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REVIEW ARTICLE

The essential roles of m⁶A modification in osteogenesis and common bone diseases



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KEYWORDS

Bone diseases; m⁶A modification; Osteogenesis; Regulatory role; Stem cells Abstract N6-methyladenosine (m⁶A) is the most prevalent modification in the eukaryotic transcriptome and has a wide range of functions in coding and noncoding RNAs. It affects the fate of the modified RNA, including its stability, splicing, and translation, and plays an important role in post-transcriptional regulation. Bones play a key role in supporting and protecting muscles and other organs, facilitating the movement of the organism, ensuring blood production, etc. Bone diseases such as osteoarthritis, osteoporosis, and bone tumors are serious public health problems. The processes of bone development and osteogenic differentiation require the precise regulation of gene expression through epigenetic mechanisms including histone, DNA, and RNA modifications. As a reversible dynamic epigenetic mark, m⁶A modifications affect nearly every important biological process, cellular component, and molecular function, including skeletal development and homeostasis. In recent years, studies have shown that m⁶A modification is involved in osteogenesis and bone-related diseases. In this review, we summarized the proteins involved in RNA m⁶A modification and the latest progress in elucidating the regulatory role of m⁶A modification in bone formation and stem cell directional differentiation. We also discussed the pathological roles and potential molecular mechanisms of m⁶A modification in bone-related diseases like osteoporosis and osteosarcoma and suggested potential areas for new strategies that could be used to prevent or treat bone defects and bone diseases.

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Introduction

Bone is mineralized connective tissue composed of extracellular matrix and cell populations including osteoblasts. osteoclasts, osteocytes, and bone lining cells. The skeletal system accounts for about 40% of the body weight and has numerous important functions in physical support, movement, and tissue protection, as well as in blood production and immune function. Bone undergoes continuous renewal and remodeling. Bone homeostasis refers to the dynamic balance between the continuous renewal and remodeling associated with bone formation and bone resorption, being regulated by the osteoclasts and osteoblasts.^{1,2} In general, bone has extraordinary healing potential. However, the bone regeneration capacity declines with age and certain pathological changes, resulting in decreased bone density or osteoporosis. With the aging of populations worldwide. bone-related diseases such as osteoporosis, osteoarthritis, and bone defects have become a major public health concern. The key to the healing of fractures and bone defects lies in the bone's ability to regenerate. The osteogenic differentiation of stem cells is a complicated process that requires the cooperation of cell proliferation and differentiation, and the formation and deposition of a mineralized extracellular matrix.¹ Osteogenesis is regulated by numerous transcription factors, cytokines, signaling pathways, and epigenetics.² Although stem cell therapy has been widely used in the field of bone regeneration, the precise regulation of osteogenic differentiation of stem cells remains a big challenge, and a better understanding of the process would yield better treatments.

Epigenetic modifications involve heritable changes in gene expression without DNA sequence alterations and are often associated with human disease. N6-methyladenosine (m⁶A) is an abundant RNA modification that plays a crucial role in regulating RNA metabolism and covers almost the entire transcriptome. Notably, m⁶A methylation is common in eukaryotes and affects macromolecular processes like RNA maturation, splicing, transport, degradation, and translation. Although m⁶A was first reported in 1974,³⁻⁶ the introduction of methylated RNA immunoprecipitation sequencing (MeRIP-Seq) in 2012 sparked renewed interest in m⁶A research.^{7,8} Through in-depth studies of m⁶A modification in the transcriptome, it was discovered that m⁶A modification sites have a typical DRACH consensus motif, which is mainly enriched in the coding sequence (CDS) and 3' untranslated region (3' UTR).^{7,8}

The m⁶A modification is under the dynamic regulation of the m⁶A writing protein complex (methyltransferase complex, MTC), including methyltransferase like 3 (METTL3), METTL14, Wilms tumor-associated protein (WTAP), RNA binding motif protein 15 (RBM15), and zinc finger CCCH domain-containing protein 13 (ZC3H13).9-13 Other methyltransferases that add m⁶A modifications on different RNAs include METTL5-TRMT112 the complex and METTL16.9,14 Demethylation is primarily accomplished by m⁶A erasers including fat mass and obesity-associated protein (FTO) and AlkB homologue 5 (ALKBH5).^{15,16} Dynamic levels of m⁶A modification are regulated by both writers and erasers. RNA-binding proteins which can recognize m⁶A and determine RNA fates are classified as m⁶A readers and include the YT521-B homology domain family (YTHDF)¹⁷ and the IGF2 mRNA-binding protein (IGF2BP).¹⁸ The different fates of targeted RNAs are determined by m⁶A readers. Currently, m⁶A modification has been discovered to regulate RNA decay, stabilization, splicing, transport, and translation. It also regulates self-renewal, differentiation, immune response, and DNA damage response of stem cells. Accumulating evidence, including studies in our laboratory, indicates that m⁶A modification regulates a large variety of biological processes, from tumorigenesis to osteogenic differentiation.^{19,20} This article reviews m⁶A and the current understanding of its roles in bone development and bone-related diseases.

The regulation of m⁶A modification

Methyltransferases/writers write the m⁶A modification

As noted above, the writing of m⁶A is conducted by the MTC.²¹ m⁶A methyltransferase is composed of writers including METTL3, METTL5, METTL14, METTL16, WTAP, RBM15/15B, CBL proto-oncology Gene-like 1 (CBLL1), ZC3H13, and virus-like m⁶A methyltransferase-associated (VIRMA). METTL3 is a principal element of MTCs and is highly conserved among different eukaryotes. METTL14 serves a structural role for the core MTC which stabilizes and facilitates the catalytic activity of METTL3.²² WTAP functions as a subunit of m⁶A MTC that recruits the complex into the nuclear speckles.¹⁰ RBM15 and RBM15B can recruit the MTC to its target transcripts via binding to specific RNA sites for m⁶A modification.^{11,23} VIRMA preferentially localizes near the 3' UTR and stop codon for RNA methylation modifications.¹³ It has been reported that 80% of protein sequences containing ZC3H13 are low-complexity (LC) domains. The MTC is retained in the nuclear speckles due to interactions between its LC domain and ZC3H13 and WTAP, thereby enhancing its catalytic function.^{11,24} METTL16 was first reported in 2017²⁵ as an independent methyltransferase, which could adjust mRNA stability and splicing. Its binding site is inconsistent with that of the METTL3/ METTL14 MTC, and it catalyzes m⁶A modifications of small nuclear RNAs, U6 snRNAs, and other noncoding RNAs.²⁵

In addition to these proteins, there may be more undiscovered m^6A writers for mRNAs or non-coding RNAs involved in common or specific bioprocesses.

