identification of genomic biomarkers for anthracycline-induced cardiotoxicity in human iPSC-derived cardiomyocytes: an in vitro repeated exposure toxicity approach for safety assessment. *Arch Toxicol.* 2015 Nov 4. [Epub ahead of print].
- Chaudhari U, Nemade H, Gaspar JA, Hescheler J, Hengstler JG, Sachinidis A. MicroRNAs as early toxicity signatures of doxorubicin in human induced pluripotent stem cell-derived cardiomyocytes. *Arch Toxicol.* 2016 Feb 3. [Epub ahead of print].
- Drasdo D, Hoehme S, Hengstler JG. How predictive quantitative modelling of tissue organisation can inform liver disease pathogenesis. *J Hepatol.* 2014;61:951-6.
- Ghallab A, Cellière G, Henkel SG, Driesch D, Hoehme S, Hofmann U, et al. Model guided identification and therapeutic implications of an ammonia sink mechanism. *J Hepatol.* 2015 Nov 27. [Epub ahead of print].
- Godoy P, Hengstler JG, Ilkavets I, Meyer C, Bachmann A, Müller A, et al. Extracellular matrix modulates sensitivity of hepatocytes to fibroblastoid dedifferentiation and transforming growth factor beta-induced apoptosis. *Hepatology.* 2009;49:2031-43.
- Godoy P, Hewitt NJ, Albrecht U, Andersen ME, Ansari N, Bhattacharya S, et al. Recent advances in 2D and 3D in vitro systems using primary hepatocytes, alternative hepatocyte sources and non-parenchymal liver cells and their use in investigating mechanisms of hepatotoxicity, cell signaling and ADME. *Arch Toxicol.* 2013;87:1315-530.
- Godoy P, Schmidt-Heck W, Natarajan K, Lucendo-Villarin B, Szkolnicka D, Asplund A, et al. Gene networks and transcription factor motifs defining the differentiation of stem cells into hepatocyte-like cells. *J Hepatol.* 2015;63:934-42.
- Gong X, Ivanov VN, Davidson MM, Hei TK. Tetramethylpyrazine (TMP) protects against sodium arsenite-induced nephrotoxicity by suppressing ROS production, mitochondrial dysfunction, pro-inflammatory signaling pathways and programmed cell death. *Arch Toxicol.* 2015;89:1057-70.
- Grinberg M, Stöber RM, Edlund K, Rempel E, Godoy P, Reif R, et al. Toxicogenomics directory of chemically exposed human hepatocytes. *Arch Toxicol.* 2014;88:2261-87.
- Hoehme S, Brulport M, Bauer A, Bedawy E, Schormann W, Hermes M, et al. Prediction and validation of cell alignment along microvessels as order principle to restore tissue architecture in liver regeneration. *Proc Natl Acad Sci USA.* 2010;8;107:10371-6.
- Krug AK, Kolde R, Gaspar JA, Rempel E, Balmer NV, Meganathan K, et al. Human embryonic stem cell-derived test systems for developmental neurotoxicity: a transcriptomics approach. *Arch Toxicol.* 2013;87:123-43.
- Li X, Zhang R, Zhao B, Lossin C, Cao Z. Cardiotoxicity screening: a review of rapid-throughput in vitro approaches. *Arch Toxicol.* 2015 Dec 16. [Epub ahead of print].
- MacDonald JS, Robertson RT. Toxicity testing in the 21st century: a view from the pharmaceutical industry. *Toxicol Sci.* 2009;110:40-6.
- Rempel E, Hoelting L, Waldmann T, Balmer NV, Schildknecht S, Grinberg M, et al. A transcriptome-based classifier to identify developmental toxicants by stem cell testing: design, validation and optimization for histone deacetylase inhibitors. *Arch Toxicol.* 2015;89:1599-618.
- Sager PT, Seltzer J, Turner JR, Anderson JL, Hiatt WR, Kowey P, et al. Cardiovascular Safety Outcome Trials: A meeting report from the Cardiac Safety Research Consortium. *Am Heart J.* 2015;169:486-95.
- Schliess F, Hoehme S, Henkel SG, Ghallab A, Driesch D, Böttger J, et al. Integrated metabolic spatial-temporal model for the prediction of ammonia detoxification during liver damage and regeneration. *Hepatology.* 2014;60:2040-51.

Shinde V, Klima S, Sureshkumar PS, Meganathan K, Jagtap S, Rempel E, et al. Human pluripotent stem cell based developmental toxicity assays for chemical safety screening and systems biology data generation. *J Vis Exp*. 2015;17;(100):e52333.

Vartak N, Damle-Vartak A, Richter B, Dirsch O, Dahmen U, Hammad S, et al. Cholestasis-induced adaptive remodeling of interlobular bile ducts. *Hepatology*. 2015 Nov 26. [Epub ahead of print].

Villoutreix BO, Taboureau O. Computational investigations of hERG channel blockers: New insights and current predictive models. *Adv Drug Deliv Rev*. 2015;86:72-82.

Yang Y, Liu H, Liu F, Dong Z. Mitochondrial dysregulation and protection in cisplatin nephrotoxicity. *Arch Toxicol*. 2014;88:1249-56.