Demethylases/erasers removed the m⁶A modification

The m⁶A modification is dynamic and can be reversed by m⁶A demethylases, also known as m⁶A erasers, including FTO and ALKBH5.^{26,27} FTO is a nuclear protein of the AlkB family and was the first reported m⁶A demethylase.²⁶ ALKBH5 was discovered as the second m⁶A eraser. FTO and ALKBH5 are both members of a family of iron (II)/ α -ketoglutarate (α -KG)-dependent dioxygenases that recognize adenine and cytosine methylation in single-stranded DNA and RNA. A new m⁶A demethylase ALKBH3 has

recently been reported to exhibit substrate specificity only for tRNAs. $^{\rm 28,29}$

m⁶A modification is recognized via m⁶A readers

Although methylation and demethylation are accomplished by writers and erasers respectively, it is the readers that determine the functional outcome of m⁶A modification. Readers are composed of YTHDF1/2/3,³⁰ YTH domain-containing protein 1/2 (YTHDC1/2),³¹ the heterogeneous nuclear ribonucleoprotein (HNRNP) family,³⁰ eukaryotic translation initiation factor 3 (eIF3),³² and insulin-like growth factor-2 mRNA-binding protein 1/2/3 (IGF2BP1/2/ 3).¹⁸ YTHDF2 was the first identified. It recruits the CCR4-NOT complex via the binding of its N-terminal region binding to the SH domain of CNOT1 to accelerate the deadenylation and decay of m⁶A-containing RNAs.³³ In the cytoplasm. YTHDF1 interacts with initiation factors to promote the initial phase of RNA translation.³⁴ YTHDF3 promotes translation by cooperating with YTHDF1 to promote protein synthesis and affects YTHDF2-mediated mRNA decay.³⁵ YTHDC1 could promote exon inclusion, similar to SRSF3 and the opposite of SRSF10.36 YTHDC2, the fifth member of this family, is an RNA helicase whose helicase domain also contributes to RNA binding.³⁷ HNRNPA2/B1 binds to m⁶A on primary-miRNA transcripts and interacts with DGCR8 to promote primary miRNA processing.³⁸ IGF2BP recognizes m⁶A modification under both normal and stress conditions, enhancing the stability and translation of RNAs.¹⁸ eIF3 is an m⁶a reader that can initiate protein translation in a cap-independent manner when a 5' UTR m⁶a is present.³⁹

Although it is clear that m^6A readers determine the function of m^6A on RNAs at multiple levels, further studies on the precise relationships among m^6A readers are needed.

m⁶A modification regulates bone development

Bone development is regulated by various signaling pathways and epigenetics.^{40,41} Bone marrow mesenchymal stem cells (BMSCs) are cells with multi-directional differentiation potential that play a crucial part in human bone health by balancing osteogenesis and adipogenesis.^{42,43} Recently, m⁶A was reported to be involved in the pluripotent differentiadevelopment of tion and specific cell lineages,^{44–47} including the osteogenesis of BMSCs.^{48–50} For example, the expression of METTL3 was found to be negatively correlated with the adipogenesis of porcine BMSCs (pBMSCs).⁵¹ METTL3 knockout in BMSCs has been reported to increase bone loss, resulting in impaired bone formation and the generation of pathological features of osteoporosis in mice.⁴⁹ Furthermore, down-regulation of METTL3 was found to lead to decreased ALP activity and fewer mineralized nodules during the osteoblast differentiation of BMSCs, suggesting that METTL3-mediated m⁶A modification regulates the osteogenesis of BMSCs.⁵⁰ Additionally, METTL3 regulates osteoarthritis development by affecting the NF-kB and extracellular matrix synthesis pathway in chondrocytes.⁵² Down-regulation of METTL3 can also promote osteogenesis through suppression of miR-7212-5p maturation.⁵³ It has also been reported that abnormal expression of METTL3 could be an important mechanism underlying osteoporosis.⁵⁴

ALKBH5 is amplified in osteosarcoma and is highly expressed in osteosarcoma patients.⁵⁵ Recent studies have shown that ALKBH5 affects bone formation by targeting BMP2⁵⁶ and adipogenesis by targeting TRAF4.⁵⁷ FTO inhibits the osteogenic differentiation of MSCs through m⁶A demethylation.⁵⁸ FTO expression was increased during the adipogenic differentiation of BMSCs and decreased during their osteogenic differentiation.⁴⁸ FTO was also reported to enhance the stability of mRNAs which protect osteoblasts from genotoxic damage. Notably, FTO down-regulation induced age-related bone loss.⁵⁹ An m⁶A reader, YTHDF1, was reported to promote the osteogenesis of BMSCs via the translational regulation of ZNF893 (Fig. 1).⁶⁰

Through writers, erasers, and readers, m^6A modifications are widely involved in bone formation and development. It is clear that m^6A regulates bone development via both mRNAs and non-coding RNAs, with an integral role in these biological processes. However, the roles of METTL3, one of the most important writers in m^6A , in bone metabolism are complicated. It is currently unclear whether it positively or negatively affects bone development or whether there is a balance between its effects that is further regulated by other factors. Further studies are needed to clarify the roles of METTL3 and other mediators in normal bone development and various pathological conditions affecting the bone.

m⁶A affects the balance between the osteogenic and adipogenic differentiation of MSCs

It is often stated that "bone loss is fat gain", suggesting that the opposite of osteogenic differentiation is adipogenic differentiation.⁶¹ Obesity may inhibit osteogenesis, and studies have found an association between obesity and osteoporosis.⁶²⁻⁶⁴ BMSCs have the potential for both osteogenic and adipogenic differentiation,65 and both share common regulatory pathways.⁶⁶ Normally, osteogenesis and adipogenesis are in balance. Recent studies have shown that m⁶A methylation can regulate the balance between the osteogenesis and adipogenesis of MSCs. In 2018, it was reported that METTL3-mediated m⁶A modification could regulate the osteogenic differentiation fate of BMSCs by adjusting the translation efficiency of the MSC lineage distributor PTH1R.⁴⁹ Other studies have reported that METTL3-mediated m⁶A methylation is involved in the subtle regulation of the adipogenic and osteogenic differentiation of pBMSCs.49,51,67 It was also found that METTL3 could facilitate the osteogenesis of BMSCs through the LINC00657/miR-144-3p/BMPR1B axis.68 Another study reported that METLL3 could promote the osteogenesis of BMSCs by interacting with piRNA-36741.⁶⁹ However, METTL3 was also reported to inhibit osteogenesis through NF-κB signaling.70

As noted above, FTO is more inclined to induce MSCs to undergo adipogenic differentiation. It is up-regulated during adipogenesis and down-regulated during osteogenesis.⁴⁸ FTO regulates preadipocyte differentiation by adjusting the m⁶A levels, and therefore the SRSF2 binding to the splice site to



Figure 1 m⁶A reader YTHDF1 regulates ZNF839 gene translation in an m⁶A-dependent manner and therefore enhances RUNX2 gene transcription, finally potentiating osteogenesis of BMSCs.⁶⁰

control the alternative splicing of RUNX1T1.⁷¹ However, in another study, FTO was found to be up-regulated during the osteogenesis of MSCs and could promote the osteogenesis of MSCs through PPARG.⁷² In addition, ALKBH1 might regulate the fate and bone-fat balance of BMSCs, because the knockdown of ALKBH1 shifted the differentiation of BMSCs toward adipogenesis.⁷³

Studies have revealed that m⁶A could not only adjust the differentiation capacity of MSCs but can also determine their fate. The m⁶A modification usually promotes the differentiation of MSCs in one direction while inhibiting their differentiation in the other. It is likely that more lineage regulators involved in stem cell differentiation, potentially including regulators of m⁶A will be discovered in the future.

m⁶A modification impacts a variety of common bone diseases

Emerging studies have indicated that m⁶A modifications play a crucial role in various bone-related diseases. The roles of m⁶A regulators in bone development and bonerelated diseases are shown in Figure 2 and Table 1, and potential regulatory relationships are presented in Figure 3.

m⁶A modification is closely associated with the occurrence and clinical severity of osteoporosis

Osteoporosis stems from the imbalance of osteoblastmediated bone formation and osteoclast-mediated bone resorption, manifested as decreased bone density, deterioration of the bone microstructure, and excessive accumulation of bone adipose tissue, resulting in the bones becoming weak and prone to fractures.⁷⁴ In patients with osteoporosis, the BMSCs are inclined to differentiate into adipocytes, generating increased bone marrow fat and bone loss.⁷⁵ METTL3-mediated m⁶A modification can regulate the fate of BMSCs and osteoporosis.⁴⁹ Up-regulation of METTL3 could also prevent osteoporosis by affecting the translation of PTH1R in an m⁶A-dependent manner and by regulating the osteogenic differentiation of MSCs via the PTH/PTH1R pathway.⁴⁹ Down-regulation of METTL3 expression is closely associated with the m⁶A modification of RUNX2 and miR-320 precursors, inhibiting bone formation in osteoporosis or ovariectomized mouse models.⁵⁴ The METTL3 and m⁶A levels were also found to be significantly reduced in osteoporosis and oophorectomy patients. Moreover, in patients with fragility fractures, the hMSCs showed signs of accelerated methylation aging due to insufficient osteoblast activity.⁷⁶ Meanwhile, FTO also affects the osteoporotic phenotype. FTO has been reported as a regulator for the fate determination of BMSCs in osteoporosis and is elevated in the regulating axis GDF11-FTO-PPAR. The expression of FTO is increased in aging and osteoporosis.⁴⁸ Systemic FTO knockout mice exhibit retarded growth, short body length, low body weight, and low bone density immediately after birth.⁷⁷ Further studies showed that FTO could promote osteoporosis by regulating NF-κB and the MYC/PI3K/AKT pathway.^{58,78} Thus, FTO plays an important part in the pathogenesis of osteoporosis and could be a novel candidate for the prevention or treatment of osteoporosis.

However, more studies are needed to clarify which m^6A readers are involved in the m^6A -dependent regulation of osteoporosis and what the concrete outcomes of m^6A modification are in terms of bone differentiation, development, and preservation.

m⁶A modification plays important roles in the development of osteosarcoma

Osteosarcoma, a common aggressive malignancy that inhibits bone growth, often occurs in children and adolescents.^{79,80} Recent studies have reported that m⁶A modification affects the pathogenesis and progression of several cancers, including osteosarcoma. Abnormal expression of m⁶A-related molecules was related to the prognosis and development of metastasis in patients with osteosarcoma.⁸¹ Both the m⁶A and METTL3 levels were elevated in human osteosarcoma tissue and cell lines compared with normal osteoblasts.⁸¹ Further studies showed that METTL3 facilitated the occurrence of osteosarcoma by adjusting the m⁶A level of LEF1 and activating the Wnt/ β -catenin axis.⁸² METTL3 could also regulate the expression of TRAF6, DRG1, and ATAD2 through m⁶A



Figure 2 m⁶A regulators in bone development and bone diseases. Red represents promotor, green represents inhibitor, and blue indicates a controversial role. The figure was created with BioRender.com.

modification, all of which might also promote the occurrence and development of osteosarcoma.⁸³⁻⁸⁵ Other molecules involved in m⁶A have also been shown to play a role in the development of osteosarcoma. For example, RBM15 was found to promote the metastasis of osteosarcoma and decrease the survival rate of osteosarcoma patients.⁸⁶ WTAP is highly expressed in osteosarcoma tissue and promotes osteosarcoma tumorigenesis by regulating HMBOX1 mRNA stability.⁸⁷ ALKBH5 was reported to be overexpressed in osteosarcoma and ALKBH5-mediated PVT1 up-regulation through m⁶A modification promoted osteosarcoma tumorigenesis.⁸⁸ miR-451a-mediated YTHDC1 could activate the AKT/mTOR pathway via 3-phosphoinositide-dependent protein kinase 1 (PDPK1) in an m⁶Adependent manner to stimulate the progression of osteosarcoma.8

Nevertheless, few studies have focused on the impact of $m^{6}A$ -dependent non-coding RNAs on osteosarcoma, which are important in the occurrence and development of

osteosarcoma. Additionally, the role of m^6A in the tumor microenvironment is an interesting topic that remains to be researched.

m⁶A modification is involved in intervertebral disc degeneration

Intervertebral disc (IVD) degeneration is a physiological and pathological process in which the intervertebral disc gradually loses its elasticity and shock absorption functions with age, resulting in irreversible degeneration. The degeneration of the IVD includes degeneration of the annulus fibrosus, nucleus pulposus, and cartilage endplate and manifests as limited lumbar activity, low back pain, neck pain, and decreased muscle strength.^{90,91} Several studies have reported that m⁶A regulation plays a crucial part in the occurrence and progression of IVD degeneration. It was reported that WTAP expression was increased in senescent

m⁶A regulator Bone-related process/diseases Role Target Reference Writer 68 LINC00657 METTL3 Osteogenesis Promotor 69 Promotor BMP2 70 Inhibitor MYD-88/NF-κB 49 Osteoporosis Inhibitor PTH/Pth1r 54 Inhibitor RUNX2 82 Osteosarcoma Promotor LEF1 83-85 Promotor TRAF6/DRG1/ATAD2 52 Osteoarthritis Promotor NF-κB 97 Rheumatoid arthritis Inhibitor NF-_KB 105 Bone resorption circ 0008542 Promotor 103 PTPN6 METTL14 SONFH Inhibitor 87 WTAP Osteosarcoma HMBOX1 Promotor 97 IVDD Promotor IncRNA NORAD Eraser 71 RUNX1T1 **FTO** Adipogenesis Promotor 72 Osteogenesis Promotor PPARG 48 Osteoporosis PPARG Promotor 58 MYC/PI3K/AKT Promotor 78 Promotor NF-κB 56 ALKBH5 Osteogenesis Promotor BMP2 57 Adipogenesis Inhibitor TRAF4 88 Osteosarcoma Promotor PVT1 93 IVDD Promotor DNMT3B 105 circ_0008542 Bone resorption Inhibitor Reader 60 YTHDF1 **ZNF893** Osteogenesis Promotor 89 YTHDC1 Osteosarcoma Promotor PDPK1

 Table 1
 The roles of m⁶A regulators in bone development and bone-related diseases.

nucleus pulposus cells (NPCs) and promoted the m⁶A methylation of lncRNA NORAD, increasing the cellular senescence via the NORAD/PUMILIO/E2F3 axis.⁹² Another study reported that ALKBH5 expression is up-regulated in IVD degeneration and causes NPC senescence by demethylating DNMT3B transcripts.⁹³ However, more evidence is needed to determine the relationship between m⁶A modification and IVD.

m⁶A modification is involved in the pathogenesis of arthritis

Arthritis is a common chronic inflammatory disease occurring in the joints and surrounding tissues. It can be classified into dozens of separate diseases that are related to various factors such as degeneration and autoimmunity. Osteoarthritis (OA) and rheumatoid arthritis (RA) are the two main types of arthritis, both of which are characterized by clinical features like joint tenderness and swelling but have different underlying pathological mechanisms.^{94–96} METTL3 is up-regulated in OA and could regulate OA progression via the $\text{NF-}\kappa\text{B}$ pathway and extracellular matrix synthesis of chondrocytes.⁵² In patients with RA, the METTL3 expression was found to be elevated and positively correlated with the levels of CRP and ESR. Interestingly, overexpression of METTL3 could decrease the LPS-induced inflammation in macrophages via NF- κ B.⁹⁷ Another study reported that YTHDF2 knockdown could increase the expression of proinflammatory cytokines and facilitate the inflammatory response in LPS-stimulated macrophages via the MAPK and NF- κ B signaling pathways.⁹⁸

As described above, m⁶A appears to regulate OA and RA mainly through cytokines. Since the pathological mechanisms underlying both OA and RA are complex, more detailed studies on m⁶A-mediated regulation of cytokines and immune responses are required.

m⁶A modification plays an important part in periodontitis

Periodontitis is a chronic inflammatory disease of the periodontal tissue induced by multiple pathogenic species present in dental plaque. The clinical features include swollen gums, purulent discharge in the gingival pocket, absorption of alveolar bone, and loose teeth.⁹⁹ About 10.8% worldwide are suffering of people from periodontitis.¹⁰⁰ m⁶A modification was proven to play an important part in the immune microenvironment of patients with periodontitis. Notably, m⁶A regulators were found to be involved in periodontal processes, and their expression levels correlated with the immune characteristics of periodontitis.¹⁰¹ Another study reported that various m⁶A-SNPs could play an important part in the pathogenesis of periodontitis.¹⁰² However, the specific regulatory effects of m⁶A on periodontitis are still unknown and the roles of



Figure 3 m⁶A-mediated regulation in bone development and bone diseases.

 $m^{6}A$ writers, erasers, and readers in the pathogenesis of periodontitis are yet to be identified.

The functions of m⁶A modification in other bonerelated diseases

m⁶A has also been reported to be associated with steroidassociated osteonecrosis of the femoral head (SONFH), ankylosing spondylitis, and osteoclast bone resorption after dental implant placement.¹⁰³⁻¹⁰⁵ The expression levels of both m⁶A and METTL14 were down-regulated in SONFH and METTL14 could mediate the development of SONFH by regulating PTPN6.¹⁰³ TNF- α -mediated overexpression of ELMO1 promoted MSC migration in ankylosing spondylitis patients, which was mediated by METTL14-dependent m⁶A modification.¹⁰⁴ In the osseointegration microenvironment after dental implant placement, METTL3 could mediate the initiation of osteoclast bone resorption through m⁶A modification of circ_0008542 and this process could be corrected by the overexpression of ALKBH5.¹⁰⁵ Though m⁶A modification has been proven to be engaged in many bone diseases, the roles of m⁶A in other bone diseases like hyperostosis and bone cyst are unclear and further study is needed.

Conclusions and prospects

The m⁶A modification is common in eukaryotes, regulating homeostasis and normal cellular activities. In recent years, research on the involvement of m⁶A methylation in the osteogenesis of stem cells has been increasing. More and more studies have demonstrated that m⁶A methylation exists on many RNAs associated with bone development and affects the post-transcriptional regulation of RNAs. The homeostasis of m⁶A methylation is critical for osteogenic differentiation and bone growth, and disturbance of this process can lead to various bone diseases. However, the gene-specific effects of m⁶A modifications and the effects of different methylation abundance within the same gene remain to be explored. The roles of other components of m⁶A, including writers (METTL5, METTL16, KIAA1429, RBM15/15B, VIRMA, ZC3H13, and ZCCHC4) and readers (IGF2BP1, HNRNPA2B1, YTHDC1, and eIF3) in osteogenesis and bone-related diseases also need to be explored in further study. The controversial role of METTL3 in bone development also needs to be examined. There are likely other writers, erasers, and readers in m⁶A methylation that still need to be identified, some of which may shed new light on the mechanisms by which m⁶A methylation affects osteogenesis. Based on the crucial functions of m⁶A modification in regulating stem cell fates, cell differentiation, bone formation, and osteosarcoma, m⁶A modification has the potential for broad application in regenerative medicine, bone tissue engineering, and the treatment of bonerelated tumors. It is foreseeable that in the next few years, research focusing on this field will continue to increase, and m⁶A modification is well on the way to b a valid target for regulating osteogenesis and treating bone diseases.

Author contributions

YG, YS, YP, and JL conceived the idea. YG wrote the manuscript and prepared the figures and tables. YS and JL edited the manuscript.

Conflict of interests

The authors declare that they have no competing interests.

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References

- Clézardin P, Coleman R, Puppo M, et al. Bone metastasis: mechanisms, therapies, and biomarkers. *Physiol Rev.* 2021; 101(3):797–855.
- 2. Shang F, Yu Y, Liu S, et al. Advancing application of mesenchymal stem cell-based bone tissue regeneration. *Bioact Mater.* 2021;6(3):666–683.
- Desrosiers R, Friderici K, Rottman F. Identification of methylated nucleosides in messenger RNA from Novikoff hepatoma cells. *Proc Natl Acad Sci U S A*. 1974;71(10): 3971–3975.
- 4. Chandola U, Das R, Panda B. Role of the N6-methyladenosine RNA mark in gene regulation and its implications on development and disease. *Brief Funct Genomics*. 2015;14(3): 169–179.
- Wei W, Ji X, Guo X, et al. Regulatory role of N⁶-methyladenosine (m⁶ A) methylation in RNA processing and human diseases. J Cell Biochem. 2017;118(9):2534–2543.
- Zhao BS, Roundtree IA, He C. Post-transcriptional gene regulation by mRNA modifications. Nat Rev Mol Cell Biol. 2017;18(1):31-42.
- Dominissini D, Moshitch-Moshkovitz S, Schwartz S, et al. Topology of the human and mouse m⁶A RNA methylomes revealed by m⁶A-seq. *Nature*. 2012;485(7397):201–206.
- Meyer K, Saletore Y, Zumbo P, et al. Comprehensive analysis of mRNA methylation reveals enrichment in 3' UTRs and near stop codons. *Cell*. 2012;149(7):1635–1646.
- 9. Liu J, Yue Y, Han D, et al. A METTL3-METTL14 complex mediates mammalian nuclear RNA N6-adenosine methylation. *Nat Chem Biol*. 2014;10(2):93–95.
- Ping XL, Sun BF, Wang L, et al. Mammalian WTAP is a regulatory subunit of the RNA N6-methyladenosine methyltransferase. *Cell Res.* 2014;24(2):177–189.
- Knuckles P, Lence T, Haussmann IU, et al. Zc3h13/Flacc is required for adenosine methylation by bridging the mRNAbinding factor Rbm15/Spenito to the m⁶A machinery component Wtap/Fl(2)D. *Genes Dev.* 2018;32(5-6):415-429.
- Mendel M, Chen KM, Homolka D, et al. Methylation of structured RNA by the m⁶A writer METTL16 is essential for mouse embryonic development. *Mol Cell*. 2018;71(6):986–1000.
- Lan T, Li H, Zhang D, et al. KIAA1429 contributes to liver cancer progression through N6-methyladenosine-dependent post-transcriptional modification of GATA3. *Mol Cancer*. 2019; 18(1):186.
- Zaccara S, Ries RJ, Jaffrey SR. Reading, writing and erasing mRNA methylation. *Nat Rev Mol Cell Biol*. 2019;20(10): 608–624.
- 15. Tang C, Klukovich R, Peng H, et al. ALKBH5-dependent m6A demethylation controls splicing and stability of long 3'-UTR mRNAs in male germ cells. *Proc Natl Acad Sci U S A*. 2018; 115(2):E325–E333.
- Li Y, Wu K, Quan W, et al. The dynamics of FTO binding and demethylation from the m⁶A motifs. *RNA Biol.* 2019;16(9): 1179–1189.
- 17. Zhu T, Roundtree IA, Wang P, et al. Crystal structure of the YTH domain of YTHDF₂ reveals mechanism for recognition of N6-methyladenosine. *Cell Res.* 2014;24(12):1493–1496.
- Huang H, Weng H, Sun W, et al. Recognition of RNA N6methyladenosine by IGF2BP proteins enhances mRNA stability and translation. *Nat Cell Biol.* 2018;20(3):285–295.
- **19.** Qian JY, Gao J, Sun X, et al. KIAA1429 acts as an oncogenic factor in breast cancer by regulating CDK1 in an N6-methyl-adenosine-independent manner. *Oncogene*. 2019;38(33): 6123–6141.

- 20. Song Y, Pan Y, Wu M, et al. METTL3-mediated lncRNA m⁶A modification in the osteogenic differentiation of human adipose-derived stem cells induced by NEL-like 1 protein. *Stem Cell Rev Rep.* 2021;17(6):2276–2290.
- Bokar JA, Rath-Shambaugh ME, Ludwiczak R, et al. Characterization and partial purification of mRNA N6-adenosine methyltransferase from HeLa cell nuclei. Internal mRNA methylation requires a multisubunit complex. J Biol Chem. 1994;269(26):17697–17704.
- Wang P, Doxtader K, Nam Y. Structural basis for cooperative function of Mettl3 and Mettl14 methyltransferases. *Mol Cell*. 2016;63(2):306-317.
- 23. Patil DP, Chen CK, Pickering BF, et al. M(6)a RNA methylation promotes XIST-mediated transcriptional repression. *Nature*. 2016;537(7620):369–373.
- 24. Wen J, Lv R, Ma H, et al. Zc3h13 regulates nuclear RNA m⁶A methylation and mouse embryonic stem cell self-renewal. *Mol Cell*. 2018;69(6):1028–1038.
- 25. Warda AS, Kretschmer J, Hackert P, et al. Human METTL16 is a N⁶-methyladenosine (m⁶A) methyltransferase that targets pre-mRNAs and various non-coding RNAs. *EMBO Rep.* 2017; 18(11):2004–2014.
- 26. Jia G, Fu Y, Zhao X, et al. N6-methyladenosine in nuclear RNA is a major substrate of the obesity-associated FTO. *Nat Chem Biol*. 2011;7(12):885–887.
- 27. Zheng G, Dahl JA, Niu Y, et al. ALKBH5 is a mammalian RNA demethylase that impacts RNA metabolism and mouse fertility. *Mol Cell*. 2013;49(1):18–29.
- 28. Ueda Y, Ooshio I, Fusamae Y, et al. AlkB homolog 3-mediated tRNA demethylation promotes protein synthesis in cancer cells. *Sci Rep.* 2017;7:42271.
- **29.** Yang G, Sun Z, Zhang N. Reshaping the role of m6A modification in cancer transcriptome: a review. *Cancer Cell Int*. 2020;20:353.
- Shi H, Wei J, He C. Where, when, and how: context-dependent functions of RNA methylation writers, readers, and erasers. *Mol Cell*. 2019;74(4):640–650.
- Haussmann IU, Bodi Z, Sanchez-Moran E, et al. m⁶A potentiates Sxl alternative pre-mRNA splicing for robust *Drosophila* sex determination. *Nature*. 2016;540(7632):301–304.
- Meyer KD, Jaffrey SR. Rethinking m⁶A readers, writers, and erasers. Annu Rev Cell Dev Biol. 2017;33:319-342.
- Du H, Zhao Y, He J, et al. YTHDF₂ destabilizes m(6)A-containing RNA through direct recruitment of the CCR4-NOT deadenylase complex. *Nat Commun.* 2016;7:12626.
- Wang X, Zhao BS, Roundtree IA, et al. N(6)-methyladenosine modulates messenger RNA translation efficiency. *Cell*. 2015; 161(6):1388–1399.
- Wang X, Lu Z, Gomez A, et al. N6-methyladenosine-dependent regulation of messenger RNA stability. *Nature*. 2014; 505(7481):117–120.
- Xiao W, Adhikari S, Dahal U, et al. Nuclear m(6)a reader YTHDC1 regulates mRNA splicing. *Mol Cell*. 2016;61(4): 507-519.
- Hsu PJ, Zhu Y, Ma H, et al. Ythdc2 is an N⁶-methyladenosine binding protein that regulates mammalian spermatogenesis. *Cell Res.* 2017;27(9):1115–1127.
- Alarcón CR, Goodarzi H, Lee H, et al. HNRNPA2B1 is a mediator of m(6)A-dependent nuclear RNA processing events. *Cell*. 2015;162(6):1299–1308.
- **39.** Meyer KD, Patil DP, Zhou J, et al. 5' UTR m(6)a promotes capindependent translation. *Cell*. 2015;163(4):999-1010.
- 40. Peng Y, Huang S, Wu Y, et al. Platelet rich plasma clot releasate preconditioning induced PI3K/AKT/NFκB signaling enhances survival and regenerative function of rat bone

marrow mesenchymal stem cells in hostile microenvironments. *Stem Cell Dev.* 2013;22(24):3236–3251.

- 41. Shi L, Feng L, Zhu ML, et al. Vasoactive intestinal peptide stimulates bone marrow-mesenchymal stem cells osteogenesis differentiation by activating Wnt/β-catenin signaling pathway and promotes rat skull defect repair. *Stem Cell Dev.* 2020;29(10):655–666.
- Kawai M, Devlin MJ, Rosen CJ. Fat targets for skeletal health. Nat Rev Rheumatol. 2009;5(7):365–372.
- Chen Q, Shou P, Zheng C, et al. Fate decision of mesenchymal stem cells: adipocytes or osteoblasts? *Cell Death Differ*. 2016; 23(7):1128–1139.
- **44.** Batista PJ, Molinie B, Wang J, et al. M(6)a RNA modification controls cell fate transition in mammalian embryonic stem cells. *Cell Stem Cell*. 2014;15(6):707–719.
- 45. Geula S, Moshitch-Moshkovitz S, Dominissini D, et al. Stem cells. m⁶A mRNA methylation facilitates resolution of naïve pluripotency toward differentiation. *Science*. 2015;347(6225): 1002–1006.
- Zhao BS, He C. Fate by RNA methylation: m⁶A steers stem cell pluripotency. *Genome Biol.* 2015;16(1):43.
- **47.** Ji P, Wang X, Xie N, et al. N6-methyladenosine in RNA and DNA: an epitranscriptomic and epigenetic player implicated in determination of stem cell fate. *Stem Cell Int.* 2018;2018: 3256524.
- 48. Shen GS, Zhou HB, Zhang H, et al. The GDF11-FTO-PPARγ axis controls the shift of osteoporotic MSC fate to adipocyte and inhibits bone formation during osteoporosis. *Biochim Biophys Acta, Mol Basis Dis.* 2018;1864(12):3644–3654.
- 49. Wu Y, Xie L, Wang M, et al. Mettl3-mediated m⁶A RNA methylation regulates the fate of bone marrow mesenchymal stem cells and osteoporosis. *Nat Commun.* 2018;9: 4772.
- Tian C, Huang Y, Li Q, et al. Mettl3 regulates osteogenic differentiation and alternative splicing of vegfa in bone marrow mesenchymal stem cells. *Int J Mol Sci.* 2019;20(3):551.
- 51. Yao Y, Bi Z, Wu R, et al. METTL3 inhibits BMSC adipogenic differentiation by targeting the JAK1/STAT5/C/EBPβ pathway via an m⁶A-YTHDF2-dependent manner. Faseb J. 2019;33(6): 7529–7544.
- 52. Liu Q, Li M, Jiang L, et al. METTL3 promotes experimental osteoarthritis development by regulating inflammatory response and apoptosis in chondrocyte. *Biochem Biophys Res Commun.* 2019;516(1):22–27.
- Mi B, Xiong Y, Yan C, et al. Methyltransferase-like 3-mediated N6-methyladenosine modification of miR-7212-5p drives osteoblast differentiation and fracture healing. J Cell Mol Med. 2020;24(11):6385–6396.
- 54. Yan G, Yuan Y, He M, et al. m⁶A methylation of precursor-miR-320/RUNX2 controls osteogenic potential of bone marrowderived mesenchymal stem cells. *Mol Ther Nucleic Acids*. 2020;19:421–436.
- 55. Yadav P, Subbarayalu P, Abdelfattah N, et al. Abstract 4146: N6Methyladenosine RNA demethylase ALKBH5 as a novel therapeutic target for osteosarcoma. *Cancer Res.* 2018; 78(13_Supplement):4146.
- 56. Wang HF, Kuang MJ, Han SJ, et al. BMP2 modified by the m⁶A demethylation enzyme ALKBH5 in the ossification of the ligamentum flavum through the AKT signaling pathway. *Calcif Tissue Int*. 2020;106(5):486–493.
- 57. Cen S, Li J, Cai Z, et al. $TRAF_4$ acts as a fate checkpoint to regulate the adipogenic differentiation of MSCs by activating PKM2. *EBioMedicine*. 2020;54:102722.
- Zhang X, Wang Y, Zhao H, et al. Extracellular vesicle-encapsulated miR-22-3p from bone marrow mesenchymal stem cell

promotes osteogenic differentiation via FTO inhibition. *Stem Cell Res Ther.* 2020;11(1):227.

- 59. Zhang Q, Riddle RC, Yang Q, et al. The RNA demethylase FTO is required for maintenance of bone mass and functions to protect osteoblasts from genotoxic damage. *Proc Natl Acad Sci U S A*. 2019;116(36):17980–17989.
- 60. Liu T, Zheng X, Wang C, et al. The m⁶A "reader" YTHDF₁ promotes osteogenesis of bone marrow mesenchymal stem cells through translational control of ZNF₈₃₉. *Cell Death Dis*. 2021;12(11):1–13.
- 61. James AW, Pan A, Chiang M, et al. A new function of Nell-1 protein in repressing adipogenic differentiation. *Biochem Biophys Res Commun.* 2011;411(1):126–131.
- 62. Kim HY, Kim Y. Associations of obesity with osteoporosis and metabolic syndrome in Korean postmenopausal women: a cross-sectional study using national survey data. Arch Osteoporosis. 2019;14(1):64.
- **63.** Neglia C, Argentiero A, Chitano G, et al. Diabetes and obesity as independent risk factors for osteoporosis: updated results from the ROIS/EMEROS registry in a population of five thousand post-menopausal women living in a region characterized by heavy environmental pressure. *Int J Environ Res Publ Health.* 2016;13(11):1067.
- Zhao LJ, Liu YJ, Liu PY, et al. Relationship of obesity with osteoporosis. J Clin Endocrinol Metab. 2007;92(5):1640–1646.
- **65.** Gimble JM, Robinson CE, Wu X, et al. The function of adipocytes in the bone marrow stroma: an update. *Bone*. 1996; 19(5):421-428.
- **66.** Choi YJ, Song I, Jin Y, et al. Transcriptional profiling of human femoral mesenchymal stem cells in osteoporosis and its association with adipogenesis. *Gene*. 2017;632:7–15.
- Both J, Wu T, Bras J, et al. Identification of novel candidate oncogenes in chromosome region 17p11.2-p12 in human osteosarcoma. *PLoS One*. 2012;7(1):e30907.
- Peng J, Zhan Y, Zong Y. METTL3-mediated LINC00657 promotes osteogenic differentiation of mesenchymal stem cells via miR-144-3p/BMPR1B axis. *Cell Tissue Res.* 2022;388(2): 301–312.
- Liu J, Chen M, Ma L, et al. piRNA-36741 regulates BMP2-mediated osteoblast differentiation via METTL3 controlled m6A modification. *Aging*. 2021;13(19):23361–23375.
- 70. Yu J, Shen L, Liu Y, et al. The m6A methyltransferase METTL3 cooperates with demethylase ALKBH5 to regulate osteogenic differentiation through NF-κB signaling. *Mol Cell Biochem*. 2020;463(1):203–210.
- Zhao X, Yang Y, Sun BF, et al. FTO-dependent demethylation of N6-methyladenosine regulates mRNA splicing and is required for adipogenesis. *Cell Res.* 2014;24(12):1403–1419.
- 72. Chen LS, Zhang M, Chen P, et al. The m⁶A demethylase FTO promotes the osteogenesis of mesenchymal stem cells by downregulating PPARG. *Acta Pharmacol Sin.* 2022;43(5): 1311–1323.
- **73.** Cai GP, Liu YL, Luo LP, et al. Alkbh1-mediated DNA N6methyladenine modification regulates bone marrow mesenchymal stem cell fate during skeletal aging. *Cell Prolif.* 2022; 55(2):e13178.
- Frem S, Atfi A, Razzaque MS. Anabolic effects of vitamin D and magnesium in aging bone. J Steroid Biochem Mol Biol. 2019; 193:105400.
- **75.** Rosen CJ, Ackert-Bicknell C, Rodriguez JP, et al. Marrow fat and the bone microenvironment: developmental, functional, and pathological implications. *Crit Rev Eukaryot Gene Expr.* 2009;19(2):109–124.
- 76. Del Real A, Pérez-Campo FM, Fernández AF, et al. Differential analysis of genome-wide methylation and gene expression in mesenchymal stem cells of patients with fractures and osteoarthritis. *Epigenetics*. 2017;12(2):113–122.

- 77. Gao X, Shin YH, Li M, et al. The fat mass and obesity associated gene FTO functions in the brain to regulate postnatal growth in mice. PLoS One. 2010;5(11):e14005.
- **78.** Zhuang J, Ning H, Wang M, et al. Downregulated fat mass and obesity-associated protein inhibits bone resorption and osteoclastogenesis by nuclear factor-kappa B inactivation. *Cell Signal*. 2021;87:110137.
- **79.** Endicott AA, Morimoto LM, Kline CN, et al. Perinatal factors associated with clinical presentation of osteosarcoma in children and adolescents. *Pediatr Blood Cancer*. 2017;64(6): e26349.
- **80.** Meazza C, Scanagatta P. Metastatic osteosarcoma: a challenging multidisciplinary treatment. *Expert Rev Anticancer Ther.* 2016;16(5):543–556.
- Li J, Rao B, Yang J, et al. Dysregulated m6A-related regulators are associated with tumor metastasis and poor prognosis in osteosarcoma. *Front Oncol.* 2020;10:769.
- 82. Miao W, Chen J, Jia L, et al. The m6A methyltransferase METTL3 promotes osteosarcoma progression by regulating the m6A level of LEF₁. *Biochem Biophys Res Commun.* 2019; 516(3):719–725.
- Wang J, Wang W, Huang X, et al. m6A-dependent upregulation of TRAF₆ by METTL3 is associated with metastatic osteosarcoma. J Bone Oncol. 2022;32:100411.
- 84. Ling Z, Chen L, Zhao J. m6A-dependent up-regulation of DRG1 by METTL3 and ELAVL1 promotes growth, migration, and colony formation in osteosarcoma. *Biosci Rep.* 2020;40(4): BSR20200282.
- **85.** Zhou L, Yang C, Zhang N, et al. Silencing METTL3 inhibits the proliferation and invasion of osteosarcoma by regulating ATAD2. *Biomed Pharmacother*. 2020;125:109964.
- Huang H, Cui X, Qin X, et al. Analysis and identification of m⁶A RNA methylation regulators in metastatic osteosarcoma. *Mol Ther Nucleic Acids*. 2022;27:577–592.
- 87. Chen S, Li Y, Zhi S, et al. WTAP promotes osteosarcoma tumorigenesis by repressing HMBOX1 expression in an m⁶Adependent manner. *Cell Death Dis.* 2020;11(8):659.
- Chen S, Zhou L, Wang Y. ALKBH5-mediated m⁶A demethylation of lncRNA PVT1 plays an oncogenic role in osteosarcoma. *Cancer Cell Int.* 2020;20:34.
- **89.** Cao D, Ge S, Li M. miR-451a promotes cell growth, migration and EMT in osteosarcoma by regulating YTHDC1-mediated m6A methylation to activate the AKT/mTOR signaling pathway. *J Bone Oncol*. 2022;33:100412.
- Adams MA, Roughley PJ. What is intervertebral disc degeneration, and what causes it? Spine. 2006;31(18): 2151–2161.
- **91.** Sloan Jr SR, Wipplinger C, Kirnaz S, et al. Combined nucleus pulposus augmentation and annulus fibrosus repair prevents acute intervertebral disc degeneration after discectomy. *Sci Transl Med.* 2020;12(534):eaay2380.
- 92. Li G, Ma L, He S, et al. Author Correction: WTAP-mediated m⁶A modification of lncRNA NORAD promotes intervertebral disc degeneration. *Nat Commun.* 2022;13:3572.
- 93. Li G, Luo R, Zhang W, et al. m6A hypomethylation of DNMT3B regulated by ALKBH5 promotes intervertebral disc degeneration via E4F1 deficiency. *Clin Transl Med.* 2022; 12(3):e765.
- 94. Mathew AJ, Ravindran V. Infections and arthritis. *Best Pract Res Clin Rheumatol*. 2014;28(6):935-959.
- **95.** Kretschmer J, Rao H, Hackert P, et al. The m⁶A reader protein YTHDC2 interacts with the small ribosomal subunit and the 5'-3' exoribonuclease XRN₁. *RNA*. 2018;24(10): 1339–1350.
- **96.** Marini F, Cianferotti L, Brandi ML. Epigenetic mechanisms in bone biology and osteoporosis: can they drive therapeutic choices? *Int J Mol Sci.* 2016;17(8):1329.

- **97.** Wang J, Yan S, Lu H, et al. METTL3 attenuates LPS-induced inflammatory response in macrophages via NF- *κ* B signaling pathway. *Mediat Inflamm.* 2019;2019:3120391.
- Yu R, Li Q, Feng Z, et al. m6A reader YTHDF₂ regulates LPSinduced inflammatory response. *Int J Mol Sci.* 2019;20(6):1323.
 Slots J. Periodontitis: facts, fallacies and the future. *Perio-*
- dontol. 2017;75(1):7–23, 2000.
 100. Peres MA, MacPherson LMD, Weyant RJ, et al. Oral diseases: a
- global public health challenge. Lancet. 2019;394(10194): 249–260.
- **101.** Zhang X, Zhang S, Yan X, et al. m6A regulator-mediated RNA methylation modification patterns are involved in immune microenvironment regulation of periodontitis. *J Cell Mol Med*. 2021;25(7):3634–3645.
- 102. Lin W, Xu H, Wu Y, et al. In silico genome-wide identification of m6A-associated SNPs as potential functional variants for periodontitis. J Cell Physiol. 2020;235(2):900–908.
- **103.** Cheng C, Zhang H, Zheng J, et al. METTL14 benefits the mesenchymal stem cells in patients with steroid-associated osteonecrosis of the femoral head by regulating the m6A level of PTPN6. *Aging*. 2021;13(24):25903–25919.
- 104. Xie Z, Yu W, Zheng G, et al. TNF-α-mediated m⁶A modification of ELMO1 triggers directional migration of mesenchymal stem cell in ankylosing spondylitis. *Nat Commun*. 2021;12:5373.
- **105.** Wang W, Qiao SC, Wu XB, et al. Circ_0008542 in osteoblast exosomes promotes osteoclast-induced bone resorption through m6A methylation. *Cell Death Dis.* 2021; 12(7):628.